

Cancer Stem Cells and Tumor Population Dynamics

Shekoofeh Rahanama - Zahra Maleki

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Overview

- 1 Introduction to Cancer Stem Cells
- 2 Experimental Identification of CSCs
- 3 Population Dynamics Modeling
- 4 Conclusion
- 5 References

Biology of cancer

- TNM
- Oncogene
- cancer cell structure:

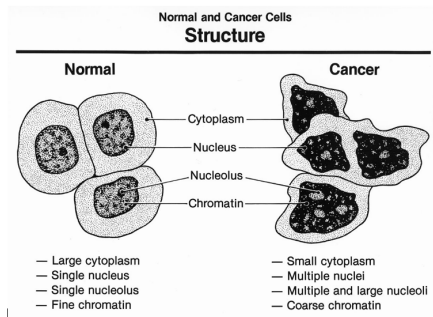


Figure: cancer cell structure

Core Hypothesis

- Tumors driven by small subpopulation: **Cancer Stem Cells (CSCs)**
- Key properties:
 - Self-renewal capability
 - Differentiation into heterogeneous cancer cells
- Cancer stem cells were first identified by John Dick in acute myeloid leukemia in the late 1990s.

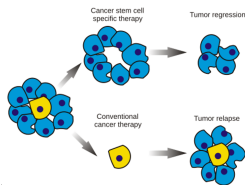


Figure:

Identification Protocol

Standard approach:

- 1 Sort cells via FACS using surface markers (e.g., CD44⁺/CD24⁻)
- 2 Transplant sorted cells into immunocompromised mice
- 3 Observe tumor formation
- 4 Re-isolate marker-positive cells from new tumor
- 5 Repeat transplantation (serial passaging)

Key evidence: Tumor formation exclusively by marker-positive cells

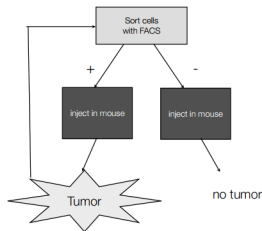


Figure: protocol used to identify CSCs

Challenges and Limitations

- **Transplantation artifacts:**

- Xeno/syngeneic grafts miss tumor microenvironment (fibroblasts, cytokines)

- **Marker reliability issues:**

- No universal marker (e.g., ALDH works in breast cancer but not melanoma)
- Marker expression may be reversible

- **Model dependence:**

- Tumor-initiating cell fraction varies with mouse strain

- **Therapeutic complications:** Targets like Notch pathway cause high toxicity

Branching Process Model

Cell types:

- CSCs: Unlimited divisions
- Cancer Cells (CCs): Finite divisions (M generations)
- Senescent cells: Death rate q

CSC division probabilities:

- Symmetric: $\text{CSC} \xrightarrow{p_2} \text{CSC} + \text{CSC}$
- Asymmetric: $\text{CSC} \xrightarrow{p_1} \text{CSC} + \text{CC}$
- Symmetric differentiation: $\text{CSC} \xrightarrow{p_0} \text{CC} + \text{CC}$

Key parameter: $\epsilon = p_2 - p_0$ (net CSC increase/generation)

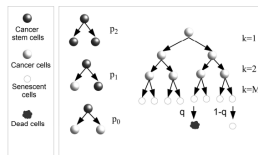


Figure: branching process for cancer stem cells

Dynamics Equations

Recursion relations (S^N : CSCs at gen. N ; C_k^N : CCs of "age" k):

$$S^N = (1 + \epsilon)S^{N-1}$$

$$C_1^N = (1 - \epsilon)S^{N-1}$$

$$C_k^N = 2C_{k-1}^{N-1} \quad (k \geq 2)$$

$$D^N = (1 - q)D^{N-1} + 2C_M^{N-1}$$

Eq. 5.2 - Only CSCs ($C_k^0 = D^0 = 0$)

$$S^N = (1 + \epsilon)^N S^0$$

$$C_k^N = \begin{cases} S^N \left(\frac{2}{1+\epsilon}\right)^{k-1} \frac{1-\epsilon}{1+\epsilon} & N > k \\ 0 & N \leq k \end{cases}$$

$$D^N = \begin{cases} S^N \frac{1-\epsilon}{\epsilon+q} \left(\frac{2}{1+\epsilon}\right)^M \left(1 - \left(\frac{1-q}{1+\epsilon}\right)^{N-M}\right) & N > M \\ 0 & N \leq M \end{cases}$$

Eq. 5.3 - Non-CSC Dynamics ($S^0 = 0, C_k^0 = D^0 = c$)

$$\hat{S}^N = 0$$

$$\hat{C}_k^N = \begin{cases} 2^k c & N < k \\ 0 & N \geq k \end{cases}$$

$$\hat{D}^N = \begin{cases} c(1-q)^N + c \frac{2}{1+q} (2^N - (1-q)^N) & N \leq M \\ c(1-q)^N \left(1 + \frac{2}{1+q} \left(\left(\frac{2}{1-q} \right)^M - 1 \right) \right) & N > M \end{cases}$$

Complete Solution (Eq. 5.4)

$$n_{\text{tot}}^N = \underbrace{S^N}_{\text{CSCs}} + \hat{S}^N + \sum_{k=1}^M \left(\underbrace{C_k^N}_{\text{CSC-derived}} + \underbrace{\hat{C}_k^N}_{\text{non-CSC}} \right) + \underbrace{D^N}_{\text{CSC-senescent}} + \underbrace{\hat{D}^N}_{\text{non-CSC senescent}}$$

Senescent cell fraction: $f_{SC}^{\infty} = \frac{1 - \epsilon}{1 + q}$

CSC fraction: $f_{CSC}^{\infty} = \frac{q + \epsilon}{1 + q} \left(\frac{1 + \epsilon}{2} \right)^M$

Interpretation:

- Normal tissue ($\epsilon = 0$): High senescence, low CSCs
- Tumors ($\epsilon > 0$): Reduced senescence, increased CSCs
- Aggressive tumors: High $\epsilon \Rightarrow$ high CSC fraction

Experimental validation: Matches melanoma growth curves (ABCG2+ vs. ABCG2- cells)

Phenotypic Switching

Mathematical models predict that depletion of CSCs induces a rapid return to the CSC state through phenotypic switching, with an overshoot effect.

Model Equations

$$\begin{aligned}\frac{dS}{dt} &= R_d (\epsilon S + p(\mu)(C + D)) \\ \frac{dC}{dt} &= R_d(1 - \epsilon)S - C(1 + p(\mu)) \\ \frac{dD}{dt} &= R_d(2C - p(\mu)D)\end{aligned}$$

Where:

S = CSC population, C = cancer cells, D = senescent cells, and $p(\mu)$ = switching probability influenced by miRNAs.

Overshoot in CSC population

The model indicates that once the CSC population is depleted, an overshoot occurs due to a feedback mechanism in the miRNA network.

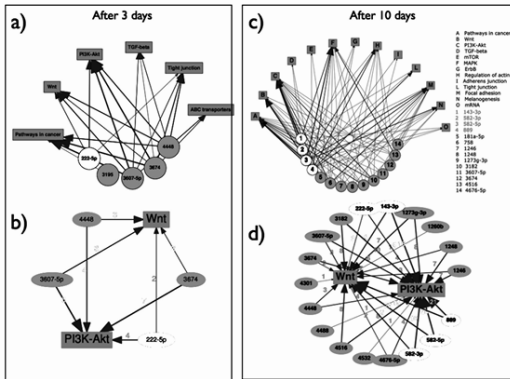







Figure: Interaction network between miRNAs and pathways during CSCswitch in melanoma

Key Takeaways

- CSCs are essential for tumor growth, but their identification and quantification remain challenging.
- Population dynamics models help explain CSC-driven tumor growth.
- Experimental and theoretical models must account for complex phenomena like phenotypic switching, which is influenced by both environmental factors and genetic regulation.

References

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-  Gupta, P.B., et al. (2011), *Stochastic state transitions give rise to phenotypic equilibrium in populations of cancer cells*.
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