When your favorite trait is not enough to explain diversification: An example of diversification linked to polyploidy and breeding system in Solanaceae

Rosana Zenil-Ferguson, ¹ J. Gordon Burleigh, ² William A. Freyman, ³ Boris Igić, ⁴ Itay Mayrose, ⁵ and Emma E. Goldberg ⁶
¹ University of Minnesota Twin Cities
² University of Florida
³ 23 and me
⁴ University of Illinois
⁵ Tel Aviv University
⁶ University of Minnesota Twin Cities
Author for correspondence: Rosana Zenil-Ferguson
Running head: Polyploidy and breeding systems Solanaceae
Keywords: Polyploidy, Breeding System, Diversification, SSE models

Abstract

Abstract text...

Introduction

Linking changes in diversification to polyploidy and breeding system has been one of the key botany questions for the past n number of years. The prevalence of polyploidy, and the multiple evolutionary changes of breeding systems in important angiosperm clades, leads to the inevitable question of the evolutionary changes.

- Diversification and polyploidy

An important debate regarding the diversification of angiosperms has been ongoing since the publication of Mayrose et al. (2011). The authors discovered that using the latest diversification model linked to two states diploid and polyploid, the net diversification of polyploids was much slower than the net diversification rate than polyploids. This result was surprising and an sparked a discussion about the long-term evolutionary consequences of polyploidy. Soltis et al. (2014) questioned if polyploidy should be regarded as an evolutionary dead-end since the net diversification of polyploids was negative, despite overwhelming evidence about the incidence of polyploidy especially at the root of highly diverse angiosperm claims (ref here). A year later, diversification models were re-tested and corrected, and still found the same pattern (Mayrose et al. 2015), to the disappointment of plenty of botanists there was no denying of the weak trend of diversification that polyploids left behind.

There are two key questions that at the time of the polyploidy debate were difficult to ask. The first is if the models used to measure the diversification of polyploids were correct, and the second question is if the models have potentially included more evidence and potential traits that are not polyploidy or other lines of evidence that could be driving the patterns. At the time, in a different context Beaulieu and O'Meara (2016) were finding an alternative solution to the first question, coming up with a new model that could represent the broad heterogeneity of the diversification process and parse out the signal between the trait of interest and the noise in diversification. Their model, the hidden state speciation and extinction model is a key component to detect whether polyploidy our something else unknown but related to it is driving the speciation and extinction patterns that we see in angiosperms.

The second question (talk here about breeding system and polyploidy, and how this could be different across clades and this is one of the reasons why we focused on Solanaceae). -Diversification and breeding systems

Goldberg and Igic 2012, different perspectives

-What we did

We investigate the relative importance of both b

Debate of Mayrose

Methods

Data

50

55

Chromosome number data were obtained for all Solanaceae taxa in the the Chromosome Counts Database (CCDB; Rice et al. 2015), and the ca. 14,000 records were cleaned semi-automatically using the CCDBcurator R package (Zenil-Ferguson et al. 2017). This large dataset includes the compilation of Solanaceae ploidy states from (Robertson et al. 2011). Species were coded as either diploid (D) or polyploid (P). For the majority of species, ploidy was assigned according to information from the original publications and the Kew Royal Botanic Gardens C-value DNA resource (Bennett and Leitch 2005). For taxa without ploidy information but with information about chromosome number, we assigned ploidy based on the multiplicity of chromosomes within the genus. For example, *Solanum betaceum* did not include information about ploidy level but it has 24 chromosomes, so because x = 12 is the base chromosome number of the *Solanum* genus (Olmstead and Bohs 2007), we assigned *S. betaceum* as diploid. Species with more than one ploidy level were assigned the smallest most frequent ploidy level recorded.

Breeding system was scored as self-incompatible (SI) or self-compatible (SC) based on results hand-curated from the literature (as in Igić et al. 2006; Goldberg et al. 2010; Robertson et al. 2011; Goldberg and Igić 2012). Most species could unambiguously be coded as either SI or SC (Raduski et al. 2012). Following previous work, we coded as SI any species with functional SI systems, even if SC or dioecy was also reported. Dioecious species without functional SI were coded as SC.

To those existing data sets, we added some additional records for chromosome number and breeding system. The Supplementary Information contains citations for the numerous original sources for all of the data. Synonymy followed Solanaceae Source (PBI *Solanum* Project 2012). Hybrids and cultivars were excluded because ploidy and breeding system are likely to be altered by domestication. As in Robertson et al. (2011), we examined closely the few species for which the merged ploidy and breeding system data indicated the presence of SI polyploids. Although SI populations frequently contain some SC individuals and diploid populations frequently contain some polyploid individuals, in no case did we find individuals that were both SI and polyploid. Because of this empirical observation and the functional explanation for whole genome duplication disabling gametophytic self-incompatibility (reviewed in Ramsey and Schemske 1998; Stone 2002), we consider only three observed character states: SI-D, SC-D, and SC-P.

Matching our character state data to the largest time-calibrated phylogeny of Solanaceae (Särkinen et al. 2013) yielded 595 species with ploidy and/or breeding system information on the tree. Binary or three-state classification of ploidy and breeding system for the 595 taxa is summarized in Table 1. We retained all of these species in each of the analyses below, because pruning away tips lacking breeding system in the ploidy-only analyses (and vice versa) would discard data that could inform the diversification models.

Models

65

We first defined a binary state speciation and extinction model (BiSSE, Maddison et al. (2007)) for polyploidy evolution where taxa was classified as diploid (D=0) and polyploid (P=1) Table 1. In a Bayesian framework, we obtained posterior probability distributions of speciation rates (λ_D , λ_P), extinction rates (μ_D , μ_P), net diversification rates ($r_D = \lambda_D - \mu_D$, $r_P = \lambda_P - \mu_P$), and relative extinction rates ($v_D = \frac{\mu_D}{\lambda_D}$, $v_D = \frac{\mu_D}{\lambda_D}$) just as previously explored in Mayrose et al. (2011). The cladogenetic changes explored in BiSSE were polyploidization rate are represented by parameter ρ and by diploidization parameter rate δ .

As a second step, we fitted a hidden state speciation and extinction model (HiSSE, Beaulieu and O'Meara (2016)) to evaluate whether the differences in diversification rates were found due to a hidden trait associated to polyploidy. As discussed by Beaulieu and O'Meara (2016) BiSSE-like models suffer from a large type I error because they fail to account as part of the null hypothesis heterogeneous diversification rate changes that do not depend on the trait of interest. By including a hidden trait in a state speciation and extinction model, HiSSE-like models address the heterogeneous background diversification changes while also parsing out the possible signal of diversification due to the trait of interest. Therefore, HiSSE model for polyploidy linked to diversification has four states: diploid and polyploid subdivided by a binary hidden trait with states A and B for which we estimated the posterior probability distributions of speciation rates (λ_{D_A} , λ_{D_B} , λ_{P_A} , λ_{P_B}), extinction rates (μ_{D_A} , μ_{D_B} , μ_{P_A} , μ_{P_B}), net diversification rates (μ_{D_A} , μ_{D_B} , μ_{P_A} , μ_{P_B}), and relative extinction(ν_{D_A} , ν_{D_B} , ν_{P_A} , ν_{P_B}). The cladogenetic changes assumed by the fitted model are polyploidization rate ρ , diploidization rate ρ , and we assumed that the changes between every hidden state are symmetrical with rate α .

For breeding system both BiSSE and a HiSSE models were fitted independently by coding data as self-compatible (SC=0) or self-incompatible (SI=1) Table 1. The BiSSE model fitted to breeding system data was done with the goal of investigating if the effect of self-incompatibility in diversification was similar to the pattern found by Goldberg and Igić (2012). However, the HiSSE model on breeding system parses out if the effect of breeding system in diversification is significant by minimizing type I error as mentioned above. For these models we assumed that self-compatibility is irreversible as discussed by Igić and Busch (2013),

so we defined the transition rate q_{IC} as the parameter for changes from self-incompatible to self-compatible state.

Next, we proposed a multivariate speciation and extinction model (MuSSE, FitzJohn (2012)) to investigate the link amongst diversification, breeding system, and polyploidy simultaneously. The MuSSE model is defined using three states self-compatible diploids (SC-D=0), polyploids that are always self-compatible (ref for this??) (P=1), and self-incompatible diploids (SI-D=2, see table Table 1) and contains ten parameters, six defining diversification (λ_{SD} , μ_{SD} , λ_P , μ_P , λ_{ID} , μ_{ID}) and the other four key cladogenetic changes that are: polyploidization of self-compatible diploids ρ_{SD} , diploidization δ , polyploidization of self-incompatible diploids ρ_{ID} , and self-incompatible to self-compatible rate q_{IC} .

100

105

115

Since the null hypothesis of the MuSSE model is that the diversification is equal and constant for all three states defined, it is possible that MuSSE can also suffer from large type I errors. In order to account for heterogeneity of the diversification rates and parse the signal in diversification coming from breeding system and polyploidy we extended MuSSE model to account for a hidden state, as Beaulieu and O'Meara (2016) did in the bivariate case. The model we proposed is a multivariate and hidden states speciation and extinction stochastic process (MuHiSSE) that allowed us to account simultaneously for the diversification rates linked to breeding system and polyploidy but also the presence of some more heterogeneity in the process. This model is analogous to the GeoHiSSE model proposed by Caetano et al. (2018) in a biogeographical context. The full MuHiSSE model has 26 parameters, however, our goal was to look for diversification rate differences so we fitted a simplified version of 17 parameters by fixing the rates amongst hidden states to be equal with parameter α and the transition rates amongst breeding system and polyploidy as defined in the MuSSE model (ρ_{SD} , δ , ρ_{ID} , q_{IC}) despite the hidden state. Using MuHiSSE we estimated twelve speciation and extinction rates (λ_{SD_A} , μ_{SD_A} , λ_{P_A} , μ_{P_A} , λ_{ID_A} , μ_{ID_B} , λ_{SD_B} , μ_{P_B} , λ_{ID_B} , μ_{ID_B}), and the net diversification and relative extinction rates associated with them.

All the models were performed using RevBayes (Höhna et al. 2016) software that performs Bayesian inference via MCMC using Metropolis-Hastings algorithm. A correction for sampling bias was done in all models by assuming that Solanaceae family has approximately 3,000 species (s = 595/3000) as the Solanaceae Source project indicates (PBI *Solanum* Project 2012). In all models, speciation and extinction parameters used log-normal prior distributions that with mean the expected net diversification rate ($\frac{(numberoftaxa)/2}{rootage}$) and standard deviation 0.5. Prior distributions for parameters defining cladogenetic changes were gamma distributed with parameters k = 0.5 and $\theta = 1$. MCMC was performed for 96 hours in the cluster at Minnesota Supercomputing Instute which allowed for 5,000 generation of burn-in and a minimum of 200,000 generations of MCMC for each of the 6 models. Convergence of the MCMC was tested

using R package coda (ref) and software Tracer (ref) to assess convergence and mixing (see supplementary information).

Results

Discussion

Acknowledgements

Literature Cited

- Beaulieu, J. M. and B. C. O'Meara, 2016. Detecting hidden diversification shifts in models of trait-dependent speciation and extinction. Syst Biol 65:583–601.
 - Bennett, M. D. and I. J. Leitch, 2005. Plant DNA C-values database.

145

150

155

160

165

- FitzJohn, R. G., 2012. Diversitree: comparative phylogenetic analyses of diversification in r. Methods Ecol Evol 3:1084–1092.
- Goldberg, E. E. and B. Igić, 2012. Tempo and mode in plant breeding system evolution. Evolution 66:3701–3709.
 - Goldberg, E. E., J. R. Kohn, R. Lande, K. A. Robertson, S. A. Smith, and B. Igić, 2010. Species selection maintains self-incompatibility. Science 330:493–495.
 - Höhna, S., M. J. Landis, T. A. Heath, B. Boussau, N. Lartillot, B. R. Moore, J. P. Huelsenbeck, and F. Ronquist, 2016. RevBayes: Bayesian phylogenetic inference using graphical models and an interactive model-specification language. Syst Biol 65:726–736.
 - Igić, B., L. Bohs, and J. R. Kohn, 2006. Ancient polymorphism reveals unidirectional breeding system shifts. Proc Natl Acad Sci USA 103:1359–1363.
 - Igić, B. and J. W. Busch, 2013. Is self-fertilization an evolutionary dead end? New Phytol 198:386–397.
 - Maddison, W. P., P. E. Midford, and S. P. Otto, 2007. Estimating a binary character's effect on speciation and extinction. Syst Biol 56:701–710.
 - Mayrose, I., S. H. Zhan, C. J. Rothfels, N. Arrigo, M. S. Barker, L. H. Rieseberg, and S. P. Otto, 2015. Methods for studying polyploid diversification and the dead end hypothesis: a reply to soltis et al. (2014). New Phytol 206:27–35.
 - Mayrose, I., S. H. Zhan, C. J. Rothfels, K. Magnuson-Ford, M. S. Barker, L. H. Rieseberg, and S. P. Otto, 2011. Recently formed polyploid plants diversify at lower rates. Science 333:1257.
 - Olmstead, R. G. and L. Bohs, 2007. A summary of molecular systematic research in Solanaceae: 1982–2006. Acta Horticulturae Pp. 255–268.
 - PBI Solanum Project, 2012. Solanaceae Source: a global taxonomic resource for the nightshade family.
 - Raduski, A. R., E. B. Haney, and B. Igić, 2012. The expression of self-incompatibility in angiosperms is bimodal. Evolution 66:1275–1283.
 - Ramsey, J. and D. W. Schemske, 1998. Pathways, mechanisms, and rates of polyploid formation in flowering plants. Annual Review of Ecology and Systematics 29:467–501.
 - Rice, A., L. Glick, S. Abadi, M. Einhorn, N. M. Kopelman, A. Salman-Minkov, J. Mayzel, O. Chay, and I. Mayrose, 2015. The chromosome counts database (CCDB) - a community resource of plant chromosome numbers. New Phytol 206:19–26.
 - Robertson, K., E. E. Goldberg, and B. Igić, 2011. Comparative evidence for the correlated evolution of polyploidy and self-compatibility in solanaceae. Evolution 65:139–155.
 - Särkinen, T., L. Bohs, R. G. Olmstead, and S. Knapp, 2013. A phylogenetic framework for evolutionary study of the nightshades (Solanaceae): a dated 1000-tip tree. BMC Evol Biol 13:214.
- Soltis, D. E., M. C. Segovia-Salcedo, I. Jordon-Thaden, L. Majure, N. M. Miles, E. V. Mavrodiev, W. Mei, M. B. Cortez, P. S. Soltis, and M. A. Gitzendanner, 2014. Are polyploids really evolutionary dead-ends (again)? a critical reappraisal of mayroseetăal. (2011). New Phytol 202:1105–1117.
 - Stone, J. L., 2002. Molecular mechanisms underlying the breakdown of gametophytic self-incompatibility. The Quarterly Review of Biology 77:17–30.
- ¹⁷⁵ Zenil-Ferguson, R., J. M. Ponciano, and J. G. Burleigh, 2017. Testing the association of phenotypes with polyploidy: An example using herbaceous and woody eudicots. Evolution 71:1138–1148.

		Models		
Туре	Total of Taxa	BiSSE/HiSSE Polyploidy	BiSSE/HiSSE Breeding System	MuSSE/MuHiSSE
Diploid Self Compatible	152	0	0	0
Diploid Self Incompatible	97	0	1	2
Diploid with unknown breeding system	219	0	(0,1)	(0,2)
Polyploid	81	1	0	1
Unknow ploidy and self compatible	34	(0,1)	0	(0,1)
Unknown ploidy and self incompatible	12	0	1	2

Table 1: Binary and three state classifications for 595 taxa with ploidy and/or breeding system data. The number of taxa in the t he sample was maximize by including tips with only ploidy or only breeding system and assigned them as uncertain in the unknown character.