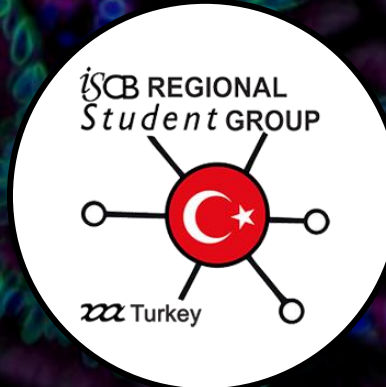


Introduction to Single-cell RNA sequencing (scRNA-seq)

Single-cell Workshop
19-20 December 2020





Single-cell sequencing

nature|**methods**

Editorial, N. M. "Method of the year 2013." Nat Methods 11 (2014): 1.



"The single-cell revolution is just starting."

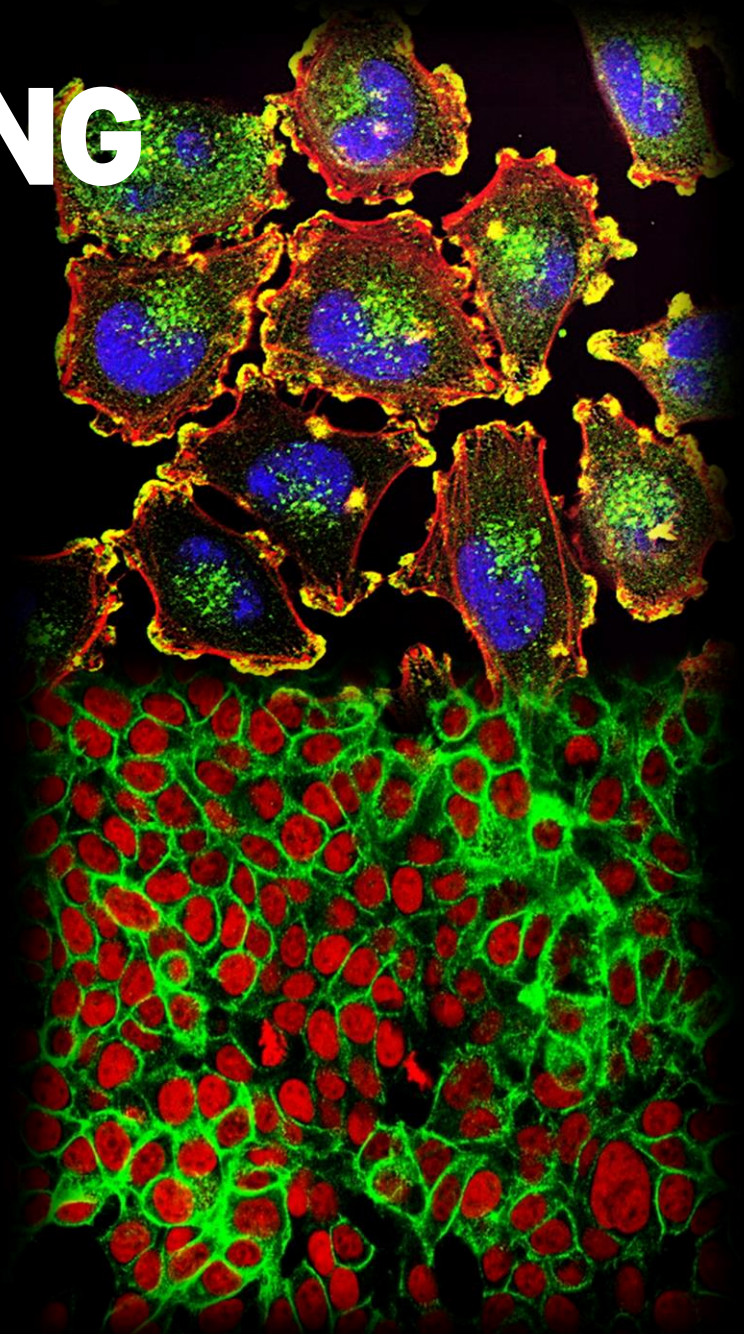
Development cell by cell

Science

Pennisi, Elizabeth. "Development cell by cell." (2018): 1344-1345.

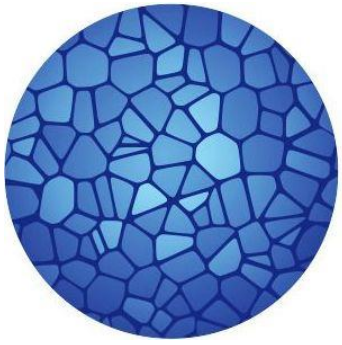
SINGLE-CELL SEQUENCING

- examines the sequence information from individual cells with optimized NGS technologies.
- higher resolution of cellular differences
- better understanding of the function of an individual cell



Bulk vs single-cell

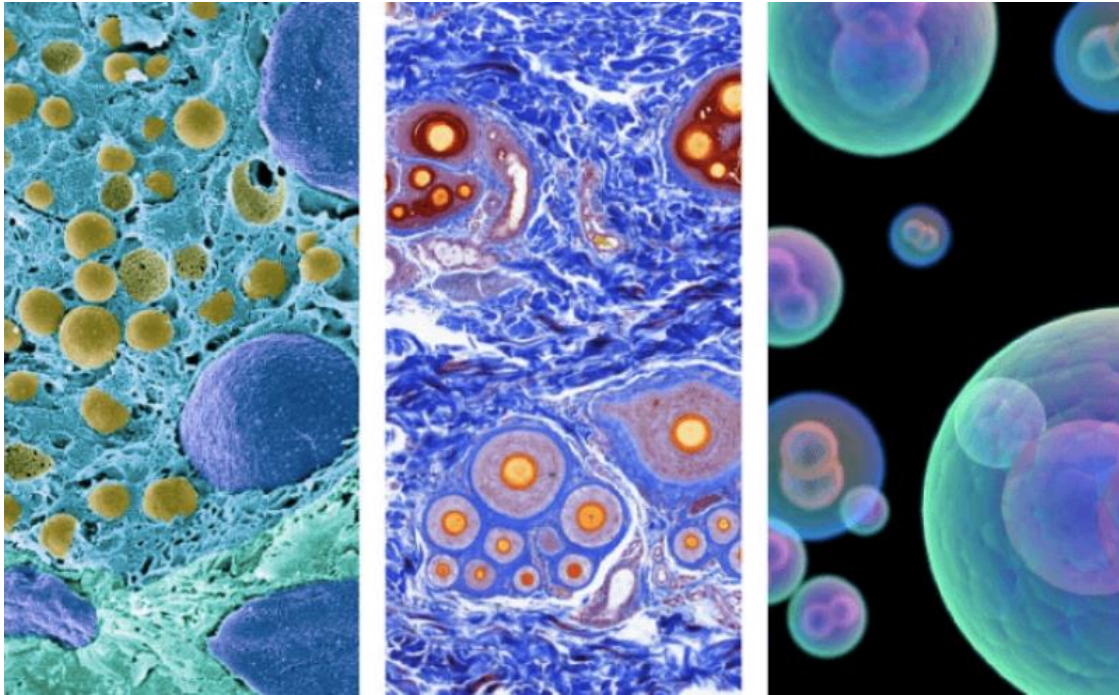




HUMAN CELL ATLAS

MISSION

To create comprehensive reference maps of all human cells—the fundamental units of life—as a basis for both understanding human health and diagnosing, monitoring, and treating disease.



More specifically, a human cell atlas could:

- catalog all cell types and sub-types in the human body;
- map cell types to their location within tissues and within the body;
- distinguish cell states;
- capture the key characteristics of cells during transitions, such as activation or differentiation;
- trace the history of cells through a lineage

Transcriptomics

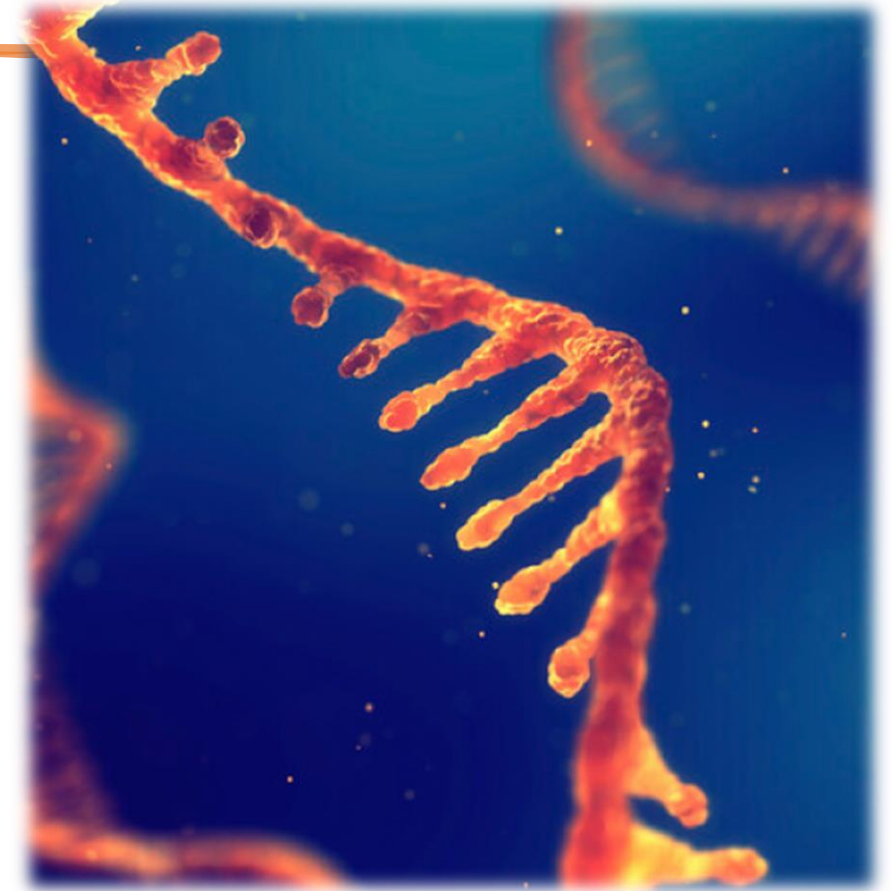
Transcriptome:

- the sum of all of the RNA transcripts
- mostly refers to mRNAs

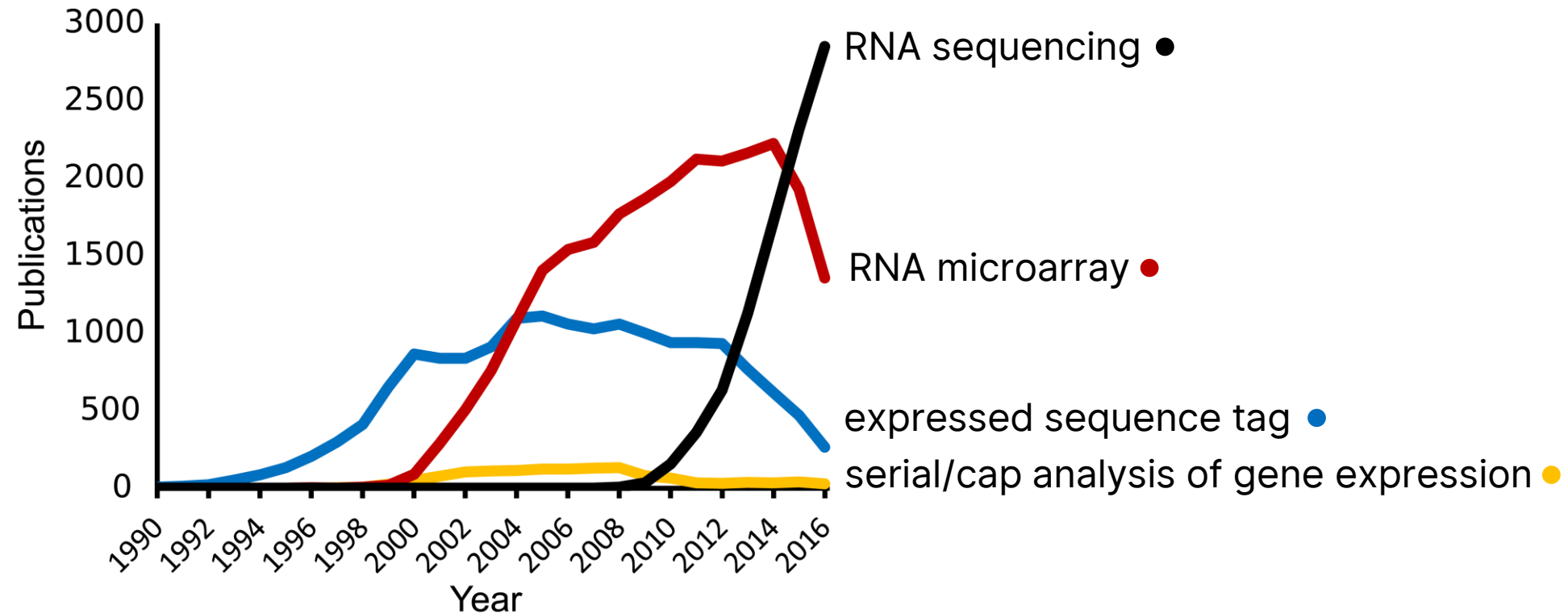
- The study of the transcriptome using high-throughput techniques.
- A snapshot in time of the total transcripts present in a cell.

Main aims of transcriptomics

- to classify all types of transcripts (mRNAs, ncRNAs, sRNAs...)
- to determine the transcriptional structure of genes (e.g. splicing patterns and post-transcriptional modifications)
- to quantify the changing expression levels of each transcript in different organisms/conditions/timepoints

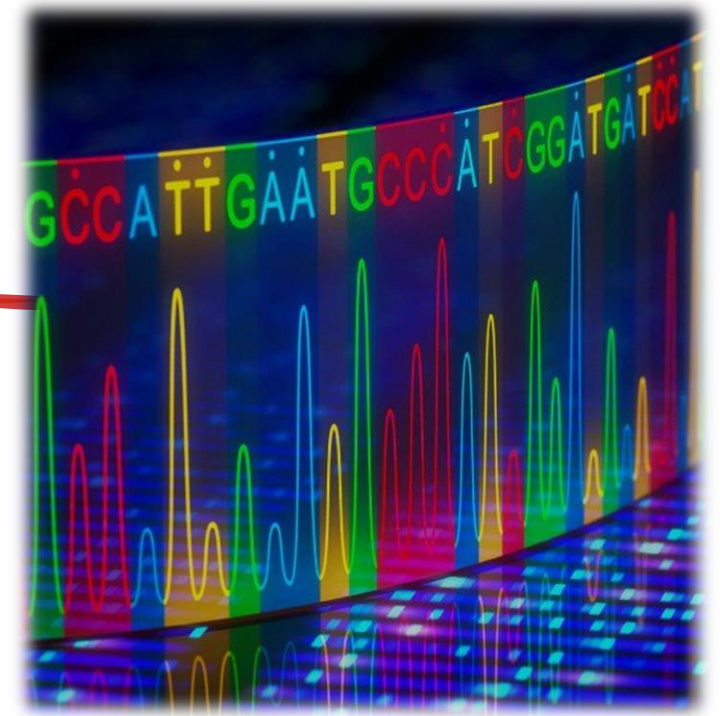


Transcriptomics method use over time



RNA sequencing (RNA-seq)

- A technique to measure the quantity of RNA molecules by using next-generation sequencing (NGS) technology.



Advantages

- wider dynamic range,
- more accuracy,
- higher sensitivity,
- ability to discover SNPs and rare mutations, previously unrecognized gene isoforms, microbial RNAs, and regulatory micro-RNAs

Disadvantages

- biases introduced during cDNA library construction and sequence alignment,
- lack of standardization between sequencing platforms and read depth,
- higher start-up costs

Single-cell RNA sequencing (scRNA-seq)

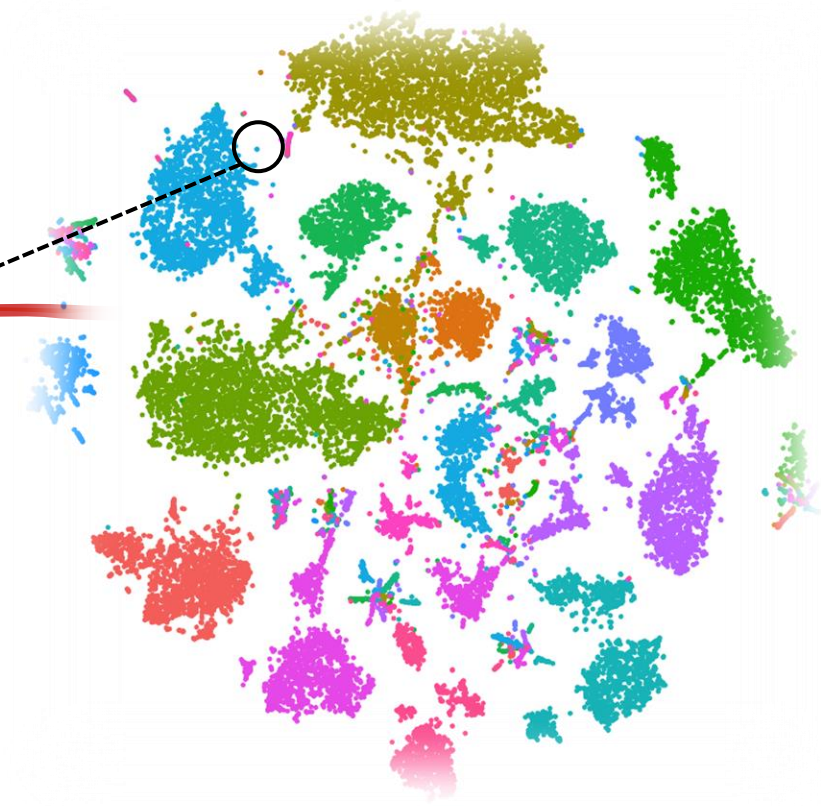
- A promising and highly trending approach to study the transcriptomes of individual cells.

Advantages

- can detect heterogeneity,
- can distinguish a small number of cells,
- can portray cell maps

Disadvantages

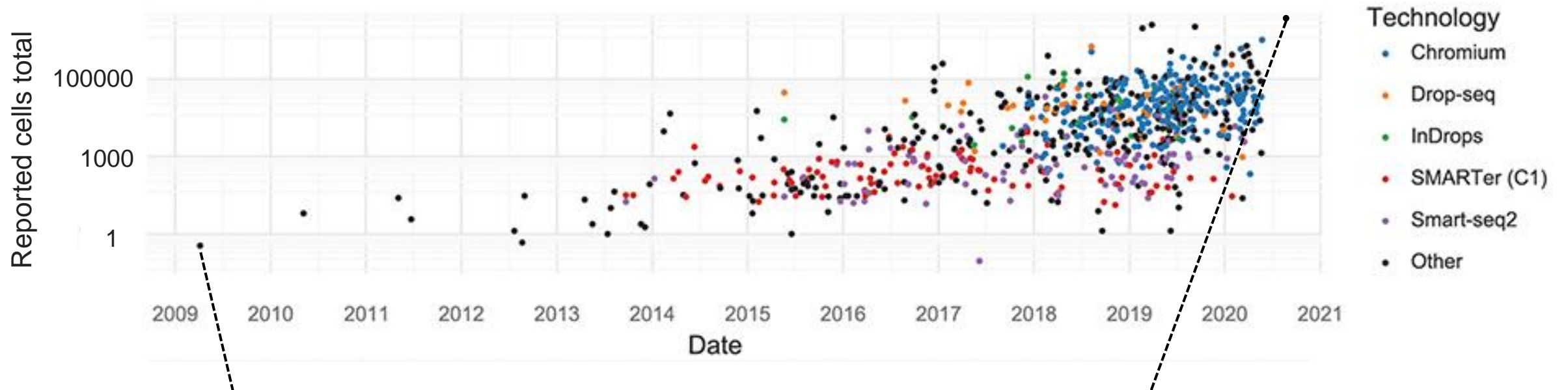
- bias of transcript coverage,
- low capture efficiency,
- higher level of technical noise,
- dropout events



Common Applications

- describing population diversity
- tracing cell lineages
- classifying cell types
- genomic profiling of rare cells

The number of cells measured in scRNA-seq experiments according to scRNA-seq protocols

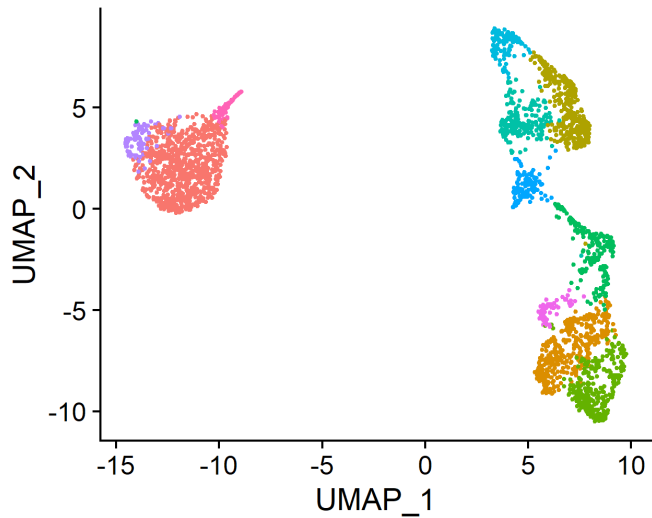


April 2009: 1 cell by Tang et al.

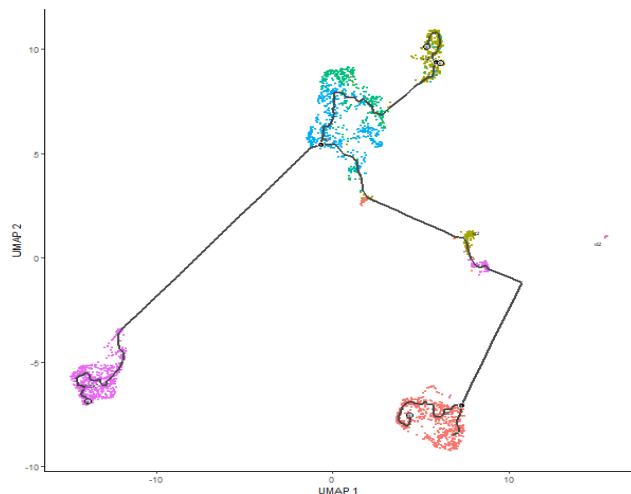
Tang, Fuchou, et al. "mRNA-Seq whole-transcriptome analysis of a single cell." *Nature methods* 6.5 (2009): 377-382.

November 2020: ~4 million cells by Shendure Lab

Cao, Junyue, et al. "A human cell atlas of fetal gene expression." *Science* 370.6518 (2020).

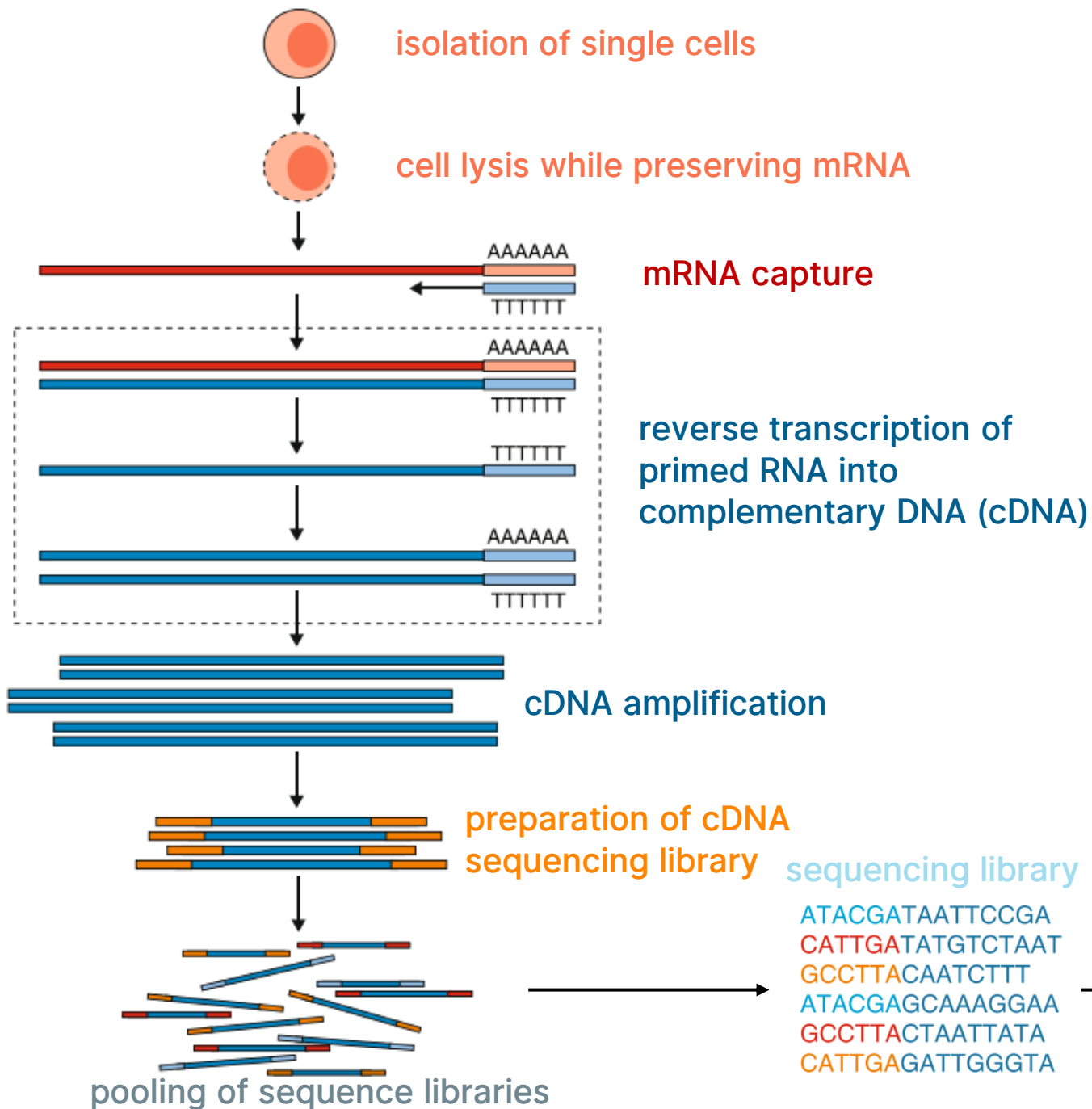


Why to perform scRNA-seq?



- to assess transcriptional similarities and differences within a population of cells
 - heterogeneity analysis
 - identification of rare cell populations
- to examine the single cells where each one is unique
 - individual T lymphocytes expressing highly diverse T-cell receptors
 - neurons within the brain
 - cells within an early-stage embryo
- to trace lineage and developmental relationships between heterogeneous cellular states
 - embryonal development
 - differentiation of cancer cells, myoblasts or stem cells

General workflow of scRNA-seq experiments

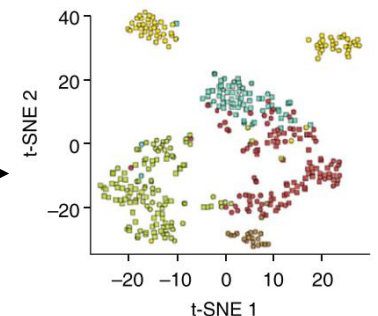


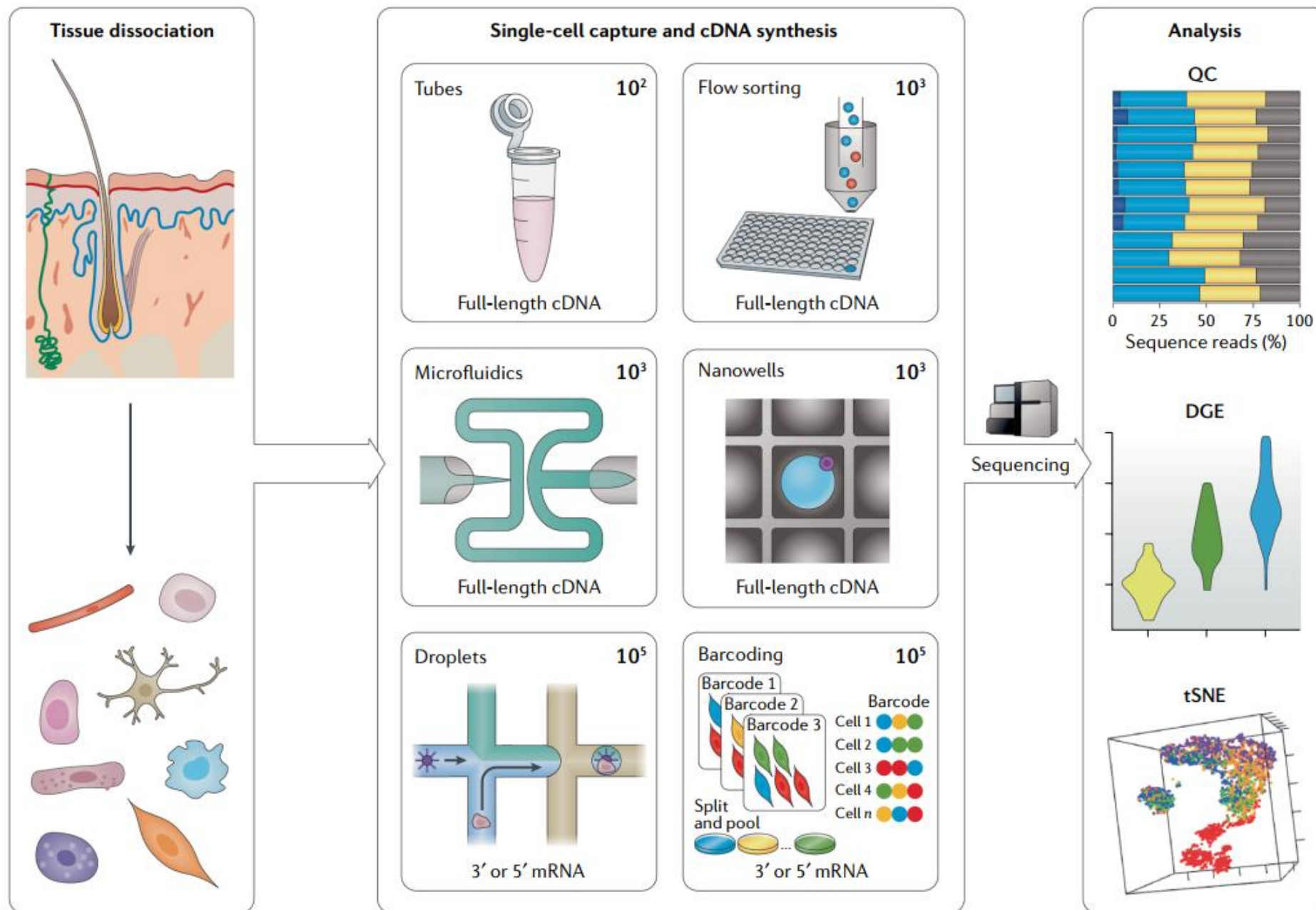
COUNT MATRIX

	Cell1	Cell2	...	CellN
Gene1	3	2	.	13
Gene2	2	3	.	1
Gene3	1	14	.	18
...
...
...
GeneM	25	0	.	0

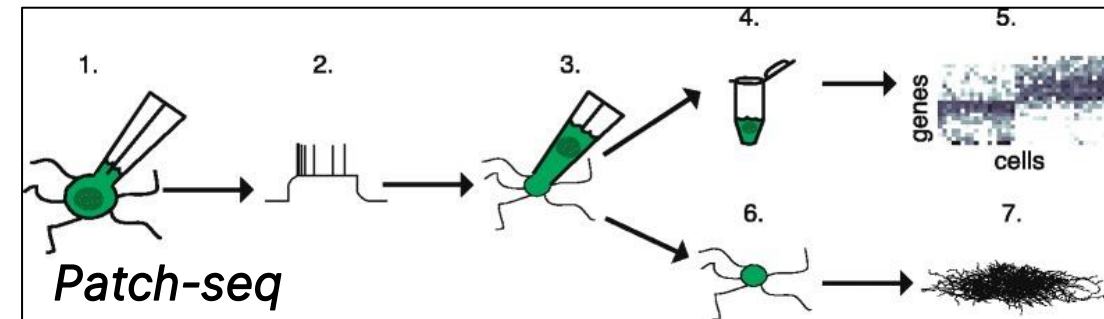
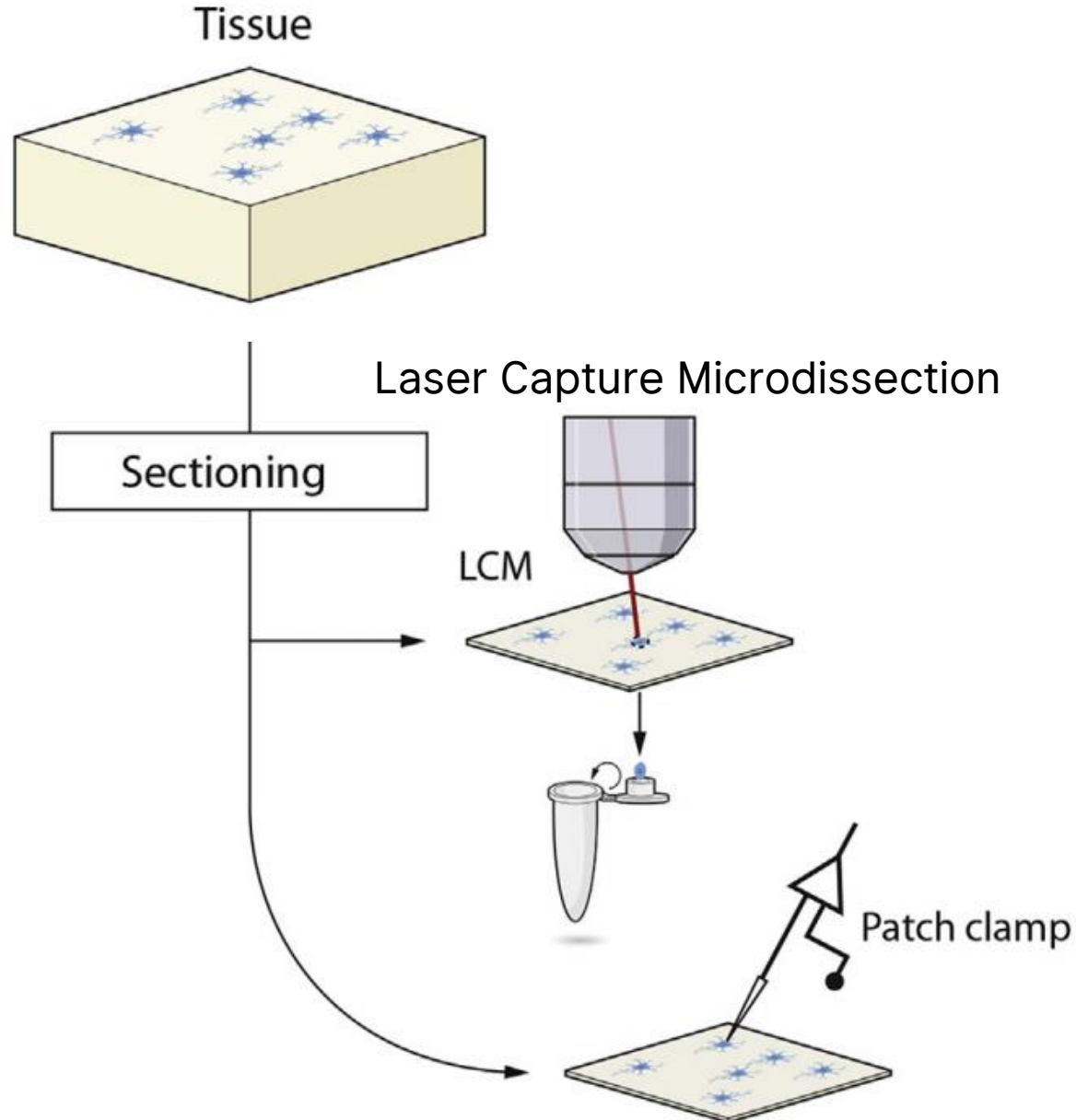
use of bioinformatic tools to assess quality and variability

use of specialized tools to analyse and present the data

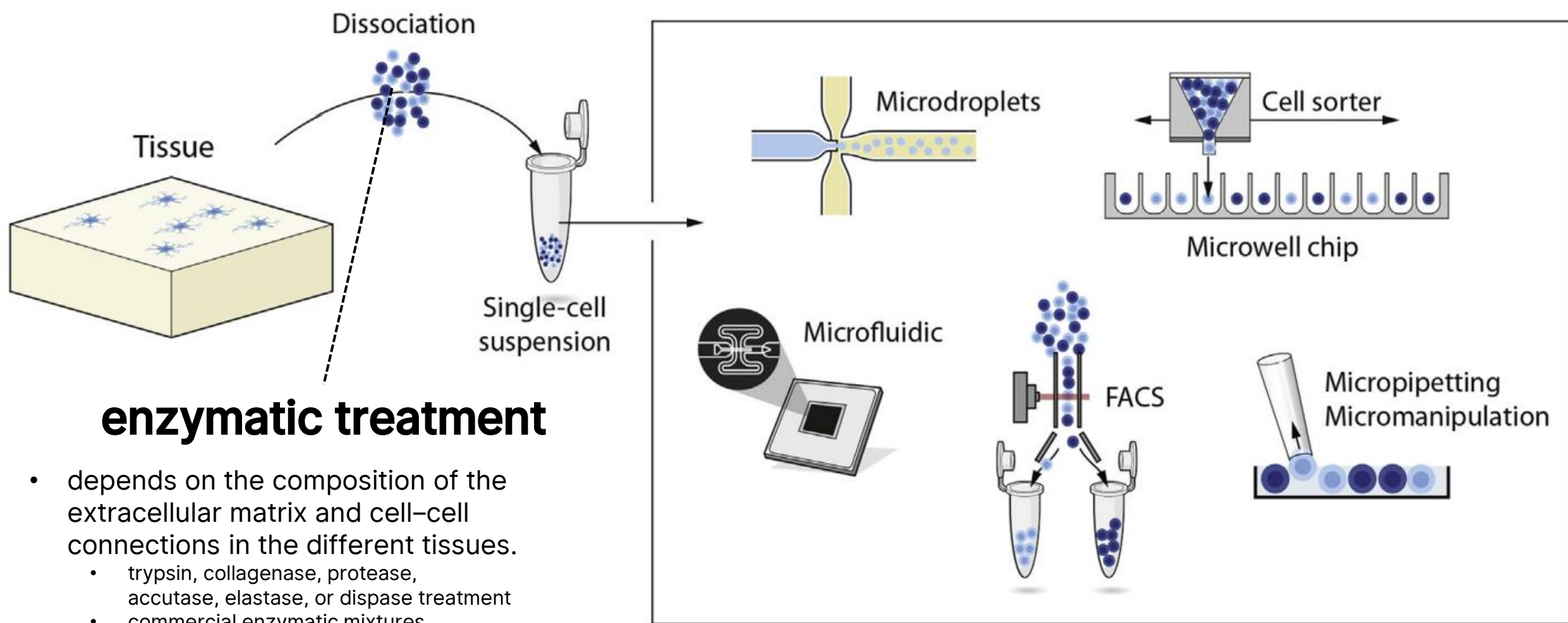




Isolation of single cells



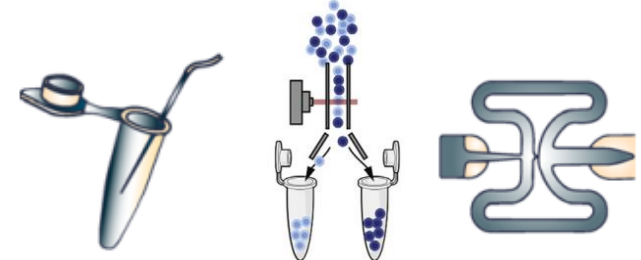
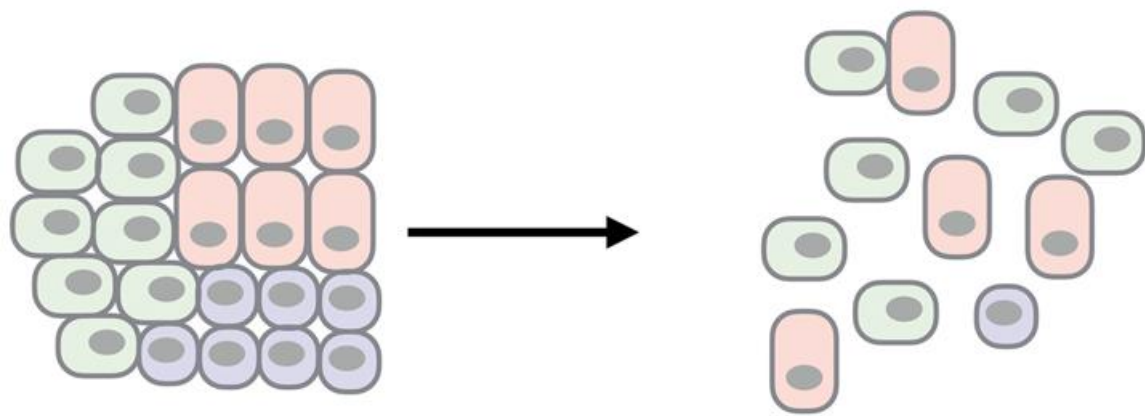
- an unbiased strategy to characterize and classify cell types
 - by integrating information about each cell's morphology, physiology, and gene expression into a common framework.
- can be used as a complementary method to 'annotate' cell type classification based primarily on scRNA-seq of dissociated neurons
 - 30-40 Patch-seq samples per day with 2-3 people
 - ~\$21/cell



enzymatic treatment

- depends on the composition of the extracellular matrix and cell-cell connections in the different tissues.
 - trypsin, collagenase, protease, accutase, elastase, or dispase treatment
 - commercial enzymatic mixtures (TrypLE Express and Liberase Blendzyme 3)
- cellular stressor for mature neurons
 - can degrade surface proteins
 - may alter gene expression.

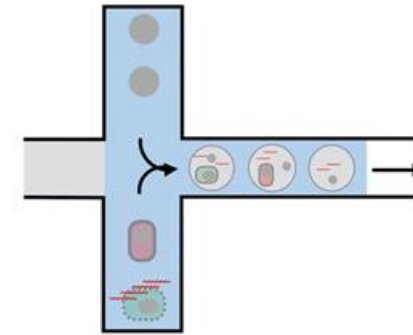
Isolation of single cells



more controlled, more flexible,
lower-throughput, more expensive



Well-based

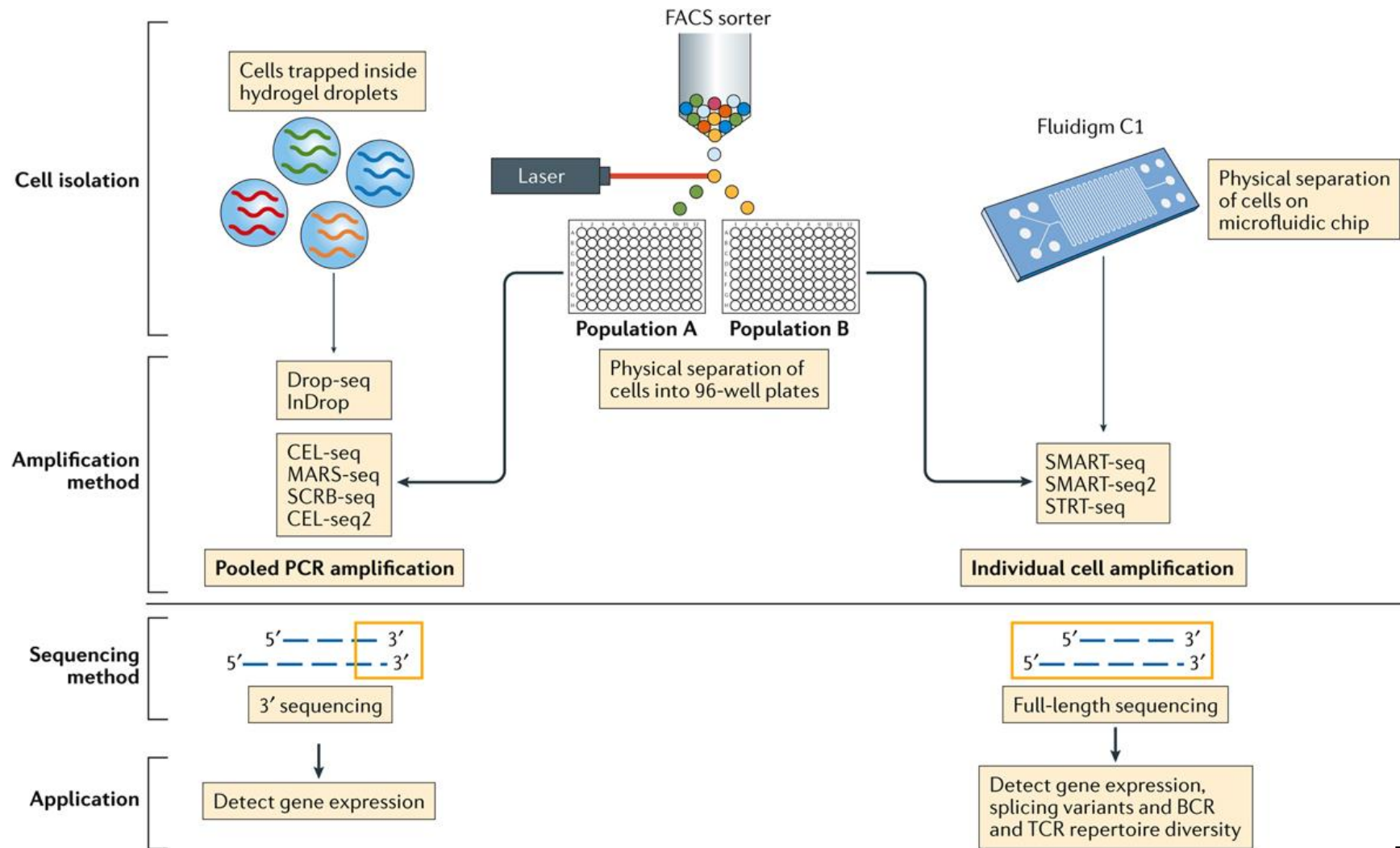


Droplet-based

higher possibility of doublets,
efficient, high-throughput, cheaper

Isolation of single cells

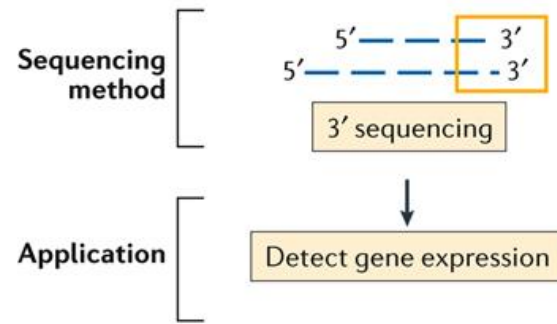
Single-cell Methods



Full-length sequencing

Tag-based (3' (or 5')-end) sequencing:

Nature Reviews | Immunology

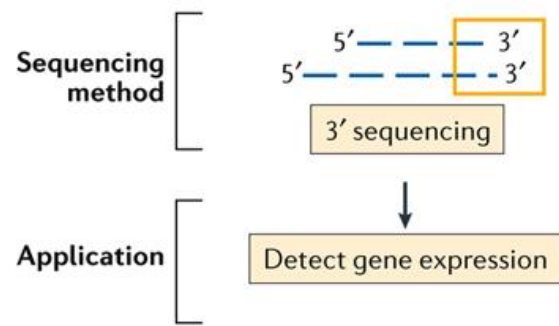


Tag-based (3' (or 5')-end) sequencing:

- **Barcode:**
identifies independent cells
- **UMI:**
identifies independent molecules

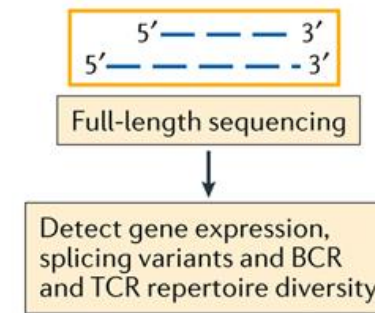
- **cell-specific barcodes:** non-random nucleotide sequences
 - determines which cell the read originated from
 - allowing for multiplexing the cDNA amplification
 - increasing the throughput of scRNA-seq library generation by one to three orders of magnitude

- **unique molecular identifiers (UMIs):** random nucleotide sequences
 - tag individual mRNA molecules
 - allow for the distinction between original molecules and amplification duplicates that derive from the cDNA or library amplification
 - allow more accurate quantification
 - used to collapse PCR duplicates



Tag-based (3' (or 5')-end) sequencing:

- enables the incorporation of cell-specific barcodes and **unique molecular identifiers (UMIs)**
- Larger number of cells sequenced allows better identity of cell type populations
- Cheaper per cell cost
- Best results with > 10,000 cells
- **ERCC spike-ins:** broad concentration of synthetic RNAs of various lengths and GC content,
 - often used to measure biases in RNA-seq experiments and for quality control or normalization.
 - to better account for technical variability due to random dropout events during library preparation.



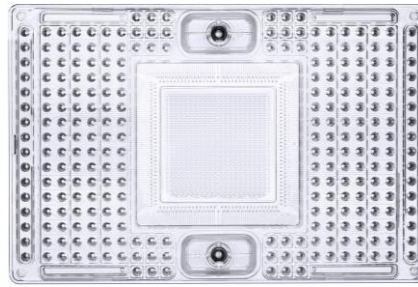
Full-length sequencing:

- tries to achieve a uniform gene body read coverage and increases the number of mappable reads.
 - detection of isoform-level differences
 - identification of allele-specific differences
- deeper sequencing of a smaller number of cells
- best for samples with low number of cells
- not possible to multiplex and pool all samples for one tube preparation of Illumina sequenceable libraries
- more expensive and troublesome
- not compatible with high-throughput methods
- no UMIs

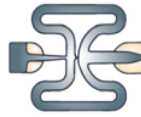


FLUIDIGM®

Fluidigm C1 System



- Developed in early 2000s
- 96/800 cells at once
- difficult to integrate with experiments that involve long processing times
- high cost of the microfluidic cartridges

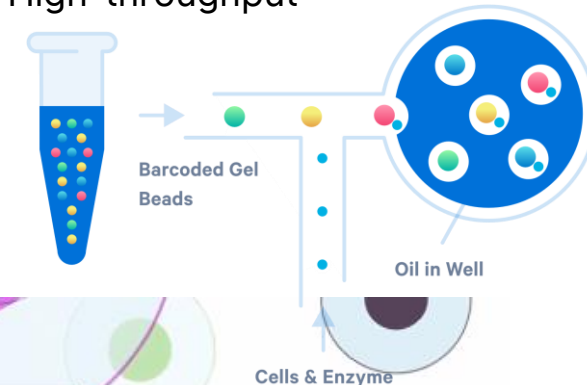


10x GENOMICS®

Chromium Single Cell Gene Expression Solution



- Developed in 2016.
- 1–8 samples in one run,
 - 10,000 cells per sample.
- High-throughput

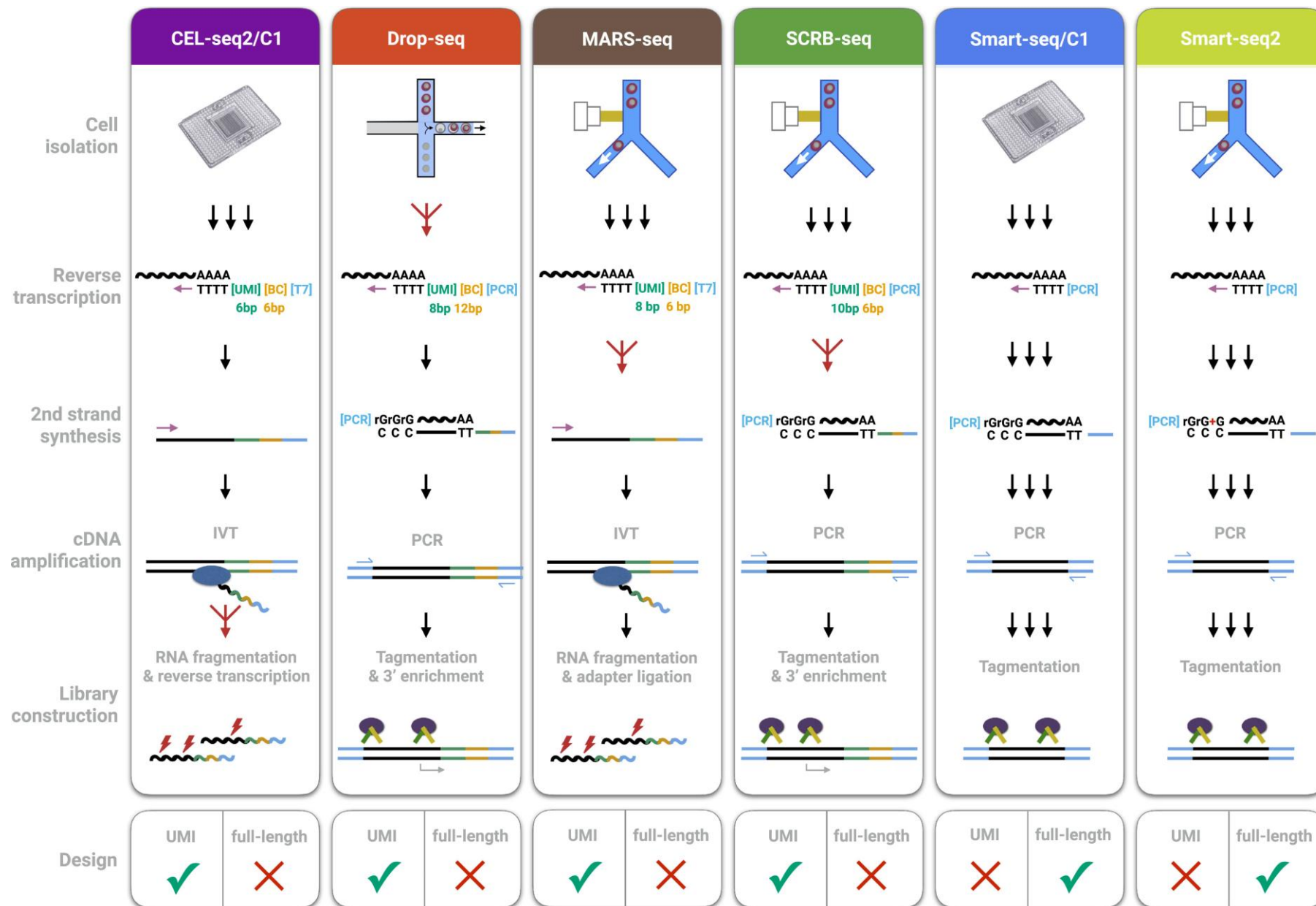


The key to our system is our ability to generate large numbers of GEMs.

VIDEO CAN BE FOUND IN RESOURCES LIST

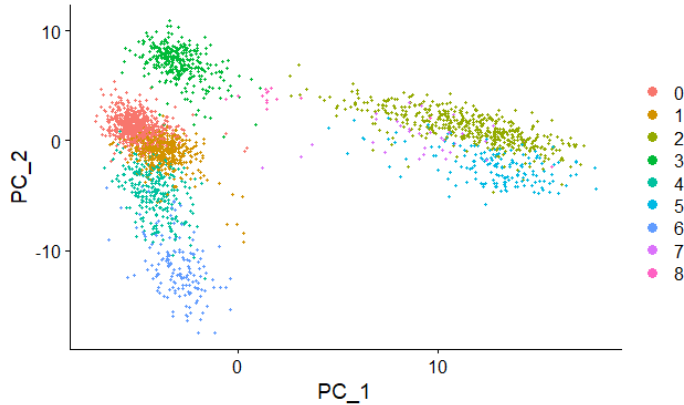
Single T Cell

Functionalized Gel Bead



Methods	Transcript coverage	UMI possibility	Strand specific	References
Tang method	Nearly full-length	No	No	Tang et al., 2009
Quartz-Seq	Full-length	No	No	Sasagawa et al., 2013
SUPeR-seq	Full-length	No	No	Fan X. et al., 2015
Smart-seq	Full-length	No	No	Ramskold et al., 2012
Smart-seq2	Full-length	No	No	Picelli et al., 2013
MATQ-seq	Full-length	Yes	Yes	Sheng et al., 2017
STRT-seq and STRT/C1	5'-only	Yes	Yes	Islam et al., 2011, 2012
CEL-seq	3'-only	Yes	Yes	Hashimshony et al., 2012
CEL-seq2	3'-only	Yes	Yes	Hashimshony et al., 2016
MARS-seq	3'-only	Yes	Yes	Jaitin et al., 2014
CytoSeq	3'-only	Yes	Yes	Fan H.C. et al., 2015
Drop-seq	3'-only	Yes	Yes	Macosko et al., 2015
InDrop	3'-only	Yes	Yes	Klein et al., 2015
Chromium	3'-only	Yes	Yes	Zheng et al., 2017
SPLiT-seq	3'-only	Yes	Yes	Rosenberg et al., 2018
sci-RNA-seq	3'-only	Yes	Yes	Cao et al., 2017
Seq-Well	3'-only	Yes	Yes	Gierahn et al., 2017
DroNC-seq	3'-only	Yes	Yes	Habib et al., 2017
Quartz-Seq2	3'-only	Yes	Yes	Sasagawa et al., 2018

Dimension Reduction Methods for scRNA-seq



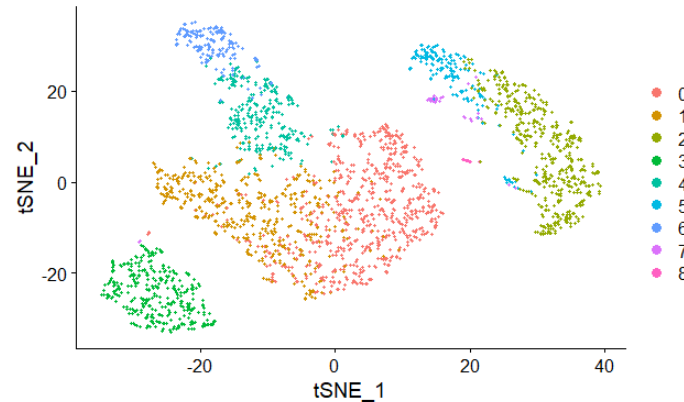
PCA (Principal Component Analysis)

LINEAR

- performs an orthogonal transformation of the original dataset to create a set of new, uncorrelated variables or principal components (linear combinations of variables in the original dataset.)

Pros: • highly interpretable and computationally efficient

Cons: • very inappropriate due to highly non-linear structure



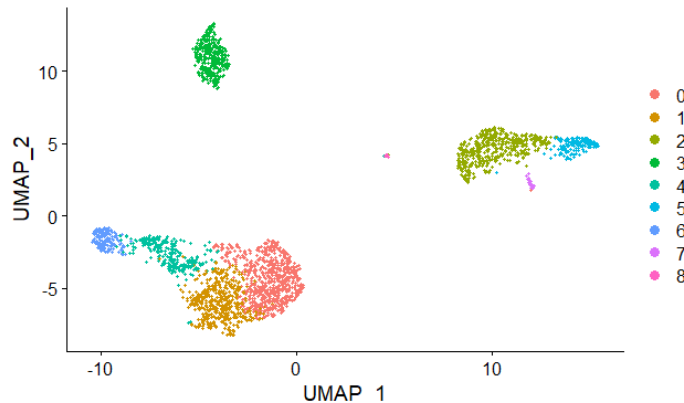
tSNE (t-Stochastic Neighbourhood Embedding)

LINEAR

- graph based and non-linear technique, projects high dimensional data onto 2D or 3D components

Pros: • designed to preserve the local structures

Cons: • stochastic method
• global structure not preserved
• only limited to data exploration or visualisation.
• computationally expensive



UMAP (Uniform Manifold Approximation and Projection)

NON-LINEAR

- graph based and principally similar to t-SNE where it constructs a high dimensional graph representation of the data, then optimizes a low-dimensional graph to be as structurally similar as possible.

Pros: • computationally efficient
• better preservation of the global structure
• can be used as a pre-processing step

Cons: • false detection of noise
• accuracy of global structure

END OF THE SESSION

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