

Sequence Analysis

Supplementary Material for "MESA: automated assessment of synthetic DNA fragments and simulation of DNA synthesis, storage, sequencing, and PCR errors"

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Abstract

Summary: The development of *de novo* DNA synthesis, PCR, DNA sequencing, and molecular cloning gave researchers unprecedented control over DNA and DNA-mediated processes. To reduce the error probabilities of these techniques, DNA composition has to adhere to method-dependent restrictions. To adhere to such restrictions, a synthetic DNA fragment is often adjusted manually or by using custom-made scripts. In this paper, we present MESA (*MOSLA Error Simulator*), a web application for the assessment of DNA fragments based on limitations of DNA synthesis, amplification, cloning, sequencing methods, and biological restrictions of host organisms. Furthermore, MESA can be used to simulate errors during synthesis, PCR, storage, and sequencing processes.

Availability: MESA is available at mesa.mosla.de, with the source code available at github.com/umr-ds/mesa_dna_sim.

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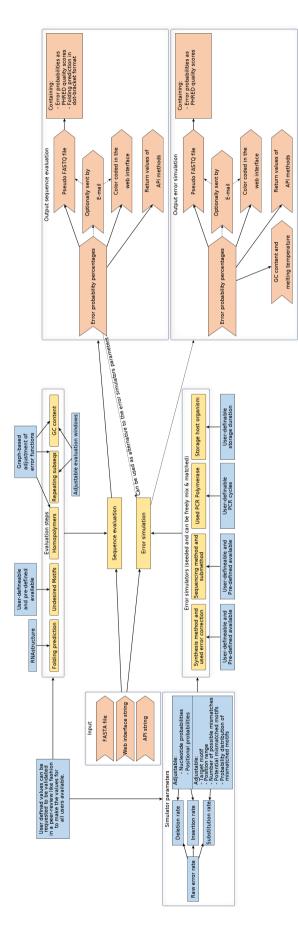


Fig. 1. Workflow of MESA: yellow rectangles denote methods used by the web application and are available in the API, orange arrows denote input / output and blue rectangles denote user-adjustable parameters.

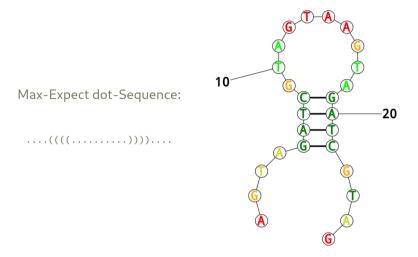


Fig. 2. A predicted secondary structure using the MaxExpect (Reuter and Mathews, 2010) algorithm in dot-bracket format (left) and the same structure as PNG (right).

Table 1. Color codings

| Color | Error source | Error type |
|-------|--------------|-----------------------|
| | Synthesis | Insertion |
| | Synthesis | Deletion |
| | Synthesis | Mismatch/Substitution |
| | Storage | Insertion |
| | Storage | Deletion |
| | Storage | Mismatch/Substitution |
| | Sequencing | Insertion |
| | Sequencing | Deletion |
| | Sequencing | Mismatch/Substitution |
| | PCR | Insertion |
| | PCR | Deletion |
| | PCR | Mismatch/Substitution |

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Table 2. Pre-defined DNA synthesis error profiles

| Method | Error-correction | Reference |
|------------------|-------------------|--------------------------|
| CSO ¹ | ErrASE | Kosuri and Church (2014) |
| CSO^1 | MutS | Kosuri and Church (2014) |
| CSO^1 | Consensus Shuffle | Kosuri and Church (2014) |
| $MBOP^2$ | OH^3 | Kosuri and Church (2014) |
| $MBOP^2$ | HTLH^4 | Kosuri and Church (2014) |
| $MBOP^2$ | ErrASE | Kosuri and Church (2014) |
| $MBOP^2$ | NB^5 | Kosuri and Church (2014) |
| $MBOP^2$ | NGS^6 | Kosuri and Church (2014) |

1: Column synthesized oligos, 2: Microarray based oligo pools, 3: Oligo hybridization based error correction, 4: High-temperature ligation / hybridization based error correction, 5: Nuclease based error correction, 6: NGS based error correction

Table 3. Pre-defined DNA sequencing error profiles

| M | lethod | Submethods | Reference |
|---|---------|--------------------------|--------------------------------|
| P | lumina | Single-Read & Paired-End | Schirmer <i>et al.</i> (2016) |
| | acBio | Subread & CCS | Weirather <i>et al.</i> (2017) |
| | anopore | 1D & 2D | Weirather <i>et al.</i> (2017) |

Table 4. Pre-defined PCR error rates

| Polymerase | Reference |
|------------|-------------------------|
| Taq | McInerney et al. (2014) |
| Pfu | McInerney et al. (2014) |
| Pwo | McInerney et al. (2014) |
| Phusion | McInerney et al. (2014) |
| | |

Table 5. Pre-defined mutation rates and -spectra

| Host organism | Reference |
|---|--|
| E. coli S. cerevisiae M. musculus H. sapiens | Lee et al. (2012) and Sung et al. (2016) Drake et al. (1998) and Sung et al. (2016) Drake et al. (1998) and Sung et al. (2016) Nachman and Crowell (2000) and Sung et al. (2016) |

Table 6. Pre-defined in-vitro depurination rates

| pН | Temperature (Kelvin) | Reference |
|----|----------------------|------------------|
| 8 | 293.15 | An et al. (2014) |
| 8 | 253.15 | An et al. (2014) |
| 8 | 193.15 | An et al. (2014) |
| 7 | 293.15 | An et al. (2014) |
| 7 | 253.15 | An et al. (2014) |
| 7 | 193.15 | An et al. (2014) |

Table 7. REST API methods

| Endpoint | Description | Input |
|-------------------|--|--|
| /api/homopolymer | Calculates homopolymer error probabilities | sequence (string), homopolymer_error_prob (dictionary), asHTML (boolean) |
| /api/gccontent | Calculates GC error probabilities | sequence (string), gc_windowsize (integer), gc_error_prob (dictionary), asHTML (boolean) |
| /api/kmer | Calculates error probabilities based on the occurrences of repeating subsequences/kmers | sequence (string), kmer_windowsize (integer), kmer_error_prob (dictionary), asHTML (boolean) |
| /api/subsequences | Calculates error probabilities based on the occurrences of undesired subsequences | sequence (string), enabledUndesiredSeqs (list of dictionaries), asHTML(boolean) |
| /api/fasta_all | Wraps do_multiple() which works with FASTA files, calls do_all() for every sequence of the FASTA file in another thread and sends the UUIDs of all results per email | UUID (string), key (string), sequence_list (list) |
| /api/max_expect | Calculates the secondary structure with the lowest free energy | sequence (string) |
| /api/getIMG | Loads the image with the given id and type from the Redis server and returns it if possible | ID (string), type (string) |
| /api/all | Loads the results from the Redis server with the UUID if possible and enables send_mail if the given sequence is longer than 1000 characters. Starts another thread for every request and uses all parameters of the request to calculate the different error probabilities and results | UUID (string), send_mail (boolean), sequence (string), email (string), all parameters oft he request |

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References

An, R., Jia, Y., Wan, B., Zhang, Y., Dong, P., Li, J., and Liang, X. (2014). Non-enzymatic depurination of nucleic acids: Factors and mechanisms. PLoS ONE, 9(12), e115950. Drake, J. W., Charlesworth, B., Charlesworth, D., and Crow, J. F. (1998). Rates of spontaneous mutation. Genetics, 148, 1667–1686.

Kosuri, S. and Church, G. M. (2014). Large-scale de novo DNA synthesis: technologies and applications. *Nature Methods*, **11**(5), 499–507. Lee, H., Popodi, E., Tang, H., and Foster, P. L. (2012). Rate and molecular spectrum of spontaneous mutations in the bacterium escherichia coli as determined by whole-genome sequencing. Proceedings of the National Academy of Sciences, 109(41), E2774–E2783.

McInerney, P., Adams, P., and Hadi, M. Z. (2014). Error rate comparison during polymerase chain reaction by dna polymerase. *Molecular Biology International*, **2014**, 1–8. Nachman, M. W. and Crowell, S. L. (2000). Estimate of the mutation rate per nucleotide in humans. *Genetics*, **156**, 297–304. Reuter, J. S. and Mathews, D. H. (2010). RNAstructure: software for RNA secondary structure prediction and analysis. *BMC Bioinformatics*, **11**(1).

Schirmer, M., D'Amore, R., Ijaz, U. Z., Hall, N., and Quince, C. (2016). Illumina error profiles: resolving fine-scale variation in metagenomic sequencing data. BMC Bioinformatics, 17(1).

Sung, W., Ackerman, M. S., Dillon, M. M., Platt, T. G., Fuqua, C., Cooper, V. S., and Lynch, M. (2016). Evolution of the insertion-deletion mutation rate across the tree of

life. *G3: Genes, Genomes, Genetics*, **6**(8), 2583–2591.

Weirather, J. L., de Cesare, M., Wang, Y., Piazza, P., Sebastiano, V., Wang, X.-J., Buck, D., and Au, K. F. (2017). Comprehensive comparison of pacific biosciences and oxford nanopore technologies and their applications to transcriptome analysis. *F1000Research*, **6**, 100.