

# Genetic Analysis of Egyptian Glandless Cotton<sup>1</sup>

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## ABSTRACT

The Egyptian cotton (*Gossypium barbadense* L.) strain, 'Bahtim 110', is glandless due to the expression of a dominant allele at a single locus while the glandless condition found in Upland cotton (*G. hirsutum* L.) is due to the expression of recessive alleles at two loci. The gene conditioning the glandless character in the Egyptian strain was found to be an allele at the  $G_l^1$  locus and was designated  $G_l^1$ . Potential use of this gene in glandless cottonseed breeding programs is discussed.

**Additional index words:** *Gossypium barbadense* L., *G. hirsutum* L., Linkage, Monosomes, Gossypol, Cottonseed breeding.

McMICHAEL (1959) reported the identification of a mutant in cotton in which the aerial plant parts were devoid of the lysigenous pigment glands typical of *Gossypium* spp. The glandless phenotype was determined by the combined action of the recessive genes  $gl_2$  and  $gl_3$  in the homozygous state (McMichael, 1960). Egyptian workers reported the identification of a single dominant mutation in *G. barbadense* L., following the irradiation of 'Giza 45' seed with <sup>32</sup>P, that imparted glandlessness. A strain of cotton homozygous for this new allele was released as 'Bahtim 110' (Afifi et al., 1965; 1966). Seed designated as Bahtim 110 were distributed to U.S. cotton workers. We began genetic analyses independently, but we combined our efforts when we became aware of each other's activities. This paper reports the combined results of these genetic analyses.

## MATERIALS AND METHODS

J.A. Lee performed genetic analyses in the *G. barbadense* background. The tester stocks that were used were derived from 'Sea Island' with the genotypes  $2(G_l^1G_l^1)$ ,  $2(G_l^1gl_3)$ , and  $2(gl_2G_l^1)$ , and 'Pima S-4' derived  $2(gl_2G_l^1)$  and  $2(gl_2gl_3)$ . R.J. Kohel transferred the glandless mutant to *G. hirsutum* by backcrossing to TM-1 (Kohel et al., 1970). Three and four backcrosses were completed with the glandless mutant "isoline" that was used in this study. All tester lines were *G. hirsutum*, except for one involving the doubled haploid experimental strain of *G. barbadense*, 3-79. The *G. hirsutum* standard, TM-1, has the genotype  $2(G_l^1G_l^1)$ . The tester,  $2(gl_2gl_3)$ , and the monosome for chromosome 12, were TM-1 isolines. The monomeric  $2(G_l^1gl_3)$  was obtained from J.A. Lee. Linkage analyses utilized T582 and T586, recessive and dominant multiple marker tester lines, respectively (Kohel, 1978).

## RESULTS AND DISCUSSION

### Inheritance

Crosses of the glandless Bahtim 110 [ $2(G_l^1)$ ] to normally glanded Sea Island (SI) and 3-79 produced  $F_1$  plants with reduced glandedness on the cotyledons. The  $F_2$  segregation ratios confirmed that the segre-

gation was that of a single partially dominant gene (Table 1).

Crosses of the glandless Bahtim 110 to normally glanded *G. hirsutum*, TM-1, produced a variable response. Cotyledonary classification for presence or absence of glands based on that established by Lee (1962) for the segregation of the  $gl_2$  and  $gl_3$  genes was used. Initially it was not clear whether the variable segregation pattern was due to the instability of the interspecific hybrid population, genetic modifiers, or a less distinct phenotypic expression associated with this mutant than that reported for  $gl_2$  and  $gl_3$  by Lee (1962). With additional backcrosses (BC) and progeny testing (Table 1) it was established that the segregation was that of a single partially dominant gene and that the heterozygote had a range of expression of glandedness. Cotyledons of heterozygous plants had either marginal or reduced glandedness while hypocotyls had either reduced or no glands. Subsequent heterozygous progeny from both the marginal and reduced glandedness classes gave both types of phenotypic expression in the cotyledons. Therefore, the phenotype of the heterozygote is described as sparse cotyledonary glandedness to include this range of variability, and to distinguish it from the phenotypic classes associated with segregation of the recessive  $gl_2$  and  $gl_3$  genes.

### Allelism Tests

Once the mode of inheritance of the Bahtim 110 glandless mutant reported by Afifi et al. (1965,1966)

**Table 1. Segregation of glandless Bahtim 110,  $2(G_l^1)$ , following crosses with normally glanded *G. barbadense* and *G. hirsutum* cottons.**

Phenotype	Segregation		Proposed ratio	Chi-square and probability
No. plants				
<i>G. barbadense</i>				
	$[2(G_l^1) \times SI]F_1$ , $[3-79 \times 2(G_l^1)]F_1$			
	a	b	Pooled	
Glandless	50	42	92	1
Reduced	112	88	200	2
Glanded	56	46	102	1
			394	
<i>G. hirsutum</i>				
	$[TM-1 \times 2(G_l^1)]F_1$			
	a	b	Pooled	
Glandless	17	47	64	1
Sparse	9	90	99	2
Glanded	5	34	39	1
			202	
	$TM-1[TM-1 \times 2(G_l^1)]S, BC$			
	a	b	c	Pooled
Glandless	23	287	74	384
Sparse	53	552	110	715
Glanded	24	306	53	383
				1482
	$TM-1[TM-1 \times 2(G_l^1)]BC$			
	a	b	c	Pooled
Sparse	12	15	50	77
Glanded	14	10	56	80
				157

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**Table 2. Allelic tests of glandless Bahtim 110,  $2(Gl^e)$ , with  $gl_2$  and  $gl_3$  *G. barbadense* and *G. hirsutum* testers.**

Segregation				Proposed ratio	Chi-square and probability	
Phenotype	No. plants					
<i>G. barbadense</i>						
	$[2(Gl^e) \times 2(gl_1Gl_3)]F_1$					
	SI	S-4	Pooled			
Glandless	133	82	215	3	$X^2 = 0.41$	
Glanded	52	26	78	1	$P = 0.70-0.50$	
			293			
	$[2(Gl^e) \times 2(Gl_1gl_3)]F_1$					
Glandless	143			10	$X^2 = 3.32$	
Sparse	37			2	$P = 0.20-0.10$	
Glanded	71			4		
	251					
	$[2(Gl^e) \times 2(gl_1gl_3, S-4)]F_1$					
Glandless	195			13	$X^2 = 1.37$	
Sparse	33			2	$P = 0.50-0.30$	
Glanded	11			1		
	239					
<i>G. hirsutum</i>						
	$[2(Gl_1gl_3) \times 2(Gl^e), TM-1]F_1$					
Glandless	73			10	$X^2 = 2.83$	
Sparse	9			2	$P = 0.30-0.20$	
Glanded	22			4		
	104					
	$[2(gl_1gl_3), TM-1 \times 2(Gl^e)]F_1$					
	a	b	c	Pooled		
Glandless	42	64	149	255	13	$X^2 = 8.90$
Sparse	7	11	37	55	2	$P = 0.02-0.01$
Glanded	6	13	11	30	1	
				340		
	(Monosome $12 \times 2(Gl^e), TM-1$ )					
	F <sub>1</sub>	F <sub>2</sub>				
Glandless (monosome)	6	all glandless (7 plants)				
Glanded (disome)	17	segregation (1:2:1)			$X^2 = 0.12$	
					$P = 0.95-0.90$	

was found to express in both *G. barbadense* and *G. hirsutum*, tests were made to determine the relation, if any, with the  $gl_2$  and  $gl_3$  loci. Bahtim 110 was crossed to the Sea Island monomerics,  $2(Gl_2gl_3)$  and  $2(gl_2Gl_3)$ , and Pima S-4 monomeric,  $2(gl_2Gl_3)$ . In both monomeric cross combinations the  $F_1$  seedlings had glandless cotyledons. Segregation of the  $F_2$  from the crosses of the  $Gl_3$  monomeric,  $2(gl_2Gl_3)$ , combination fit a 3 glandless to 1 glanded ratio (Table 2). This segregation suggested that the Bahtim 110 glandless mutant gene was an allele at the  $Gl_2$  locus.

With the information from the  $Gl_3$  monomeric  $F_2$  and knowing the phenotypes of both monomerics  $\times$  Bahtim 110  $F_1$ 's, our postulated ratio in the Bahtim 110  $\times$   $Gl_2$  monomeric,  $2(Gl_2gl_3)$ ,  $F_2$  was 10 glandless : 2 sparse : 4 glanded cotyledonary classes. The data fit this expected ratio (Table 2).

With the assumption that glandless Bahtim 110 also carries a  $Gl_3$  allele, one can predict that crosses with the recessive glandless,  $2(gl_2gl_3)$ , would produce a seedling with cotyledons that were glandless in the  $F_1$  and in the  $F_2$  would have the phenotypic ratio of 13 glandless : 2 sparse : 1 glanded. Segregation of Bahtim 110  $\times$  Pima S-4 glandless,  $2(gl_2gl_3)$ ,  $F_2$  fit this expected ratio (Table 2).

The glandless Bahtim 110 allele,  $Gl^e$ , transferred to *G. hirsutum* (TM-1), was utilized in a similar series of tests with *G. hirsutum* testers. Crosses of  $2(Gl^e)$  with the monomeric,  $2(Gl_2gl_3)$ , produced  $F_1$  seedlings with

**Table 3. Detailed analysis of  $Gl_2^e$  and  $N_1$  linkage.**

Segregation							
Phenotype	No. plants				Chi-square analysis		
$[2(Gl_2N_1) \times 2(Gl_2^eN_1)]2(Gl_2N_1)$	a	b	c	Pooled	Source	Chi-square	P
$Gl_2^eN_1$	49	24	14	87	$Gl_2^e$ vs. $Gl_2$	1.00	0.50-0.30
$Gl_2^eN_1$	13	10	10	33	$N_1$ vs. $n_1$	2.78	0.10-0.05
$Gl_2N_1$	23	10	5	38	Linkage	30.61	<0.01
$Gl_2N_1$	32	13	22	67	Recombination percent =		
				225			31.56 $\pm$ 3.10

glandless cotyledons, and the  $F_2$  segregated with the ratios of the phenotypic classes 10 glandless : 2 sparse : 4 glanded (Table 2).

The Bahtim 110 allele,  $Gl^e$ , was crossed to the *G. hirsutum* glandless isolate,  $2(gl_2gl_3)$ , from its native background and after each of two backcrosses to TM-1. The segregation in the resulting three *G. hirsutum*  $F_2$  seedling populations did not fit the 13 glandless : 2 sparse : 1 glanded phenotypic ratio as well as in the *G. barbadense*  $F_2$  population (Table 2). However, all the other data show that  $Gl^e$  is an allele of  $Gl_2$  and these data do not refute that finding.

The Bahtim 110 mutant is a partially dominant mutant allele at the  $Gl_2$  locus. This single dominant allele mimics the same control of gland expression as that obtained by the combined effects of the duplicate recessive genes  $gl_2$  and  $gl_3$  discovered by McMichael (1959). We propose the gene symbol  $Gl_2^e$  to designate this new dominant allele at the  $Gl_2$  locus.

### Linkage and Monosome Analyses

Two additional tests were performed with the  $Gl_2^e$  TM-1 isolate. It was crossed with the monosome for chromosome 12 for verification of chromosome location, and with the multiple marker lines, T582 and T586, for linkage analysis. When the  $Gl_2^e$  isolate was crossed onto the monosome, the resulting  $F_1$  seedlings segregated into glandless and sparse cotyledonary glandedness classes. When transplanted to the field the glandless plants expressed the monosome 12 phenotype and the sparse glanded plants expressed the normal disomic phenotype (Table 2).

When the crosses for the linkage analysis were begun, allelism of  $Gl_2^e$  to the  $Gl_2$  locus was not known. The only genetic marker in the multiple marker lines that is resident on the same chromosome as  $Gl_2$ , chromosome 12, is  $N_1$  (Naked seed). Therefore the linkage relation between  $Gl_2^e$  and  $N_1$  is the test combination of primary interest. The linkage analysis results and detailed analysis of the  $Gl_2^e-N_1$  linkage (Table 3) revealed a significant  $Gl_2^eN_1$  linkage with 32% recombination. Recombination percentage between  $gl_2-N_1$  was  $37.61 \pm 4.66$  (Kohel, unpublished).

### Practical Implications

The  $Gl_2^e$  allele offers a new alternative genetic system to glandless cottonseed breeding programs. Use of the  $Gl_2^e$  allele provides greater ease and simplicity in genetic manipulation because it is a single dominant gene as opposed to the previously used genetic system that involved the duplicate recessives  $gl_2$  and  $gl_3$ . The single partially dominant gene would result in easier recognition in segregating populations and

a resulting decrease in population size and number of generations required for transference to different backgrounds.

The  $Gl_2^e$  allele may also be combined with  $gl_3$  to solve another problem of producers of glandless cottonseed. The very low tolerance requirements for seed gossypol require stringent quality control. One source of contamination is through outcrossing with glanded cottons that produce glanded  $F_1$  seeds. A glandless cotton that combined the  $Gl_2^e$  and  $gl_3$  genes,  $2(Gl_2^e gl_3)$ , when crossed with a normally glanded cotton,  $2(Gl_2 Gl_3)$ , would produce glandless  $F_1$  seeds; the seedling genotype would be  $Gl_2^e Gl_2 Gl_3 gl_3$ . This system could be useful in preventing contamination due to chance outcrossing with glanded plants in production fields.

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