

Interacting associations between ploidy, breeding system, and lineage diversification

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Abstract

The effect of polyploidy in diversification remains a contentious issue. On the one hand, recent studies that found that polyploids have slower speciation rates and higher extinction rates than diploids left scientist wondering if polyploidy is truly an evolutionary dead-end. On the other hand, botanist have found strong
5 molecular support of multiple polyploidy events at the root of highly diverse clades which challenges the evolutionary dead-end conclusions reached by modeling approaches. We re-investigate the role of polyploidy in speciation and extinction from a new modeling perspective considering that patterns found in diversification models can be misleading and incorrectly attributed to polyploidy when other observed and unobserved plant traits are responsible of shaping diversification. Using statistically robust comparative
10 phylogenetic approaches, we show that it is possible to detect whether the contribution of polyploidy to speciation and extinction is significant under the presence of other potential traits also affect diversification. We use the phylogeny, polyploidy, and breeding system data of 595 Solanaceae species to understand the contribution of polyploidy to diversification. We ask if Solanaceae polyploids are evolutionary dead-ends, and whether breeding system or some other unobserved traits are responsible of the patterns of diversification
15 observed in the phylogeny.

Introduction

Species accumulate around the tree of life at different rates. The search for traits that explain these differences has been accelerated by dramatic increases in phylogenetic data and, despite some setbacks (Maddison and FitzJohn 2015; Rabosky and Goldberg 2015), advances in analytical methods (Maddison et al. 2007; 20 FitzJohn et al. 2009; Goldberg and Igić 2012; Beaulieu and O’Meara 2016). In these studies, it is common to identify a single focal trait and investigate its association with rates of speciation and extinction.

This is problematic because the context in which traits occur can lead to complex interactions, perhaps with other traits that make up an organism or its environment. Additionally, a lack of methods that allow for simultaneous inference of effects of other traits, which also influence diversification, can lead to 25 incorrect conclusions about the focal trait (Rabosky and Goldberg 2015; Beaulieu and O’Meara 2016). One approach to this problem is to allow for a ‘hidden’ or unspecified trait in the analysis (Beaulieu and O’Meara 2016). This can be effective, but it leaves behind the question of whether the hidden trait corresponds to a real trait that should be sought, or whether it is an approximation of some unknowable heterogeneity, as well as whether interactions between the known and hidden trait were modeled appropriately. Alternatively, we can 30 simultaneously consider the effects of more than one known trait, especially in systems where multiple traits are suspected to influence diversification, and where interactions between those traits are well understood. In this paper, we focus on two of the best studied traits in flowering plants, ploidy and breeding system, and investigate their individual and joint effects on diversification.

Polyploidization multiplies the genomic content of cells, and it consequently has the potential to 35 affect many or all traits and a great variety of evolutionary processes. It is also a mutation that occurs commonly in plants, as it is widespread at both population and lineage scales. The prevalence of variation in chromosome number, and especially ploidy, has been broadly considered a salient feature of flowering plants for nearly a century (Stebbins 1938). The recent dramatic increase in the scale of available genome sequences uncovered ancient rounds of whole-genome duplications, and subsequent diploidization, across 40 angiosperms (Lynch and Conery 2000; Vision et al. 2000). Nearly all lineages of flowering plants are thought to have undergone at least one or two rounds of polyploidization. The prevalence of polyploidy, its variation across clades, and its large effects on genotypes and phenotypes raises the hypothesis that ploidy plays an important role in shaping rates of speciation and extinction. This has led to a debate: Mayrose et al. (2011, 2015) found a lower rate of lineage diversification for polyploids, while Soltis et al. (2014) argued 45 that polyploidy is not an evolutionary dead end because whole genome duplications occurred at the base of many successful angiosperm clades, and Landis et al. (2018) reported higher diversification rates following

some whole genome duplications. Our study revisits this question, additionally incorporating the process of diploidization. Furthermore, we consider the interaction between polyploidy and other traits.

From the beginning, study of this variable cytogenetic property considered correlations with other traits (Stebbins 1938). Some of these associations are driven by indirect or ecological effects, such as the tendency for polyploidy, and other traits, to be found at higher latitudes or marginal habitats. Other correlations may have a more direct, causal connection. Among the many changes associated with polyploidization, perhaps the most prominent is the association between polyploidy and propensity for self-fertilization (Stebbins 1950). In some cases, the evidence for a correlated shift in mating system along with polyploidization appears limited and sometimes contradictory (Barringer 2007; Barrett 2008; Husband et al. 2008). In other cases, however, polyploidy is not only a suspected correlate of breeding systems but indeed a causal link (Stout and Chandler 1942; Lewis 1947). Doubled number of alleles in pollen is thought to effect disruption of the genetic mechanisms in gametophytic self-incompatibility systems, which prevent self-fertilization (Entani et al. 1999; Tsukamoto et al. 2005; Kubo et al. 2010). This creates a correlation between polyploidy and self-compatibility by precluding the existence of self-incompatible polyploids. In clades with these systems, it is thus natural to consider the simultaneous macroevolution of ploidy and breeding system.

Breeding system shifts—changes in the collection of physiological and morphological traits that determine the likelihood that any two gametes unite—are remarkably common and affect the distribution and amount of genetic variation in populations (Stebbins 1974; Barrett 2013). In particular, self-incompatibility (SI) systems cause a plant to reject its own pollen, and their loss, yielding self-compatibility (SC), is one of the most replicated transitions in flowering plant evolution. Previous phylogenetic analyses have reported higher rates of diversification for SI than for SC lineages (Goldberg et al. 2010; de Vos et al. 2014), but they have not considered the possibility of other correlated traits driving this pattern. Given that changes in ploidy and breeding systems may be causally related and have profound effects on the fate of lineages, it seems particularly profitable to examine possible interactions in their macroevolutionary effects. This includes their joint influence on lineage diversification, and also potential patterns in the order of their transitions. For example, do losses of SI more commonly occur tied to polyploidization, or without a ploidy shift? Do polyploids arise more commonly from SI or SC diploids? Robertson et al. (2011) found that the pathway from SI diploids to SC polyploids is dominated by loss of SI followed later by polyploidization over long timescales, but proceeds in one step via polyploidization of SI species over short timescales. We revisit this question with a greatly improved phylogeny and methods that allow for diversification rate differences.

Here, we use extensive data on ploidy and breeding system in Solanaceae to investigate the associations of these two traits with lineage diversification. Considering each trait separately indicates that each

is connected to diversification differences. Considering them jointly, however, reveals that the ploidy connection is removed by incorporating breeding system. We further show that the general results are robust to allowing for diploidization and a hidden trait, and something about pathways. Our results emphasize the importance of considering traits not only in isolation, especially when there are strong correlations between them.

Methods

Data

Chromosome number data were obtained for all Solanaceae taxa in the Chromosome Counts Database (CCDB; Rice et al. 2015), and the ca. 14,000 records were cleaned semi-automatically using the CCDBcurator R package (Rivero et al. 2019). This large dataset includes the compilation of Solanaceae ploidy states from Robertson et al. (2011). Species were coded as either diploid (D) or polyploid (P). For the majority of species, ploidy was assigned according to information from the original publications and the Kew Royal Botanic Gardens C-value DNA resource (Bennett and Leitch 2005). For taxa without ploidy information but with information about chromosome number, we assigned ploidy based on the multiplicity of chromosomes within the genus. For example, *Solanum betaceum* did not include information about ploidy level but it has 24 chromosomes, so because $x = 12$ is the base chromosome number of the *Solanum* genus (Olmstead and Bohs 2007), we assigned *S. betaceum* as diploid. Species with more than one ploidy level were assigned the smallest and most frequent ploidy level recorded. Breeding system was scored as self-incompatible (I) or self-compatible (C) based on results curated from the literature and original experimental crosses (as compiled in Igić et al. 2006; Goldberg et al. 2010; Robertson et al. 2011; Goldberg and Igić 2012). Most species could unambiguously be coded as either I or C (Raduski et al. 2012). Following previous work, we coded as I any species with functional I systems, even if C or dioecy was also reported. Dioecious species without functional I were coded as C.

To those existing data sets, we added some additional records for chromosome number and breeding system. The Supplementary Information contains citations for the numerous sources for the added data. Resolution of taxonomic synonymy followed the conventions provided in Solanaceae Source (PBI *Solanum* Project 2012). Hybrids and cultivars were excluded because ploidy and breeding system can be affected by artificial selection during domestication. Following the reasoning outlined in Robertson et al. (2011), we examined closely the few species for which the merged ploidy and breeding system data indicated the presence of self-incompatible polyploids. Although SI populations frequently contain some SC individuals, and diploid populations frequently contain some polyploid individuals, in no case did we find a convincing

case of a naturally occurring SI and polyploid population. The single instance of an SI and polyploid individual appears to be an allopolyploid hybrid of *Solanum oplocense* Hawkes x *Solanum gourlayii* Hawkes, reported by Camadro and Peloquin 1981. Under exceedingly rare circumstances, it is possible for polyploids containing multiple copies of S-loci to remain SI, so long as they express a single allele at the S-locus (discussed in Robertson et al. 2011). Because of the resulting absence of SI and polyploid populations, as well as the linked functional explanation for disabling of gametophytic self-incompatibility systems with non-self recognition, following whole genome duplication (reviewed in Ramsey and Schemske 1998; Stone 2002), we consider only three observed character states: self-incompatible diploids (ID), self-compatible diploids (CD), and polyploids which are always self-compatible (CP).

Matching our character state data to the largest time-calibrated phylogeny of Solanaceae (Särkinen et al. 2013) yielded 595 species with ploidy and/or breeding system information on the tree. Binary or three-state classification of ploidy and breeding system for the 595 taxa is summarized in Fig. 1. We retained all of these species in each of the analyses below because pruning away tips lacking breeding system in the ploidy-only analyses (and vice versa) would discard data that could inform the diversification models. A total of 405 taxa without any information about breeding system or polyploidy were excluded. Tips without trait data are much less informative for diversification parameters linked to trait values. Including this many more species would have prohibitively slowed our analyses, especially those implementing the most complex models.

Models for ploidy and diversification

To investigate the association between ploidy level and diversification, we first defined a binary state speciation and extinction model (BiSSE, Maddison et al. 2007) in which taxa were classified as diploid (D) or polyploid (P) (Fig. 1). We call this the *D/P ploidy* model. In a Bayesian framework, we obtained posterior probability distributions of speciation rates (λ_D , λ_P), extinction rates (μ_D , μ_P), net diversification rates ($r_D = \lambda_D - \mu_D$, $r_P = \lambda_P - \mu_P$), and relative extinction rates ($v_D = \mu_D/\lambda_D$, $v_P = \mu_P/\lambda_P$) associated with each state. This analysis explores the same question as Mayrose et al. (2011, 2015), but our analyses differ because we include not only polyploidization (parameter ρ , the transition rate from *D* to *P*), but also diploidization (parameter δ , the transition rate from *P* to *D*).

Our second model assesses the signal of diversification due to ploidy differences while also parsing out the heterogeneity of diversification rates due a possible unobserved trait. BiSSE-like models can suffer from a large false discovery rate because they fail to account for diversification rate changes that do not directly depend on the trait of interest (Rabosky and Goldberg 2015; Beaulieu and O’Meara 2016). Diversification rate differences explained by something (trait) other than ploidy, are accommodated by adding

a hidden state (HiSSE model; Beaulieu and O’Meara 2016). In this model, each of the observed diploid and polyploid states is subdivided by a binary hidden trait with states *A* and *B*. We call this the *D/P+A/B ploidy and hidden state* model. We estimated the posterior probability distributions of speciation rates (λ_{D_A} , λ_{D_B} , λ_{P_A} , λ_{P_B}), extinction rates (μ_{D_A} , μ_{D_B} , μ_{P_A} , μ_{P_B}), net diversification rates (r_{D_A} , r_{D_B} , r_{P_A} , r_{P_B}), and
145 relative extinction rates (v_{D_A} , v_{D_B} , v_{P_A} , v_{P_B}). In this model polyploidization rate ρ and diploidization rate δ are also included, and changes between hidden states are symmetrical with rate α .

Models for breeding system and diversification

To assess the effects of breeding system in the diversification process, we first fit model in which the states are self-incompatible (I) or self-compatible (C). This is the same as the analysis of Goldberg et al. (2010),
150 save for an updated phylogeny (Särkinen et al. 2013). We call this BiSSE model the *I/C breeding system* model. To parse out the effect of breeding system on diversification, while allowing for the possibility of heterogeneous diversification rates unrelated to breeding system, we subdivided each of those states into hidden states *A* and *B*. We call this HiSSE model the *I/C+A/B breeding system and hidden state model*.

For all breeding system models, we allow transitions from *I* to *C* (at rate q_{IC}) but not the reverse.
155 Within Solanaceae, self-incompatibility is homologous in all species in which S-alleles were cloned, and controlled crosses performed. All species sampled to date, possess a non-self recognition, RNase-based, gametophytic self-incompatibility (shared even shared even with other euasterid families; Ramanauskas and Igić 2017). Furthermore, species that are distantly related within this family carry closely-related alleles, with deep trans-specific polymorphism, at the S-locus, which controls the SI response (Ioerger et al. 1990;
160 Igić et al. 2006). This represents very strong evidence that the SI mechanism, and our *I* state is ancestral to the Solanaceae, and did not arise independently within the family ($q_{CI} = 0$).

Models for ploidy, breeding system, and diversification

If ploidy and breeding system each influence lineage diversification individually, it is logical to examine their possible joint effects. We thus fit a multi-state model that includes both traits (MuSSE, FitzJohn
165 2012). The three states in this model are self-incompatible diploids (ID), self-compatible diploids (CD), and polyploids, which are always self-compatible (CP). As explained above, we did not include a state for self-incompatible polyploids because they are not observed in the data, and that trait combination state is mechanistically predicted not to occur. We call this the *ID/CD/CP ploidy and breeding system* model. The model has 10 parameters, six for diversification in each state (λ_{ID} , λ_{CD} , λ_{CP} for speciation, μ_{ID} , μ_{CD} , μ_{CP} for extinction) and four for transitions between states (ρ_I , ρ_C for polyploidization transitions from *ID* and
170 *CD* to *CP*, respectively; δ for diploidization from *CP* to *CD*; q_{IC} for loss of self-incompatibility without

polyploidization, from *ID* to *CD*). The total rate of loss of self-incompatibility, i.e., transitions out of *ID*, is $q_{IC} + \rho_I$. Diploidization from *CP* to *ID* is not allowed because it would represent a simultaneous regain of SI.

The ID/CD/CP model could potentially capture similar dynamics as earlier models, if the effects of the hidden state in D/P+A/B were effectively caused by breeding system (or its correlates), and the hidden state in I/C+A/B was effectively caused by ploidy. There is also the potential, however, for a hidden factor to be influencing diversification beyond both of our focal traits, and this could again mislead inferences. We therefore added a hidden trait layer on top of our three-state model (analogous to Caetano et al. 2018; Herrera-Alsina et al. 2018; Huang et al. 2018). We refer to this as the *ID/CD/CP+A/B* model. A fully parameterized version of this model would have 26 rate parameters (Herrera-Alsina et al. 2018). Because our goal was to look for diversification rate differences associated with ploidy and breeding system rather than the specific effects of the hidden states, we fitted a simplified version with 16 parameters. The reduction in parameter space is achieved by fixing the rates for transitions among hidden states to be equal with rate α , and fixing the transition rates between observed states to be independent of the hidden state (rates ρ_I , ρ_C , δ , q_{IC} as defined for the ID/CD/CP model). There are additionally twelve diversification rate parameters (λ_{ID_A} , λ_{ID_B} , λ_{CD_A} , λ_{CD_B} , λ_{CP_A} , λ_{CP_B} , μ_{ID_A} , μ_{ID_B} , μ_{CD_A} , μ_{CD_B} , μ_{CP_A} , μ_{CP_B}).

Diploidization as an exploratory hypothesis

For all four models that consider ploidy changes, we allowed diploidization. Previous modeling approaches (Mayrose et al. 2011) have argued against inferring diploidization rates when using ploidy data that comes from classifications based on chromosome number multiplicity or chromosome number change models like chromEvol (Mayrose et al. 2010). These types of classifications do not allow for a ploidy reversion. Where indicated, the classification of ploidy for the data used in our models was based on chromosome multiplicity at the genus level. However, the majority of the ploidy classifications were adopted from original studies with alternative sources of information (e.g., geographic distribution, genus ploidy distribution) where ploidy was defined by authors that found evidence for it. Since it is not clear whether diploidization can be detected under alternative ploidy classifications or even classifications based on chromosome number multiplicity at the genus level, we also fit the models without diploidization in order to test whether the conclusions about diversification are sensitive to including diploidization. As discussed by Servedio et al. (2014), the presence or absence of a hypothesis can have an exploratory goal. In our case the diploidization parameter (or its absence, $\delta = 0$) in our models is an opportunity to explore an assumption that might be important but that is not the single definitive process to understand the interactions among polyploidy,

breeding system, and diversification.

Statistical inference under the models

205 Parameters for each of the 10 diversification models were estimated using custom code in the RevBayes (Höhna et al. 2016) environment. Code for analyses and key results is available at <https://github.com/roszenil/solploidy>. We included a correction for incomplete sampling in all analyses, based on assuming that the Solanaceae family has approximately 3,000 species ($s = 595/3000$) as estimated by the Solanaceae Source project (PBI *Solanum* Project 2012). For all 10 models, we assumed that speciation and
210 extinction parameters had log-normal prior distributions with means equal to the expected net diversification rate (number of taxa/ $[2 \times \text{root age}]$) and standard deviation 0.5. Priors for parameters defining trait changes were assumed to be gamma distributed with parameters $k = 0.5$ and $\theta = 1$. For each model, an MCMC chain was run for 96 hours in the high-performance computational cluster at the Minnesota Supercomputing Institute, which allowed for 5,000 generations of burn-in and a minimum of 200,000 generations of MCMC
215 for each of the 6 models. For each model, convergence and mixing of the MCMC was tested using the R library coda and the software package Tracer (see supplementary information for convergence plots).

Model selection

We calculated the marginal likelihood for each of the 10 models in RevBayes (Höhna et al. 2016). Marginal likelihoods were calculated using 50 stepping stone steps under the methodology of Xie et al. (2010). Each
220 stepping stone step was found by calculating 500 generations of burn-in followed by a total of 1,000 MCMC steps (Table 1). The calculation of each marginal likelihood ran for 24 hours on a high-performance computational cluster.

Using the marginal likelihood values, we calculated thirteen different Bayes factors. Six compared the models of polyploidy against each other (D/P and D/P+A/B, each with or without diploidization), one
225 compared the breeding system models (I/C and I/C+A/B), and six compared the models with both traits (ID/CD/CP and ID/CD/CP+A/B, each with or without diploidization) (Table 2). Other comparisons between these models are not valid because the input data are different under the different state space codings (Fig. 1). In essence, the D/P, I/C, and ID/CD/CP state spaces are not lumpable with respect to one another (Tarasov 2018).

Traits and diversification

We found that when considered alone, ploidy and breeding system each showed significant associations with net diversification differences. When considered together, however, the effect of breeding system dominated the effect of ploidy, although hidden factors played an important role as well.

235 For ploidy alone (D/P model), we found a greater net diversification rate for diploids than for polyploids, in agreement with (Mayrose et al. 2011, 2015). This result holds with (Fig. 2A) or without (Fig. 3A) the diploidization parameter, though including diploidization shifts the net diversification rate of polyploids to be non-negative. Incorporating a hidden state, however, removes the clear separation in diversification between diploids and polyploids (D/P+A/B model, Fig. 2B, Fig. 3B). Thus, differences in net diversification
240 are better explained by an unknown factor than by ploidy.

For breeding system alone, we found a larger net diversification rate for SI and for SC species, in agreement with (Goldberg et al. 2010) (I/C model, Fig. 2C). When a hidden state is added to the model (I/C+A/B), the large net diversification difference persists for one of the hidden states but is removed for the other (Fig. 2D). Thus, differences in net diversification are best explained by both breeding system and an
245 unknown factor. The transition rate from SI to SC is $q_{IC} = 0.3$ for both the I/C and the I/C+A/B models.

For ploidy and breeding system together (ID/CD/CP model), the net diversification rate for SI diploids (ID) was greater than for either SC diploids or SC polyploids, with or without diploidization (Fig. 2E, Fig. 3C). It thus appears that the difference in net diversification with breeding system persists when ploidy is included in the model, but not the reverse. The association of ploidy with net diversification
250 (Fig. 2A, Fig. 3A) appears to be driven by the subset of diploids that are SI; among SC species, net diversification rates for diploids and polyploids are similar. When a hidden state is included (IC/CD/CP+A/B model), the same general pattern remains when diploidization is excluded (Fig. 3D), although the higher net diversification rate of ID is less clear within one of the hidden states. With diploidization, the net diversification rate of ID is still greater than CD within each hidden state, but diversification for P is highly uncertain
255 and perhaps bimodal.

Model selection

In Table 1 we listed the marginal likelihood values calculated in log scale ($\log(P(\mathbf{X}|M))$) for each of the models tested. The different components included and excluded for each model as a summary of the diagrams from Fig. 2 are indicated in each model. From the marginal likelihoods in log scale, the Bayes
260 factors in $\log\text{-scale}(BF(M_0, M_1) = \log(P(\mathbf{X}|M_0) - P(\mathbf{X}|M_1)))$ were calculated as shown in table Table 2.

After testing every single pair of polyploidy models (1-4) we found overwhelming evidence that the best polyploidy model is always the D/P+A/B, that is the model with hidden state and diploidization, that is Bayes factors were always greater than 2 when D/P+A/B was the first model in the Bayes factor equation or negative when D/P+A/B was the second model in Bayes factor equation.

For the two models following the evolution of breeding system, the I/C+A/B is the best choice between models 3 and 4 (BF(IC, IC+A/B)=-39.5<0). The four models that follow the diversification linked to both polyploidy and breeding system (models 5-8) where compared in pairs. Bayes factors show decisive evidence in favor of the IC/P/CD+A/B over the rest, meaning that the model that has a hidden state and diploidization is chosen over the ones that lack either of both of those options Table 2. Therefore, the models that were chosen were always the ones containing the hidden trait, or diploidization parameter δ when ploidy state was considered.

Diploidization as an exploratory hypothesis

For the two models that only consider diploid and polyploid state (D/P and D/P+A/B) the effect of removing parameter δ was that net diversification rate of polyploids can be negative with high probability (Fig. 3AB). However, in the models that simultaneously considered polyploidy and breeding system, removing the δ parameter did not have much of an effect on the location of the posterior distribution of polyploid net diversification rate but on the credible interval width and overlap with net diversification of diploids. When diploidization was absent, the net diversification of polyploids overlaps with the net diversification of self-compatible diploids but not with the one of self-incompatible diploids (Fig. 3CD). For all models, the posterior distribution of diploidization rate is more uncertain than the one of polyploidization rates (see supplementary information figures).

Pathways to polyploidy

Polyploidization rates from self-compatible diploid ρ_{CD} and from self-incompatible diploid ρ_{ID} are not significantly different (see supplementary information figures). However, the distribution of ρ_{ID} seems to always consider faster values compare to the distribution of ρ_{CD} . This result is highlighted in ancestral reconstructions of the ID/CD/CP model where transitions from ID to CP are more probable than transitions from CD to CP.

Discussion

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Literature Cited

- Barrett, S. C., 1988. The evolution, maintenance, and loss of self-incompatibility systems, vol. Plant reproductive ecology: patterns and strategies, Pp. 98–124. Oxford University Press Oxford.
- 295 ———, 2008. Major evolutionary transitions in flowering plant reproduction: an overview.
- , 2013. The evolution of plant reproductive systems: how often are transitions irreversible? Proceedings of the Royal Society B: Biological Sciences 280:20130913.
- Barringer, B. C., 2007. Polyploidy and self-fertilization in flowering plants. American Journal of Botany 94:1527–1533.
- 300 Beaulieu, J. M. and B. C. O’Meara, 2016. Detecting hidden diversification shifts in models of trait-dependent speciation and extinction. Syst Biol 65:583–601.
- Bennett, M. D. and I. J. Leitch, 2005. Plant DNA C-values database.
- Caetano, D. S., B. C. O’Meara, and J. M. Beaulieu, 2018. Hidden state models improve state-dependent diversification approaches, including biogeographical models. Evolution 72:2308–2324.
- 305 Camadro, E. and S. J. Peloquin, 1981. Cross-incompatibility between two sympatric polyploid *Solanum* species. Theoretical and Applied Genetics 60:65–70.
- Entani, T., S. Takayama, M. Iwano, H. Shiba, F.-S. Che, and A. Isogai, 1999. Relationship between polyploidy and pollen self-incompatibility phenotype in *Petunia hybrida* Vilm. Bioscience, Biotechnology, and Agrochemistry 63:1882–1888.
- 310 FitzJohn, R. G., 2012. Diversitree : comparative phylogenetic analyses of diversification in r. Methods Ecol Evol 3:1084–1092.
- FitzJohn, R. G., W. P. Maddison, and S. P. Otto, 2009. Estimating trait-dependent speciation and extinction rates from incompletely resolved phylogenies. Systematic Biology 58:595–611.
- Goldberg, E. E. and B. Igić, 2012. Tempo and mode in plant breeding system evolution. Evolution 66:3701–3709.
- 315 Goldberg, E. E., J. R. Kohn, R. Lande, K. A. Robertson, S. A. Smith, and B. Igić, 2010. Species selection maintains self-incompatibility. Science 330:493–495.
- Herrera-Alsina, L., P. van Els, and R. S. Etienne, 2018. Detecting the dependence of diversification on multiple traits from phylogenetic trees and trait data. Systematic biology .
- 320 Höhna, S., M. J. Landis, T. A. Heath, B. Boussau, N. Lartillot, B. R. Moore, J. P. Huelsenbeck, and F. Ronquist, 2016. Revbayes: Bayesian phylogenetic inference using graphical models and an interactive model-specification language. Systematic Biology 65:726–736.
- Huang, D., E. E. Goldberg, L. Chou, and K. Roy, 2018. The origin and evolution of coral species richness in a marine biodiversity hotspot. Evolution 72:288–302.
- 325 Husband, B. C., B. Ozimec, S. L. Martin, and L. Pollock, 2008. Mating consequences of polyploid evolution in flowering plants: Current trends and insights from synthetic polyploids. International Journal of Plant Sciences 169:195–206.
- Igić, B., L. Bohs, and J. R. Kohn, 2006. Ancient polymorphism reveals unidirectional breeding system shifts. Proc Natl Acad Sci USA 103:1359–1363.
- 330 Iogerger, T. R., A. G. Clark, and T. H. Kao, 1990. Polymorphism at the self-incompatibility locus in Solanaceae predates speciation. Proceedings of the National Academy of Sciences USA 87:9732–9735.
- Kubo, K.-i., T. Entani, A. Takara, N. Wang, A. M. Fields, Z. Hua, M. Toyoda, S.-i. Kawashima, T. Ando, A. Isogai, et al., 2010. Collaborative non-self recognition system in s-rnase-based self-incompatibility. Science 330:796–799.
- 335 Landis, J. B., D. E. Soltis, Z. Li, H. E. Marx, M. S. Barker, D. C. Tank, and P. S. Soltis, 2018. Impact of whole-genome duplication events on diversification rates in angiosperms. American Journal of Botany 105:348–363.

Lewis, D., 1947. Competition and dominance of incompatibility alleles in diploid pollen. *Heredity* 1:85–108.

340 Lynch, M. and J. S. Conery, 2000. The evolutionary fate and consequence of duplicate genes. *Science* 290:1151–1155.

Maddison, W. P. and R. G. FitzJohn, 2015. The unsolved challenge to phylogenetic correlation tests for categorical characters. *Systematic Biology* 64:127–136.

Maddison, W. P., P. E. Midford, and S. P. Otto, 2007. Estimating a binary character's effect on speciation
345 and extinction. *Syst Biol* 56:701–710.

Mayrose, I., M. S. Barker, and S. P. Otto, 2010. Probabilistic models of chromosome number evolution and the inference of polyploidy. *Systematic Biology* 59:132–144.

Mayrose, I., S. H. Zhan, C. J. Rothfels, N. Arrigo, M. S. Barker, L. H. Rieseberg, and S. P. Otto, 2015. Methods for studying polyploid diversification and the dead end hypothesis: a reply to soltis et al. (2014).
350 *New Phytol* 206:27–35.

Mayrose, I., S. H. Zhan, C. J. Rothfels, K. Magnuson-Ford, M. S. Barker, L. H. Rieseberg, and S. P. Otto, 2011. Recently formed polyploid plants diversify at lower rates. *Science* 333:1257.

Olmstead, R. G. and L. Bohs, 2007. A summary of molecular systematic research in Solanaceae: 1982–2006. *Acta Horticulturae* Pp. 255–268.

355 PBI *Solanum* Project, 2012. Solanaceae Source: a global taxonomic resource for the nightshade family.

Rabosky, D. L. and E. E. Goldberg, 2015. Model inadequacy and mistaken inferences of trait-dependent speciation. *Systematic Biology* 64:340–355.

Raduski, A. R., E. B. Haney, and B. Igić, 2012. The expression of self-incompatibility in angiosperms is bimodal. *Evolution* 66:1275–1283.

360 Ramanauskas, K. and B. Igić, 2017. The evolutionary history of plant t2/s-type ribonucleases. *PeerJ* 5:e3790.

Ramsey, J. and D. W. Schemske, 1998. Pathways, mechanisms, and rates of polyploid formation in flowering plants. *Annual Review of Ecology and Systematics* 29:467–501.

Rice, A., L. Glick, S. Abadi, M. Einhorn, N. M. Kopelman, A. Salman-Minkov, J. Mayzel, O. Chay, and
365 I. Mayrose, 2015. The chromosome counts database (CCDB) - a community resource of plant chromosome numbers. *New Phytol* 206:19–26.

Rivero, R., E. B. Sessa, and R. Zenil-Ferguson, 2019. Eyechrom and ccdb curator: Visualizing chromosome count data from plants. *Applications in Plant Sciences* P. e01207.

Robertson, K., E. E. Goldberg, and B. Igić, 2011. Comparative evidence for the correlated evolution of
370 polyploidy and self-compatibility in solanaceae. *Evolution* 65:139–155.

Särkinen, T., L. Bohs, R. G. Olmstead, and S. Knapp, 2013. A phylogenetic framework for evolutionary study of the nightshades (Solanaceae): a dated 1000-tip tree. *BMC Evol Biol* 13:214.

Servedio, M. R., Y. Brandvain, S. Dhole, C. L. Fitzpatrick, E. E. Goldberg, C. A. Stern, J. Van Cleve, and
375 D. J. Yeh, 2014. Not just a theory?the utility of mathematical models in evolutionary biology. *PLoS biology* 12:e1002017.

Soltis, D. E., M. C. Segovia-Salcedo, I. Jordon-Thaden, L. Majure, N. M. Miles, E. V. Mavrodiev, W. Mei, M. B. Cortez, P. S. Soltis, and M. A. Gitzendanner, 2014. Are polyploids really evolutionary dead-ends (again)? a critical reappraisal of mayrose et al.(2011). *New Phytologist* 202:1105–1117.

380 Stebbins, G. L., 1938. Cytological characteristics associated with the different growth habits in the dicotyledons. *American Journal of Botany* 25:189–198.

———, 1950. *Variation and Evolution in Plants*. Columbia University Press, New York.

———, 1974. *Flowering plants: evolution above the species level*. Belknap Press of Harvard University Press, Cambridge, Mass.

Stone, J. L., 2002. Molecular mechanisms underlying the breakdown of gametophytic self-incompatibility.
The Quarterly Review of Biology 77:17–30.

Stout, A. B. and C. Chandler, 1942. Hereditary transmission of induced tetraploidy and compatibility in
fertilization. Science 96:257–258.

Tarasov, S., 2018. Integration of anatomy ontologies and evo-devo using structured markov models suggests
a new framework for modeling discrete phenotypic traits. BioRxiv P. 188672.

Tsukamoto, T., T. Ando, H. Watanabe, E. Marchesi, and T.-h. Kao, 2005. Duplication of the s-locus f-box
gene is associated with breakdown of pollen function in an s-haplotype identified in a natural population
of self-incompatible petunia axillaris. Plant Molecular Biology 57:141–153.

Vision, T. J., D. G. Brown, and S. D. Tanksley, 2000. The origins of genomic duplications in *Arabidopsis*.
Science 290:2114–2117.

de Vos, J. M., C. E. Hughes, G. M. Schneeweiss, B. R. Moore, and E. Conti, 2014. Heterostyly acceler-
ates diversification via reduced extinction in primroses. Proceedings of the Royal Society B: Biological
Sciences 281:20140075.

Xie, W., P. O. Lewis, Y. Fan, L. Kuo, and M.-H. Chen, 2010. Improving marginal likelihood estimation for
bayesian phylogenetic model selection. Systematic biology 60:150–160.

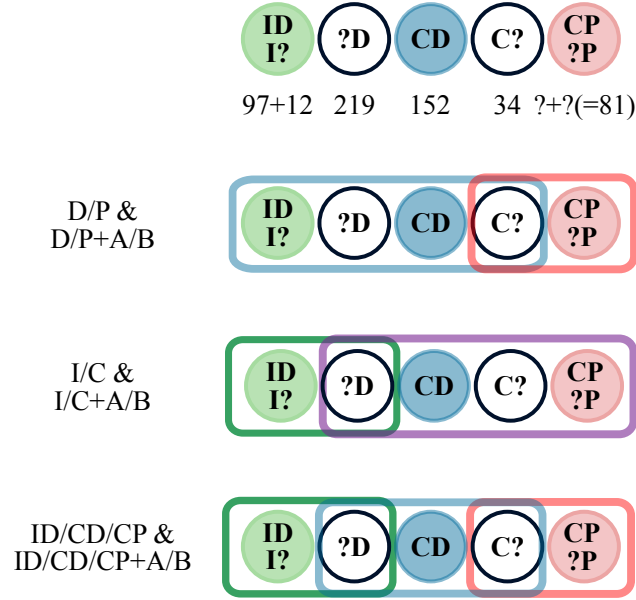


Figure 1: Character states used for each of the models. Each species retained on the tree belonged to one of five possible categories, depending on whether ploidy and/or breeding system were known. The number of species in each is shown under the corresponding circles in the top row. These categories were then grouped in a manner appropriate to the states of each model. For example, there are 34 species that are self-compatible and of unknown ploidy; these are coded as either *D* or *P* in the D/P models (uncertain, or consistent with either state), as *C* in the I/C models, and as either *CD* or *CP* in the ID/CD/CP models. In all cases, species were coded as either *A* or *B* in the hidden state models.

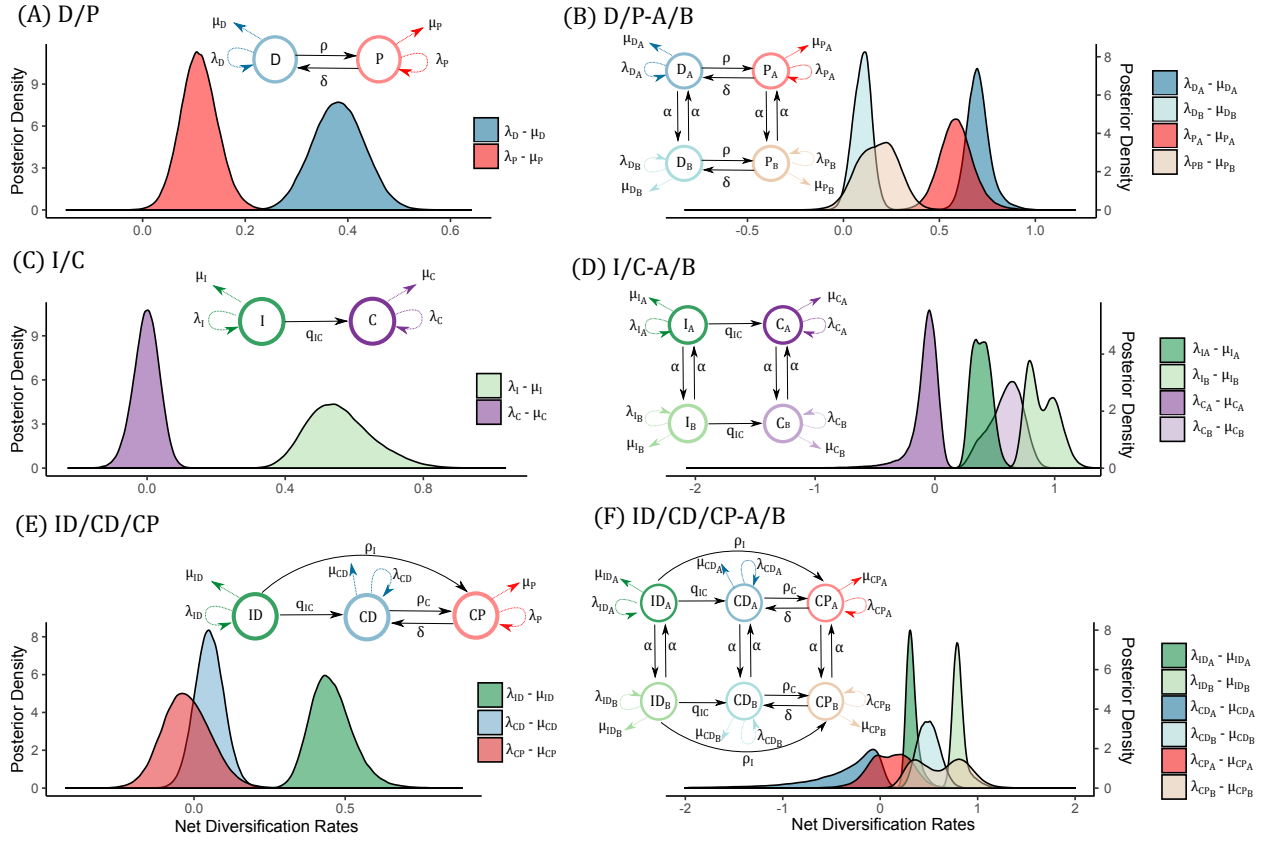


Figure 2: Net diversification rates for all models that include diploidization.

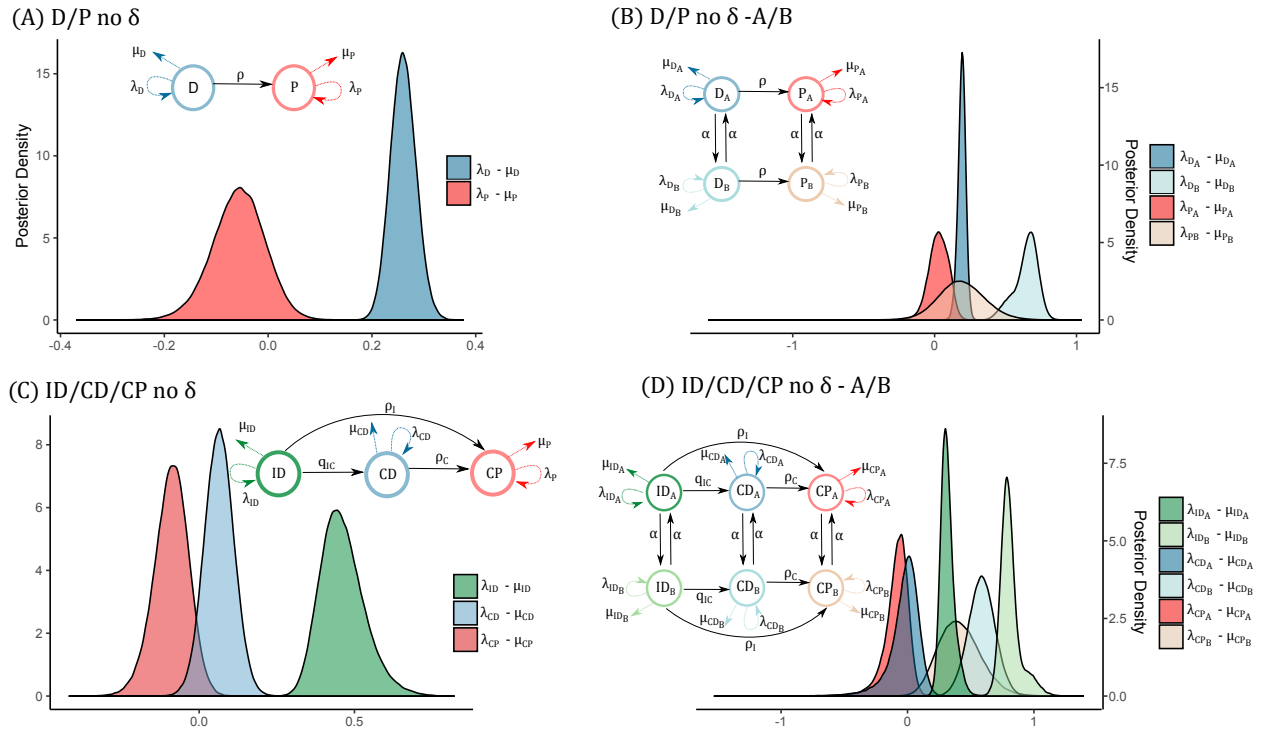


Figure 3: Net diversification rates for all models that do not include diploidization.

Model	Model Type	Ploidy	Diploidization	Breeding System	Hidden State	Parameters	Marginal Log- Likelihood
1. D/P	BiSSE	Yes	Yes	No	No	6	-1182.93
2. D/P no δ	BiSSE	Yes	No	No	No	5	-1193.66
3. D/P- A/B	HiSSE	Yes	Yes	No	Yes	11	-1145.69
4. D/P no δ -A/B	HiSSE	Yes	No	No	Yes	10	-1150.99
5. I/C	BiSSE	No	No	Yes	No	5	-1194.80
6. I/C-A/B	HiSSE	No	No	Yes	Yes	10	-1155.37
7. ID/P/CD	MuSSE	Yes	Yes	Yes	No	10	-1344.50
8. ID/P/CD no δ	MuSSE	Yes	No	Yes	No	9	-1345.87
9. ID/P/CD-A/B	MuHiSSE	Yes	Yes	Yes	Yes	16	-1300.35
10.ID/P/CD no δ -A/B	MuHiSSE	Yes	No	Yes	Yes	15	-1303.55

Table 1: Marginal likelihoods for the ten models examined. States encoded with letter D: diploid, P: polyploid, C: self-compatible, I: self-incompatible, A: first state of hidden trait, B: second state of hidden trait, δ : re-diploidization.

Ploidy Models					Breeding System Models		Ploidy and Breeding System Models					
	1	2	3	4		5	6		7	8	9	10
1. D/P	·	10.72	-37.24	-31.94	5. I/C	·	-39.43	7. ID/P/CD	·	1.36	-44.15	-40.95
2. D/P no δ	·	·	-47.97	-42.66	6. I/C-A/B	·	·	8. ID/P/CD no δ	·	·	-45.51	-42.31
3. D/P- A/B	·	·	·	5.30				9. ID/P/CD-A/B	·	·	·	3.2
4. D/P no δ -A/B	·	·	·	·				10. ID/P/CD no δ -A/B	·	·	·	·

Table 2: Bayes factors in log scale. We compare every possible pair. Number models as indicated in Table 1.

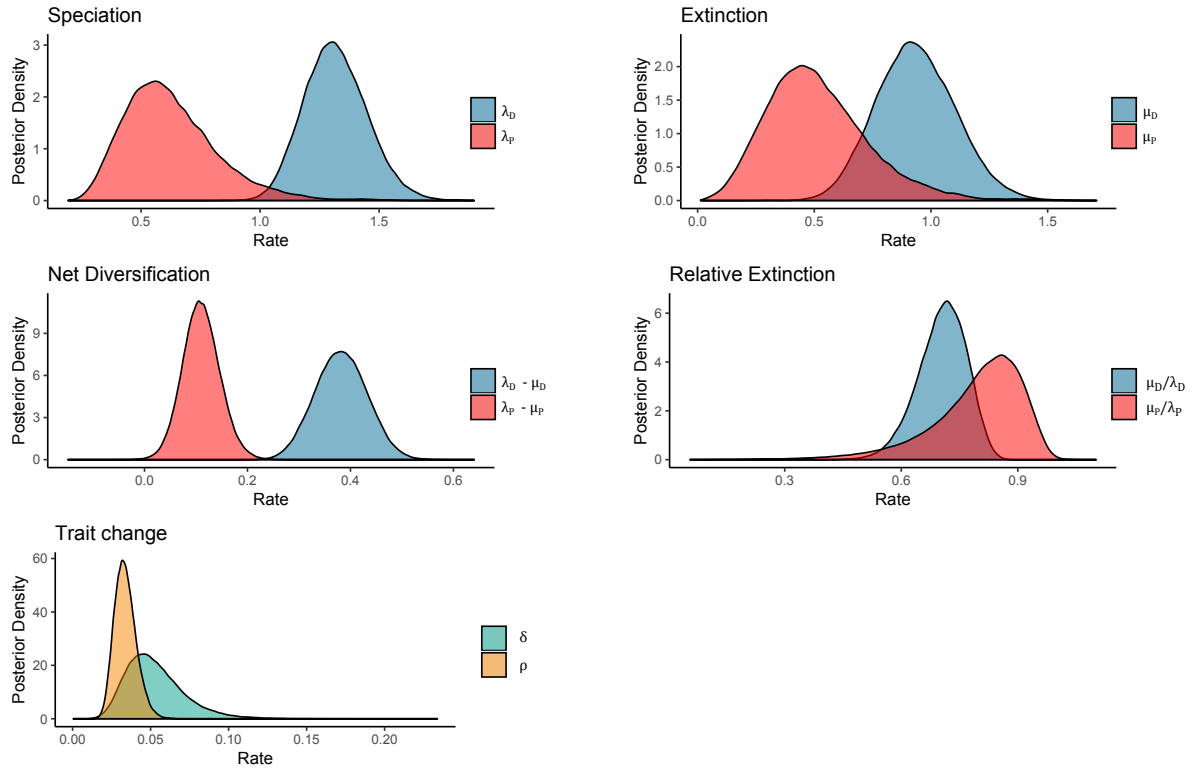


Figure S1: Posterior distribution for each of the parameters in the D/P polyploidy model

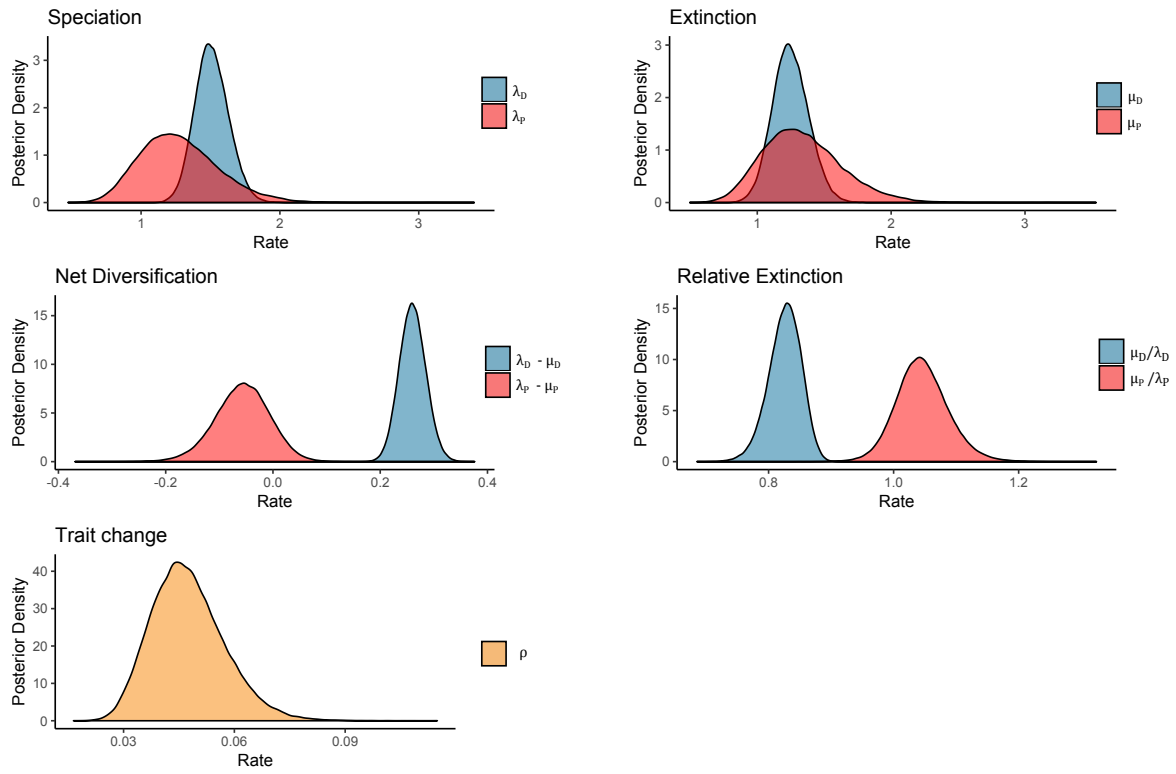


Figure S2: Posterior distribution for each of the parameters in the D/P no δ polyploidy model

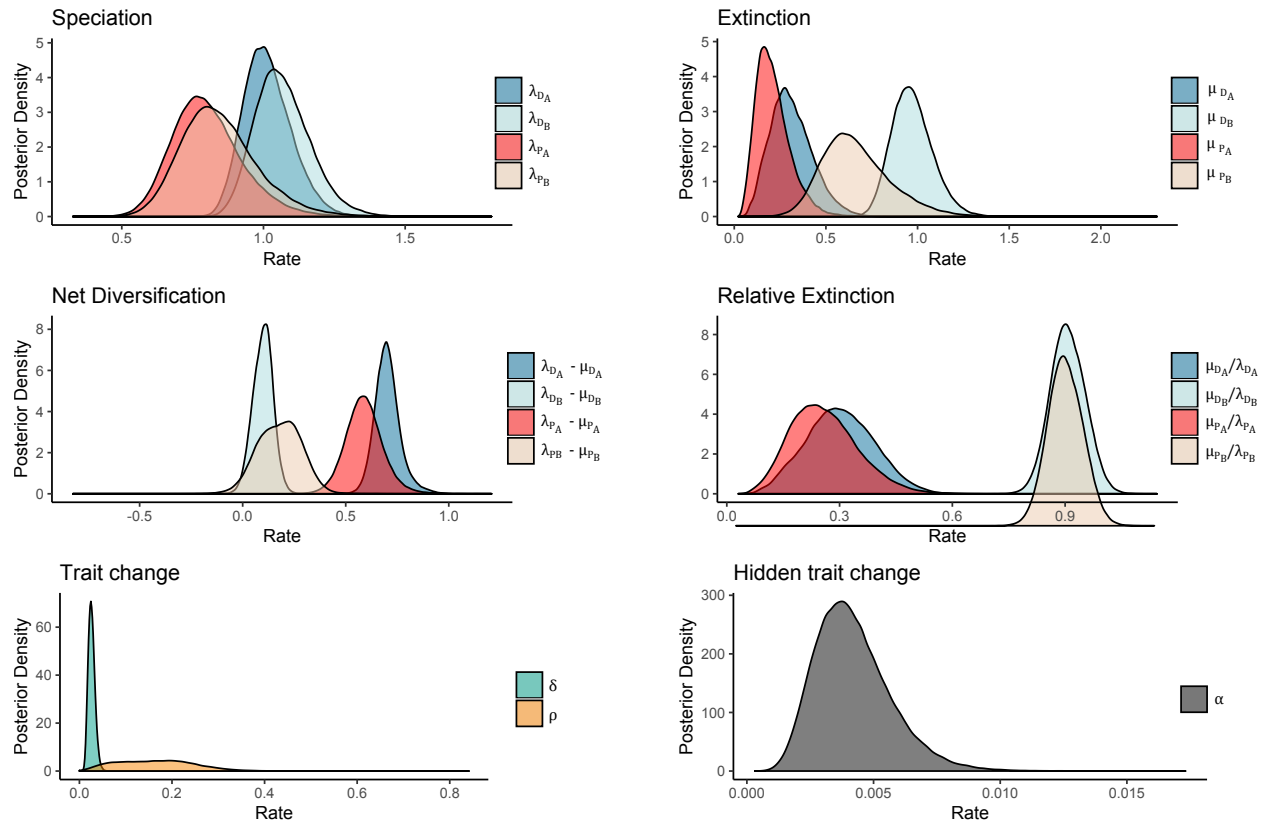


Figure S3: Posterior distribution for each of the parameters in the D/P-A/B polyploidy model

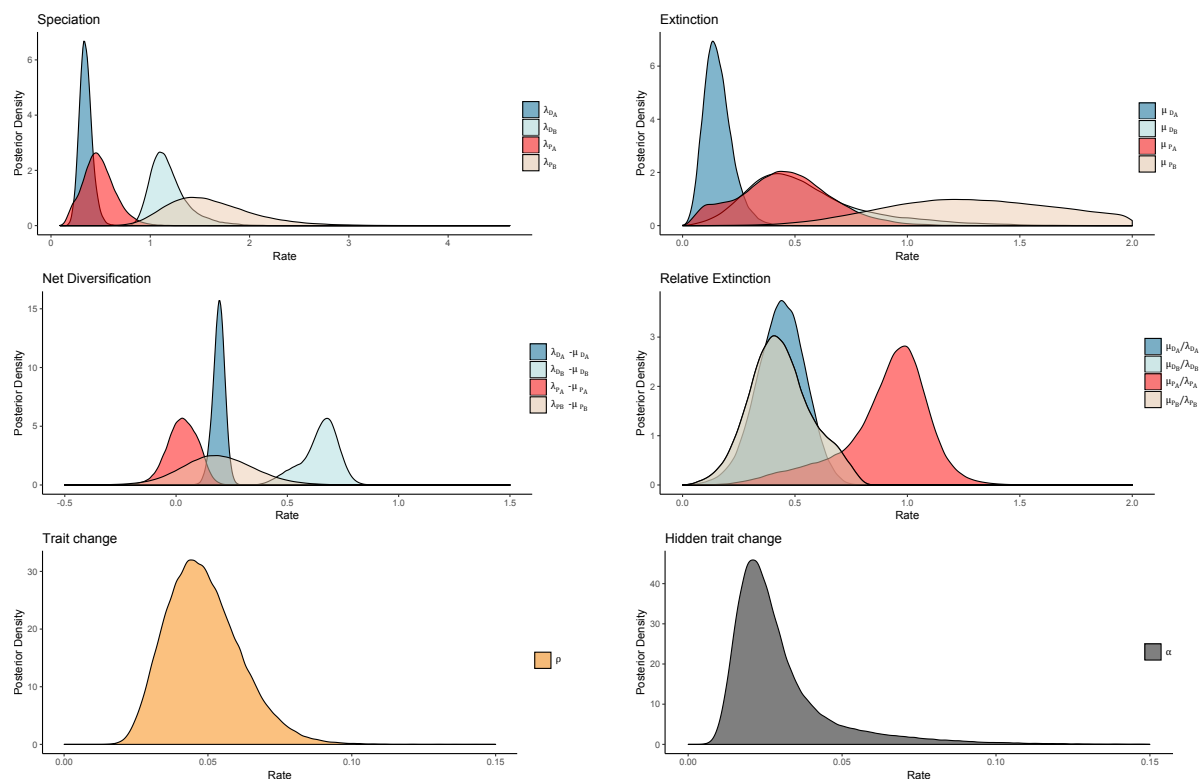


Figure S4: Posterior distribution for each of the parameters in the D/P no δ -A/B polyploidy model

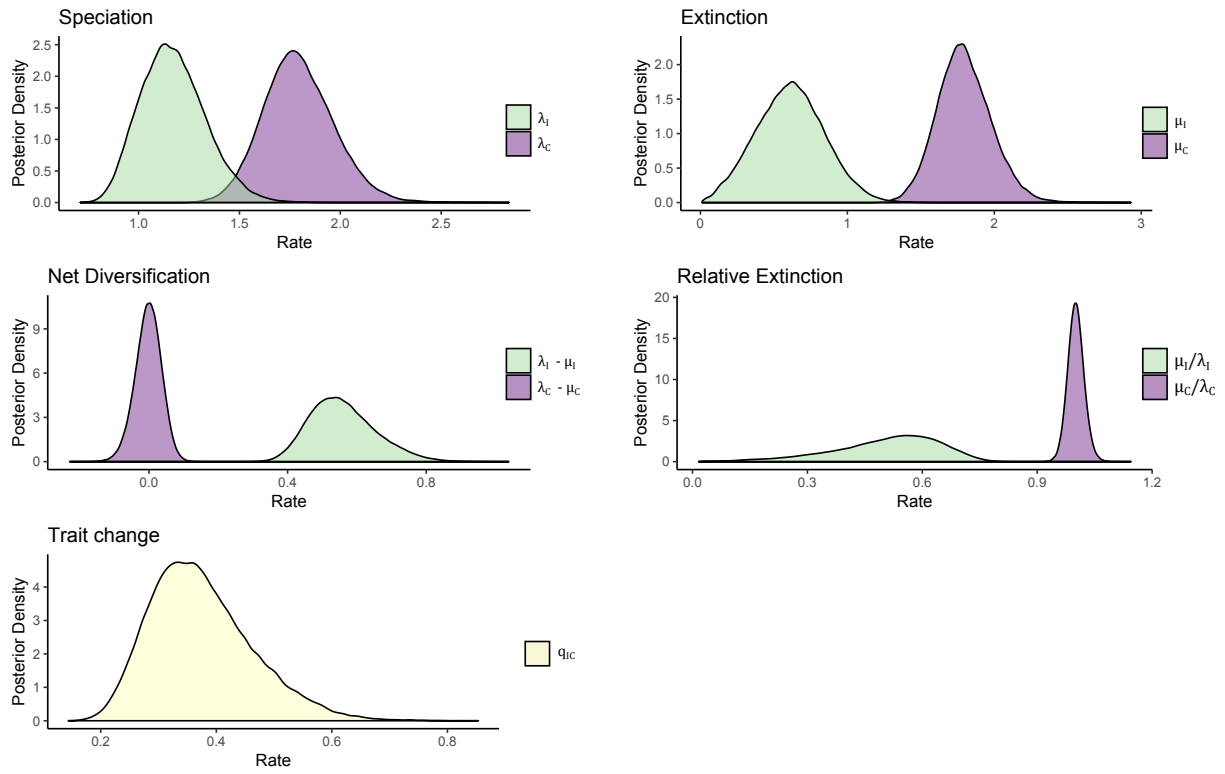


Figure S5: Posterior distribution for each of the parameters in the I/C breeding system model

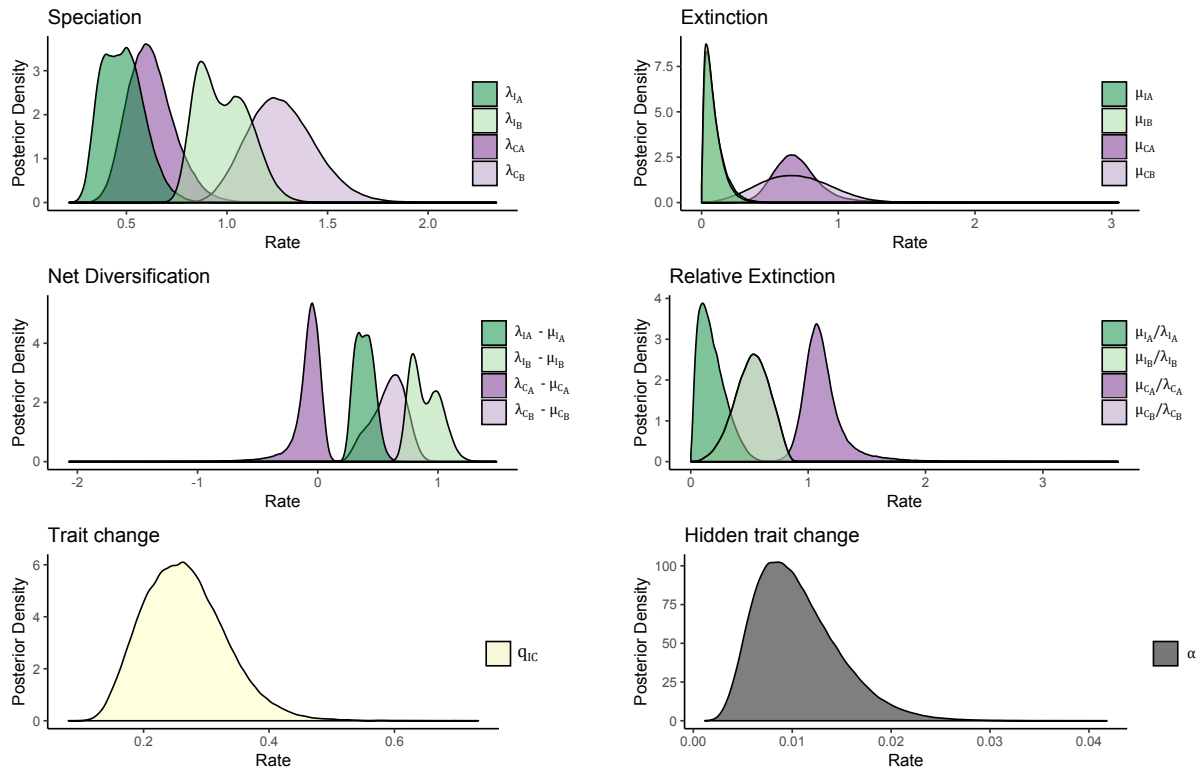


Figure S6: Posterior distribution for each of the parameters in the I/C-A/B breeding system model

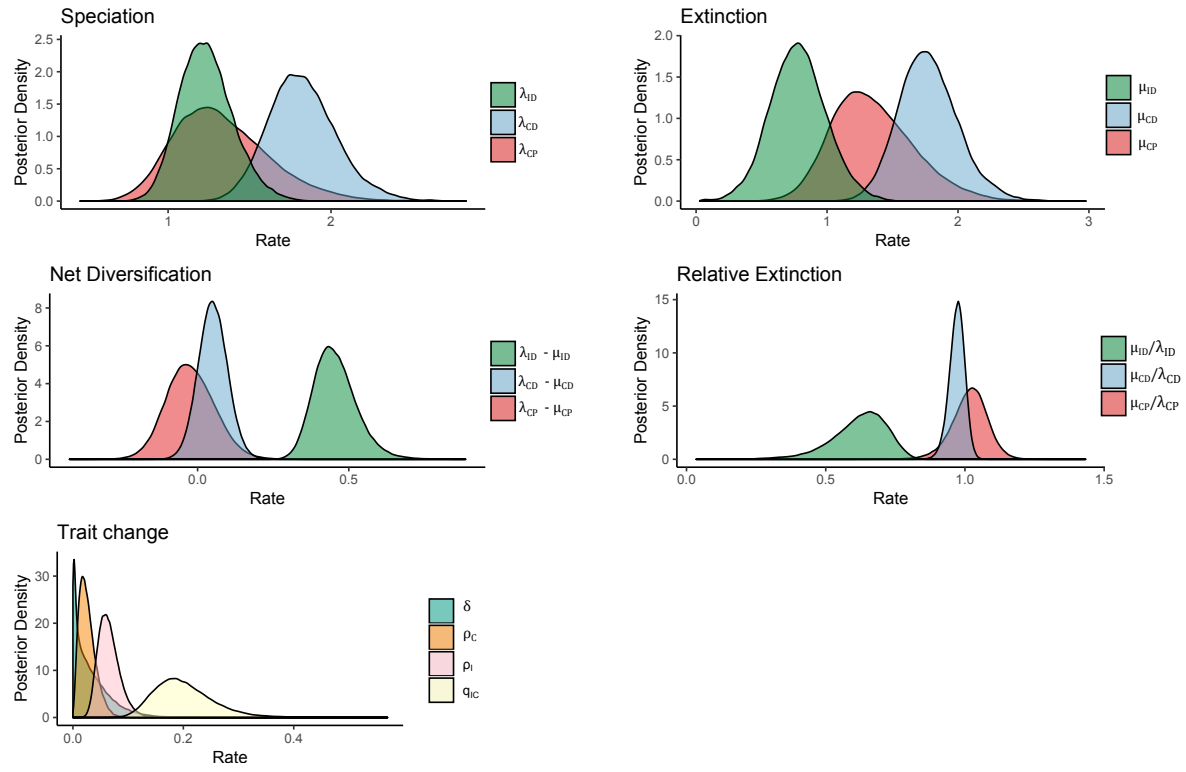


Figure S7: Posterior distribution for each of the parameters in the ID/CD/CP polyploidy and breeding system model

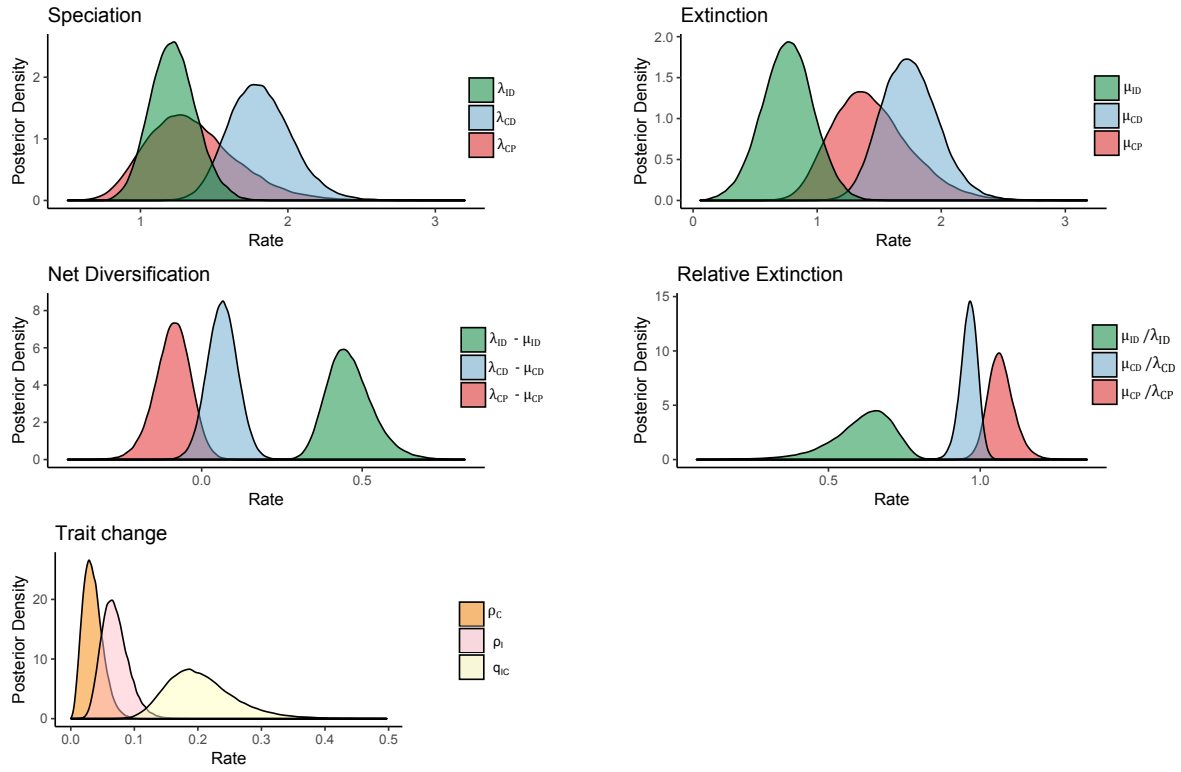


Figure S8: Posterior distribution for each of the parameters in the ID/CD/CP no δ polyploidy and breeding system model

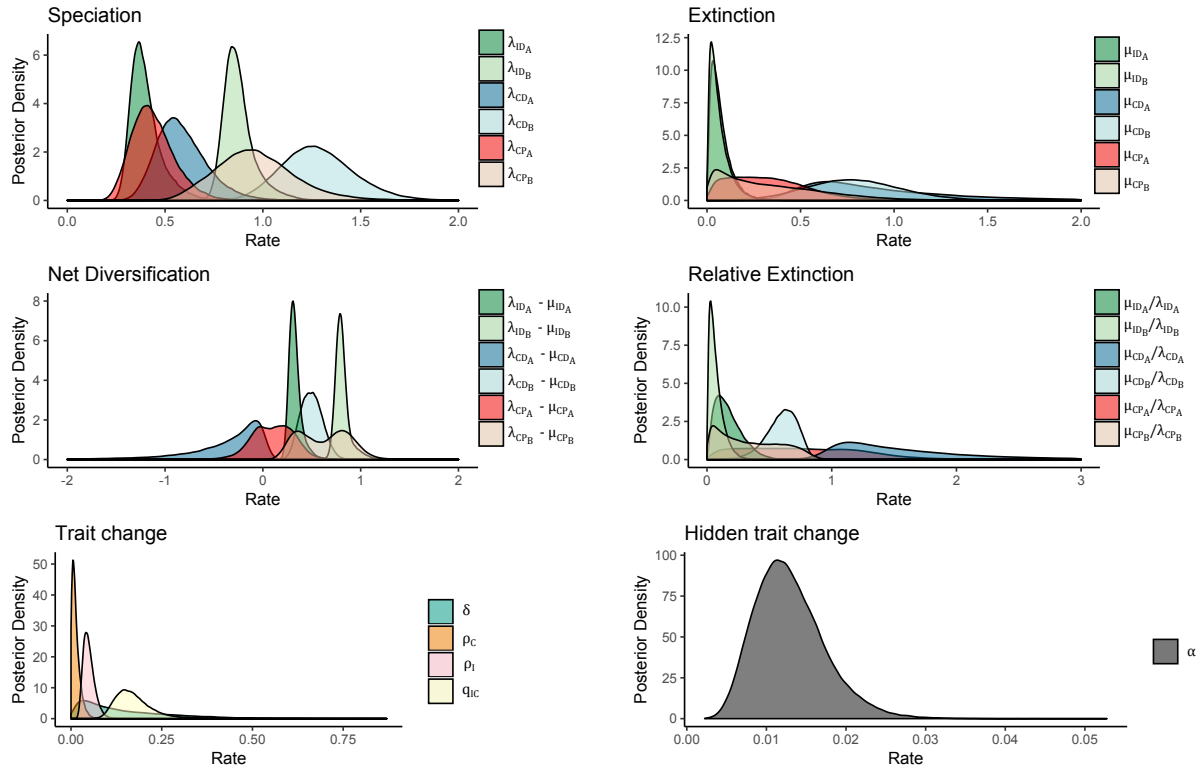


Figure S9: Posterior distribution for each of the parameters in the ID/CD/CP-A/B polyploidy and breeding system model

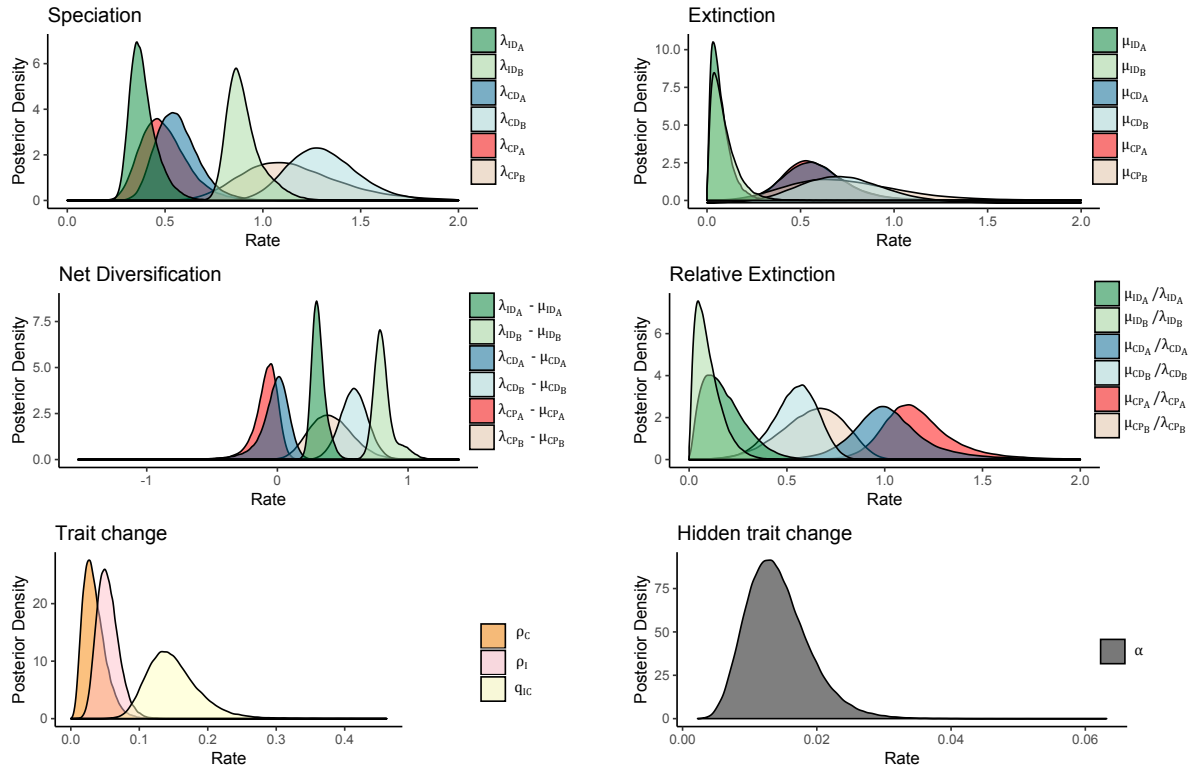


Figure S10: Posterior distribution for each of the parameters in the ID/CD/CP no δ - A/B polyploidy and breeding system model

	D/P	D/P+A/B	I/C	I/C+A/B	ID/CD/CP	ID/CD/CP+A/B
r_D	0.382	0.698, 0.100	—	—	—	—
r_P	0.109	0.587, 0.182	—	—	—	—
r_I	—	—	0.550	0.386, 0.877	—	—
r_C	—	—	-0.001	-0.059, 0.606	—	—
r_{ID}	—	—	—	—	0.449	0.318, 0.789
r_{CD}	—	—	—	—	0.050	-0.248, 0.494
r_{CP}	—	—	—	—	-0.027	0.110, 0.634
ρ	0.033	0.026	—	—	—	—
ρ_I	—	—	—	—	0.063	0.047
ρ_C	—	—	—	—	0.024	0.011
δ	0.050	0.162	—	—	0.022	0.107
q_{IC}	—	—	0.364	—	0.194	0.164

	D/P	D/P+A/B	ID/CD/CP	ID/CD/CP+A/B
r_D	0.260	0.193, 0.658	—	—
r_P	-0.056	0.030, 0.187	—	—
r_I	—	—	—	—
r_C	—	—	—	—
r_{ID}	—	—	0.455	0.309, 0.797
r_{CD}	—	—	0.065	-0.006, 0.587
r_{CP}	—	—	-0.088	-0.074, 0.403
ρ	0.047	0.047	—	—
ρ_I	—	—	0.067	0.053
ρ_C	—	—	0.033	0.032
q_{IC}	—	—	0.198	0.145

Table S1: Median rate estimates for all models. Units are per million years. Two comma-separated numbers refer to the A and B hidden states, and — means the parameter was not present in the model. The upper section is for models with diploidization, and the lower section is for models without. The supplemental figures show the corresponding distributions of parameter estimates.