Lab 3 Bioinformatics

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Assignment 1

1.1

In Chapter 3 we read the sequences and use clustal function for multiple sequence alignment and we use mafft function for alignemnt of the sequences. Then we just name the sequences and save the data.

In Chapter 5 we compute the distance matrices for according to six different models. With two of the obtained distances we compute nj trees. We use one of the leaves as root and perform bootsraping to see how reliable the tree is. We did not manage to make work this line "phyml.sylvia <- phymltest("sylvia.txt", execname ="~/phyml")" and therefore we could not perform analysis related to it. Using the function chronopl with cross validation we estimated the node ages and saved this as sylvia.chrono.

In Chapter 6 we are using the ace function to see the properties of geographical range. With this we can see the likelihoods of the range. With information from geo_range we can fill the leaves with corresponding colors and using the likelihood we can fill the nodes with the colors corresponding to the likelihood

1.2

```
gr_sym$rates

## [1] 3.983947 0.000000 11.884956

gr_sym$se

## [1] 2.319376 NaN 5.916586

gr_er$rates

## [1] 5.988848

gr_er$se

## [1] 2.230071

gr_ard$rates

## [1] 2.967695 0.000000 0.000000 0.000000 7.202527
```

NaN

We used 3 different models for analyzing the geographical range variable. In the results we can see the maximum likelihood estimates of the transition rates and also standard errors of these rates. First model corresponds to symmetrical model, the second one corresponds to the equal-rates model and the last one to all-rates-different model. Our goal is for sure minimize the standard errors. We can see that the lowest standard error we can get is with equal-rates model.

NaN 9.761911 2.599815

Assignment 2

[1] 2.331727 2.784387

2.1

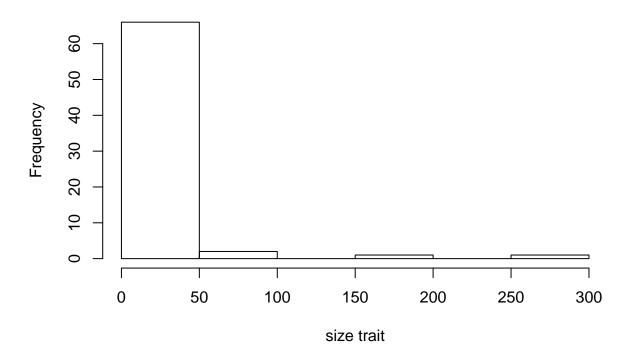
gr_ard\$se

We analysed the carnivores dataset, data(carni70), from the ade4 R package. The data is about 70 species.

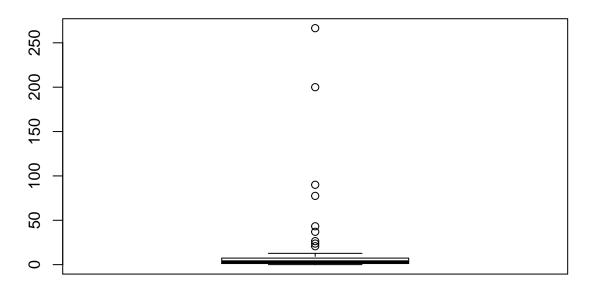
head(carni70\$tab, 10)

```
##
                            size range
## Puma concolor
                           37.01 36.00
## Herpailurus_yaguaroundi 2.59 24.00
## Leopardus_wiedii
                            3.20 21.75
## Leopardus_pardalis
                            7.90 24.50
## Oreailurus jacobita
                            3.99 1.75
## Oncifelis_colocolo
                            2.99 6.25
## Oncifelis_guigna
                            2.23
                                 1.25
## Oncifelis_geoffroyi
                           3.80 4.75
## Leopardus_tigrinus
                            2.19 20.25
## Lynx_rufus
                           11.20 13.50
```

Histogram of Size

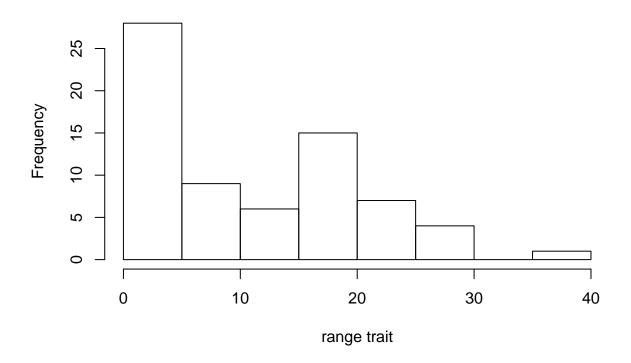




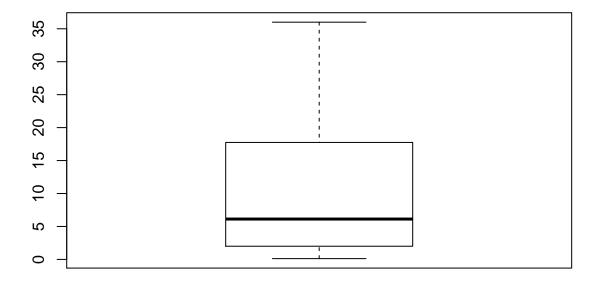


The distribution of the size trait is skewed to the right. In other words, the majority of the 70 species have small sizes. Some of the species have outlier sizes.

Histogram of Range

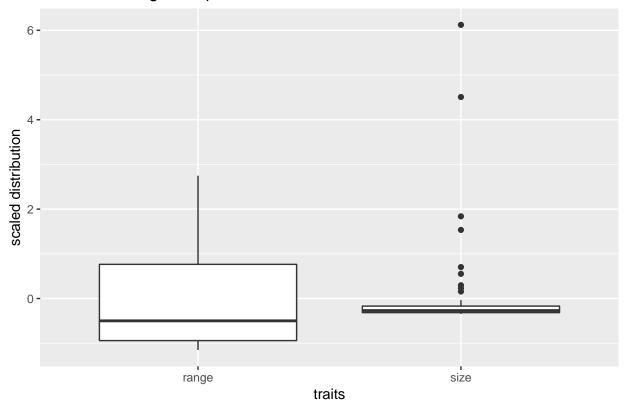


Range



The distribution of the range trait is also generally more balanced however with evident skewness to the right. There are fewer and fewer species with longer ranges.

Size and Range compared on the same scale



After scaling the data and comparing the distributions of size and range traits on the same scale with boxplots, it is clear that the size trait is more skewed towards low values than the range trait.

```
rownames(carni70_2[which.max(carni70_2[,"size"]), ])

## [1] "Ursus_arctos"

rownames(carni70_2[which.min(carni70_2[,"size"]), ])

## [1] "Mustela_nivalis"

rownames(carni70_2[which.max(carni70_2[,"range"]), ])

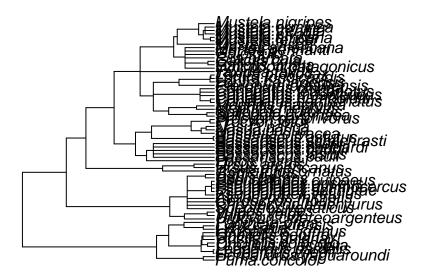
## [1] "Puma_concolor"

rownames(carni70_2[which.min(carni70_2[,"range"]), ])
```

The species "Ursus_arctos" has the biggest size while "Mustela_nivalis" the smallest size. The species "Puma_concolor" has the biggest range while the species "Bassariscus_pauli" has the smallest range

[1] "Bassariscus_pauli"

```
tree_phylo <- ape::read.tree(text=carni70_1)
plot(tree_phylo)</pre>
```



Above is a plot of the phylogenetic tree.

2.2

2.2.1 Both traits evolve as independent Brownian motions

```
## size range
## size 73.70335 0.00000
## range 0.00000 13.54524
##
## Estimated root state
##
## size range
## theta: 38.43947 13.78439
```

Because of the assumption that the two traits follow independent processes, the off diagonal elements of the estimated rate matrix are zero. From the rate matrix, we see that the changes in the size trait along the phylogenetic tree are more frequent (at 73) than the changes of the range trait on the same tree. That explains why the histogram of the size trait is more left skewed than that of the range trait (13). This model also suggests that the ancestor(root) had a size of 38.43947 and a range of 13.78439. This model has AIC of 1186.475 and AICc of 1186.771

2.2.2 The traits evolve as a correlated Brownian motion

```
library(ouch)
tree_ouch <- ouch::ape2ouch(tree_phylo, branch.lengths = tree_phylo$edge.length)</pre>
library(mvSLOUCH)
mvSLOUCH::BrownianMotionModel(tree_ouch, data = as.matrix(carni70$tab))
## $ParamsInModel
## $ParamsInModel$Sxx
##
               size
                        range
## size 171.415584 0.00000
           1.327048 26.01045
##
  range
  $ParamsInModel$vX0
##
##
              [,1]
          8.434762
## size
## range 11.084473
##
##
## $ParamSummary
## $ParamSummary$StS
##
               size
## size 29383.3025 227.4768
           227.4768 678.3047
## range
##
## $ParamSummary$LogLik
   [1] -656.7365
##
## $ParamSummary$dof
## [1] 5
##
## $ParamSummary$m2loglik
## [1] 1313.473
##
```

\$ParamSummary\$aic

```
## [1] 1323.473
##
## $ParamSummary$aic.c
## [1] 1323.921
##
## $ParamSummary$sic
## [1] 1338.181
##
## $ParamSummary$bic
## [1] 1338.181
##
## $ParamSummary$RSS
## [1] 140
```

Under the assumption that the two traits follow correlated Brownian motion processes, the ancestor (root) had a size of 8.434762 and a range of 11.084473. Even under this model, the size trait changes more frequently than the range trait, with 29383.3025 for size versus 678.3047 for range. The difference between size and range is clearly indicated in the skewnesses of histograms. This model has AIC of 1323.473 and AIC.c of 1323.921

2.2.3 independent Ornstein Uhlenbeck processes

```
## species in the matrix are assumed to be in the same order as in the phylogeny, otherwise specify row
##
## -- Summary results --
## LogLikelihood:
                      -248.9895
## AIC:
             503.979
## AICc:
             504.3427
## 3 parameters
##
## Estimated theta values
## OU1 11.08609
##
## ML alpha values
##
    0.2295954
##
##
## ML sigma values
##
##
    38.05329
```

The following is one of the mathematical representations of a Ornstein-Uhlenbeck processes sourced from the Wikipedia page on Ornstein-Uhlenbeck process

Representation via a stochastic differential equation [edit]

An Ornstein-Uhlenbeck process, x_t , satisfies the following stochastic differential equation:

$$dx_t = heta(\mu - x_t)\,dt + \sigma\,dW_t$$

where heta>0, μ , and $\sigma>0$ are parameters and W_t denotes the Wiener process.

This shows that the OU process comprises a Wiener process component (Brownian motion component.) When we modeled the changes in both the size and range traits, we obtained theta and sigma values. The theta and sigma values help us compare the two traits. The size trait has a theta value of 38.43947 which is bigger than the theta value for range of 11.08609. Given the above shown differential question, this implies that the Ornstein-Uhlenbeck change process of size trait is bigger than that of the range trait. Additionally concerning the sigma values, the size trait has a bigger sigma value of 73.70246 as compared to that of the range trait of 38.05329. The sigma value is the Wiener process drift term. That means that the change in the size trait takes longer to reach equilibrium (or stabilise) as compared to the change in the range trait. The size trait has an AIC of 654.5278 and AICc of 654.8914. The range trait has an AIC of 503.979 and an AICc of 504.3427. The sum of the AIC for both traits is 1158.5068 and that for AICc is 1159.2341

2.2.4 traits evolve as a bivariate Ornstein-Uhlenbeck process

```
mvMORPH::mvOU(tree_phylo, data=carni70$tab, model = c("OU1"),
             diagnostic = TRUE, echo = TRUE)
## row names of the data matrix must match tip names of your phylogeny!
## successful convergence of the optimizer
## a reliable solution has been reached
## -- Summary results --
## LogLikelihood: -572.6063
## AIC:
        1161.213
## AICc:
           1162.312
## 8 parameters
##
## Estimated theta values
## size range
## OU1 37.55833 11.42602
## ML alpha values
## _____
##
           size
## size 8.345549e-05 -0.003859286
## range -3.859286e-03 0.233577603
## ML sigma values
## _____
          size range
## size 73.766441 5.859783
## range 5.859783 38.247492
```

This model does not give us notable new information from the earlier approaches. However, it is worth noting that the model has AIC of 1161.213 and AICc of 1162.312

2.2.5 size evolves as a Brownian motion and range as an Ornstein Uhlenbeck

The model has AIC of 1295.603 and AIC.c of 1296.235. The R output of this model is included in the Appendix.

comparison of the above five models

```
## 2 Both traits as a correlated Brownian motion 1323.473
## 3 both traits as independent Ornstein Uhlenbeck processes 1158.507
## 4 both traits as bivariate Ornstein-Uhlenbeck processes 1161.213
## 5 size as Brownian motion and range as Ornstein Uhlenbeck 1295.603
```

From Wikipedia,

"AIC estimates the relative information lost by a given model: the less information a model loses, the higher the quality of that model."

Appendix

```
library(ade4)
library(tidyverse)
library(ape)
library(mvMORPH)
library(ouch)
library(mvSLOUCH)
library(ape)
###
### Chapter 3
###
x \leftarrow paste("AJ5345", 26:49, sep = "")
x \leftarrow c("Z73494", x)
sylvia.seq <- read.GenBank(x)</pre>
sylvia.clus <- clustal(sylvia.seq) ## storing to the matrix</pre>
library(phyloch)
## Martin ..... Please change the following lines if you knit to pdf ....
# sylvia.maff <- mafft(sylvia.seq, path = #"C:/Users/smelo/Downloads/mafft/mafft-win/mafft")
sylvia.maff <- mafft(sylvia.seq,</pre>
    path = "C:/Users/rsseg/Downloads/mafft-win/mafft")
#sylvia.maff <- mafft(sylvia.seq)</pre>
identical(sylvia.clus[x, ], sylvia.maff[x, ])
taxa.sylvia <- attr(sylvia.seq, "species")</pre>
names(taxa.sylvia) <- names(sylvia.seq)</pre>
rm(sylvia.seq)
taxa.sylvia[1] <- "Sylvia_atricapilla"</pre>
taxa.sylvia[24] <- "Sylvia_abyssinica"</pre>
sylvia.eco <- read.table("sylvia_data.txt")</pre>
str(sylvia.eco)
rownames(sylvia.eco)
save(sylvia.clus, taxa.sylvia, sylvia.eco,
```

```
file = "sylvia.RData")
###
### Chapter 5
###
syl.K80 <- dist.dna(sylvia.clus, pairwise.deletion = TRUE)</pre>
syl.F84 <- dist.dna(sylvia.clus, model = "F84", p = TRUE)</pre>
syl.TN93 <- dist.dna(sylvia.clus, model = "TN93", p = TRUE)
syl.GG95 <- dist.dna(sylvia.clus, model = "GG95", p = TRUE)
round(cor(cbind(syl.K80, syl.F84, syl.TN93, syl.GG95)), 3)
syl.JC69 <- dist.dna(sylvia.clus, model = "JC69", p = TRUE)</pre>
syl.raw <- dist.dna(sylvia.clus, model = "raw", p = TRUE)</pre>
layout(matrix(1:2, 1))
plot(syl.JC69, syl.raw)
abline(b = 1, a = 0) # draw x = y line
plot(syl.K80, syl.JC69)
abline(b = 1, a = 0)
layout(matrix(1:3, 1))
for (i in 1:3) {
  s <- logical(3); s[i] <- TRUE
  x <- sylvia.clus[, s]
  d <- dist.dna(x, p = TRUE)</pre>
  ts <- dist.dna(x, "Ts", p = TRUE)
  tv <- dist.dna(x, "Tv", p = TRUE)</pre>
  plot(ts, d, xlab = "Number of Ts or Tv", col = "blue",
       ylab = "K80 distance", xlim = range(c(ts, tv)),
       main = paste("Position", i))
  points(tv, d, col = "red")
y <- numeric()
for (i in 1:3) {
 s <- logical(3); s[i] <- TRUE
  y <- c(y, dist.dna(sylvia.clus[, s], p = TRUE))
}
g <- gl(3, length(y) / 3)
library(lattice)
histogram(~ y | g, breaks = 20)
nj.sylvia.K80 <- nj(syl.K80)
nj.sylvia.GG95 <- nj(syl.GG95)
dist.topo(nj.sylvia.K80, nj.sylvia.GG95)
grep("Chamaea", taxa.sylvia, value = TRUE)
f <- function(xx) root(nj(dist.dna(xx, p=TRUE)), "AJ534526")
#tr <- f(sylvia.clus)</pre>
## same than:
tr <- root(nj.sylvia.K80, "AJ534526")</pre>
nj.boot.sylvia <- boot.phylo(tr, sylvia.clus, f, 200,
```

```
rooted = TRUE)
nj.boot.codon <- boot.phylo(tr, sylvia.clus, f, 200, 3,
                              rooted = TRUE)
nj.est <- tr
nj.est$tip.label <- taxa.sylvia[tr$tip.label]</pre>
plot(nj.est, no.margin = TRUE)
nodelabels(round(nj.boot.sylvia / 200, 2), bg = "white")
add.scale.bar(length = 0.01)
write.tree(nj.est, "sylvia_nj_k80.tre")
write.dna(sylvia.clus, "sylvia.txt")
#phyml.sylvia <- phymltest("sylvia.txt", execname = "~/phyml") ##ignore</pre>
#summary(phyml.sylvia) ##ignore
#plot(phyml.sylvia, col = "black") ##iqnore
#TR <- read.tree("sylvia.txt_phyml_tree.txt") ##iqnore</pre>
#mltr.sylvia <- TR[[28]] ##iqnore
#mltr.sylvia$tip.label <- taxa.sylvia[mltr.sylvia$tip.label] ##ignore</pre>
#mltr.sylvia <- root(mltr.sylvia, "Chamaea_fasciata") ##ignore</pre>
#plot(mltr.sylvia, no.marqin = TRUE) ##iqnore
#add.scale.bar(length = 0.01) ##ignore
#tr.ml <- drop.tip(mltr.sylvia, "Chamaea_fasciata")</pre>
tr.ml <- drop.tip(nj.est, "Chamaea_fasciata")</pre>
res <- vector("list", 9)</pre>
for (L in -4:4)
  res[[L + 5]] <- chronopl(tr.ml, 10^L, 12, 16, CV = TRUE)
Lambda <- 10^{-4:4}
CV <- sapply(res, function(x) sum(attr(x, "D2")))
plot(Lambda, CV / 1e5, log = "x")
sylvia.chrono <- res[[2]]</pre>
rts <- attr(sylvia.chrono, "rates")</pre>
summary(rts)
par(mar = c(2, 0, 0, 0))
plot(sylvia.chrono, edge.width = 100*rts, label.offset = .15)
axisPhylo()
write.tree(sylvia.chrono, "sylvia.chrono.tre")
###
### Chapter 6
###
load("sylvia.RData")
nj.est <- read.tree("sylvia_nj_k80.tre")</pre>
nj.est <- drop.tip(nj.est, "Chamaea_fasciata")</pre>
DF <- sylvia.eco[nj.est$tip.label, ]</pre>
table(DF$geo.range, DF$mig.behav)
gr_sym <- ace(DF$geo.range, nj.est, type="d", model="SYM")</pre>
gr_er <- ace(DF$geo.range, nj.est, type="d", model="ER")</pre>
gr_ard <- ace(DF$geo.range, nj.est, type="d", model="ARD")</pre>
```

```
gr_sym$rates
gr_sym$se
gr er$rates
gr_er$se
gr ard$rates
gr_ard$se
library(ade4)
data(carni70)
View(carni70)
str(carni70)
names(carni70)
carni70_1 <- carni70[[1]]</pre>
carni70_2 <- carni70[[2]]</pre>
head(carni70$tab, 10)
hist(carni70_2$size, xlab="size trait", main = "Histogram of Size")
boxplot(carni70_2$size, main="Size")
hist(carni70_2$range, xlab="range trait", main = "Histogram of Range")
boxplot(carni70 2$range, main="Range")
ggplot()+aes(x=names(carni70_2)[1], y=scale(carni70_2$size))+
  geom_boxplot()+
  geom_boxplot(aes(x=names(carni70_2[2]), y=scale(carni70_2$range)))+
  labs(y="scaled distribution", x="traits",
       title=" Size and Range compared on the same scale")
### Ursus_arctos has the biggest size
rownames(carni70_2[which.max(carni70_2[,"size"]), ])
rownames(carni70_2[which.min(carni70_2[,"size"]), ])
rownames(carni70_2[which.max(carni70_2[,"range"]), ])
rownames(carni70_2[which.min(carni70_2[,"range"]), ])
tree_phylo <- ape::read.tree(text=carni70_1)</pre>
plot(tree_phylo)
mvMORPH::mvBM(tree_phylo, carni70$tab, model="BM1",
             param = list(constraint = "diagonal"))
library(ouch)
tree_ouch <- ouch::ape2ouch(tree_phylo, branch.lengths = tree_phylo$edge.length)</pre>
library(mvSLOUCH)
mvSLOUCH::BrownianMotionModel(tree_ouch, data = as.matrix(carni70$tab))
mvMORPH::mvOU(tree_phylo, data=carni70$tab$size, model = c("OU1"),
              diagnostic = FALSE, echo = TRUE)
mvMORPH::mvOU(tree_phylo, data=carni70$tab$range, model = c("OU1"),
              diagnostic = FALSE, echo = TRUE)
mvMORPH::mvOU(tree_phylo, data=carni70$tab, model = c("OU1"),
               diagnostic = TRUE, echo = TRUE)
mvslouchModel(tree_ouch, kY=1,
      data = as.matrix(cbind(carni70$tab$range,carni70$tab$size)))
```

```
model1=c("Both traits evolve as independent Brownian motions", 1186.475, 1186.771)
model2=c("Both traits as a correlated Brownian motion", 1323.473, 1323.921)
model3=c("both traits as independent Ornstein Uhlenbeck processes", 1158.5068, 1159.2341)
model4=c("both traits as bivariate Ornstein-Uhlenbeck processes",
         1161.213, 1162.312)
model5=c("size as Brownian motion and range as Ornstein Uhlenbeck",
         1295.603, 1296.235)
comparison matrix <- rbind(model1, model2, model3, model4, model5)</pre>
comparison_matrix <- as.data.frame(comparison_matrix)</pre>
comparison_matrix[,2] <- as.numeric(as.character(comparison_matrix[,2]))</pre>
comparison_matrix[,3] <- as.numeric(as.character(comparison_matrix[,3]))</pre>
colnames(comparison_matrix) <- c("model", "AIC", "AIC.c")</pre>
rownames(comparison_matrix) <- NULL</pre>
print(comparison_matrix[,1:2])
# [1] "Starting point of heuristic search procedure : "
          \boldsymbol{A}
                    Syy
# -0.3038808 -1.1454798
# $`FinalFound`
# $`FinalFound`$`HeuristicSearchPointFinalFind`
       \boldsymbol{A}
                      Syy
                               LogLik
#
     2.339259
               3.712662 -641.801747
# $`FinalFound`$ParamsInModel
# $`FinalFound`$ParamsInModel$`A`
           V1
# V1 10.37354
# $`FinalFound`$ParamsInModel$B
# V1 -0.6884966
# $`FinalFound`$ParamsInModel$mPsi
       reg.1
# V1 10.17529
# $`FinalFound`$ParamsInModel$mPsi0
   [,1]
# V1
# $`FinalFound`$ParamsInModel$vYO
         [,1]
# V1 10.73511
# $`FinalFound`$ParamsInModel$vXO
```

```
# [,1]
# V2 8.434762
# $`FinalFound`$ParamsInModel$Syy
#
# V1 40.96271
# $`FinalFound`$ParamsInModel$Syx
# V1 O
# $`FinalFound`$ParamsInModel$Sxy
# V2 0
# $`FinalFound`$ParamsInModel$Sxx
# V2 171.4156
#
# $`FinalFound`$ParamSummary
# $`FinalFound`$ParamSummary$`phyl.halflife`
# $`FinalFound`$ParamSummary$`phyl.halflife`$`directions`
     [,1]
# [1,]
# $`FinalFound`$ParamSummary$`phyl.halflife`$halflives
# eigenvalues 10.37354269
# halflife 0.06681875
# %treeheight 6.68187524
# $`FinalFound`$ParamSummary$`phyl.halflife`$halflifeLowerbounds
# [1] 0.06681875
#
# $`FinalFound`$ParamSummary$expmtA
# V1 3.124839e-05
# $`FinalFound`$ParamSummary$optimal.regression
# V1 0.06637044
# $`FinalFound`$ParamSummary$mPsi.rotated
      reg.1
# V1 10.17497
# $`FinalFound`$ParamSummary$mPsiO.rotated
# [,1]
# V1 O
# $`FinalFound`$ParamSummary$cov.matrix
```

```
# V1 V2
# V1 191.5953 1762.193
# V2 1762.1927 29383.302
# $`FinalFound`$ParamSummary$corr.matrix
  V1
# V1 1.0000000 0.7426952
# V2 0.7426952 1.0000000
# $`FinalFound`$ParamSummary$conditional.cov.matrix
#
# V1 85.91207
# $`FinalFound`$ParamSummary$conditional.corr.matrix
# V1 1
# $`FinalFound`$ParamSummary$stationary.cov.matrix
# V1 87.1148
# $`FinalFound`$ParamSummary$stationary.corr.matrix
# V1 1
#
# $`FinalFound`$ParamSummary$optima.cov.matrix
# V1 129.4345
# $`FinalFound`$ParamSummary$optima.corr.matrix
# V1 1
# $`FinalFound`$ParamSummary$cov.with.optima
# V1 116.9575
# $`FinalFound`$ParamSummary$corr.with.optima
# V1 0.7426952
#
# $`FinalFound`$ParamSummary$evolutionary.regression
# V1 0.05997259
# $`FinalFound`$ParamSummary$StS
         V1
                 V2
# V1 1677.944
                0.0
# V2 0.000 29383.3
# $`FinalFound`$ParamSummary$LogLik
# [1] -641.8017
```

```
# $`FinalFound`$ParamSummary$dof
# [1] 6
# $`FinalFound`$ParamSummary$m2loglik
# [1] 1283.603
# $`FinalFound`$ParamSummary$aic
# [1] 1295.603
# $`FinalFound`$ParamSummary$aic.c
# [1] 1296.235
# $`FinalFound`$ParamSummary$sic
# [1] 1313.253
# $`FinalFound`$ParamSummary$bic
# [1] 1313.253
# $`FinalFound`$ParamSummary$RSS
# $`FinalFound`$ParamSummary$RSS$`RSS`
# [1,] 139.8423
# $`FinalFound`$ParamSummary$RSS$R2
           [,1]
# [1,] 0.0177938
\# $`FinalFound`$ParamSummary$trait.regression
# $`FinalFound`$ParamSummary$trait.regression[[1]]
# V1 0.05997259
#
#
# $`FinalFound`$ParamSummary$confidence.interval
# $`FinalFound`$ParamSummary$confidence.interval$`regression.summary`
\# $`FinalFound`$ParamSummary$confidence.interval$`regression.summary`$`B.regression.confidence.interval
    Lower.end Estimated.Point Upper.end
# V1 -1.888154
                    -0.6884966 0.5111613
\# \$ `FinalFound` \$ParamSummary\$confidence.interval\$ `regression.summary` \$mPsi.regression.confidence.interv
# Lower.end Estimated.Point Upper.end
# V1 6.944177
                 10.17529 13.40641
#
# $`FinalFound`$LogLik
# [1] -641.8017
# $MaxLikFound
# $MaxLikFound$`HeuristicSearchPointMaxLik`
```

```
# A Syy LogLik
#
  2.339259
               3.712662 -641.800203
# $MaxLikFound$ParamsInModel
# $MaxLikFound$ParamsInModel$`A`
         V1
# V1 10.37354
# $MaxLikFound$ParamsInModel$B
# V1 -0.6792388
#
# $MaxLikFound$ParamsInModel$mPsi
# req.1
# V1 10.15824
# $MaxLikFound$ParamsInModel$mPsi0
# [,1]
# V1 O
# $MaxLikFound$ParamsInModel$vYO
# V1 10.72571
# $MaxLikFound$ParamsInModel$vXO
#
    [,1]
# V2 8.434762
# $MaxLikFound$ParamsInModel$Syy
# V1
# V1 40.96271
# $MaxLikFound$ParamsInModel$Syx
# V2
# V1 O
# $MaxLikFound$ParamsInModel$Sxy
# V2 0
# $MaxLikFound$ParamsInModel$Sxx
# V2 171.4156
# $MaxLikFound$ParamSummary
# $MaxLikFound$ParamSummary$`phyl.halflife`
# $MaxLikFound$ParamSummary$`phyl.halflife`$`directions`
# [,1]
# [1,] 1
# $MaxLikFound$ParamSummary$`phyl.halflife`$halflives
```

```
# eigenvalues 10.37354269
# halflife 0.06681875
# %treeheight 6.68187524
# $MaxLikFound$ParamSummary$`phyl.halflife`$halflifeLowerbounds
# [1] 0.06681875
#
# $MaxLikFound$ParamSummary$expmtA
# V1 3.124839e-05
# $MaxLikFound$ParamSummary$optimal.regression
#
# V1 0.065478
# $MaxLikFound$ParamSummary$mPsi.rotated
# req.1
# V1 10.15793
# $MaxLikFound$ParamSummary$mPsiO.rotated
# [,1]
# V1 O
# $MaxLikFound$ParamSummary$cov.matrix
          V1
# V1 188.6378 1738.498
# V2 1738.4977 29383.302
# $MaxLikFound$ParamSummary$corr.matrix
           V1
# V1 1.0000000 0.7384302
# V2 0.7384302 1.0000000
# $MaxLikFound$ParamSummary$conditional.cov.matrix
#
# V1 85.77755
# $MaxLikFound$ParamSummary$conditional.corr.matrix
# V1
# V1 1
# $MaxLikFound$ParamSummary$stationary.cov.matrix
# V1
# V1 86.94816
{\tt\#\$MaxLikFound\$ParamSummary\$stationary.corr.matrix}
# V1 1
# $MaxLikFound$ParamSummary$optima.cov.matrix
# V1 125.977
```

```
# $MaxLikFound$ParamSummary$optima.corr.matrix
# V1 1
# $MaxLikFound$ParamSummary$cov.with.optima
# V1 113.8334
# $MaxLikFound$ParamSummary$corr.with.optima
#
# V1 0.7384302
#
# $MaxLikFound$ParamSummary$evolutionary.regression
# V1 0.05916618
# $MaxLikFound$ParamSummary$StS
          V1
# V1 1677.944
# V2 0.000 29383.3
# $MaxLikFound$ParamSummary$LogLik
# [1] -641.8002
# $MaxLikFound$ParamSummary$dof
# [1] 6
# $MaxLikFound$ParamSummary$m2loglik
# [1] 1283.6
# $MaxLikFound$ParamSummary$aic
# [1] 1295.6
# $MaxLikFound$ParamSummary$aic.c
# [1] 1296.232
# $MaxLikFound$ParamSummary$sic
# [1] 1313.25
# $MaxLikFound$ParamSummary$bic
# [1] 1313.25
# $MaxLikFound$ParamSummary$RSS
# $MaxLikFound$ParamSummary$RSS$`RSS`
# [1,] 139.8697
# $MaxLikFound$ParamSummary$RSS$R2
# [1,] 0.01778443
#
#
```

```
\# $MaxLikFound$ParamSummary$trait.regression
# $MaxLikFound$ParamSummary$trait.regression[[1]]
# V1 0.05916618
#
#
\# $MaxLikFound$ParamSummary$confidence.interval
# $MaxLikFound$ParamSummary$confidence.interval$`regression.summary`
# $MaxLikFound$ParamSummary$confidence.interval$`regression.summary`$`B.regression.confidence.interval`
# Lower.end Estimated.Point Upper.end
# V1 -1.867116
                 -0.6792388 0.5086384
\# $MaxLikFound$ParamSummary$confidence.interval$`regression.summary`$mPsi.regression.confidence.interval
# Lower.end Estimated.Point Upper.end
# V1 6.883083
                 10.15824 13.4334
#
#
# $MaxLikFound$LogLik
# [1] -641.8002
```