

Algorithms for the Correction of Photobleaching

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Preface

Ways to read

This thesis may be read on the web at <https://rorynolan.github.io/phdthesis/>. If you are reading this on the web now but would like a PDF version, click on the download symbol at the top left of the page (to the right of the **A**) and select PDF.

Apportion of credit

For clarity, I include the following reliable rules of thumb:

- Molecular biology was done by Maro Iliopoulou and Luis Alvarez.
- Imaging was done by Maro Iliopoulou, Luis Alvarez and Sergi Padilla-Parra.

- The idea that correction for bleaching was the crucial step for FFS analysis was formulated by Luis Alvarez, Sergi Padilla-Parra and I.
- I formulated the solutions for how to correctly correct for bleaching, i.e. the automatic parameter choice and the Robin Hood algorithm.
- I wrote all of the software and maintain all of it.
- All FFS analysis was performed using my software. The software was used to analyse data by Maro Iliopoulou, Luis Alvarez, Sergi Padilla Parra and I.
- Structural modelling was done by Thomas Bowden and Yasunori Watanabe.
- On all papers where I am the first listed author, I wrote the paper, taking suggested amendments and augmentation from other listed authors. The NSMB paper ([Iliopoulou et al., 2018](#)) was written by Sergi; I also made significant contributions to the writing of that manuscript but my main role in that project was in data analysis.

Publications

I hereby list my publications. These can all be downloaded from <https://github.com/rorynolan/phdthesis/tree/master/papers>, where the naming convention is JournalYEAR.pdf.

First author

- R. Nolan and S. Padilla-Parra. “filestrings: An R package for file and string manipulation”. In: *Journal of Open Source Software* 2.14 (2017).
- R. Nolan and S. Padilla-Parra. “exampletestr—An easy start to unit testing R packages”. In: *Wellcome Open Research* 2 (2017).
- R. Nolan, L. Alvarez, J. Elegheert, et al. “nandb—number and brightness in R with a novel automatic detrending algorithm”. In: *Bioinformatics* 33.21 (2017).
- R. Nolan, M. Iliopoulou, L. Alvarez, et al. “Detecting protein aggregation and interactions in live cells: a guide to Number and Brightness”. In: *Methods* (2017).
- R. Nolan and S. Padilla-Parra. “ijtiff: An R package providing TIFF I/O for ImageJ users”. In: *Journal of Open Source Software* 3.23 (2018).
- R. Nolan, L. Alvarez, S. C. Griffiths, et al. “Calibration-Free In-Vitro Quantification of Protein Homo-Oligomerization Using Commercial Instrumentation and Free, Open Source Brightness Analysis Software”. In: *Journal of Visualized Experiments* 137 (2018).

Co-first author

- M. Iliopoulou, R. Nolan, et al. “A dynamic three step mechanism drives the HIV-1 pre-fusion reaction”. In: *Nat. Struct. Mol. Biol.* 25.9 (2018).

Other

- D. M. Jones, L. A. Alvarez, R. Nolan, et al. “Dynamin-2 stabilizes the HIV-1 fusion pore with a low oligomeric state”. In: *Cell reports* 18.2 (2017).
- G. M. Jakobsdottir, M. Iliopoulou, R. Nolan, et al. “On the whereabouts of HIV-1 cellular entry and its fusion ports”. In: *Trends in molecular medicine* (2017).
- Q. F. Wills, E. Mellado-Gomez, R. Nolan, et al. “The nature and nurture of cell heterogeneity: accounting for macrophage gene-environment interactions with single-cell RNA-Seq”. In: *BMC genomics* 18.1 (2017).

In preparation

- The *Robin Hood* algorithm (described in section 3.9) will be submitted to a journal after the submission of this thesis but before the end of my PhD contract.

Acronyms used in this thesis

Acronym	Term
ACF	AutoCorrelation Function
ALEX	Alternating Laser EXcitation
APD	Avalanche Photo-Diode
BMC	BioMed Central
CCD	Charge-Coupled Device
ccN&B	cross-correlated Number and Brightness
CCR5	C-Chemokyne Receptor 5
CD4	Cluster of Differentiation 4
CMOS	Complementary Metal–Oxide Semiconductor
CXCR4	C-X-C motif Chemokyne Receptor 4
DNA	Deoxyribinucleic Acid
dSTORM	direct STochastic Optical Reconstruction Microscopy
EM	Electron Microscopy
EMCCD	Electron-Multiplied Charge-Coupled Device
FCS	Fluorescence Correlation Spectroscopy
FFS	Fluorescence Fluctuation Spectroscopy
FIJI	FIJI Is Just ImageJ
FKBP	FK Binding Protein

FRET	Forster Resonance Energy Transfer
GaAsP	Gallium Arsenide Phosphide
GFP	Green Fluorescent Protein
HIV	Human Immunodeficiency Virus
HOMO	Highest Occupied Molecular Orbital
HXB2	Subtype B2 HIV-1 isolate
HyD	Hybrid Detector
I/O	Input/Output
JR-FL	JR (patient name) Frontal Lobe
LOMO	Lowest Occupied Molecular Orbital
MSD	Mean Squared Displacement
mTFP	monomeric Turquoise Fluorescent Protein
N&B	Number and Brightness
NSM	Non-Stationary Mean
NSMB	Nature Structural and Molecular Biology
NSV	Non-Stationary Variance
PCF	Pair Correlation Function
PDB	Protein DataBase
PDF	Portable Document Format

PhD	Philosophiae Doctor
PMT	Photon Multiplier Tube
QE	Quantum Efficiency
RNA	Ribonucleic Acid
sCMOS	scientific Complementary Metal–Oxide Semiconductor
SP	Scanning Platform
TIFF	Tagged Image File Format
TIR	Total Internal Reflection
TIRF	Total Internal Reflection Fluorescence
TZM-bl	Stable cell line expressing CD4 and CCR5

Mathematical symbols used in this thesis

Symbol	Meaning
θ	angle of incident light ray
B	apparent brightness of entities
N	apparent number of entities
G	autocorrelation
φ	concentration

corr	correlation
θ_c	critical angle
crosscor	cross-correlation
B_{cc}	cross-correlated brightness
σ_{cc}^2	cross-variance
\tilde{x}	deviation of x from the expected value μ of X
D	diffusion constant
J	diffusive flux
ϵ	entity brightness
E	expectation operator
τ	exponential smoothing parameter
j	frame number
S_0	highest occupied molecular orbital
I	intensity
l	length parameter of boxcar
T_1	lowest energy triplet state
S_1	lowest unoccupied molecular orbital
msd	mean squared displacement
μ	mean; expected value

n	moment number; number of entities; number of replicates
\mathbb{N}	natural numbers (excluding zero)
\mathbb{N}_0	natural numbers (including zero)
d	number of dimensions
α	number of swaps
p	particle position; pixel position
x	position in space; horizontal position; instance of random variable X
k	positive real number
X	random variable
\mathbb{R}	real numbers
n_2	refractive index of cover slip
n_1	refractive index of sample on cover slip
τ_D	residence time of particle (in confocal volume)
i	slice number
\exp	the exponential function
t	time
Var	variance function
σ^2	variance in intensity
y	vertical position

Acknowledgements

Thanks to my parents Kate and Dave who were good enough to house me rent-free for 18 years. They encouraged me academically after it became clear that I was not the next Kevin Kilbane. Thanks to my wife Naomi who puts up with me now that my parents have had enough of me. Thanks to my brother Brendan who taught me (perhaps wrongly) that maths is cool.

Thanks to my supervisor Sergi Padilla-Parra who had many good ideas of what to study, for his intelligent input and generosity with time. Moreover, thanks to him for emphasising that it's possible to work too hard. It's largely thanks to his attitude that my time in Oxford was so enjoyable.

Thanks to all of my lab mates: Dan, Keith, Xenia, Raquel, Luis, Margarita, Marina, Maro, Marina, Maria, Chad and Irene; it was interesting and fun to spend time with you all. I'll look back upon my time in the lab fondly, I hope you can say the same.

Thanks to my friends Kathrin and Zammy from my PhD programme; we shared the journey to becoming doctors and come out of the other end with lifelong friendships as well as new titles.

Thanks to my college friends Bas, Jessica, Francis, Sarah, Andy, Frankie, Jonny,

Suze, Walid, Sophia, Martin, Max, Maj, Nirmalie, Luiza, LD and many more. It was a privilege to have so many good friends at once.

There are plenty more people I should thank, but I'm tired so I'm going to stop now.

Abstract

The measured intensity (ideally in units of photon counts) of a fluorescent sample over time constitutes a time-series called an *intensity trace*. The idea of *fluorescence fluctuation spectroscopy* (FFS) is to extract information from intensity traces.

Photobleaching is the phenomenon of the breaking of fluorophores (light emitters) over time. Photobleaching causes fluorescent signal to diminish over time. This changes the intensity trace, introducing a downward trend.

Many quantitative methods in FFS implicitly assume that there is no bleaching in the data. Hence, data with significant levels of photobleaching must be corrected prior to the application of equations and algorithms in these fields. This correction is often termed *detrending*, since its aim is to remove the downward trend in the data introduced by photobleaching.

Previous detrending methods have two main caveats:

1. They rely on either fitting or smoothing, both of which approximate data as continuous. This is inappropriate for fluorescence intensity data, which is count data (i.e. discrete, not continuous).
2. They require the user to choose a detrending parameter. The choice of this parameter is crucial to the success or failure of the detrending routine, yet instructions on how to choose it did not exist.

The work in this thesis solves problems 1 and 2 above by means of an automatic (no user input required) parameter finding routine and a new detrending algorithm which treats the data as discrete and avoids fitting and smoothing, thereby avoiding the approximation of non-continuous data as continuous.

These advancements are then used in a study investigating the stoichiometry of the interaction of the HIV-1 virus' envelope with its cellular receptors and co-receptors over time. This is the first study of its kind in live cells and it was facilitated by the advances in detrending presented in this thesis.

Chapter 1

Introduction

1.1 Fluorescence

Definition 1.1. *Fluorescence* is the emission of photons related to the process of *relaxation* for a given molecule from an electronically *excited* state (S_1) to the *ground* state (S_0).

These two electronic states (S_0 and S_1) of a molecule are defined as the highest occupied molecular orbital (S_0 , HOMO) and the lowest unoccupied molecular orbital (S_1 , LUMO). Each electronic state has various associated *vibrational* states between which *non-radiative* transitions can occur. An electron can be *excited*

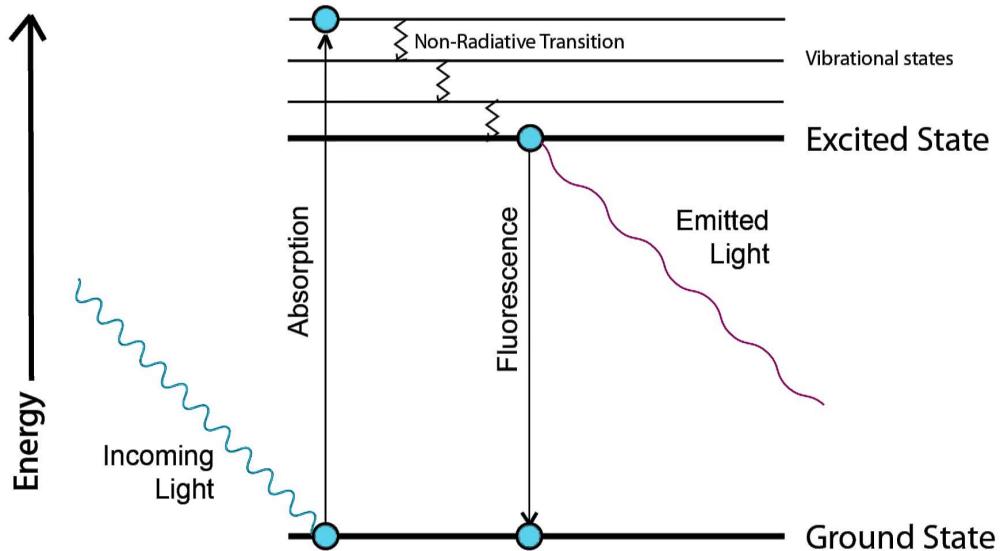


Figure 1.1: Jablonski diagram of the process of fluorescence. Absorption causes excitation, relaxation causes the emission of light.

from the ground state S_0 into the higher energy excited state S_1 by absorption of a photon of the appropriate energy. The electron will remain in the excited state (possibly undergoing non-radiative transitions through vibrational states) for a period of nanoseconds before *relaxing* to the lower energy ground state, losing energy by means of emitting a photon of that energy. This is shown by means of a *Jablonski* diagram (figure 1.1).

Definition 1.2. The *fluorescence lifetime* of an electron is the amount of time it remains in the excited state before returning to the ground state.

Definition 1.3. A *fluorophore* is a fluorescent chemical compound that can re-emit light upon light excitation.

Fluorophores under constant excitation emit light one photon at a time according to Poisson statistics.

1.1.1 Phosphorescence

Definition 1.4. Excited electrons can undergo a *forbidden* transition into the *triplet* state T_1 , where they remain for a period of milliseconds before relaxing to the ground state with the emission of a photon. This is *phosphorescence*.

1.1.2 Photobleaching

In reality, an incident photon can *break* the fluorophore such that it is subsequently unable to emit light. A fluorophore to which this has happened is said to be *photobleached*.

1.2 Confocal light microscopy

All of the images used in my PhD were collected on a confocal microscope. This type of microscope guarantees that only in-focus light is collected at the detector.

See figure 1.2.

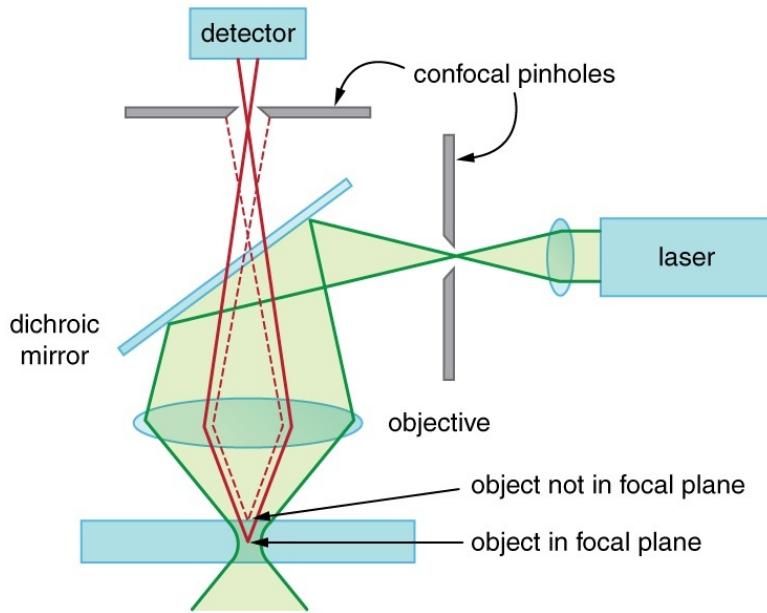


Figure 1.2: Confocal microscope light path showing how out of focus light does not make it to the detector.²

Definition 1.5. The *confocal volume* is the *in-focus* volume within a sample that is efficiently detected using a system designed with confocal optics ([FCSXpert, 2018](#)).

An image is acquired on a confocal microscope by scanning this apparatus across a sample, collecting one pixel at a time.

1.2.1 Detectors

There are many types of confocal microscope detectors. I will discuss the most common ones here. Most rely on the photoelectric effect ([Einstein, 1905a](#)), a

phenomenon whereby incident light upon a material causes electrons to dissociate from that material.

Definition 1.6. The *quantum efficiency* (QE) of a detector is the fraction of fluorescent signal that is reported by the detector.

1.2.1.1 Photon multiplier tubes

Photon multiplier tubes (PMTs) use the photoelectric effect. Dissociated electrons are accelerated through a potential difference towards a cathode where their accumulated kinetic energy is used to release more electrons. These are then accelerated in vacuum towards another cathode and so on for a predefined number of these accelerations until at last the electrons are discharged into an anode where the current is measured ([Hammamatsu, 2007](#)); see figure 1.3. This current corresponds to the incident light intensity.

The QE of standard PMTs is approximately 25% for blue-green light. This is because many incident photons on the multi-alkali material of regular PMT photocathodes fail to free any electrons. This problem gets worse at higher wavelengths where photons have lower energy. PMTs also suffer significantly from thermal noise (whereby dissociated electrons are created due to heat energy).

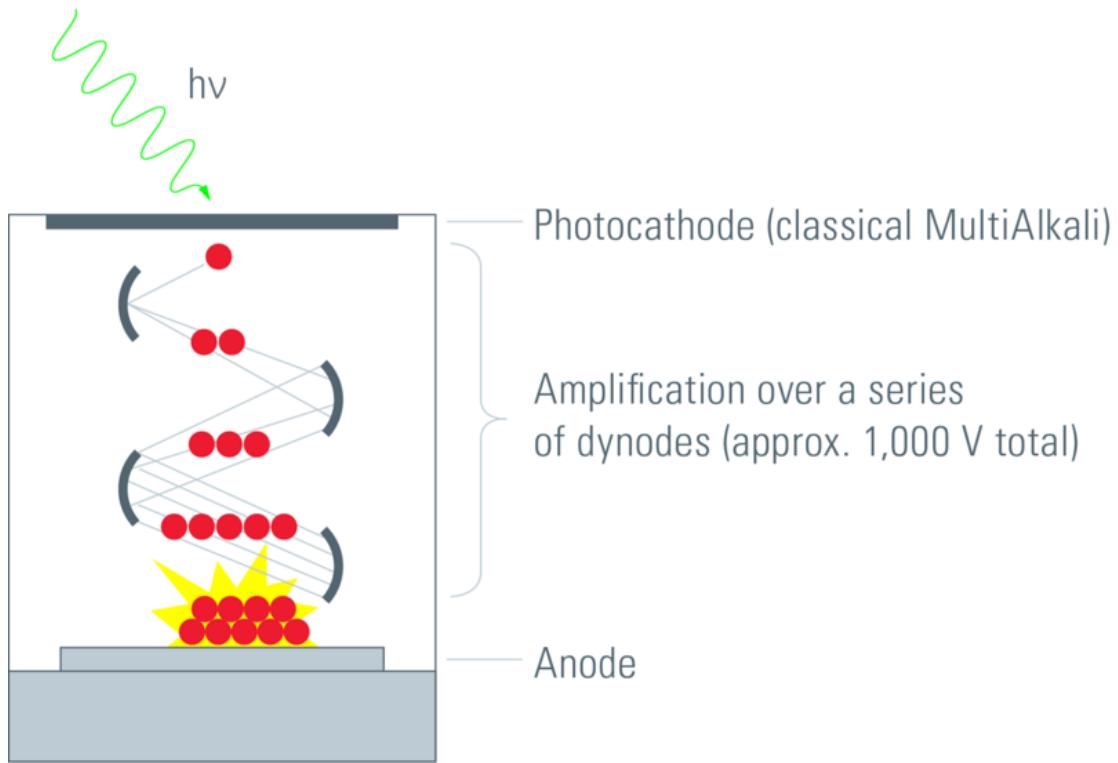


Figure 1.3: PMT detector setup. Electrons are dissociated by photons at the photocathode, accelerated towards various other cathodes where more and more are freed and then finally they arrive at the anode where the current is measured (Leica, 2012).

1.2.1.2 Avalanche photo-diodes

Definition 1.7. *Avalanche multiplication* in semiconductors occurs when free electrons which are moving in an electric field across the semiconductor collide with bound electrons, freeing them to move and indeed to free more bound electrons. This compounding increase in free electrons in the electric field leads to an increase in the electric current.

Avalanche photo-diodes (APDs) are semiconductors that exploit the photoelectric effect together with avalanche multiplication (the photoelectric effect starts the avalanche) to convert light into measurable electric current; see figure 1.4. They are somewhat analogous to PMTs, using the avalanche effect in place of the series of cathodes.

The QE of APDs can be as high as 45%, however their dynamic range is low and they can only function with low-intensity light. APDs are less prone to thermal noise than PMTs.

1.2.1.3 Hybrid detectors

APDs have better sensitivity and lower noise, however PMTs have a larger dynamic range. In hybrid detectors (HyDs), PMT and APD technology is combined

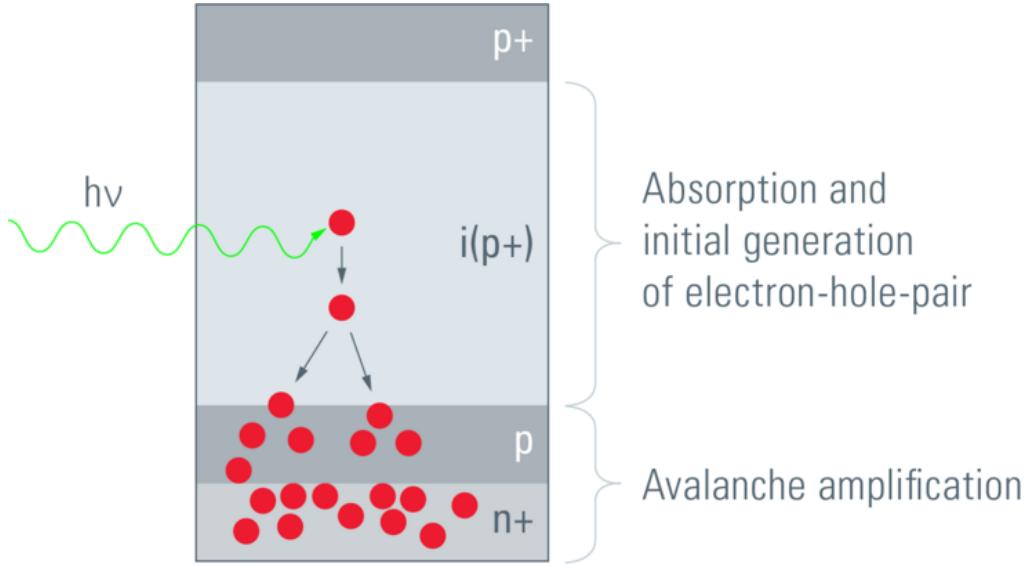


Figure 1.4: APD detector setup. The incident photon dissociates an electron. Given the electric field across the semiconductor, this electron is accelerated, initiating an avalanche (Leica, 2012).

to get the best of both worlds. With HyDs, photons are converted to electrons at the photocathode, then accelerated in vacuum towards a semiconductor where they initiate an electron avalanche. See figure 1.5. The difference between HyDs and APDs is this acceleration step.

HyDs with GaAsP (gallium arsenide phosphide) photocathodes achieve a quantum efficiency of up to 50% (Leica, 2012). They also suffer least from detector afterpulsing, a phenomenon which can cause real signal pulses to be followed by a feedback pulse at a later time (Zhao et al., 2003).

With hybrid detectors, the electric current pulse caused by a photon striking the

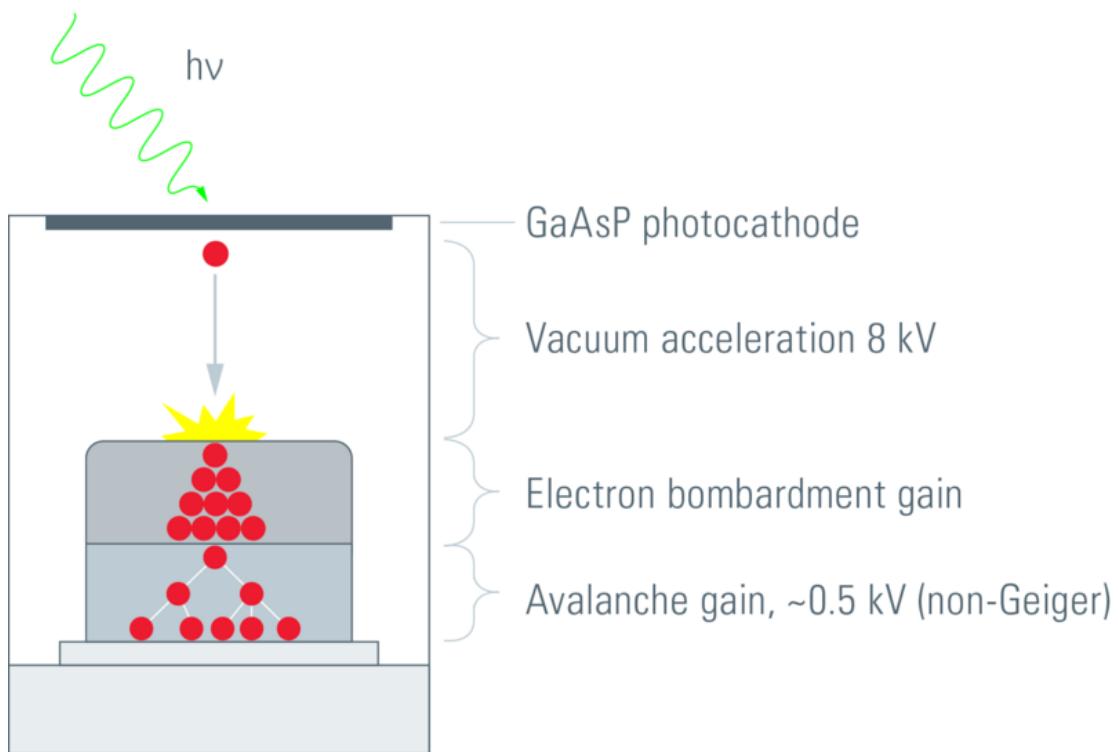


Figure 1.5: HyD detector setup. A photon dissociates an electron from the photocathode, this electron is then accelerated PMT-style towards an APD-style semiconductor setup, triggering an avalanche (Leica, 2012).

photocathode is strong (well above background) and sharp (the pulse is very short-lived). This means that it is easy to detect the arrival of single photons at the photocathode. This enables HyDs to have a *photon-counting* mode whereby the readout is in units of photons (not electron current). This is a much more physically relevant readout — given that we are interested in measuring fluorescence — which is very convenient for many biological applications, not least fluorescence fluctuation spectroscopy (FFS, discussed in section 1.5). For this reason, the HyD was the detector of choice for all of the acquisitions in my thesis.

Remark. Henceforth, intensity counts will be assumed to be in units of photons.

1.3 Intensity traces

Consider figure 1.6. A fluorophore enters the confocal volume, is excited there and emits photons which are collected by the detector. When the fluorophore is not in the confocal volume (start and end), no photons are detected. When the particle is in the confocal volume, photons it emits are collected at the detector. Different numbers of photons are collected per unit time (per ms here).

Definition 1.8. The time-series of intensity counts in a given pixel or image is referred to as its *intensity trace*.

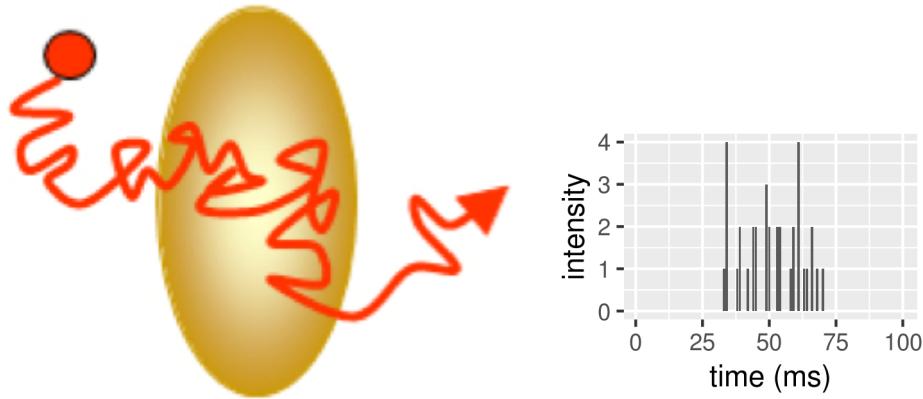


Figure 1.6: Left: a fluorophore diffusing through the confocal volume (Padilla-Parra, 2009). Right: the intensity trace due to this fluorophore. This intensity trace has mean 0.35 and variance 0.69.

1.4 Diffusion

Definition 1.9. *Diffusion* is the net movement of molecules or atoms from a region of high concentration (or high chemical potential) to a region of low concentration (or low chemical potential) as a result of random motion of the molecules or atoms.

Fick's first law of diffusion relates the concentration to the diffusive flux, assuming a steady state. It describes how the flux goes from high to low concentration areas, with magnitude proportional to the concentration gradient. The law is:

$$J = -D \frac{d\varphi}{dx} \quad (1.1)$$

where

- J is the diffusive flux
- D is the diffusion constant
- x is the position in space
- φ is the concentration at position x .

Definition 1.10. *Brownian motion* is the random motion of particles suspended in a fluid resulting from their collision with the fast-moving molecules in the fluid (Einstein, 1905b).

Definition 1.11. In an ensemble of N particles, each of which occupies position $x_i(t)$ at time t , the *mean squared displacement* of the particles in the ensemble is

$$\text{msd}(t) = \frac{1}{N} \sum_{i=1}^N [x_i(t) - x_i(0)]^2 \quad (1.2)$$

For free Brownian motion in 1 dimension, the expected value of the MSD is $\text{msd}(t) = 2Dt$, where D is the diffusion constant. For free Brownian motion in d dimensions, the expected value of the MSD is $\text{msd}(t) = 2dDt$, reflecting the fact that free Brownian motion in d dimensions is one-dimensional free Brownian motion happening simultaneously and independently in each individual dimension.

These definitions are the formally correct ones. In biophysics, however, both of

these processes (diffusion and Brownian motion) are most often referred to as *diffusion*. I will follow this convention.

1.4.1 Anomalous diffusion

Anomalous diffusion is a diffusion process whereby equation (1.2) no longer holds and the relationship between MSD and time becomes

$$\text{msd}(t) = 2dDt^\alpha \quad (1.3)$$

where $\alpha \neq 1$. $\alpha > 1$ is known as *superdiffusion* (faster than normal) and $\alpha < 1$ is known as *subdiffusion* (slower than normal).

1.4.2 Diffusion in the presence of barriers

If barriers to diffusion are present, neither equations (1.2) and (1.3) hold. The pair correlation function approach (section 1.6.3.1) can be used to image barriers to diffusion (Digman and Gratton, 2009).

1.5 Fluorescence fluctuation spectroscopy

Broadly, fluorescence fluctuation spectroscopy (FFS) is the analysis of the intensity fluctuation of a fluorescence signal (Chen et al., 1999). This very often takes the form of moment analysis (Qian and Elson, 1990). Briefly, moment analysis is an attempt to extract data from a distribution of values using its *moments*. The first moment of a distribution is its mean value, the second moment is its variance. The n^{th} moment of a random variable X with expected value $E[X] = \mu$ for $n > 1$ is $E[(X - \mu)^n]$.

Intensity traces can be viewed as distributions with moments. For example, the intensity trace in figure 1.6 has mean 0.35 and variance 0.69.

1.5.1 Number and brightness

Number and brightness (N&B, Digman et al. (2008)) is an FFS technique for quantifying the oligomeric states of fluorescently labelled proteins. What follows is a mathematical description of the technique.

Definition 1.12. An *entity* is a set of molecules which are chemically bound.

Definition 1.13. The brightness ϵ of an entity is the mean number of photon detector counts it gives per unit time when in the illumination (confocal) volume.

For an image series where the i^{th} slice in the stack is the image acquired at time $t = i$, for a given pixel position (x, y) , we define $\langle I \rangle$ as the mean intensity of that pixel over the image series and σ^2 as the variance in that intensity. Define n as the mean number of entities in the illumination volume corresponding to that pixel. Assuming that all entities are mobile, we have

$$N = \frac{\langle I \rangle^2}{\sigma^2} = \frac{\epsilon n}{1 + \epsilon} \quad (1.4)$$

$$B = \frac{\sigma^2}{\langle I \rangle} = 1 + \epsilon \quad (1.5)$$

where N and B are referred to as the *apparent number* and *apparent brightness* respectively. This gives

$$n = \frac{\langle I \rangle^2}{\sigma^2 - \langle I \rangle} \quad (1.6)$$

$$\epsilon = \frac{\sigma^2}{\langle I \rangle} - 1 \quad (1.7)$$

These relations are derived using a moment analysis technique which

was originally applied to molecules in solution (Qian and Elson, 1990).

With a scanning confocal microscope in analog mode, we must use three correction terms: 1. The proportionality constant S , which is the conversion factor between photons detected and the number of counts returned by the analog electronics. 2. The offset (bias) due to the analog electronics in the level of the background. 3. The readout noise σ_0^2 is the variance in this background signal (Dalal et al., 2008).

Then, if all entities are mobile, we have

$$N = \frac{(\langle I \rangle - \text{offset})^2}{\sigma^2 - \sigma_0^2} = \frac{\epsilon n}{1 + \epsilon} \quad (1.8)$$

$$B = \frac{\sigma^2 - \sigma_0^2}{\langle I \rangle - \text{offset}} = S(1 + \epsilon) \quad (1.9)$$

$$n = \frac{(\langle I \rangle - \text{offset})^2}{\sigma^2 - \sigma_0^2 - S(\langle I \rangle - \text{offset})} \quad (1.10)$$

$$\epsilon = \frac{\sigma^2 - \sigma_0^2}{S(\langle I \rangle - \text{offset})} - 1 \quad (1.11)$$

— Nolan et al. (2017b)³

The quantity ϵ is a relative measure of oligomeric state. That is, ϵ will be twice

³This is a formulation of N&B that I wrote in a 2017 review of the technique.

as big for dimers as it is for monomers, three times as big for trimers as it is for monomers and so on.

The way that N&B experiments to determine unknown oligomeric states are generally done is as follows:

1. For a given laser power and fluorophore with a system where all entities are known to be monomeric, measure the brightness ϵ . Call this $\epsilon_{\text{monomer}}$.
2. With the same laser power and fluorophore but now with a system where the oligomeric state is unknown, measure the brightness ϵ again. Call this $\epsilon_{\text{unknown}}$.
3. The unknown oligomeric state is equal to $\epsilon_{\text{unknown}}/\epsilon_{\text{monomer}}$.

Number and brightness has specific pixel dwell-time and frame rate requirements.

These were first articulated in my 2017 review of N&B ([Nolan et al., 2017b](#)).

Definition 1.14. The *pixel dwell time* t_{dwell} of a scanning confocal microscope is the amount of time it spends collecting photons at each given pixel before moving on to the next pixel.

It is essential that the pixel dwell time is the same for each pixel, particularly for FFS. This is something that must be carefully verified on each instrument. This can be done with a chroma-slide. The photon counts measured from a chroma-slide should follow a Poisson distribution. If instead the distribution is

super-Poissonian (has a greater variance than its mean), this is indicative of a non-constant dwell-time.

Definition 1.15. The *frame time* t_{frame} of a scanning confocal microscope is the amount of time it takes to acquire the data for all of the pixels in a whole frame. That is the length of time from the start of detection of photons from the first pixel to the end of detection of photons from the last pixel.

Definition 1.16. The mean *residence time* τ_D of a particle in the confocal volume is the average length of time that a particle that enters the confocal volume resides in that volume before exiting.

The requirement is $t_{\text{dwell}} \ll \tau_D \ll t_{\text{frame}}$. This ensures that:

1. When acquiring photons at a given pixel, the underlying configuration of entities is constant (there's not enough time for the entities to move and change their configuration).
2. When the scanner returns to a given pixel (one frame time later), the underlying configuration has changed totally since the last time the scanner was at this pixel (because so much time has passed, all of the diffusing entities have moved a lot in the meantime).

Both of these points are implicitly assumed in the derivation of the N&B equations so it is essential to get these acquisition parameters right. This is discussed at

length in [Nolan et al. \(2017b\)](#).

An important property of N&B is that if all fluorescent particles are immobile, then $B = 1$. This is because photon emission from stationary sources happens according to a Poisson distribution. Poisson distributions have variance equal to mean, this implies $\sigma^2 = \langle I \rangle$ which gives $B = \frac{\sigma^2}{\langle I \rangle} = 1$.

The N&B technique is fraught with technical difficulties. Principal of these is the problem of photobleaching. Most of my PhD focused on corrections for photobleaching. This is discussed in chapter [3](#).

1.6 Fluorescence correlation spectroscopy

Fluorescence correlation spectroscopy (FCS) is the correlation analysis of fluorescence intensity fluctuations. For this reason, FCS can be described as a subfield of FFS ([Jameson et al., 2009](#)). In practice, FFS is mostly used to refer to the non-FCS parts of the whole FFS field. I will follow that convention.

First, let us introduce some concepts from statistics.

Definition 1.17 (statistics). The *correlation* between two random variables X and Y with expected values μ_X and μ_Y and standard deviations σ_X and σ_Y is

$$\text{corr}(X, Y) = \frac{E[(X - \mu_X)(Y - \mu_Y)]}{\sigma_X \sigma_Y} \quad (1.12)$$

where E is the expectation operator.

Definition 1.18 (statistics). *Autocorrelation* $G(X; \tau)$, is the correlation of a signal X with a delayed copy of itself as a function of the delay τ .

$$G(X; \tau) = \text{corr}(X_t, X_{t+\tau}) = \frac{E[(X_t - \mu_X)(X_{t+\tau} - \mu_X)]}{\sigma_X^2} \quad (1.13)$$

Definition 1.19 (statistics). The *cross-correlation* of two series is the correlation of one with a delayed copy of the other as a function of the delay.

$$\text{crosscorr}(X, Y; \tau) = \text{corr}(X_t, Y_{t+\tau}) = \frac{E[(X_t - \mu_X)(Y_{t+\tau} - \mu_Y)]}{\sigma_X \sigma_Y} \quad (1.14)$$

For the purposes of FCS, these quantities were redefined as follows.

Definition 1.20 (FCS). The *correlation* between two random variables X and Y with expected values μ_X and μ_Y and standard deviations σ_X and σ_Y is

$$\text{corr}(X, Y) = \frac{E[(X - \mu_X)(Y - \mu_Y)]}{\mu_X \mu_Y} \quad (1.15)$$

where E is the expectation operator.

Definition 1.21 (FCS). *Autocorrelation* $G(X; \tau)$, is the correlation of a signal X with a delayed copy of itself as a function of the delay τ .

$$G(X; \tau) = \text{corr}(X_t, X_{t+\tau}) = \frac{E[(X_t - \mu_X)(X_{t+\tau} - \mu_X)]}{\mu_X^2} \quad (1.16)$$

Definition 1.22 (FCS). The *cross-correlation* of two series is the correlation of one with a delayed copy of the other as a function of the delay.

$$\text{crosscorr}(X, Y; \tau) = \text{corr}(X_t, Y_{t+\tau}) = \frac{E[(X_t - \mu_X)(Y_{t+\tau} - \mu_Y)]}{\mu_X \mu_Y} \quad (1.17)$$

The reason for these redefinitions (which just involve replacing standard deviations with means in the denominators of each expression) is that with the FCS definition, the autocorrelation has the nice property that for normal diffusion

$$G(X; 0) = \frac{1}{n} \quad (1.18)$$

where n is the mean number of fluorescent particles in the focal volume. The convenience of the statistics definitions is that there, correlations are guaranteed to be in $[-1, 1]$, with 0 representing no correlation, 1 perfect positive correlation

and -1 perfect negative correlation; this is lost with the FCS definitions: 0 still represents no correlation, but correlation values are no longer bounded, so the ideas of perfect correlation are lost.

I felt it necessary to provide these definitions for two reasons:

1. It is important for people from the fields of FCS and pure mathematics/statistics to know that they have different definitions for the same thing.
2. In FCS, it's very common for people to mistake correlation for cross-correlation. This is unfortunate, but knowing about this common mistake is essential for navigating the field in a sensible manner. It seems that when people in the FCS field correlate the signals from two separate channels, they use the term *cross*-correlation, even though they're only using correlation. I think the idea of working *across* two or more channels (and ideas such as *cross*-talk) leads to this confusion.

Remark. Henceforth, the FCS definitions of these quantities will be assumed.

1.6.1 Correlation

Co-diffusing fluorophores of different colours will necessarily induce positive correlation between the intensity traces of these two colour channels because co-diffusion means that presence of one fluorophore necessarily implies presence of

the other fluorophore. Hence, correlation may be used as a measure of co-diffusion, and co-diffusion is often interpreted as interaction (the simplest explanation for co-diffusion is that the co-diffusing entities are somehow stuck together). This idea forms the basis of much of FCS.

This does not hold for immobile particles. Immobile particles are not diffusing, so it doesn't make sense to employ a measure of diffusion.

Bleed-through of photons from one channel to another will induce artefactual correlation between the intensities of the two channels. Alternating laser excitation (ALEX, [Kapanidis et al. \(2005\)](#)) can be used to eliminate bleed through; however when using ALEX it is important to bear in mind that the two channels are no longer being acquired simultaneously. This is of particular importance for FCS. However if the instrument can go fast enough (with a laser alternation time much less than the residence time of the diffusing entities), this problem is negligible.

If the density of fluorophores under observation is too high, correlation can be practically impossible to measure. For example, the difference between 1 and 0 is much easier to detect than the difference between 11 and 10. At high density, relative differences are lower and hence harder to detect.

1.6.2 Autocorrelation

As mentioned already, the autocorrelation function (ACF) can be used to count the number of particles in the confocal volume. It can also be used to measure diffusion coefficients. The ACF is not used in my PhD.

1.6.3 Cross-correlation

The cross-correlation of intensity traces from nearby pixels can be used to measure the velocity of the movement of the labelled particles between these two pixels (Hebert et al., 2005).

1.6.3.1 Pair correlation function

The phrase *pair correlation function*⁴ was coined for this idea of cross-correlating intensity traces from nearby pixels. The PCF was used to image barriers to diffusion (Digman and Gratton, 2009) using the idea that the spatiotemporal correlation caused by particles moving from one place to another will not be present for positions p_1 and p_2 if they are on opposite sides of a barrier, because the barrier prevents travel from p_1 to p_2 . It has also been used to calculate the

⁴This was needless, it could just be called *cross-correlation function* but for the confusion about the term *cross-correlation* in the FCS field.

velocities of diffusing entities at each pixel in an image. The collection of these velocities at each pixel is known as a *diffusion tensor* (Rienzo et al., 2016). This approach is similar to that of Hebert et al. (2005).

1.6.4 Cross-correlated brightness

Cross-correlated brightness (Digman et al., 2009) molds the correlation idea (section 1.6.1) into the framework of N&B. Cross-correlated brightness isn't really a brightness at all, it's just a measure of correlation.

Suppose there are two channels with intensities I_1 and I_2 .

Definition 1.23 (cross-variance).

$$\sigma_{cc}^2 = E[(I_2 - \langle I_1 \rangle)(I_2 - \langle I_2 \rangle)] \quad (1.19)$$

Definition 1.24 (cross-correlated brightness).

$$B_{cc} = \frac{\sigma_{cc}^2}{\sqrt{\langle I_1 \rangle \langle I_2 \rangle}} \quad (1.20)$$

B_{cc} is related to $\text{corr}(I_1, I_2)$ by

$$B_{cc} = \sqrt{\langle I_1 \rangle \langle I_2 \rangle} \times \text{corr}(I_1, I_2) \quad (1.21)$$

This means that B_{cc} is just a scaled version of correlation. The need for this redefinition is unclear (but it is no harm). It does make the formula look like the brightness formula (1.5), but no such oligomeric state information can be gleaned from B_{cc} . It is merely useful as a relative measure of interaction: higher B_{cc} means more interaction, lower B_{cc} means less interaction. It is commonly used to identify interactions. Then, conventional N&B performed on each of the channels (1 and 2) can be used to measure the stoichiometry of the interaction.

Example 1.1. Protein A is labelled with a fluorophore that emits to channel 1, protein B is labelled with a fluorophore that emits to channel 2. At a given point in an image series, significant positive B_{cc} indicates interaction between protein A and protein B. From channel 1, with number and brightness we see that at this point, protein A is a dimer, whereas from channel 2, we see that protein B is a tetramer. Therefore the stoichiometry of the complex at this point is 2 units of A with 4 units of B.

Remark. Since cross-correlated brightness uses correlation but not cross-correlation, it is a prime example of the confusing naming that pervades FCS. It should be called *correlated brightness*. Rather than rename it, I will continue to refer to it as cross-correlated brightness.

1.7 Applications of FCS and FFS

FCS and FFS have been used in thousands of research projects. Here I number but a few for the sake of interest and to give biological context to these techniques.

Number and brightness (the prominent imaging FFS technique) has been used to:

- characterize the state of DNA aggregation in live cells (Mieruszynski et al., 2015)
- measure the stoichiometry of scaffold complexes in live neurons (Moutin et al., 2014)
- quantify interactions in gene expression networks (Declerck and Royer, 2013)
- measure the oligomeric state of the dynamin-2 protein at the HIV-1 fusion pore (Jones et al., 2017)
- measure the stoichiometry of the interaction of HIV-1 with its receptor and co-receptor over time in the pre-fusion process (Iliopoulou et al., 2018)

FCS has been used to:

- reveal structural and functional properties of promyelocytic leukemia nuclear bodies (Hoischen et al., 2018)
- demonstrate that HIV-1 evades antibody-dependent phagocytosis (Gach et al., 2017)

- determine the size of nanodomains ([Fenz et al., 2017](#))
- perform chromatographic measurements ([Kisley and Landes, 2014](#))
- quantify interactions of membrane proteins ([Ly et al., 2014](#))

Chapter 2

Instrumentation and Software

2.1 Instrumentation

All of the images used in this thesis were acquired on a Leica SP8 confocal microscope¹ equipped with hybrid detectors (section 1.2.1.3) in photon counting mode.

¹<https://www.Leica-microsystems.com/products/confocal-microscopes/details/product/Leica-tcs-sp8>

2.2 Software programs, languages and tools

2.2.1 C++

C++ (Stroustrup, 2013) is a general-purpose programming language optimized for performance (speed), efficiency (with use of computer resources) and flexibility. I used it for its speed, since many of the algorithms that I developed were quite computationally intensive and hence speed was an important consideration.

2.2.2 R

R (R Core Team, 2016) is a programming language and free software environment for statistical computing and graphics. I use R primarily as a wrapper for my C++ code to make my algorithms more user-friendly. R is best used with the *RStudio* integrated development environment.²

2.2.3 ImageJ

ImageJ (Rueden et al., 2017) is an open source image processing program designed for scientific multidimensional images. It is the preferred image viewing

²<https://www.rstudio.com>

and analysis software in the community. I have written my software in C++ and R because they are easier for developers, but I still intend to translate my image-related algorithms ImageJ plugins. ImageJ is best used via the FIJI ([Schindelin et al., 2012](#)) distribution.

2.2.4 Git and GitHub

Git is a free and open source distributed version control system designed to handle everything from small to very large projects with speed and efficiency.³ GitHub is a web-based hosting service for version control using Git. All of the computer code used during my thesis can be found on my GitHub at <https://github.com/rorynolan>. The vast majority of my time during my thesis was spent writing code so this GitHub account is the best record of the work that I have done.

³<https://git-scm.com/>

2.3 My software packages

2.3.1 `filesstrings`

`filesstrings` is an R package providing convenient functions for moving files, deleting directories, and a variety of string operations that facilitate manipulating file names and extracting information from strings. The motivation for making this package was to facilitate the use of file names for metadata. This is very common in microscopy, e.g. a file name like `well1_cell1_before_drug_addition.tif` is often seen. Using file names for metadata like this is a good idea, however if the naming or the extraction of data from these names is inconsistent, analysis becomes a nightmare and less reproducible. `filesstrings` provides a consistent means of working with such file names. This package was peer reviewed and published ([Nolan and Padilla-Parra, 2017b](#)). It gets 2,000 downloads per month which amounts to 30,000 since it was first released.

2.3.2 `examplerestr`

Definition 2.1. *Unit testing* is a software testing method by which individual units of source code, sets of one or more computer program modules together

with associated control data, usage procedures, and operating procedures, are tested to determine whether they are fit for use.

Unit testing is a tool to verify that software is performing as intended. It is a great way to discover bugs in software. `exempltestr` is an R package which makes it easier for R package developers to write unit tests for their packages. It helped me to eradicate many bugs in all of my packages. Interestingly, `exempltestr` was used to unit test and eradicate bugs in itself! This package was peer reviewed and published in 2017. ([Nolan and Padilla-Parra, 2017a](#)) It gets 500 downloads per month which amounts to 10,000 since it was first released.

2.3.3 `ijtiff`

An R package for general purpose tagged image file format (TIFF) input and output (I/O). This is currently the only such package with read and write support for TIFF files with floating point (real-numbered) pixels, and the only package that can correctly import TIFF files that were saved from *ImageJ* ([Rueden et al., 2017](#)). R has millions of users worldwide so this TIFF I/O capability is a basic need for masses of people. `ijtiff` gets 500 downloads per month which amounts to 5,000 since it was first released.

This package is part of the *rOpenSci* project. rOpenSci is a non-profit initiative

founded to make scientific data retrieval reproducible.⁴ This package was peer reviewed and published (Nolan and Padilla-Parra, 2018).

2.3.4 `autothresholdr`

`autothresholdr` provides the *ImageJ* (Rueden et al., 2017) *Auto Threshold* plugin (Landini et al., 2016) functionality to R users. It gets 600 downloads per month which amounts to 10,000 since it was first released.

2.3.5 `detrendr`

`detrendr` is an R package for detrending images (correcting for photobleaching). It provides all detrending algorithms mentioned in section 3. The detrending is done in C++ in the background for speed but it is wrapped in an R package for ease of use. It gets 300 downloads per month which amounts to 4,000 since it was first released.

⁴<https://ropensci.org/about>

2.3.6 nandb

`nandb` is an R package for number and brightness analysis. This was published along with a paper about detrending for number and brightness (Nolan et al., 2017a). It gets 300 downloads per month which amounts to 4,000 since it was first released.

2.3.7 brownded

`brownded` is an R software package (<https://github.com/rorynolan/brownded>) for simulating bounded Brownian motion in any number of dimensions, where *bounded* Brownian motion is Brownian motion in an d -dimensional box where the particles collide elastically (without loss of energy) within the boundaries of the box. `brownded` allows specification of the number of dimensions, the number of particles, the size of the box and the diffusion coefficient of the particles.

`brownded` also facilitates the simulation of images created from fluorescent particles undergoing bounded Brownian motion. It allows specification of the time at which each image should be taken, the pixel size and the brightness of the particles. Each fluorescent particle contributes photon counts to its pixel of residence at that time according to a Poisson process.

Finally, `brownded` facilitates the synthetic bleaching of fluorescent particles, so bleaching can be investigated with images produced with `brownded`.

Chapter 3

Photobleaching Correction

3.1 Introduction to photobleaching

In the ideal case, an *incident* photon of appropriate wavelength is absorbed by a fluorophore, promoting the fluorophore to an excited state; subsequently, the fluorophore relaxes down to its ground state by emitting a photon. In reality, it is possible that the incident photon can *break* the fluorophore with the result that it will no longer emit light. This breaking is referred to as *photobleaching* (or *bleaching* for short). Bleaching causes a diminution in the number of effective fluorophores, which is a direct cause of a loss of fluorescent signal.

Many quantitative methods in fields such as fluorescence fluctuation spectroscopy (FFS) and fluorescence correlation spectroscopy (FCS) implicitly assume that there is no bleaching in the data. Hence, data (image series) with significant levels of photobleaching must be corrected prior to the application of equations and algorithms in these fields. A main focus of this thesis is on how to correct fluorescent image series for the effects of bleaching, given that bleaching *does* occur. There is no attempt to understand *why* and/or *how* photobleaching occurs.

Remark. All of the current literature mentions bleaching correction as being purely for correcting the problem of non-stationary mean (NSM), neglecting the problem of non-stationary variance (NSV). Figure 3.5 shows that NSM and NSV go hand-in-hand. Correction for NSM is referred to as trend removal or *detrending*. Hence, the terms *detrending* and *bleaching correction* have come to be used interchangeably. I will follow this convention and use the term *detrending* from now on to mean correction for NSM and possibly also NSV. Starting at section 3.3, the focus is on correcting for NSM. Discussion of correction for NSV starts in section 3.7.

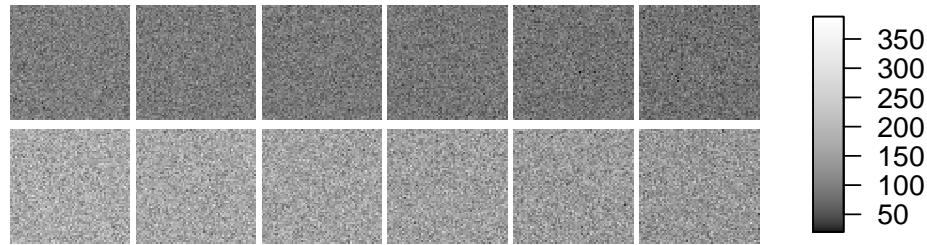


Figure 3.1: Frames 1, 100, 200, 300, 400 and 500 from simulated image series. $\epsilon = 4$ (top) and $\epsilon = 7$ (bottom) with 15% (top) and 20% (bottom) bleaching.

3.2 The effects of bleaching in FCS and FFS

We simulate two image series with 500 frames, 64x64 pixels per frame (pixel size $1 \mu\text{m}$), each with 100,000 diffusing ($D = 100 \mu\text{m}^2/\text{s}$) fluorescent particles which are uniformly distributed at the beginning. In the first image series (`img1`), these have brightness $\epsilon = 4$ and in the second (`img2`) they have brightness $\epsilon = 7$. See figure 3.1. We bleach these by 15% and 20% to create `img1_bleached` and `img2_bleached` respectively.

It may not be obvious that these image series are subject to bleaching from figure 3.1, but we can see it more clearly in figure 3.2.

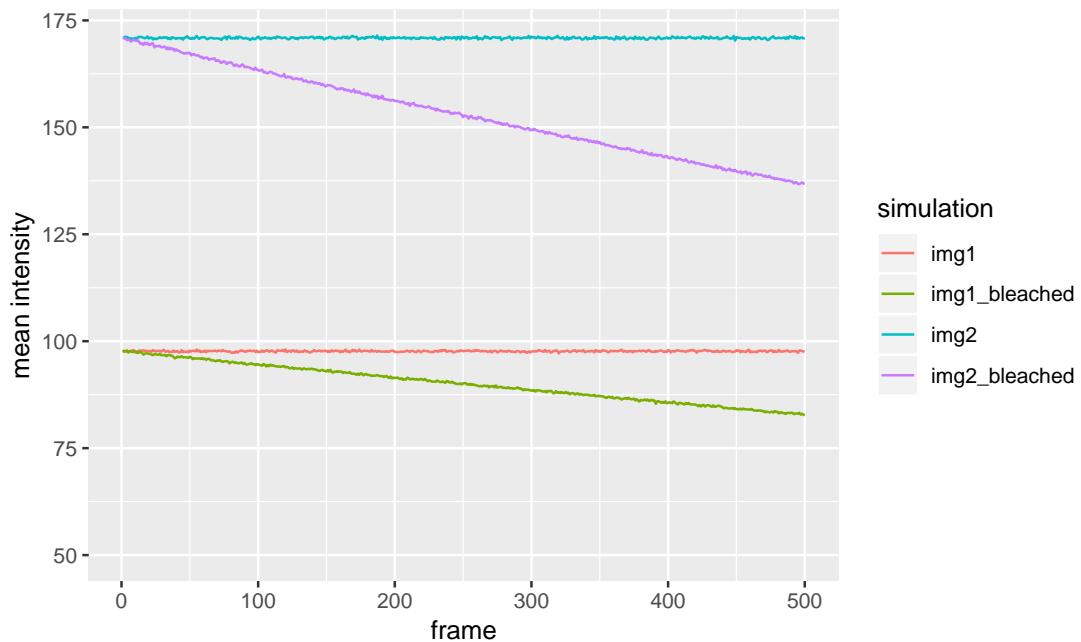


Figure 3.2: Mean intensity profiles of the simulated image series with and without bleaching. Simulation 1 was bleached by 15% and simulation 2 by 20%. The bleaching is such that the mean intensity decreases according to a single exponential decay (even though it looks quite linear).

3.2.1 FCS

The unrelated images `img1` and `img2` have a tiny median cross-correlated brightness of $B_{cc} = 0.0036$, signifying no significant correlation, as one would expect. However, the bleached images `img1_bleached` and `img2_bleached` have a significant $B_{cc} = 0.3686$. This shows that bleaching is introducing correlation between otherwise unrelated images. Since correlation is used as a proxy for hetero-interaction, bleaching can make it appear as though there is interaction when in fact there is not.

3.2.1.1 Why does bleaching introduce correlation

Let's take another look at the formula for correlation:

Definition 3.1 (FCS). The *correlation* between two random variables X and Y with expected values μ_X and μ_Y and standard deviations σ_X and σ_Y is

$$\text{corr}(X, Y) = \frac{E[(X - \mu_X)(Y - \mu_Y)]}{\mu_X \mu_Y} \quad (3.1)$$

So correlation is answering the question: *Do X and Y deviate from their mean at the same time and in the same or opposite direction?* If there is no pattern in their deviations from mean, then the correlation will be zero. If they deviate

from their means in the same direction at the same time, there will be a positive correlation; if they deviate from their means in the opposite direction at the same time, there will be a negative correlation. Correlation is all about measuring whether deviation from mean in two different series is synchronised or not.

Let's create two unrelated intensity traces X and Y , both of length 500. X will be Poisson($\lambda = 50$) and Y will be Poisson($\lambda = 30$).

```
#>
#> Pearson's product-moment correlation
#>
#> data: X and Y
#> t = 1.0731, df = 498, p-value = 0.2837
#> alternative hypothesis: true correlation is not equal to 0
#> 95 percent confidence interval:
#> -0.03982597 0.13515369
#> sample estimates:
#>       cor
#> 0.04803237
```

There's no pattern in how X and Y deviate from their means, indeed a Pearson correlation test of X and Y returns an insignificant p -value of 0.28.

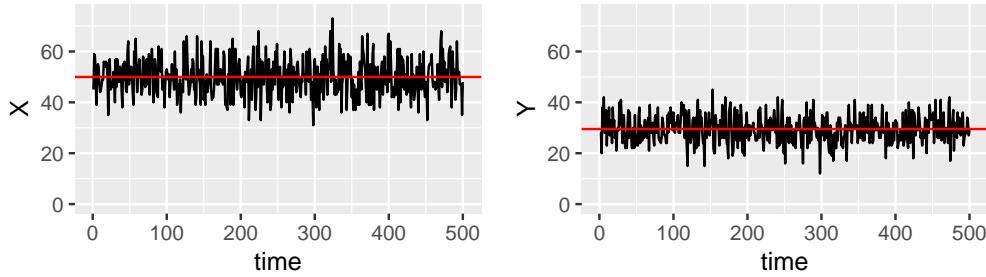


Figure 3.3: Unrelated intensity traces X and Y with their means in red.

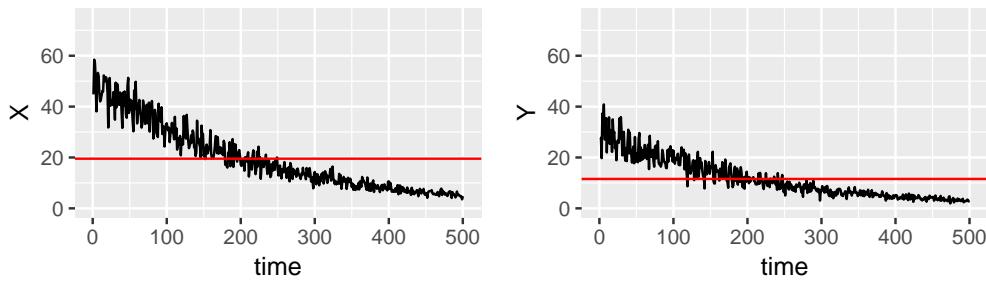


Figure 3.4: Unrelated intensity traces X and Y with their means in red.

Now let's bleach them both by 90%.

Now, there is visible correlation. At the start of X and Y , both are consistently above their mean and at the end, both are consistently below. They are simultaneously above and below respectively, so there will be positive correlation. This is backed up by a Pearson correlation test which returns a significant p -value of less than 10^{-200} .

Remark. Bleaching also introduces autocorrelation for the same reason. The series X had no autocorrelation before bleaching ($p = 0.23$), whereas after bleaching, X has significant autocorrelation ($p < 10^{-200}$).

3.2.2 FFS

Definition 3.2 (median mean pixel intensity). The *median mean pixel intensity* of an image series is found by taking the mean intensity of each pixel in the image series and then taking the median of those means. It can be thought of as a summary statistic for the pixel intensity of the image series.

Definition 3.3 (median pixel intensity variance). The *median pixel intensity variance* of an image series is found by taking the variance in the intensity of each pixel in the image series and then taking the median of those variances. It can be thought of as a summary statistic for the variance in the pixel intensity of the image series.

In FFS, one is always interested in the mean and variance of pixel values. `img1` has a median mean pixel intensity of 98 and a median pixel intensity variance of 487. The mean brightness is $\epsilon = 3.9959$ (very close to 4, as expected since the image series was simulated with brightness $\epsilon = 4$). For `img1_bleached`, we find a median mean pixel intensity of 90 and a median pixel intensity variance of 468. The mean brightness is $\epsilon = 4.2026$. Hence, bleaching has altered both the means and variances of the pixels, resulting in a change in calculated brightness. The non-stationary mean frame intensity introduced by bleaching decreases the mean but increases the variance. The loss in signal has the effect of slightly decreasing

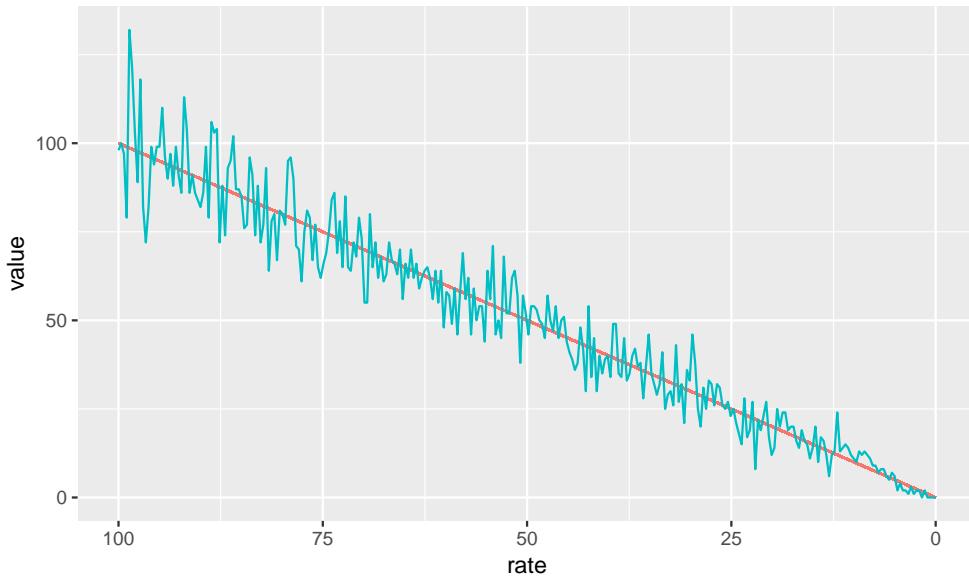


Figure 3.5: A decrease in the Poisson rate (e.g. for emission of photons) leads to a decrease in the mean (blue line) but also a decrease in fluctuations around the mean. Notice that towards the right where rate is low, fluctuations around the mean are at their smallest.

the variance: with Poisson statistics (such as photon-emission), a loss of signal (photons) leads directly to a loss in variance. This is a subtle point not discussed anywhere in the literature; it is shown in figure 3.5.

3.3 Exponential fitting detrending

Naively, one could assume that bleaching of fluorophores takes place at some constant rate. This would mean that the intensity of the image would die-off according to an exponential decay. In figure 3.6, we fit an exponential decay to

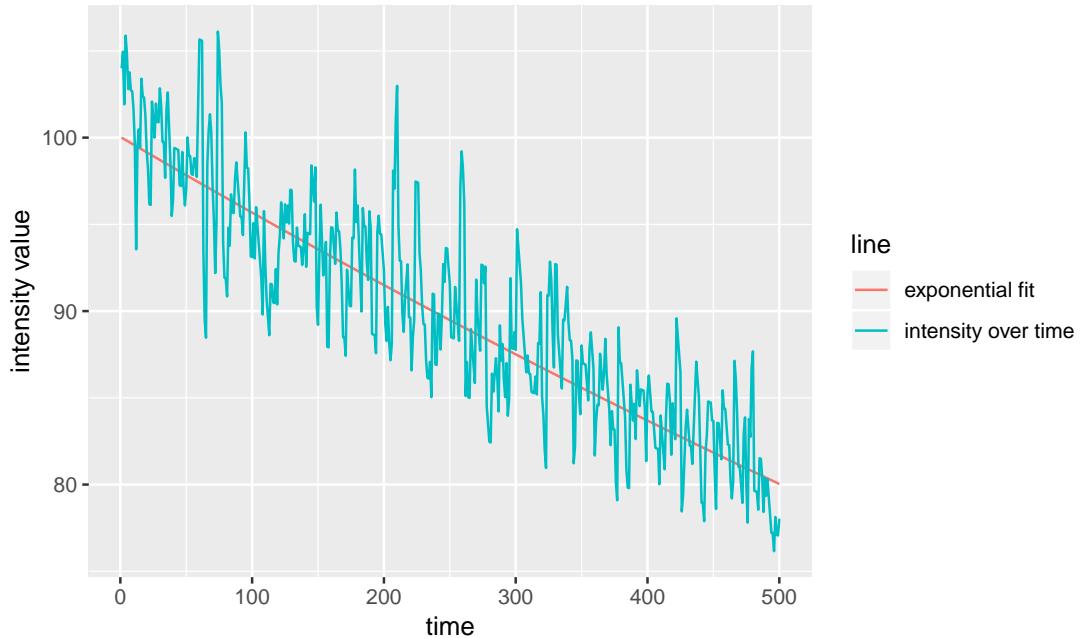


Figure 3.6: Exponential fit to intensity trace of image which is subject to bleaching at a constant rate.

such ideal data.

Having fit the data, one may record the deviations from the fitted line as the *fluctuations* and replace these fluctuations about a straight line, which is placed at the mean of the original series; for figure 3.6 above, this mean is 90. Figure 3.7 shows the corrected series.

We can see that here, in the ideal case where the naive assumptions of the exponential decay fitting approach hold, this approach works quite well. Let us now examine the case where these assumptions don't hold because there are other long-term fluctuations e.g. due to cell movement. We add these other fluctuations

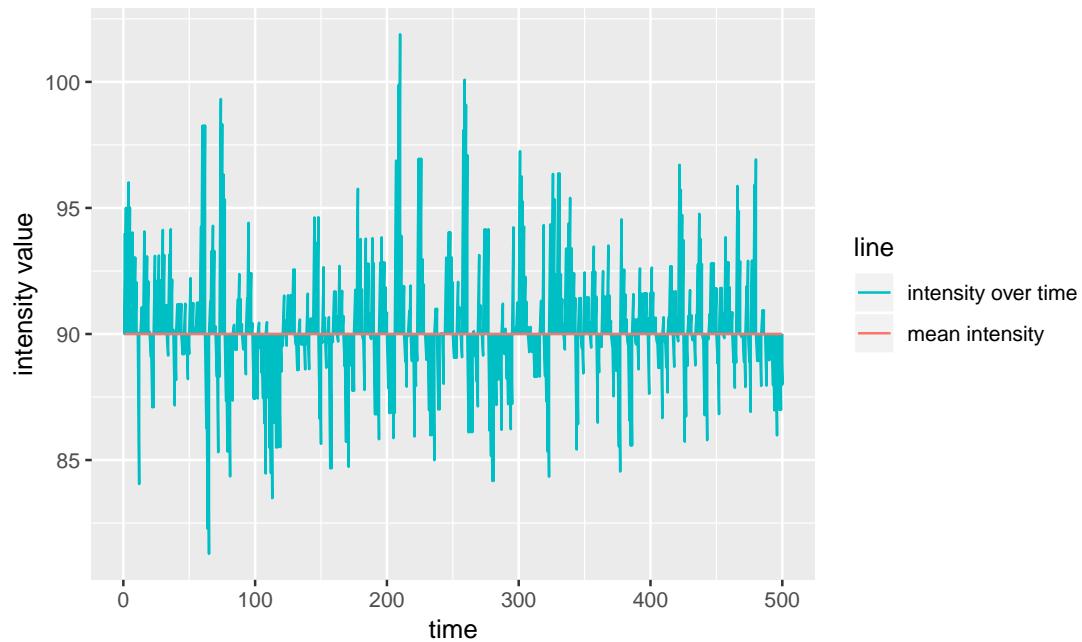


Figure 3.7: The blue line from figure 3.6 has changed to a straight horizontal line cutting the y axis at the mean intensity of the original intensity trace. The fluctuations about the blue line that existed in figure 3.6 are preserved here. As an example, see the large downward fluctuation at $t \approx 50$ seconds in both figures.

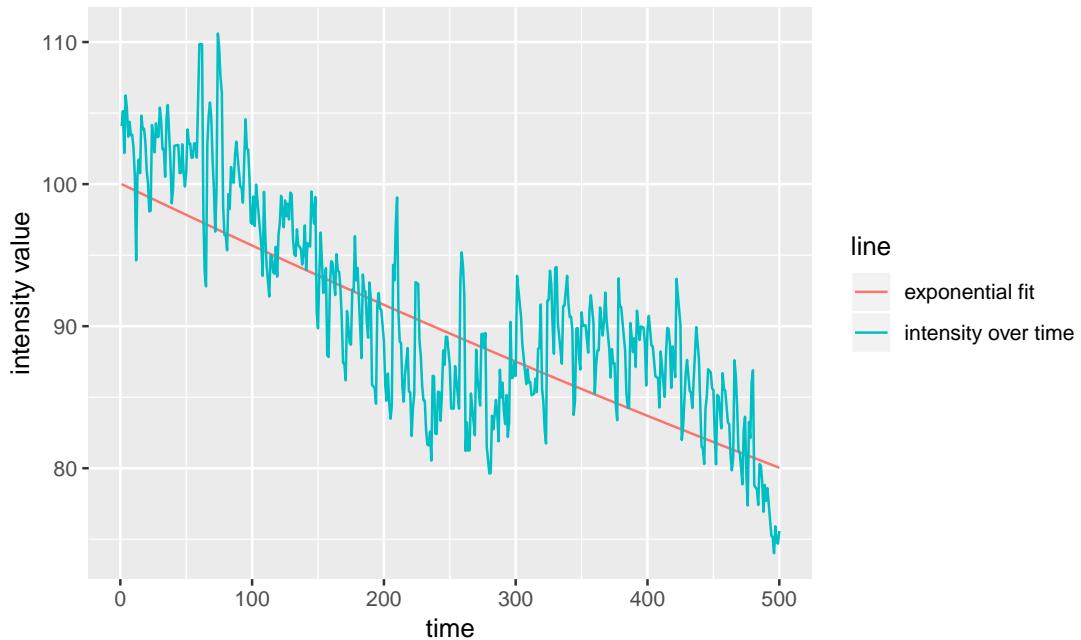


Figure 3.8: An exponential decay with added sinusoidal variance, fit with a simple exponential decay.

as a gentle sinusoid. See figure 3.8.

One can see by eye that this is not a good fit for the data. This has disastrous consequences for the detrended series, shown in figure 3.9.

One can see in figure 3.9 that the exponential fit detrend failed to remove the sinusoidal trend in the data (even though it *did* remove the exponential decay component). We have now seen that exponential fitting detrending is appropriate when the decay has a particular form, but is otherwise not fit for use. This is a problem common to all fitting approaches to detrending, even the more flexible

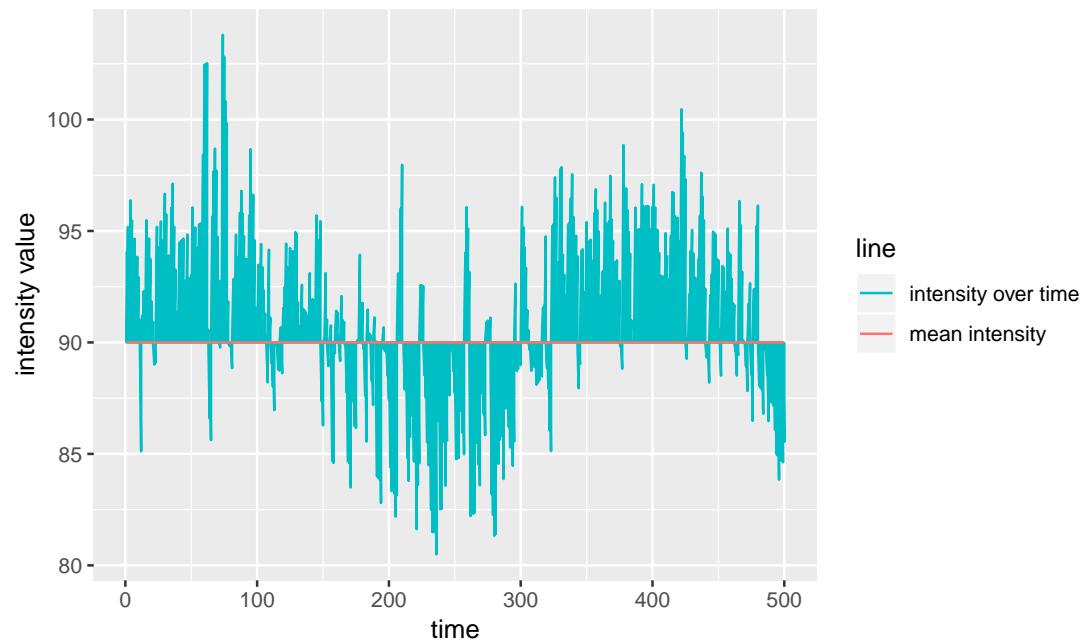


Figure 3.9: Result of exponential fitting detrending applied to a decay with a long-term sinusoidal trend component.

types like polynomial detrending (Chan et al., 1977). For this reason alone, for the purpose of detrending, fitting approaches should be avoided.

3.4 Boxcar smoothing detrending

A common approach to obtaining the line from which to measure deviations/fluctuations (as in the red line in figure 3.6) is to *smooth* the time series, i.e. construct the line by taking a *local average* at each point. This is often referred to as *boxcar smoothing* because it can be visualized as drawing a box around a neighborhood of points, taking their average as the smoothed value at that point and then moving the box onto the next series of points and repeating the procedure; see figure 3.10.

The length of the boxcar is equal to $2l + 1$ for natural numbered parameter $l \in \mathbb{N}$. This ensures that the length of the boxcar is always odd) which means it can always be centered upon a point). Hence the allowable lengths of a boxcar are 3, 5, 7, 9, etc.

The boxcar parameter l has a large effect on the type of smoothing achieved. This can be seen in figure 3.11 where boxcar smoothing is applied to the trace in figure 3.6. The traces for $l = 1$ and $l = 3$ are far too wiggly (not smooth enough); the

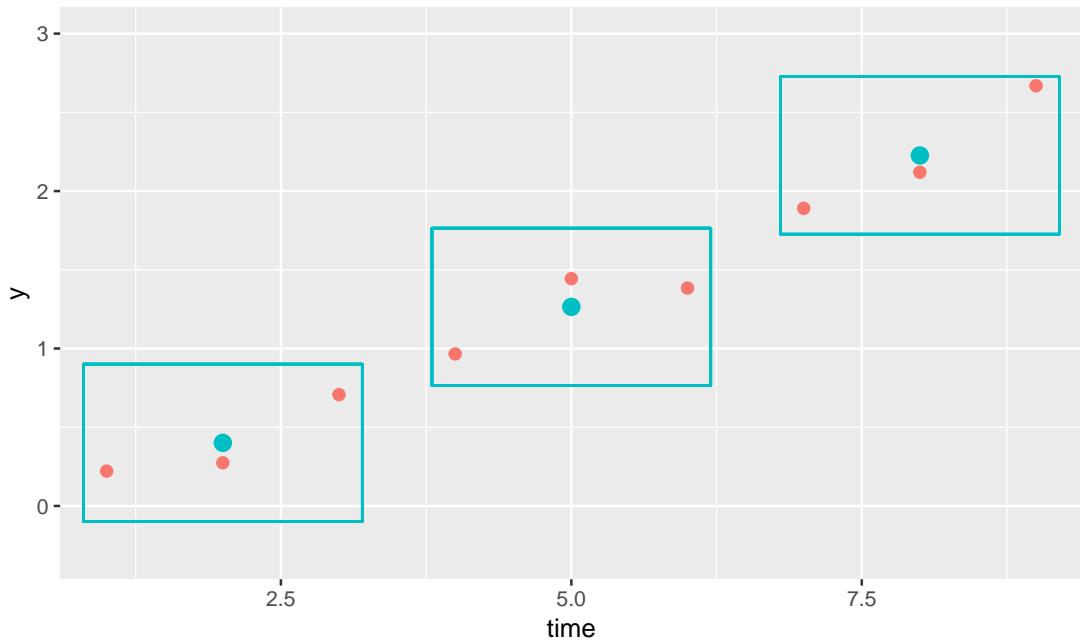


Figure 3.10: The original time series is depicted by the red dots. The blue rectangles represent the *boxcar*. This boxcar is said to be of length 3 because it is wide enough to encompass 3 points at a time. The boxcar is centered on a point and then the smoothed value at that point (blue dot) is calculated as the mean value of all points within the boxcar. In reality, every point gets a smoothed value which means that the boxcar *overlaps* but in this figure—for the sake of clarity—they are not overlapped.

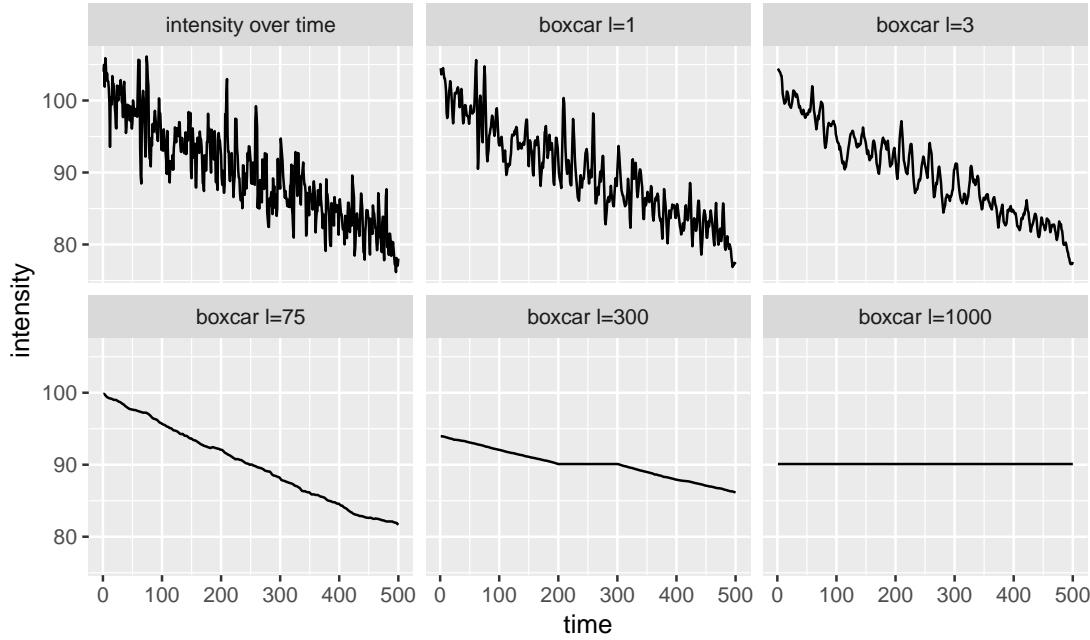


Figure 3.11: The original intensity trace is shown in the top-left. The other panels show the result of boxcar smoothing for $l = 1, 3, 75, 300$ and 1000 . $l = 1$ and $l = 3$ are not smooth enough, $l = 75$ looks like it might be OK although it is slightly wiggly. $l = 300$ and $l = 1000$ are over-smoothed.

trace for $l = 75$ is better but perhaps still slightly wiggly; finally, the traces for $l = 300$ and $l = 1000$ are too close to straight horizontal lines (too smooth).

This begs the question: what is the correct smoothing parameter l ?

3.5 The correct smoothing parameter for detrending

Figure 3.11 shows that the choice of boxcar size is crucial because different sizes lead to very different *smoothed* lines. The most common choice in the community is to choose $l = 10$ (Laboratory for Fluorescence Dynamics, 2018). There is no justification for this choice.

In section 1.5.1, we learned that for immobile particles, the expected brightness is $B = 1$. This fact can be used to solve for the appropriate choice of l to use for detrending a specific image series.

Definition 3.4. The *mean intensity profile* of one channel of an image series is obtained by calculating the mean intensity of each frame in that image series.

The mean intensity profile can be used to visualize the bleaching of an image series. If the fluorophores are bleaching, the mean intensity should be decreasing over time. To proceed with solving for the appropriate l , we need to make one assumption; this is that any two image series with the same mean intensity profile are appropriately detrended with the same detrending parameter l . This assumption seems reasonable, however there is no need to debate its validity because later, detrending with the solved-for parameter l will be evaluated with simulated

data and compared to the standard $l = 10$. If this assumption is bad, then the performance of the detrending that relies on it should also be bad. With this assumption in hand, solving for l proceeds as follows:

1. Simulate an image series with immobile particles only which has the same mean intensity profile as the acquired real data.
2. Given that the simulated series is of immobile particles only, once properly detrended, it should have $B = 1$.
3. The l for which the detrended series has mean brightness closest to 1 is the most appropriate for the simulated data.
4. By the assumption above, this l is the most appropriate for the real data.

Mathematically, this can be expressed as

$$l = \operatorname{argmin}_{\tilde{l}} |1 - (\text{mean brightness of simulated series detrended with parameter } \tilde{l})| \quad (3.2)$$

In fact, what I have done here is to give a general method for solving for any detrending parameter α :

$$\alpha = \operatorname{argmin}_{\tilde{\alpha}} |1 - (\text{mean brightness of simulated series detrended with parameter } \tilde{\alpha})|$$
(3.3)

This will be useful later when other detrending regimes with their own parameters are introduced.

3.6 Exponential smoothing detrending

Exponential smoothing is a slight alteration to boxcar smoothing. The idea is that when computing a local average, points nearer to the point of interest should have greater weights. The weights fall off with distance $|t|$ from the point of interest according to $\exp(-\frac{|t|}{\tau})$ where the parameter τ is a positive real number. This function is visualized in figure 3.12. For small values of τ , only values very close to the point of interest have importance when calculating the local average. For larger values of τ , further values also have importance (but closer values always have higher weights). In this sense, increasing the value of τ has a similar effect to increasing the value of l for the boxcar in that further away points are taken into account.

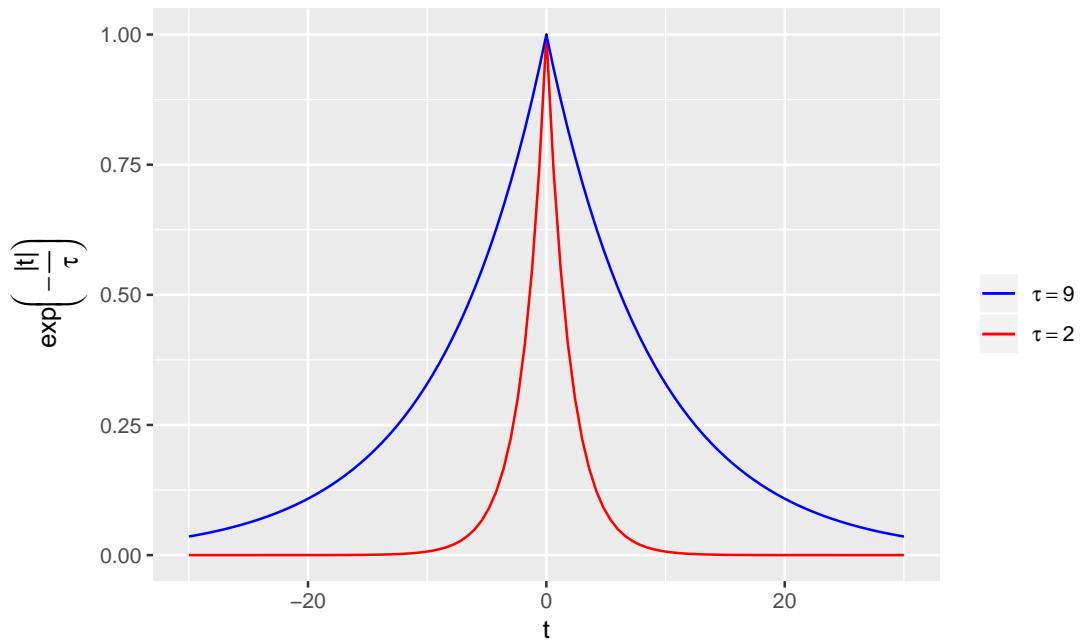


Figure 3.12: The function $\exp(-\frac{|t|}{\tau})$ visualized with $\tau = 2$ and $\tau = 9$. For $\tau = 2$, points at distance $|t| = 10$ have approximately zero weight, whereas for $\tau = 9$, these points have significant weight.

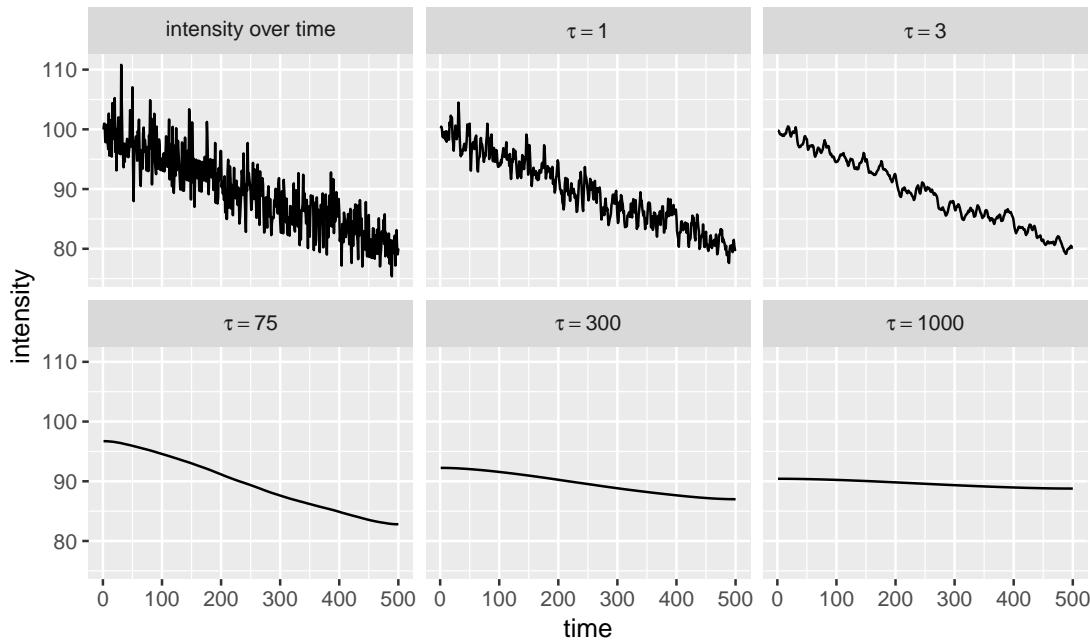


Figure 3.13: The original intensity trace is shown in the top-left. The other panels show the result of exponential smoothing for $\tau = 1, 3, 75, 300$ and 1000 .

In figure 3.13, exponential smoothing with different parameters τ is applied to the trace in figure 3.6. The results are similar to those in figure 3.11.

Heuristically, exponential smoothing detrending seems favorable to boxcar detrending because the idea that points further away from the point of interest are less important (but still somewhat important) when computing the local average is reasonable. Indeed, this was the method proposed in the original number and brightness paper (Digman et al., 2008). For this reason, exponential smoothing was the method of choice for my paper where the method of choosing the correct detrending parameter was published (Nolan et al., 2017a).

3.7 Correcting for non-stationary variance

Definition 3.5. The variance of a random variable X is the expected value of the squared deviation of X from its mean μ :

$$\text{Var}(X) = E[(X - \mu)^2] \quad (3.4)$$

All of this chapter so far has focused on correcting for non-stationary mean. As shown in figure 3.5, as the mean decreases, so too does the variance. For an instance x of the random variable X with expected value $E[X] = \mu$, $x - \mu$ is the *deviation* of x from μ . If we write x as $x = \mu + \tilde{x}$, then we get the deviation $x - \mu = (\mu + \tilde{x}) - \mu = \tilde{x}$, so \tilde{x} is the deviation. For a given point in figure 3.5, its deviation is its distance from the red line. For positive real number k , making the transformation $\tilde{x} \rightarrow \sqrt{k}\tilde{x}$ i.e. $x \rightarrow \mu + \sqrt{k}\tilde{x}$ causes the variance (i.e. the *squared deviation*) to be transformed as $\text{Var}(X) \rightarrow k \times \text{Var}(X)$. Hence, we have a way to modify the variance of a time series as a whole by modifying the deviation of each time point from the mean. For months, I toyed with this idea as a solution of correcting for non-stationary variance. However, in reality the contribution to the variance in intensity at a given pixel is down to both Poisson photon statistics and fluorophore movement. This combination of factors makes it very difficult

to ascertain the amount by which the variance should be altered. I eventually abandoned my efforts to alter the variance like this in favor of the *Robin Hood* detrending algorithm (section 3.9) which includes correction for non-stationary variance as an intrinsic part of its detrending routine.

3.8 Caveats of fitting and smoothing approaches

Both fitting and smoothing approaches to detrending have serious caveats. Fitting approaches assume that the fluorescence intensity decay has a certain form. Unpredictable issues such as cell movement mean that no particular decay form can be assumed. Smoothing methods do not perform well at the edges of time series that they are applied to. They also require the user to choose a smoothing parameter. The problem of how to best choose this parameter was solved recently (Nolan et al., 2017a), but this method has not been widely adopted. Most importantly, both fitting and smoothing fail when the data cannot be approximated as mathematically continuous (fitted and smoothed lines are continuous approximations of data). Fluorescence intensity data at low intensities—where most pixel values are either 0 or 1—are quasi-binary¹ and hence a continuous approximation does not make sense (see figure 3.14). This means that neither fitting nor smooth-

¹By *quasi-binary*, I just mean that almost all values are 0 or 1.

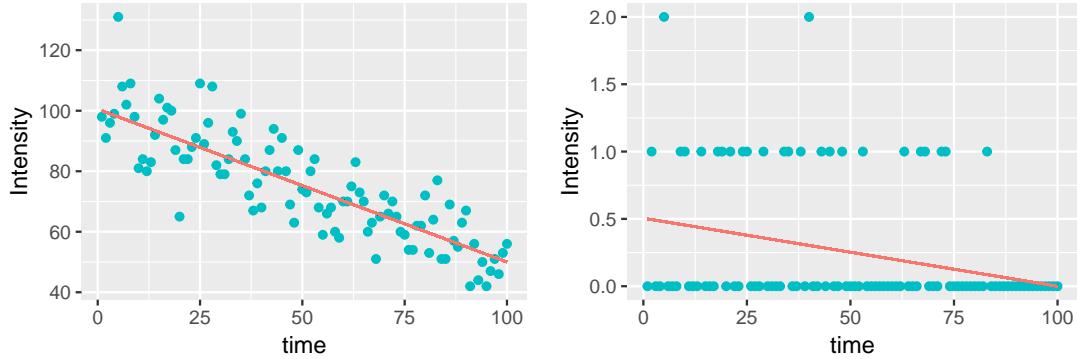


Figure 3.14: Left: for high ($\gg 1$) intensity values, the line is a satisfactory approximation of the data, representing it well. Right: for low (quasi-binary) intensity values, the line is not a good approximation for the data and indeed no line or curve could represent the data well.

ing are applicable detrending methods at low intensities. This is the crucial caveat of these methods because, when bleaching is a problem, it is common to reduce laser power to reduce bleaching, which leads directly to lower intensity images. With fitting and smoothing techniques, it may sometimes be advisable to increase the laser power to achieve higher intensities such that the detrending routines will function properly. This means one may need to bleach more in order to be able to correct for bleaching. This farcical situation necessitates a new detrending technique which can function at low intensities.

3.9 Robin Hood detrending

Intensity images in units of photons are count data. This means that the values are all natural numbers, i.e. elements of $\mathbb{N}_0 = \{0, 1, 2, 3, \dots\}$. Fitting and smoothing give real-numbered values (elements of \mathbb{R}), which must then be transformed back into count data (elements of \mathbb{N}_0), normally by rounding. This means that fitting and smoothing methods of detrending push values through $\mathbb{N}_0 \rightarrow \mathbb{R} \rightarrow \mathbb{N}_0$ (the move back $\rightarrow \mathbb{N}_0$ is necessary because calculations like Qian and Elson's moment analysis assume photon count data, which is necessarily in \mathbb{N}_0). When current methods were failing to properly detrend low-intensity images, I began to wonder was it necessary to go through the real numbers \mathbb{R} , given that the start and end points were the natural numbers \mathbb{N}_0 ?

Consider figure 3.15. There is a bleached and unbleached version of an intensity trace. Suppose that our real data is the bleached trace, but we *wish* it looked like the unbleached trace. You may wonder why the unbleached trace is not at the starting intensity of the bleached series. For reasons that will become clear, the Robin Hood algorithm can only place the detrended image at the mean intensity of the original image. This is not a problem because the issue with bleaching in FCS and FFS is mainly that the changing signal leads to incorrect calculations, not that the loss in signal leads to a critical lack of information (photons). Indeed,

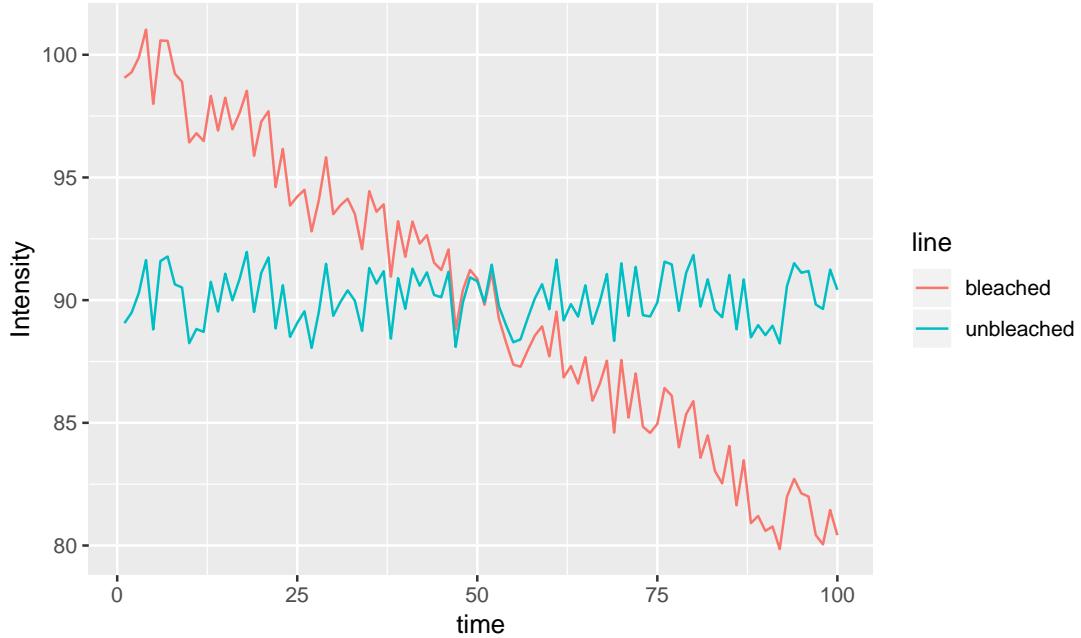


Figure 3.15: Bleached and unbleached intensity traces.

a feature of the Robin Hood algorithm is that it preserves the mean intensity of the real data on a pixel-by pixel basis.

To get to the unbleached intensity trace from the bleached intensity trace, intensity must be subtracted from time-points with too much intensity and added to time points with too little intensity. This can be done by *taking* counts from frames with too much intensity and *giving* them to frames with too little intensity. In this way, no counts are gained or lost, they are just moved around the image series. See figure 3.16. Counts are passed from one frame to another *along* a given pixel, i.e. if a count is taken from pixel at position p in some frame i , it must be given

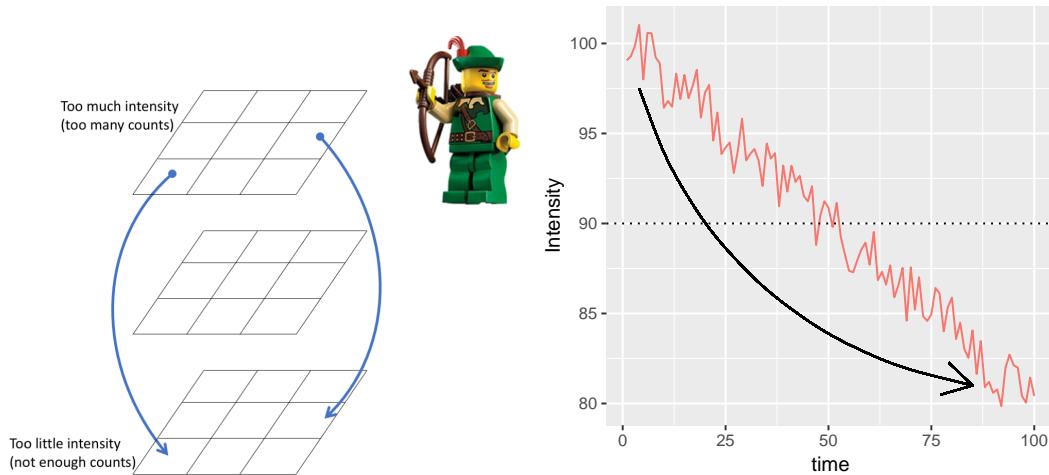


Figure 3.16: Robin Hood: counts are taken from frames of higher intensity (usually closer to the start of the image series) and given to frames of lower intensity (usually closer to the end of the image series).

to a pixel at the same position p in some other frame j . It is this condition that ensures that the mean intensity images of the original and detrended image series are the same.

To determine how many swaps need to be made to detrend a given image series, equation (3.3) can be used, with α being the number of swaps.

The random gifting of counts from higher to lower intensity frames has the effect of temporally redistributing mean intensity but *also* variance in intensity. With photon statistics (which follow a Poisson distribution), random counts provide both mean and variance. This is in contrast to all previous methods which consist of determining local deviation and adding it to a *fixed* global mean: this provides no temporal redistribution of variance.

3.10 A comparison of detrending methods

To compare the various detrending methods, I use the following workflow:

1. Simulate a number $N = 100,000$ of particles diffusing with known diffusion rate. Simulations were done with the `brownded` software package (section [2.3.7](#)).
2. Simulate photon emission from these particles with chosen brightness ϵ and create an image series from this, being careful to (virtually) sample at a rate appropriate for number and brightness analysis.
3. Bleach the simulation with a chosen constant bleaching rate.²
4. Simulate photon emission from the bleached simulation (bleached particles don't emit photons) with the same brightness ϵ and create an image series.
5. Detrend the bleached image series.
6. Evaluate the detrending algorithm by measuring how close the brightness of the detrended bleached image series is to the known simulated brightness.

For all combinations of brightnesses of $\epsilon = 0.001, 0.01, 0.1, 1, 10$ and bleaching fractions of 0%, 1%, 5%, 10%, 15%, 20%, 25%, 30%, 20 images of 64x64 pixels and 5,000 frames were simulated using 100,000 fluorescent diffusing particles.³

²It would be good to test with non-constant bleaching rates too, this wasn't done due to time constraints.

³The simulation took 3 weeks.

These were detrended with the following detrending routines:⁴

1. Boxcar with $l = 10$ (`boxcar10`, the most common detrending routine).
2. Exponential smoothing with automatically chosen parameter τ (`autotau`).
3. Robin Hood with automatically chosen swaps (`robinhood`).

The performance was evaluated using the *mean relative error*.

Definition 3.6. For a given brightness and bleaching fraction,

$$\text{mean relative error} = \frac{|(\text{calculated brightness after detrending}) - (\text{true brightness})|}{(\text{true brightness})} \quad (3.5)$$

Figure 3.17 shows the results. Before I discuss them, note that the common brightnesses that we see are in the range $\epsilon = 0.003$ to $\epsilon = 0.1$.

The most striking thing about figure 3.17 is that the best choice in all cases is to not detrend at all! This is an interesting result and seems to render all detrending routines worthless. However, when working with real data, not detrending does not work well at all. This will be shown in chapter 4. This indicated that my simulations are unrealistic. This is probably because with real data, bleaching is likely not taking place at a constant, steady rate and other factors such as cell

⁴The detrending took 2 weeks.

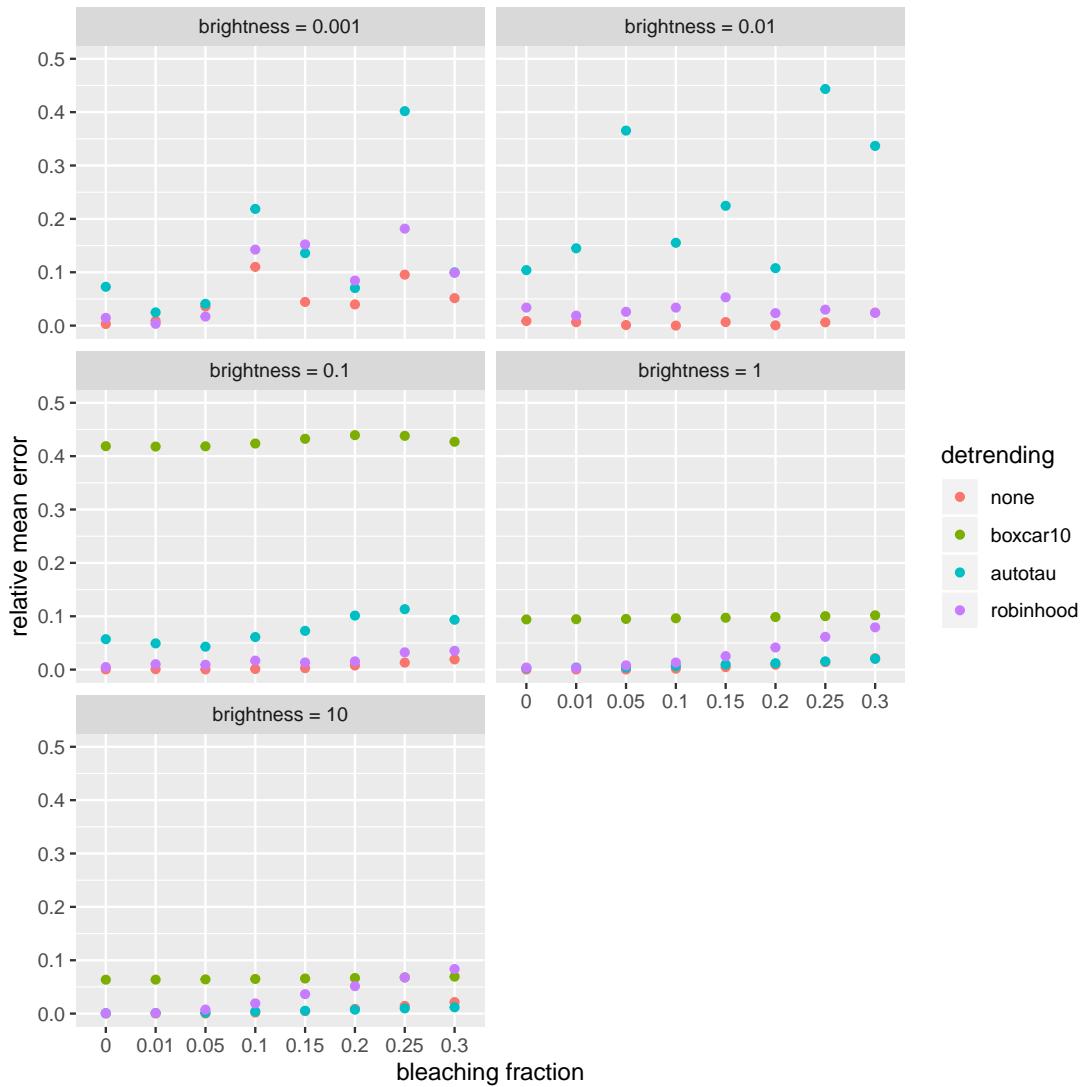


Figure 3.17: A comparison of different detrending methods with various brightnesses and bleaching fractions (steady, constant-rate bleaching), including the results of not detrending at all.

movement or/and laser power fluctuations are contributing to medium and long term intensity fluctuations and these have a detrimental effect on calculations if not detrended out. It would be possible to study this by mimicking real bleaching profiles with simulations (see section 6.2).

The worst performer by far is `boxcar10`. For example, at $\epsilon = 0.1$, it makes an error of worse than 40% and for $\epsilon = 0.001, 0.01$, its error is worse than 50%, so it does not even appear on the plot. This is good evidence that arbitrarily choosing the parameter l is very bad practice. For realistic brightnesses (≤ 0.1), `robinhood` is the best with errors almost always lower than 5%. `autotau` also performs very well, with errors almost always less than 10%. At the lowest brightness $\epsilon = 0.001$, all methods are somewhat erratic. That is because at this extremely low brightness, there is a critical lack of information (photons) for the algorithms to work with. Finally, at unrealistically high brightnesses of $\epsilon = 1, 10$, `autotau` begins to perform well because at these high photon counts, the caveats of smoothing have totally disappeared. However, I cannot explain the degradation in the performance of `robinhood` in this case. Fortunately, there is no need to dwell on this, as this situation ($\epsilon = 1, 10$) does not arise in practice because available fluorophores are not this bright.

Chapter 4

Applications

4.1 Dimerization of FKBP12

4.1.1 Introduction

Myristoylated FKBP12 is known to dimerize upon addition of the dimerizing drug AP1510 (Amara et al., 1997). As a test application of exponential smoothing detrending (section 3.6) with automatically chosen parameter τ , we used this system with number and brightness to verify this dimerization. We tested this in 20 Cos7 cells with mClover-labelled FKBP12 (Nolan et al., 2017a).

4.1.2 Experimental results and discussion

We found a brightness increase in ϵ of ≈ 1.6 -fold using the *automatic* detrending method. Assuming that the population was all monomers before ligand addition, the ≈ 1.6 -fold increase upon addition suggests that dimerization had occurred (see figure 4.1), however, we expected the increase to be ≈ 2 -fold upon dimerization. There could be many reasons for this discrepancy.

1. FKBP is endogenously expressed in Cos7 cells, so depending on the level of native expression (not labelled) and the level of expression of our transfected FKBP (labelled), it could be that many cases of what is actually a dimer is behaving like a monomer because only one of the pair is labelled.
2. Recently, a paper came out ([Dunsing et al., 2018](#)) explaining how the assumption that all fluorophores emit signal is invalid (because many never function properly) and that because of this, oligomeric state changes calculated from brightness must be adjusted by a correction factor specific to the fluorophore. Unfortunately, this study did not characterize mClover, so we do not know its correction factor to be able to apply it. Any such correction would bring us higher than 1.6.

It is possible that all of the above reasons are valid and are present to different degrees in different images. This could be a cause of the heterogeneity of fold-

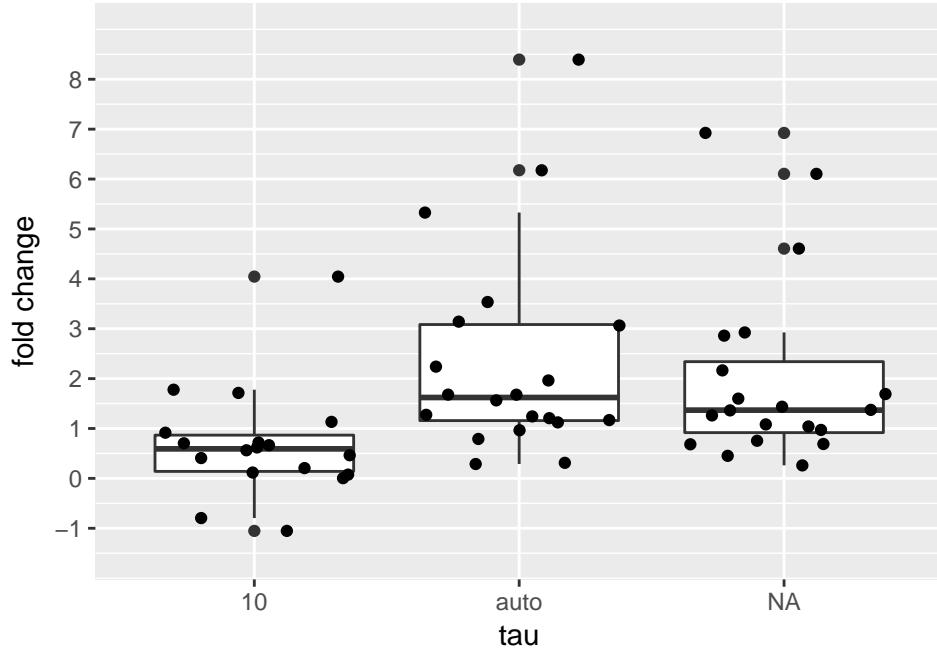


Figure 4.1: The fold changes in brightness ϵ upon addition of AP1510 drug shown for different detrending routines ($\tau = 10$, auto τ and no detrend).

changes in ϵ measured from single images. This heterogeneity means that many replicates are needed in number and brightness experiments in order to converge upon the true value of the fold change.

A pair of cells from this study together with their brightness statistics are shown in figure 4.2. Notice how there is no discernible change in intensity before and after addition of the drug, but there is a discernible change in brightness B , best seen using the histogram of pixel brightnesses.

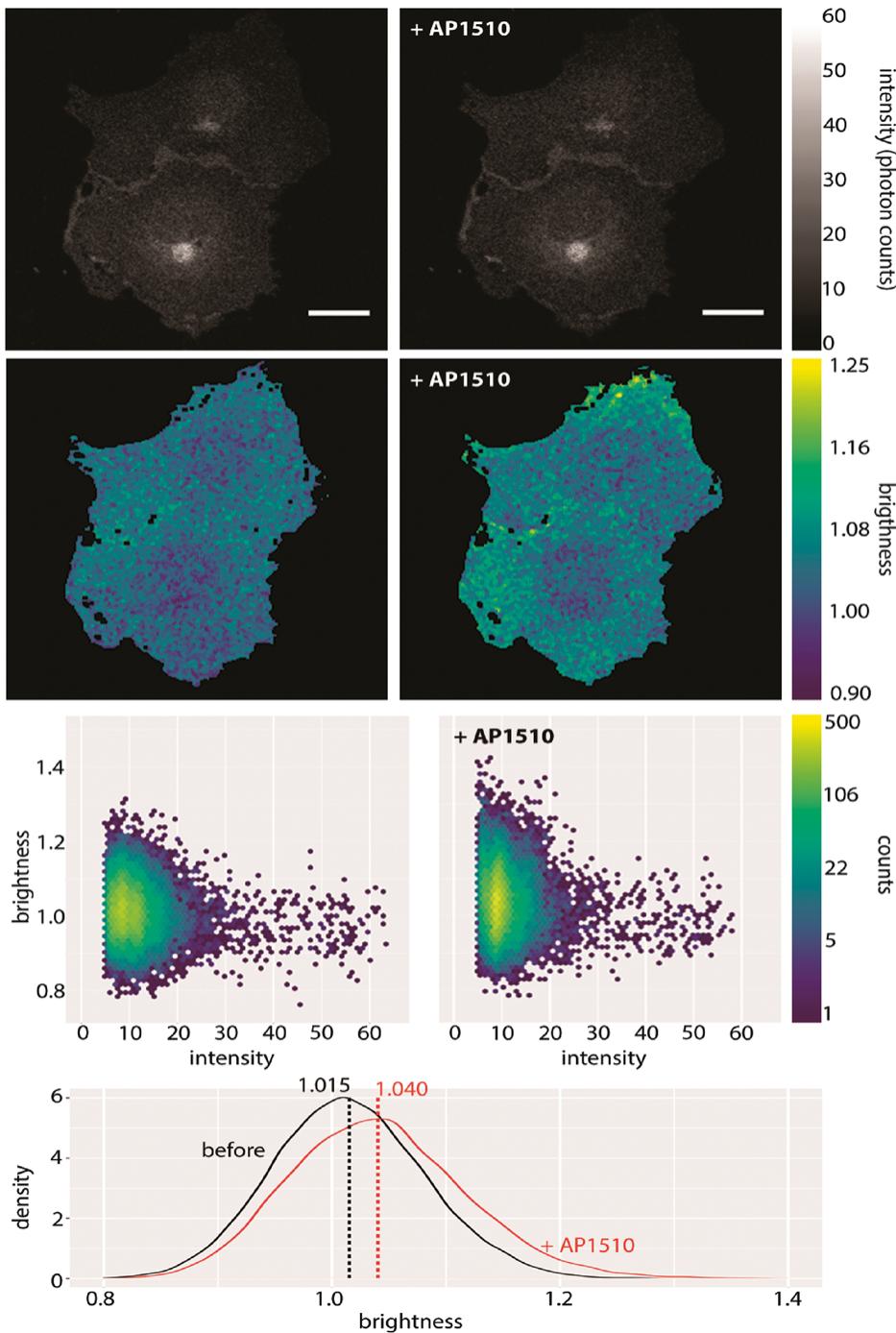


Figure 4.2: mClover-labelled myristoylated FKBP12 before and after application of 50nM AP1510. Shown here are intensity (first row), brightness (second row), a plot of intensity versus brightness (third row) and brightness histograms (fourth row). Notice how the change in brightness upon addition of the drug is seen most clearly by comparing the brightness histograms. The vertical lines in the histogram plot show the means of those histograms. Brightness here refers to B. Scale bar 20 μ m. (Nolan et al., 2017a)

4.2 *In vitro* number and brightness

4.2.1 Introduction

In our research group, we believe that the most practical quantitative method for measuring homo-dimerization *in vivo* and *in vitro* is N&B (Digman et al., 2008) because it is calibration-free and does not require specialized instrumentation. There are many examples of the application of N&B *in vivo* (the original N&B paper has over 250 citations, most of which are *in vivo* applications) but none *in vitro*. Hence, we published a protocol (Nolan et al., 2018) detailing how N&B can be applied *in vitro*. This time, we used FKBP12F36V which is an FKBP mutant with a new dimerizing drug AP20187 (known colloquially as the *BB dimerizer*). The fact that FKBP12F36V is an FKBP12 mutant and that BB only dimerizes this mutant means that the previous issue of the interaction of endogenous and exogenous proteins is eliminated.

4.2.2 Experimental results and discussion

In this experiment, the FKBP12F36V was labelled with mVenus. We found that the brightness doubled from $\epsilon = 0.005$ to $\epsilon = 0.010$ upon addition of the drug. See figure 4.3. This analysis was done with exponential smoothing detrending with

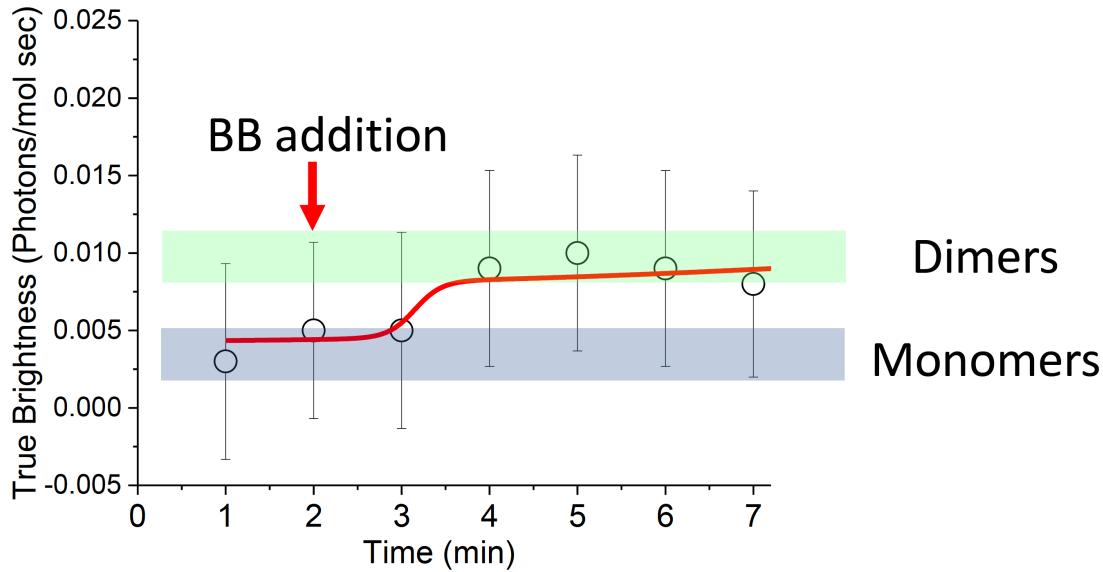


Figure 4.3: Dimerization of FKBP12F36V upon BB addition is seen by a brightness doubling from $\epsilon = 0.005$ to $\epsilon = 0.010$ over a period of minutes.

automatically chosen parameter τ . Without detrending, the pre-BB brightness was calculated as $\epsilon = 0.026$, whereas the post-BB brightness was calculated as $\epsilon = 0.010$ again, which would lead one to conclude that the oligomeric state decreased upon BB addition. This is a nonsensical result given that the FKBP12F36V is monomeric to begin with. This shows that detrending is absolutely necessary and that neglecting this step can lead to incorrect results.

Remark. This paper also included an important correction to the equation for brightness ϵ when analog equipment is used. The correct equation is

$$\epsilon = \frac{\sigma^2 - \sigma_0^2}{S(\langle I \rangle - \text{offset})} - 1 \quad (4.1)$$

The S in the denominator was omitted in the original paper (Dalal et al., 2008) and this error was reproduced in our N&B review (Nolan et al., 2017b).

4.3 HIV-1 receptor stoichiometry

4.3.1 Introduction

4.3.1.1 HIV-1 cell entry

HIV-1 infects many cell types (e.g. CD4 T cells, macrophages, dendritic cells) and has different modes of entry for each cell type and indeed possibly more than one mode of entry in any given cell type (Jakobsdottir et al., 2017). Endocytosis is thought to be a common entry route (Miyauchi et al., 2009), particularly in macrophages (Marechal et al., 2001) and Dendritic cells (Ménager and Littman, 2016). In CD4 T cells, HIV-1 has been shown to fuse at the plasma membrane without needing endocytosis (Herold et al., 2014).

Entry of HIV into any cell involves the initial binding of the CD4 receptor on that cell by the HIV-1 virus. Subsequently, a co-receptor (often CCR5 or CXCR4) is used in the fusion process (Jakobsdottir et al., 2017). The question of how many receptors and co-receptors are required to facilitate fusion (the *stoichiometry* of

the interaction of HIV-1 with its receptor and co-receptor—possibly different for different cell types) had not been answered.

4.3.1.2 The use of number and brightness to study HIV-1 cell entry

Our main motivation for studying N&B in the first place was that we thought it was a valuable method to study the process of HIV-1 fusion in live cells. N&B was first used in our research group to study the oligomeric state of dynamin at the HIV-1 fusion pore in TZM-bl cells (Jones et al., 2017). This study concluded that dynamin-2 stabilizes the HIV-1 fusion pore with a low oligomeric state.

Following on from this, we wanted to study the stoichiometry of the interaction of HIV-1 with its receptor (CD4) and co-receptor (CCR5 or CXCR4) upon the engagement of the virus with the cell and to follow this interaction stoichiometry up to the point of fusion; see figure 4.4.

Entry of HIV-1 into a host cell requires an initial interaction between the viral-envelope glycoprotein spike complex—Env—with cell surface displayed CD4 and co receptors (Jakobsdottir et al., 2017). Although structural studies have revealed the intra-molecular basis for CD4 receptor and CXCR4/CCR5 co-receptor-induced conformational changes to the HIV-1 Env during host cell entry (Ozorowski et al.,

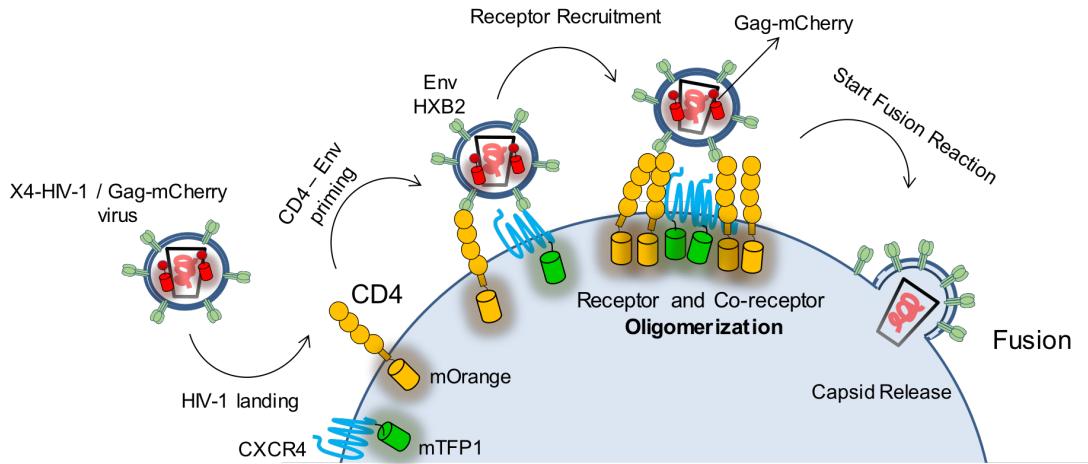


Figure 4.4: The HIV-1 envelope glycoprotein Env must bind the receptor (CD4) and form a complex with the co-receptor (CXCR4 or CCR5, this figure shows an X4-tropic virus and co-receptor) to initiate the fusion process. Labeling the viral Gag protein with mCherry, the receptor with mTFP1 and the co-receptor with mOrange, it is possible to follow these three players in the fusion reaction and to quantify their interaction.

2017), little is known about how the inter-molecular dynamics and stoichiometry of this process culminates in fusion with the host cell membrane in live cells (Brandenberg et al., 2015). This is due to the difficulty of working with live cells and the lack of temporal resolution of the techniques commonly employed (i.e. crystallography and cryo-EM).

— Iliopoulou et al. (2018)

We saw N&B as the ideal technique to probe this stoichiometry temporally. With our microscope, we could acquire 100 frames per 1.7 minutes, therefore, using

each consecutive sequence of 100 frames to create a brightness image, we could obtain 1 brightness image every 1.7 minutes and use this to calculate this temporal stoichiometry.

4.3.2 Experimental setup

Cos7 cells were transfected with labelled receptor (CD4) and co-receptor (CXCR4 or CCR5) (these cells do not express these proteins endogenously). Virus was added at time $t = 0$ and imaging proceeded for a number of minutes at 100 frames per 1.7 minutes. Alternating laser excitation (ALEX, [Kapanidis et al. \(2005\)](#)) was used to eliminate the possibility of channel bleed-through. See figure 4.5.

4.3.3 Analysis

The virus channel was used to locate the virus at a given point in time. The receptor and co-receptor were used to calculate brightness and cross-correlated brightness every 100 frames (every 1.7 min). The brightness was used to determine the number of receptor and co-receptor units involved in a complex. The cross-correlated brightness was used to delimit whether or not the receptor and co-receptor units were together in the *same* complex. See figure 4.6, this is the

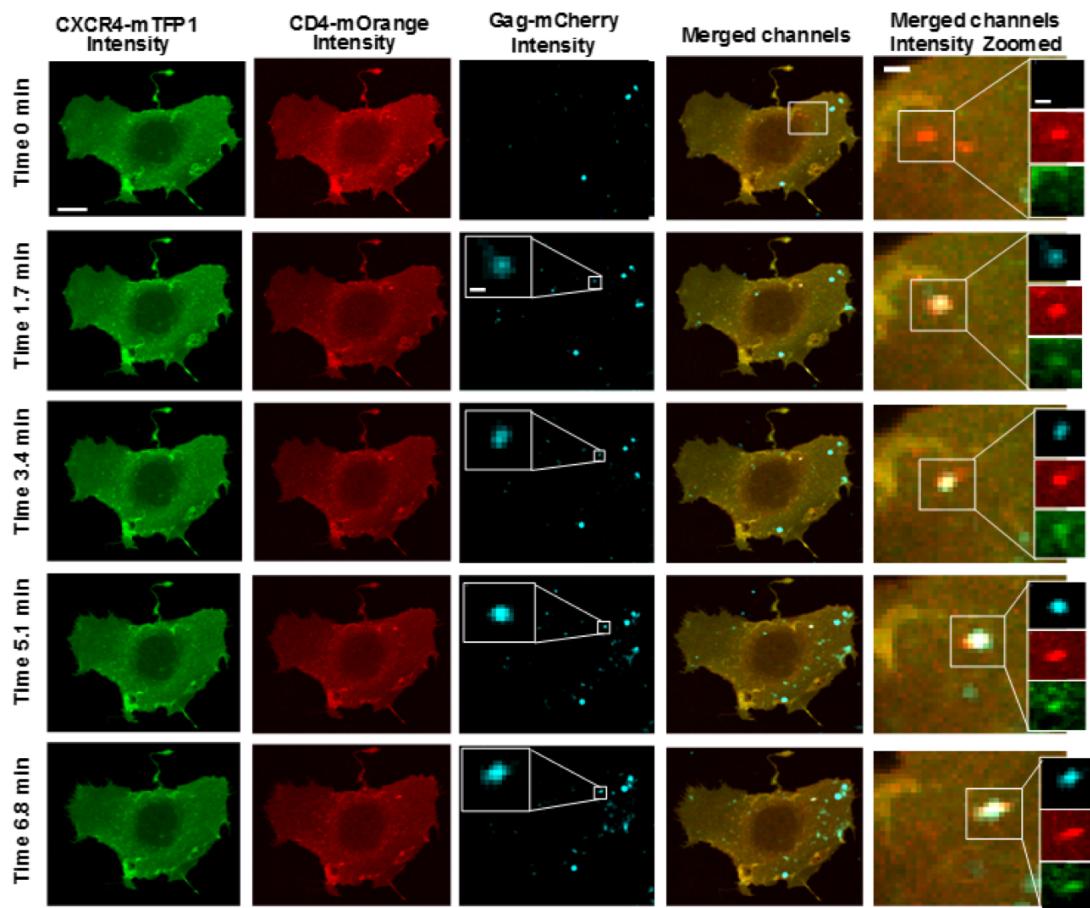


Figure 4.5: Intensity images from the virus, receptor and co-receptor. Every 100th frame is shown. A virus which lands at $t \approx 1.7$ minutes is highlighted.

corresponding brightness and cross-correlated brightness image of figure 4.5. Notice that once the virus lands, the oligomeric state of the receptor and co-receptor increases. We also see significant positive cross-correlated brightness in this area, indicating that the virus has triggered a complex of receptor and co-receptor.

4.3.4 Results

Figure 4.7 shows the results of the analysis detailed in figures 4.5 and 4.6 for $n = 10$ cases where virus triggered receptor and co-receptor complexes in the X4-tropic setting and $n = 12$ in the R5-tropic setting. A three-step pre-fusion process is hypothesized for each.

Our studies support a dynamic three step model for both HIV_{HXB2} and HIV_{JR-FL} (figure 4.8). For X4 tropic virions, Env – CD4 interactions induce CXCR4 dimerization, CD4 then engages with two Env (shown by 3 color TIRF-dSTORM microscopy) to generate a hexamer that might serve as a scaffold to stabilise a final 4 CD4 – 1/2 CXCR4 conformation, with a single Env. We speculate that for HIV_{HXB2}, step 2 is crucial to culminate the fusion reaction and there could be an anchoring domain and a fusion domain that undergoes gp120 disassembly leading to 6 helix bundle formation. For R5-tropic virions, Env – CD4

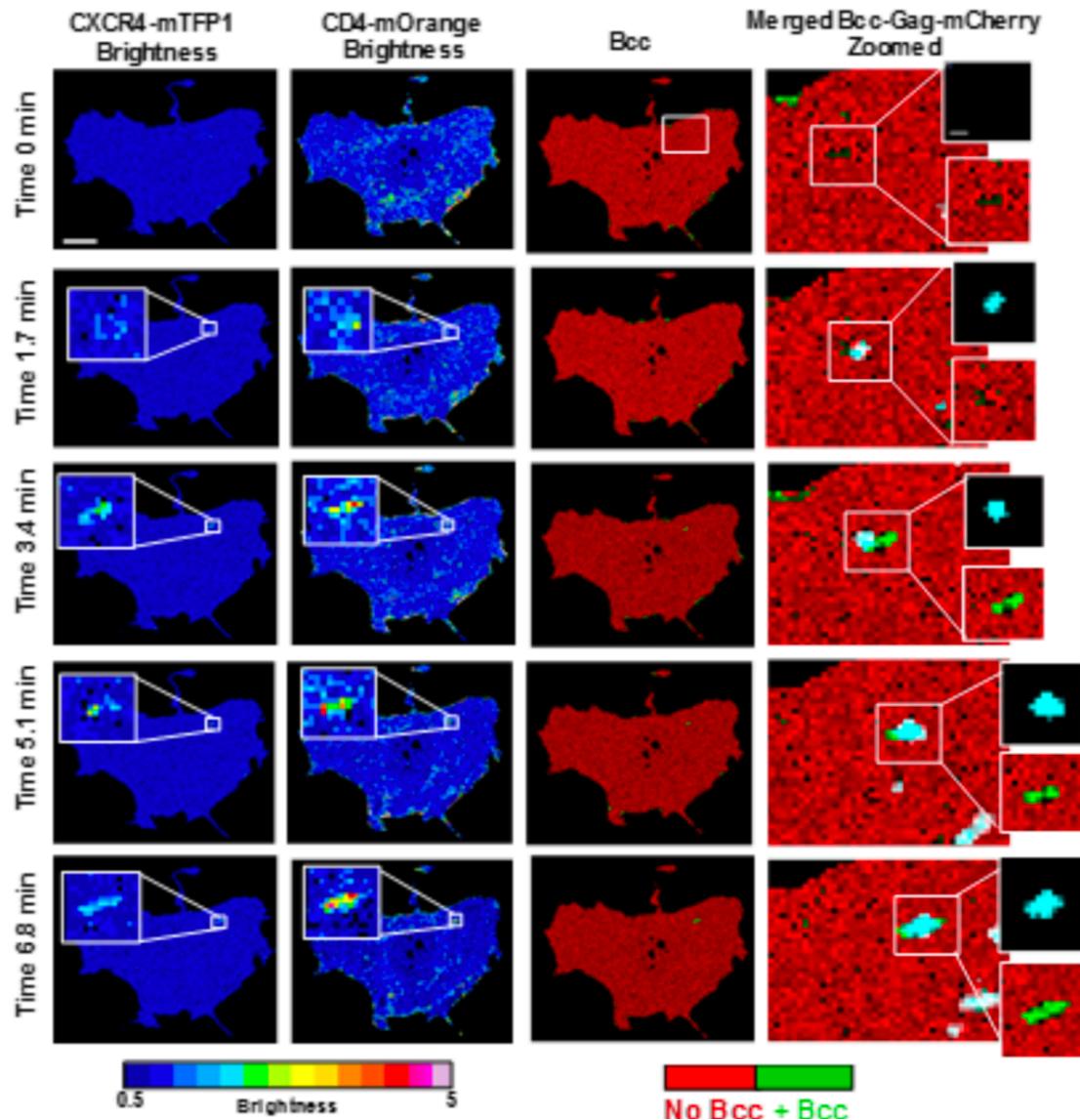


Figure 4.6: Brightness images of receptor and co-receptor and cross-correlated brightness image of the interaction between the two.

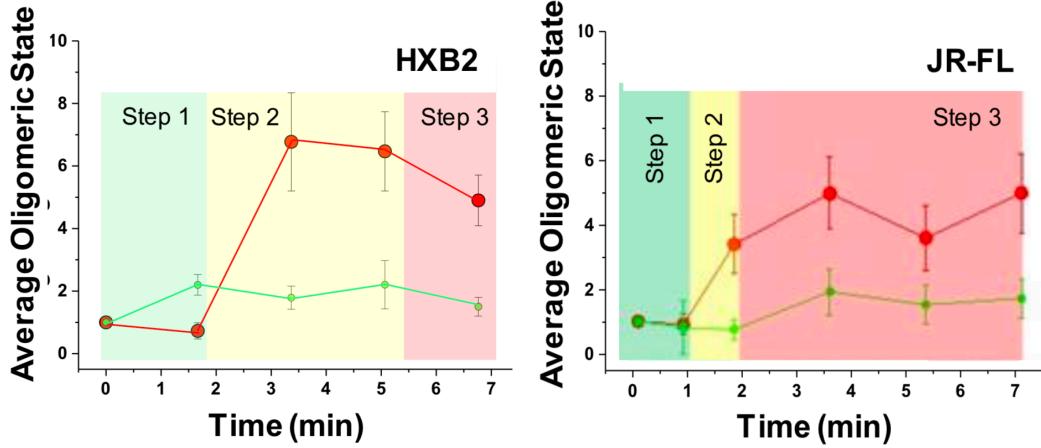


Figure 4.7: Number of receptor and co-receptor units involved in complexes with virus over time, obtained by brightness analysis. Left panel: HIV_{HXB2}. Right panel: HIV_{JR-FL}.

interactions form the previously described asymmetric pre-hairpin intermediate (Munro et al., 2014) (Kwon et al., 2015) (Ma et al., 2018), following binding and oligomerisation of 2 additional CD4 molecules with concomitant CCR5 dimerization. After this, the secondary intermediate leads to the final fusion competent complex with a total of 4 ± 0.3 CD4, 2 ± 0.3 CCR5 and 1 JR-FL Env. Interestingly, dSTORM experiments show an average stoichiometry for step 3 of 3.4 ± 0.1 CD4, 1.5 ± 0.2 CCR5.

Our data indicate that both HXB2 Env and JR-FL Env start with an asymmetric intermediate bound to a single CD4, as previously suggested. Our models also support the existence of important differences in the entry mechanisms of X4 and R5 strains. In the X4 strains,

CXCR4 dimerization (Tan et al., 2013) (Qin et al., 2015) occurs prior to CD4 hexamer formation and following initial Env – CD4 recognition (Liu et al., 2017). For R5 tropic JR-FL, CCR5 dimerization (Qin et al., 2015) occurs after Env-CD4 complexation and recruitment of two additional CD4 molecules (Wu et al., 1996) around the complex.

— Iliopoulou et al. (2018)

For the X4 and R5-tropic cases, structural modelling of the hypothesis has been done to visualize our findings and to assess their viability; this produced figure 4.8.

4.3.5 Conclusion

Time-resolved N&B enabled us to answer questions about the interaction between HIV-1 and its receptor and co-receptor in live cells which up to now could not be answered. We addressed this problem with the N&B technique. Successful application of N&B relies on being able to correct for bleaching reliably (as seen in section 4.2). This was impossible before the advent of my bleaching correction algorithms, therefore their development was a key step in making this work possible.

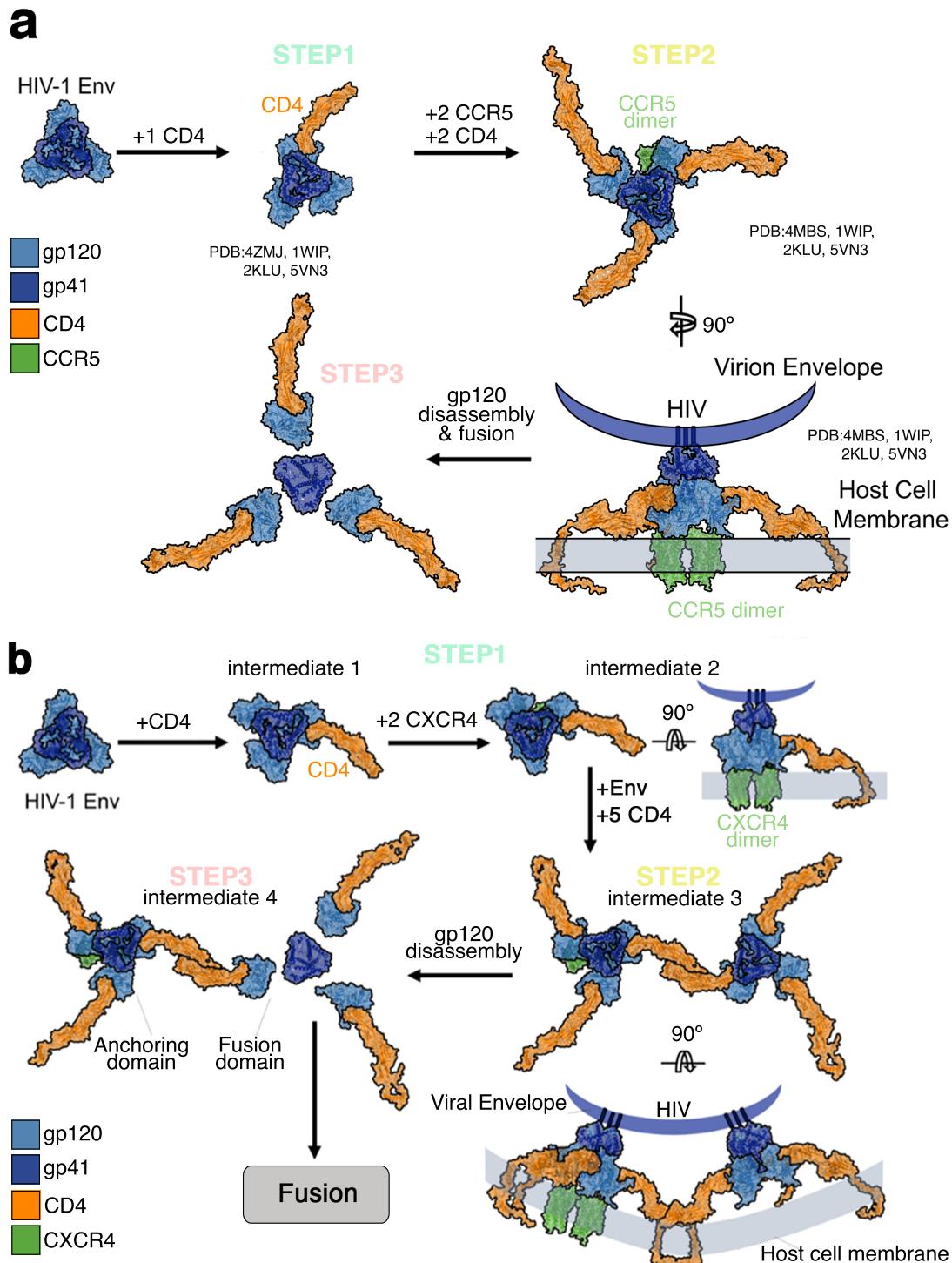


Figure 4.8: Model of pre-fusion reaction sequences for the R5 (a) and X4 (b) tropic cases.

Chapter 5

Discussion

5.1 Fluorescence fluctuation spectroscopy

Even with photon-counting detectors, FFS is a difficult technique. There are many pitfalls: the acquisition settings (dwell time and frame rate) must be correct and the correct settings for these parameters depend on the residence time τ_D of the protein of interest, which is non-trivial to measure. The acquired data must be checked to ensure there is not an excess of photobleaching and if there is not, there will inevitably still be some bleaching, so this must be corrected for. Until now, it was practically impossible to perform this correction correctly, because all methods required the user to select a vital correction parameter (τ or l) without

providing any instructions as to how the parameter should be chosen. I have solved this problem such that now image series can be safely detrended by novice users, as this parameter is chosen for them in the background. Now, detrending an image `img.tif` with the *Robin Hood* method is as simple as typing the command `img_detrend_rh("img.tif")` in my software. This should make FFS techniques safer and easier to use, opening FFS techniques up to more users, however the expertise required is still such that FFS may struggle to expand from the domain of microscopy and biophysics into a more commonly used biological technique.

5.2 The evolution of detrending algorithms

Previously, FFS detrending methods were based on smoothing methods taken from the field of time-series analysis. The fact that these smoothing methods required a choice of smoothing parameter was ignored by sticking to the arbitrary choice of $l = 10$ for this parameter.

My work in investigating the significance of this smoothing parameter found that this arbitrary choice was totally inappropriate. The use of simulated image series and the fact that immobile particles have brightness $B = 1$ opened up a means of solving for the correct choice of this parameter without the need for user input. This was the first set of *automatic detrending* methods, whereby to detrend, the

users task was as simple as clicking a **detrend** button.

Still, using smoothing approaches to detrend low-intensity data is problematic because it involves approximating very discrete time series with continuous functions; this is unwise and unnecessary. The *Robin Hood* idea of giving photon counts directly from one pixel to another in an image series circumvents the need for smoothing. The detrending process can be simplified from $\mathbb{N}_0 \rightarrow \mathbb{R} \rightarrow \mathbb{N}_0$ to $\mathbb{N}_0 \rightarrow \mathbb{N}_0$. Conveniently, the automatic parameter finding approach used in the smoothing approaches to detrending can readily be extended to *Robin Hood* detrending.

There are no obvious caveats to the *Robin Hood* bleaching correction method. However, it is vital to bear in mind that one should always try to avoid the source of error in the first place rather than rely on correction methods.

5.3 Applications of the new detrending techniques

5.3.1 FKBP

The FKBP applications of these detrending techniques in Cos-7 cells (Nolan et al., 2017a) and in-vitro were mainly to demonstrate that N&B used with these detrending techniques is a reliable method to measure oligomerization. The in vitro study (Nolan et al., 2018) was particularly interesting because it was the first in vitro application of N&B.

5.3.2 HIV-1 receptor stoichiometry

The study of HIV-1 receptor stoichiometry (Iliopoulou et al., 2018) is a *real-life* application of N&B and ccN&B, made possible by the automatic detrending algorithm. We have shown that this kind of fine-grained information about the process of HIV-1 fusion can be measured on a temporal basis in live cells. Whilst this alone is very exciting, it paves the way for similar studies to be done with HIV-1 in different cell types and indeed for other virus fusion processes to be probed in this way.

5.3.3 Multiplexing with structural biology

Our collaboration with structural biologists (Iliopoulou et al., 2018) is a demonstration of how live cell fluorescence microscopy and structural biology can be complementary. The sub-molecular insight from structural biology is not available from live cell fluorescence microscopy, while the dynamic information from live cell fluorescence microscopy cannot be gotten from structural biology.

5.3.4 Fluorescence fluctuation spectroscopy

FFS has many applications. Indeed the original N&B paper (Digman et al., 2008) alone has over 250 citations. All of these and future FFS studies require detrending to be reliable. *Robin Hood* detrending is the answer for this. A major challenge will be making *Robin Hood* visible, available and easy to use for the community. This means that the algorithm must be peer-reviewed, made available in all of the major free imaging softwares (ImageJ, python, R) and very well documented: a good manual is essential with any software package.

Chapter 6

Future plans

6.1 Translate software to *ImageJ*

Whilst I really like R, the fluorescence community does not use it, which has been a major barrier to the use of my algorithms by others. I will code my detrending algorithms as *ImageJ* plugins and also as python modules so that more of the community have easy access to them.

6.2 Study real data bleaching profiles with simulations

It was mentioned in section 3.10 that it would be possible to study the effect of real bleaching as opposed to simulated ideal bleaching and why it is more necessary to detrend in the real data case, by mimicking real bleaching profiles with simulations. This is something I would like to do. It would be difficult because it would require the collection and cataloging of a diverse set of real data bleaching profiles from various biological samples. This would be the bottleneck because I have already written the simulation and analysis pipelines.

6.3 Robin Hood for single point FCS

Right now, Robin Hood works only for imaging FFS/FCS because it has a step where it uses the mean intensity of a frame. With single point, there are no frames to take the mean of. This problem can be overcome with the use of smoothing splines. I will work on this when the thesis work is done.

6.4 Robin Hood publication

Once *Robin Hood* has been implemented in *ImageJ* and more realistic bleaching problems have been studied, *Robin Hood* should be published.

6.5 Revisit old N&B data with new detrending algorithm

Having claimed that Robin Hood detrending is significantly better than previous detrending methods, it makes sense to revisit old results found using old detrending methods to see how much they would be changed with the application of this new detrending algorithm. The main difficulty in such a project would be obtaining the old data. To this day, most microscopy publications do not have their associated images in a centralised repository, so the only way to get this data is to request it from the authors.

6.6 Compare FRET with FCS

I have the idea that when the question “Do these proteins interact?” is answered by Forster resonance energy transfer (FRET, Förster (1948)), it should also be answerable by FCS. I would like to try to reproduce some standard FRET results with FCS. I am particularly interested to find out if there are instances where one technique succeeds in detecting interaction and the other fails, and why this might be. For example, it might be possible that for a given interacting pair of proteins, it is impossible to label them such that the FRET couple is close enough for FRET to be detected, but that this interaction is detectable by FCS. These techniques have been compared before (Sahoo and Schwille, 2011), but not by trying to reproduce previous work. In addition, attempts by new groups to reproduce work that is accepted in the literature are always interesting (Baker, 2016).

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