crlmm to downstream data analysis

VJ Carey

April 7, 2009

1 Running CRLMM on a nontrivial set of CEL files

We work with the 90 CEU samples hybridized to Affy 6.0 chips, which are assumed to be in the current directory. First, we identify the files and run crlmm. The results will be saved to the variable crlmmResult.

```
> library(crlmm)
> celFiles <- list.celfiles()</pre>
> celFiles[1:4]
[1] "NAO6985_GW6_C.CEL" "NAO6991_GW6_C.CEL" "NAO6993_GW6_C.CEL"
[4] "NAO6994_GW6_C.CEL"
> if (!exists("crlmmResult")) {
      if (file.exists("crlmmResult.rda"))
           load("crlmmResult.rda")
      else {
           crlmmResult <- crlmm(celFiles)</pre>
          save(crlmmResult, file = "crlmmResult.rda")
      }
+ }
  This is currently a SnpSet object.
> class(crlmmResult)
[1] "SnpSet"
attr(,"package")
[1] "Biobase"
```

2 Adding information to a *SnpSet*

We will use the GGdata package to obtain extra information on the samples. This will be later used when building an eSet extension to store the genotyping results.

```
> library(GGdata)
> if (!exists("hmceuB36")) data(hmceuB36)
> pd <- phenoData(hmceuB36)</pre>
> ggn <- sampleNames(pd)</pre>
> preSN <- sampleNames(crlmmResult)
> simpSN <- gsub("_.*", "", preSN)</pre>
> if (!all.equal(simpSN, ggn)) stop("align GGdata phenoData with crlmmResult read")
   The additional information obtained from GGdata can be easily combined to what
is already available on crlmmResult.
> sampleNames(crlmmResult) <- simpSN
> phenoData(crlmmResult) <- combine(pd, phenoData(crlmmResult))
> dim(calls(crlmmResult))
[1] 906600
                90
> dim(confs(crlmmResult))
[1] 906600
                90
> calls(crlmmResult)[1:10, 1:2]
               NA06985 NA06991
SNP_A-2131660
                     2
                              2
SNP_A-1967418
                     3
                              3
SNP_A-1969580
                     3
                              3
SNP_A-4263484
                     2
                              1
SNP_A-1978185
                     1
                              1
SNP_A-4264431
                     1
                              1
                     3
                              3
SNP_A-1980898
SNP_A-1983139
                              1
                     1
                     2
                              2
SNP_A-4265735
SNP_A-1995832
                     2
                              3
> confs(crlmmResult)[1:10, 1:2]
               NA06985 NA06991
SNP_A-2131660
                 10561
                         11574
                 12517
SNP_A-1967418
                         14866
```

```
SNP_A-1969580
                  7632
                           7606
SNP_A-4263484
                 15621
                          20059
SNP_A-1978185
                 14030
                          18021
SNP_A-4264431
                 17792
                          17235
SNP_A-1980898
                  7640
                           7642
SNP_A-1983139
                 14127
                           8974
SNP_A-4265735
                  8976
                           9153
SNP_A-1995832
                 10336
                          17920
```

3 Coercing to snp.matrix as a prelude to a GWAS

```
> library(snpMatrix)
> crlmmSM <- as(t(calls(crlmmResult)) - 1, "snp.matrix")
> crlmmSM

A snp.matrix with 90 rows and 906600 columns
Row names: NA06985 ... NA12892
Col names: SNP_A-2131660 ... SNP_A-8574011
```

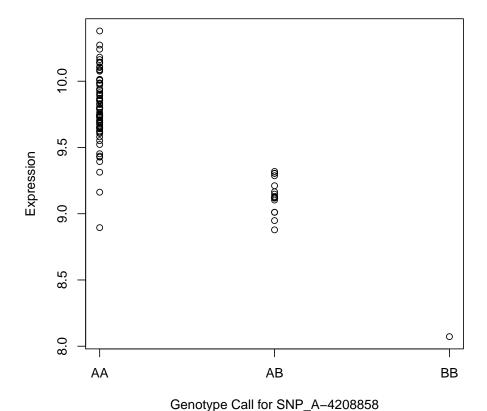
4 Conducting a GWAS

We want to find SNP for which rare allele count is predictive of expression of CPNE1. We will use expression data available from GGdata. This is a very naive analysis.

```
> library(illuminaHumanv1.db)
> rmm <- revmap(illuminaHumanv1SYMBOL)</pre>
> mypr <- get("CPNE1", rmm)</pre>
> ex <- as.numeric(exprs(hmceuB36)[mypr[1], ])</pre>
> subjdata <- pData(hmceuB36)</pre>
> subjdata[["ex"]] <- ex</pre>
> gwas <- snp.rhs.tests(ex ~ male, data = subjdata, snp.data = crlmmSM,
      family = "gaussian")
> ok <- which(p.value(gwas) < 1e-10)
> gwas[ok, ]
               Chi.squared Df
                                    p.value
SNP_A-4208858
                  53.62528 1 2.426168e-13
SNP_A-2022241
                  42.67385 1 6.467116e-11
SNP_A-2039695
                  46.23796 1 1.047283e-11
SNP_A-2047882
                  48.35134 1 3.563006e-12
SNP_A-2108011
                  46.23796 1 1.047283e-11
SNP_A-2125946
                  41.91646 1 9.525721e-11
```

```
SNP_A-2171015
                 46.23796
                            1 1.047283e-11
                            1 3.212681e-11
SNP_A-2184991
                 44.04283
SNP_A-2216659
                 48.35134
                            1 3.563006e-12
SNP_A-2220183
                 53.62528
                            1 2.426168e-13
SNP_A-2231089
                 46.23796
                            1 1.047283e-11
SNP_A-2231469
                  53.62528
                            1 2.426168e-13
SNP_A-2275065
                 53.62528
                            1 2.426168e-13
SNP_A-1806237
                 48.35134
                            1 3.563006e-12
SNP_A-1912540
                 46.23796
                            1 1.047283e-11
SNP_A-1921183
                            1 3.563006e-12
                 48.35134
SNP_A-8611599
                  51.91596
                            1 5.792757e-13
                            1 5.792757e-13
SNP_A-8699268
                 51.91596
```

```
> plot(ex ~ calls(crlmmResult)["SNP_A-4208858", ], xlab = "Genotype Call for SNP_A-4208858", ]  
+ ylab = "Expression", xaxt = "n")  
> axis(1, at = 1:3, labels = c("AA", "AB", "BB"))
```



5 Session Info

This vignette was created using the following packages:

```
> sessionInfo()
```

R version 2.9.0 Under development (unstable) (2009-02-08 r47879) $x86_64$ -unknown-linux-gnu

locale:

LC_CTYPE=en_US.UTF-8; LC_NUMERIC=C; LC_TIME=en_US.UTF-8; LC_COLLATE=en_US.UTF-8; LC_MONETAF

attached base packages:

- [1] splines stats graphics grDevices utils datasets methods
- [8] base

other attached packages:

- [1] GGdata_0.99.3 illuminaHumanv1.db_1.1.3 GGBase_3.2.11
- [4] RSQLite_0.7-1 DBI_0.2-4 snpMatrix_1.7.5
- [7] survival_2.34-1 GSEABase_1.5.2 graph_1.21.4
- [10] annotate_1.21.3 AnnotationDbi_1.5.15 Biobase_2.3.11
- [13] crlmm_1.0.78

loaded via a namespace (and not attached):

- [1] affyio_1.11.3 cluster_1.11.12 genefilter_1.23.2
- [4] mvtnorm_0.9-4 preprocessCore_1.5.3 tools_2.9.0
- [7] XML_1.99-0 xtable_1.5-4