

# 1 The Michaelis-Mentil Model

Image from Biostats Book

Where does this come from?

## 1.1 Some background

In biochemistry, Michaelis-Menten kinetics is one of the simplest and best-known models of enzyme kinetics. It is named after German biochemist Leonor Michaelis and Canadian physician Maud Menten. The reaction can be illustrated as



where  $E$  is the enzyme,  $S$  is the substrate,  $ES$  is the enzyme binding to the substrate, and  $P$  is a product.

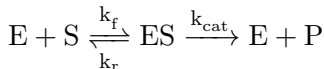
**Question:** How can we model this?

## 1.2 Building a Mathematical Model



- $\frac{d[E]}{dt}$  depends on ??
- $\frac{d[S]}{dt}$  depends on ??
- $\frac{d[ES]}{dt}$  depends on ??
- $\frac{d[P]}{dt}$  depends on ??

This leads to the following system of differential equations:



$$\frac{d[S]}{dt} = -k_f[E][S] + k_r[ES] \quad (1)$$

$$\frac{d[E]}{dt} = -k_f[E][S] + k_r[ES] + k_{cat}[ES] \quad (2)$$

$$\frac{d[ES]}{dt} = k_f[E][S] - k_r[ES] - k_{cat}[ES] \quad (3)$$

$$\frac{d[P]}{dt} = k_{cat}[ES] \quad (4)$$

## 2 Simplifying the Model

Now we make some simplifications:

- If the enzyme is conserved, then  $[E] + [ES] = [E]_0$  is a constant.
- In situations where  $\frac{d[P]}{dt}$  is constant (3) and (4) imply that  $\frac{d[ES]}{dt} = 0$ . This is often used as an approximation when  $\frac{d[P]}{dt}$  is nearly constant.

- From this it follows that

$$k_f[E][S] = (k_r + k_{cat})[ES]$$

$$k_f([E]_0 - [ES])[S] = (k_r + k_{cat})[ES]$$

$$([E]_0 - [ES])[S] = \frac{k_r + k_{cat}}{k_f}[ES]$$

$$[E]_0[S] = \left( \frac{k_r + k_{cat}}{k_f} + [S] \right) [ES]$$

$$[ES] = \frac{[E]_0[S]}{\frac{k_r + k_{cat}}{k_f} + [S]}$$

$$k_{cat}[ES] = \frac{k_{cat}[E]_0[S]}{\frac{k_r + k_{cat}}{k_f} + [S]}$$

$$v = \frac{d[P]}{dt} = \frac{k_{cat}[E]_0[S]}{k_f + k_r + [S]}$$

- Ignoring the meaning of the constants (for the moment) and focusing on the form, we now have the following sort of relationship between  $v$  and  $[S]$

$$v = \frac{\alpha[S]}{\beta + [S]} \quad (5)$$

That is, the “velocity” of the reaction (rate at which the product is produced) is determined by the concentration of the substrate and constants that do not depend on  $v$  or  $[S]$ .

Equation (5) is not in our favorite linear form, but we can use transformations to get into a linear form:

$$\frac{1}{v} = \frac{\beta + [S]}{\alpha[S]} = \frac{1}{\alpha} + \frac{\beta}{\alpha} \frac{1}{[S]}$$

### 3 Using Data to Fit the Model

This now provides an experimental way to estimate the constants  $\alpha$  and  $\beta$  (and from them to infer things about  $k_f$ ,  $k_r$ , and  $k_{cat}$ .) We can gather data providing values of  $v$  and  $[S]$  and fit  $\frac{1}{v}$  as a linear function of  $\frac{1}{[S]}$ . Let's do that using some data from a lab conducted by students at Calvin college.



```
mm <-
```

```
read.csv("http://www.calvin.edu/~rpruim/data/cal  
summary(mm)
```

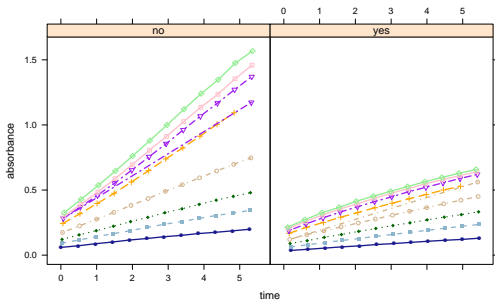
	batch	time	absorbance
SML-1	A1 : 12	Min. :0.00	Min. :0.036
SML-10	B2: 12	1st Qu.:1.37	1st Qu.:0.188
SML-11	B3: 12	Median :2.74	Median :0.332
SML-12	B4: 12	Mean :2.74	Mean :0.430
SML-13	B5: 12	3rd Qu.:4.10	3rd Qu.:0.557
SML-14	B6: 12	Max. :5.47	Max. :1.568
(Other)	:120		

## 3.1 Dealing with Indirect Measurements

The measurements are a bit indirect. The amount of product is inferred from the absorbance, and  $v$  is inferred by the rate of change in absorbance, which should be a roughly linear function of **time** (in minutes) for each value of  $[S]$  (**substrate**, mM peroxide) if our assumptions are true. Notice too that the experiment was run with and without an inhibitor.

Our first step is to estimate the values of  $v$  for each level of **substrate** by fitting a simple linear model and determining the slope. We begin by plotting the data to confirm that our linearity assumptions seem appropriate.

```
xyplot(absorbance ~ time | inhibitor, data =  
mm, groups = factor(substrate),  
type = c("p", "l"))
```



Not bad for student-collected data.

Now we need to estimate all those slopes. We can do this all at once with a clever choice of model. For the following analysis, we'll use only the data without the inhibitor.

```
mm$S <- factor(mm$substrate)
mmno <- subset(mm, inhibitor == "no")
slope.model <- lm(absorbance ~ time * S, data
= mmno)
summary(slope.model)
```

Call:

```
lm(formula = absorbance ~ time * S, data = mmno)
```

Residuals:

Min	1Q	Median	3Q	Max
-0.14643	-0.00300	0.00011	0.00270	0.04913

Coefficients:

	Estimate	Std. Error	t value	Pr(> t )
(Intercept)	0.06214	0.01075	5.78	1.4e-07
time	0.02641	0.00345	7.66	3.7e-11
S0.2	0.03173	0.01523	2.08	0.04041
S0.3	0.05935	0.01526	3.89	0.00021
S0.5	0.10578	0.01528	6.92	1.0e-09

S1	0.15577	0.01562	9.97	1.1e-15
S1.5	0.20656	0.01516	13.63	< 2e-16
S2	0.19780	0.01535	12.88	< 2e-16
S3	0.22729	0.01538	14.78	< 2e-16
time:S0.2	0.02179	0.00488	4.47	2.6e-05
time:S0.3	0.04235	0.00488	8.68	3.7e-13
time:S0.5	0.08312	0.00488	17.04	< 2e-16
time:S1	0.15245	0.00523	29.14	< 2e-16
time:S1.5	0.17090	0.00468	36.54	< 2e-16
time:S2	0.19665	0.00488	40.31	< 2e-16
time:S3	0.21396	0.00488	43.86	< 2e-16

Residual standard error: 0.0198 on 80 degrees of

Multiple R-squared: 0.998, Adjusted R-squared: 0.  
F-statistic: 2.56e+03 on 15 and 80 DF, p-value:

```
coef(slope.model)
```

(Intercept)	time	S0.2	S0.3
0.0621	0.0264	0.0317	0.0594
S1.5	S2	S3	time:S0.2
0.2066	0.1978	0.2273	0.0218
time:S1	time:S1.5	time:S2	time:S3
0.1525	0.1709	0.1966	0.2140

```
mmSlopes <- data.frame(S =  
as.numeric(levels(mmno$S)), v =  
coef(slope.model)["time"] +  
  c(`time:S0.1` = 0,  
coef(slope.model)[paste("time:S",  
levels(mmno$S)[-1],  
  sep = "")]))
```

mmSlopes

	S	v
time:S0.1	0.1	0.0264
time:S0.2	0.2	0.0482
time:S0.3	0.3	0.0688
time:S0.5	0.5	0.1095



```
time:S1    1.0  0.1789
time:S1.5  1.5  0.1973
time:S2    2.0  0.2231
time:S3    3.0  0.2404
```

## 3.2 Fitting with Ordinary Least Squares

Now we can fit our Michaelis-Menten model:

```
mmModel <- lm((1/v) ~ I(1/S), mmSlopes)
summary(mmModel)
```

Call:

```
lm(formula = (1/v) ~ I(1/S), data = mmSlopes)
```

Residuals:

Min	1Q	Median	3Q	Max
-0.544	-0.239	0.119	0.209	0.449

Coefficients:

	Estimate	Std. Error	t value	Pr(> t )
(Intercept)	2.5915	0.1973	13.1	1.2e-05
I(1/S)	3.5408	0.0468	75.6	3.6e-10

Residual standard error: 0.41 on 6 degrees of fr

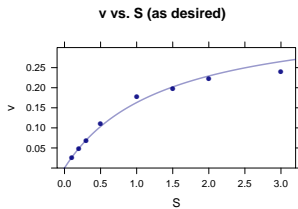
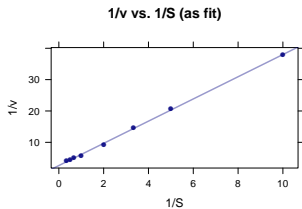
Multiple R-squared: 0.999, Adjusted R-squared: 0.  
F-statistic: 5.72e+03 on 1 and 6 DF, p-value: 3

```
alpha.hat <- 1/coef(mmModel)[1]  
beta.hat <- coef(mmModel)[2] * alpha.hat  
c(alpha.hat = alpha.hat, beta.hat = beta.hat)
```

alpha.hat.(Intercept)	beta.hat.I(1/S)
0.386	1.366

```
f <- makeFun(mmModel)  
xyplot(1/v ~ 1/S, mmSlopes, main = "1/v vs.  
1/S (as fit)")
```

```
g <- makeFun(f(1/x) ~ x)
plotFun(g(x) ~ x, add = TRUE, col = "navy",
alpha = 0.4)
xyplot(v ~ S, mmSlopes, ylim = c(0, 0.3),
main = "v vs. S (as desired)")
plotFun(1/f(S) ~ S, add = TRUE, col = "navy",
alpha = 0.4)
```



Doing this we see some issues. Although there is a nice tight fit of the transformed data to the least squares regression line, the fit doesn't look nearly as good after back-transforming to the original scales. In particular, errors are much larger for larger values of  $S$ . Or thought about the other way around, the way we have fit the model has forced the fit to be extremely good for

small values of  $S$  at the cost of allowing much poorer fits for larger values of  $S$ . This is because the transformations transform the scale for the residuals as well as for the inputs. Note too that the distribution of values of  $1/S$  is not nearly as uniform as it is for  $S$ .

### **3.3 Fitting with Nonlinear Least Squares**

We can do better if we use nonlinear least squares instead of transforming and using least squares. In nonlinear least squares we fit a parameterized function of arbitrary form by determining (well, estimating anyway) the values of the parameters that minimize the

sum of the squares of the residuals

$$SS(\alpha, \beta) = \sum_{i=1}^n (v_i - f(\alpha, \beta; S_i))^2$$

This approach will be less forgiving of such large residuals for large values of  $S$ . We lose something in the exchange, however. It is no longer the case that simple closed-form formulas exist for the estimates. This means that we will need to rely on numerical estimation. Furthermore the estimators are not guaranteed to be unbiased.

# we provide nls() with our estimates from above as a starting point.

```
model.nls <- nls(v ~ alpha * S/(beta + S),  
data = mmSlopes, start = list(alpha =  
alpha.hat,  
beta = beta.hat))  
summary(model.nls)
```

Formula:  $v \sim \alpha * S / (\beta + S)$

Parameters:

	Estimate	Std. Error	t value	Pr(> t )
alpha	0.3297	0.0163	20.24	9.4e-07



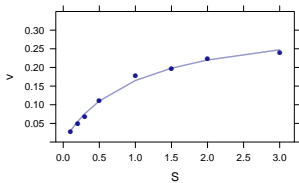
beta	0.9979	0.1191	8.38	0.00016
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Residual standard error: 0.00782 on 6 degrees of freedom

Number of iterations to convergence: 5

Achieved convergence tolerance: 6.84e-06

```
xyplot(v ~ S, mmSlopes, ylim = c(0, 0.35))  
ladd(panel.xyplot(mmSlopes$S,  
predict(model.nls), type = "l", col = "navy",  
alpha = 0.4))
```



As expected, the fit is much better for larger values of  $S$  and we've lost very little for smaller values of  $S$ . Now that we have (two sets of) estimates for  $\alpha$  and  $\beta$ , we should pause a moment to see what these parameters tell us about the chemistry. Recall Equation

(5)

$$v = \frac{\alpha[S]}{\beta + [S]}$$

As  $[S]$  increases,

$$\frac{\alpha[S]}{\beta + [S]} \nearrow \alpha$$

so  $\alpha$  gives the horizontal asymptote representing the maximum velocity. And if  $[S] = \beta$ , we get

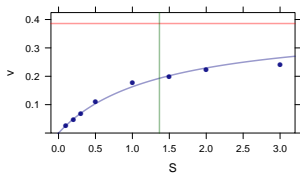
$$v = \frac{\alpha[S]}{[S] + [S]} = \frac{\alpha}{2} ,$$

so  $\beta$  is the value of  $[S]$  that gives half the maximum velocity.

```
xyplot(v ~ S, mmSlopes, ylim = c(0, 1.1 *  
alpha.hat), main = "linear least squares  
fit")  
plotFun(1/f(S) ~ S, add = TRUE, col = "navy",  
alpha = 0.4)  
ladd(panel.abline(h = alpha.hat, col = "red",  
alpha = 0.4))  
ladd(panel.abline(v = beta.hat, col =  
"darkgreen", alpha = 0.4))  
xyplot(v ~ S, mmSlopes, ylim = c(0, 1.1 *  
alpha.hat), main = "non-linear least squares
```

```
fit")  
ladd(panel.xyplot(mmSlopes$S,  
predict(model.nls), type = "l", col = "navy",  
alpha = 0.4))  
ladd(panel.abline(h = coef(model.nls)[1], col  
= "red", alpha = 0.4))  
ladd(panel.abline(v = coef(model.nls)[2], col  
= "darkgreen", alpha = 0.4))
```

**linear least squares fit**



**non-linear least squares fit**

