Genome annotation: Independent study

Advisor : Dr. R. Taylor Raborn

**Dr. Volker Brendel** 

# Learning Goals

- What is genome annotation?
- How to annotate a genome?
- How to compare annotations?

### Tools used

Maker 2.31.6: Holt and Yandell (2011) MAKER2: an annotation pipeline and genome-database management tool for second-generation genome projects. BMC Bioinfo 12:491

Genometools: G. Gremme, S. Steinbiss and S. Kurtz. *GenomeTools*: a comprehensive software library for efficient processing of structured genome annotations. <u>IEEE/ACM Transactions on Computational Biology and Bioinformatics</u> 2013, 10(3):645–656

Parseval: Daniel S Standage, Volker P Brendel: ParsEval: parallel comparison and analysis of gene structure annotations. *BMC Bioinformatics* 2012, 13:187 doi:10.1186/1471-2105-13-187

Bedtools: Aaron R Quinlan, Ira M. Hall: BEDTools: a flexible suite of utilities for comparing genomic features. Bioinformatics (2010) 26 (6): 841-842.doi: 10.1093/bioinformatics/btq033

xGDBVm:

CEGMA: Genis Parra, Keith Bradnam and Ian Korf. <u>CEGMA: a pipeline to accurately annotate core genes in eukaryotic genomes.</u>" Bioinformatics, 23: 1061-1067 (2007)

Github

CEGMA: Genis Parra, Keith Bradnam and Ian Korf. <u>CEGMA: a pipeline to accurately annotate core genes in eukaryotic genomes."</u> Bioinformatics, 23: 1061-1067 (2007)

# Introduction:

## Need for genome annotation

- Genomes sequencing rate much higher than annotation
- Eukaryotic genome annotation much more complex
- Identification of genes and other features
- Many tools, many pipelines

### Gene Prediction

### Ab initio

- De novo
- Genome searched for protein coding genes
- Prokaryotic gene prediction simple
- Uses HMM
- SNAP

# **Empirical**

- Homology (Evidence based)
- Search genome for EST, protein alignment
- Dependent of assembly quality
- Matches: Complete or partial
- Genome Threader

### Daphnia pulex

- Crustacean found in ponds and lakes
- Standard organism for toxicity testing
- switch between clonal and sexual reproduction in response to environmental conditions
- 5191 scaffolds
- Genome size ~ 200 Mb
- 30,907 protein-encoding genes



# Method:

### Maker

- easy-to-use genome annotation pipeline
- de novo annotation of newly sequenced genomes
- updating existing annotations to reflect new evidence
- combine annotations, evidence, and quality control statistics for use with other GMOD programs like Gbrowse

### Maker

• Input:

Genome sequence

Transcript sequence evidence

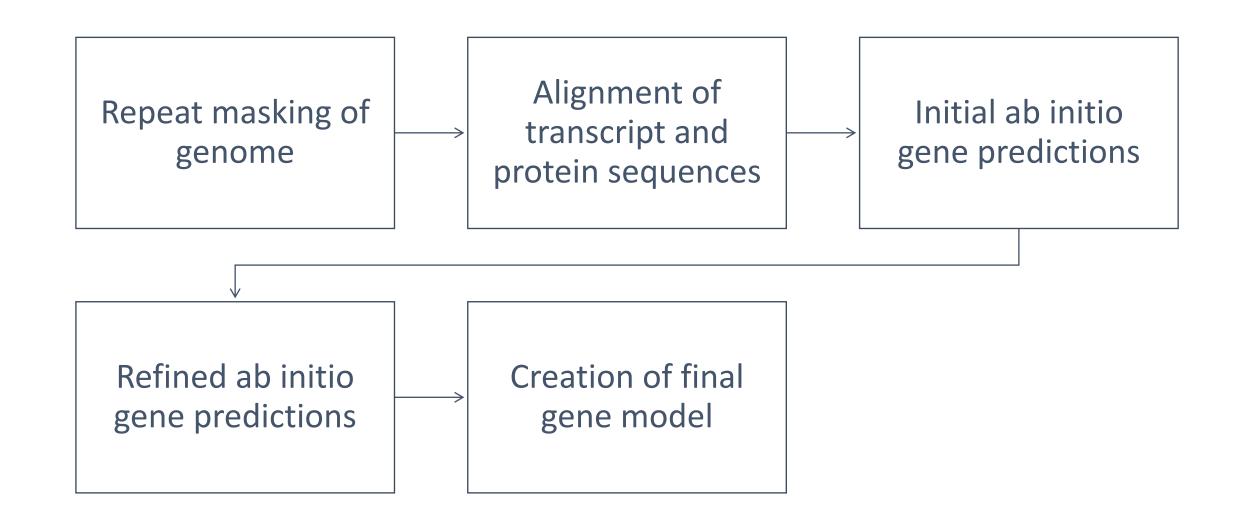
Protein sequence evidence

- Input files can be provided in several formats: FASTA, GFF
- Uses HMMs for gene prediction

# Control files

maker_exe.ctl	paths to required executables
maker_bopt.ctl	options for BLAST and Exonerate
maker_opt.ctl	Contains paths to input files and other options

# Maker pipeline



# Maker derived gene models

- Annotations with evidence is reported.
- BLAST alignment must have at least a corresponding gene predictor result
- Gene predictor result without evidence not reported
- AED score :amount of dissimilarity between evidence and annotation

### Result from Maker

GFF file for each chromosome/scaffold/contig Ab initio predictions Alignments of all transcripts and proteins MAKER gene models FASTA files with predicted transcript

## Merging the files:

### Gather the predicted proteins

cat ./\*/\*/\*.proteins.fasta > genome.predictedproteins.fasta

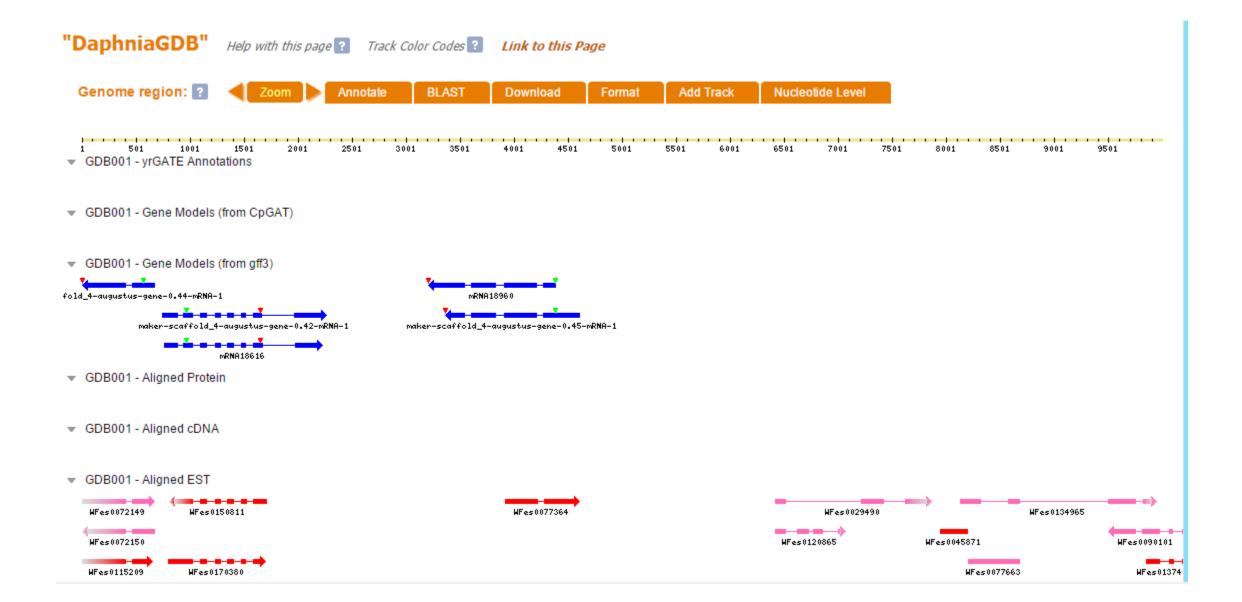
### Gather the predicted transcripts

cat ./\*/\*/\*.transcripts.fasta > genome.predictedtranscripts.fasta

### Gather the gff

gff3\_merge -d genome\_master\_datastore\_index.log

# Viewing the results:



# Results:

### Evaluation of annotation

**Gtstats** 

Parseval

Single exons BLAST

Comparison with other metazoan annotations

**CEGMA** 

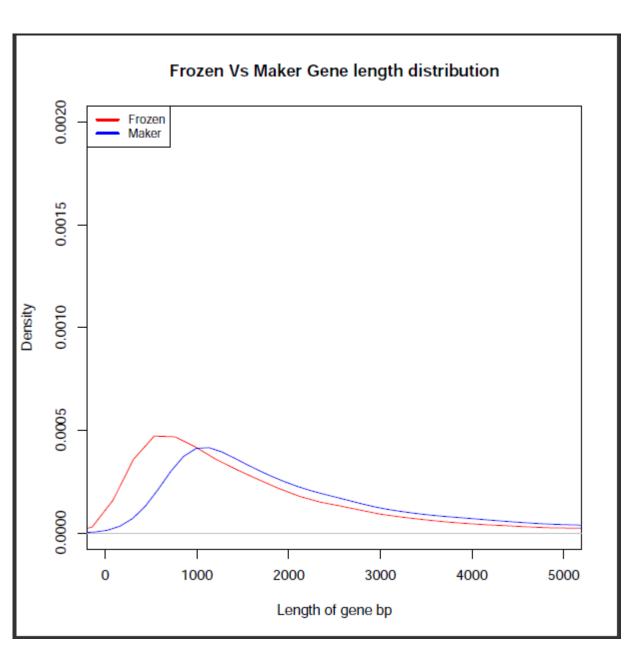
### Basic statistics:

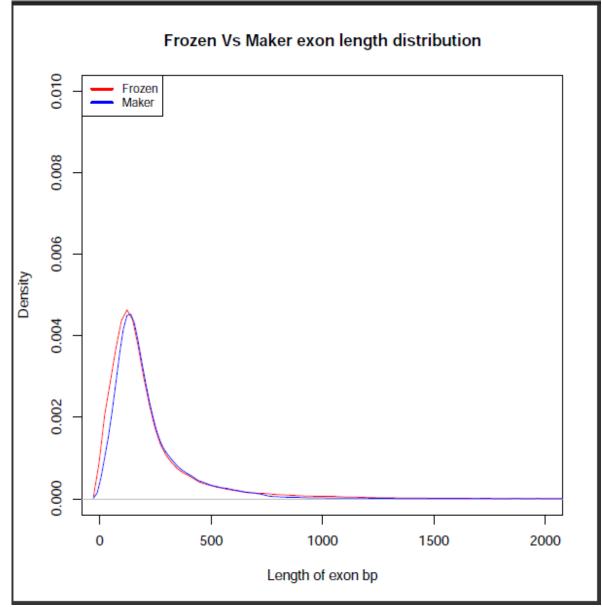
Number of	Frozen Gene Dataset	Maker annotation				
Genes	30776	11285				
Exons	145385	74868				
CDS	143218	72731				
Introns	114575	63630				
3' UTR	7961	7836				
5' UTR	7219	7026				

# Parseval comparison

Gene Loci					
Unique to FrozenGene	Unique to Maker				
17932	295				

	FrozenGene	Maker
Number of genes	30776	11285
Average per locus	1.11	0.4

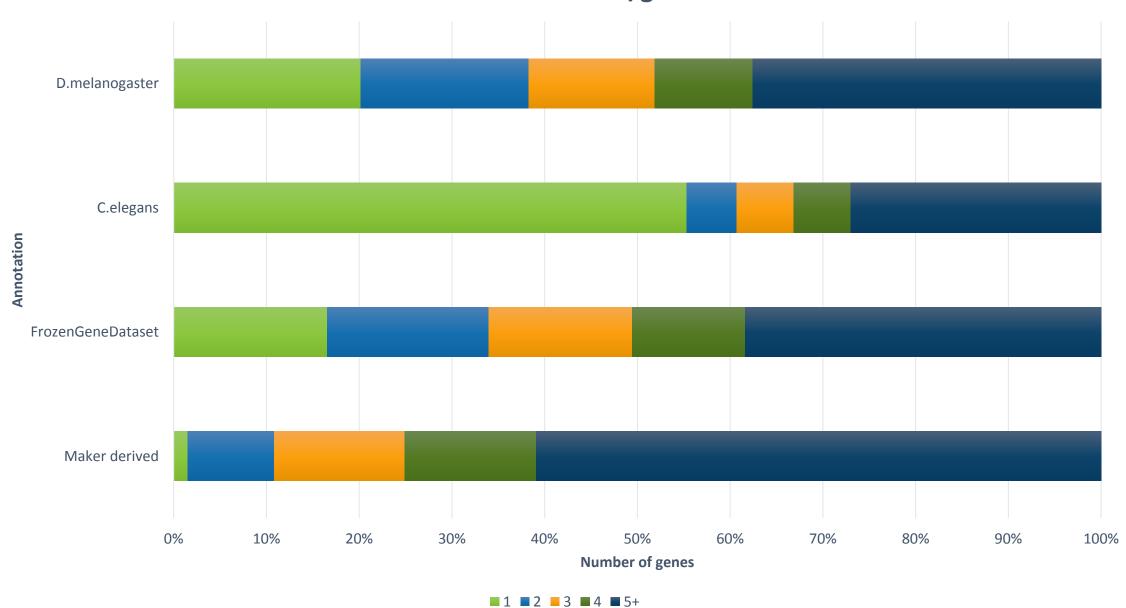


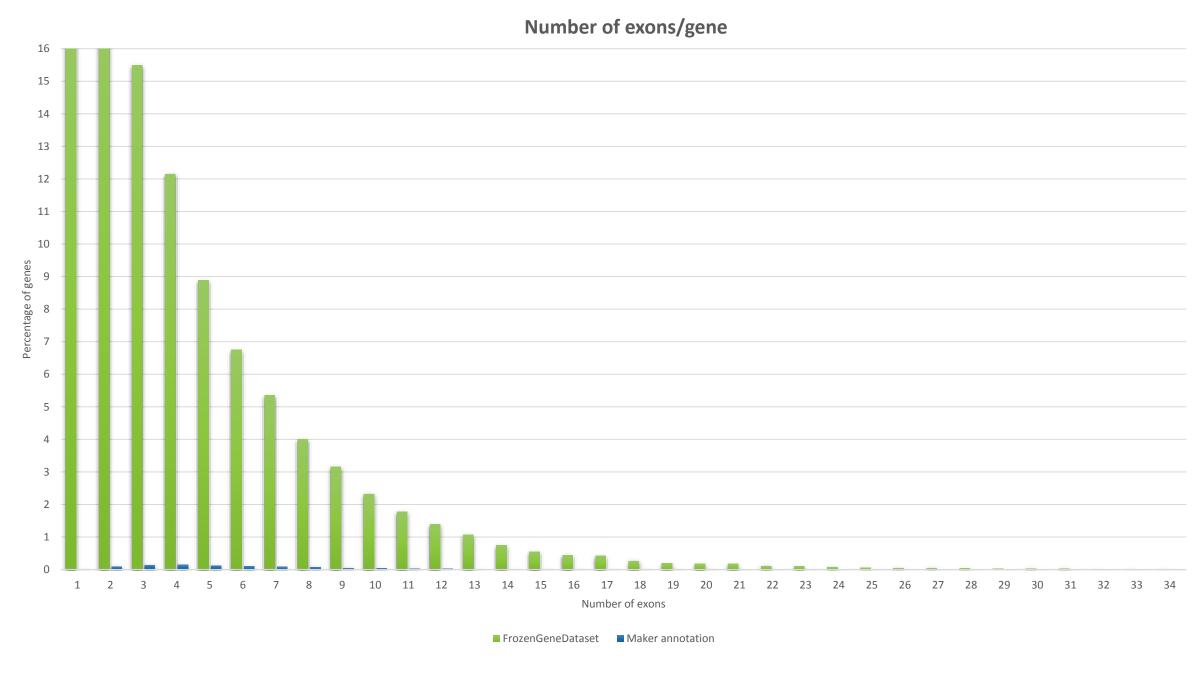


# No of exons/gene

Number of exon/gene	Frozen Gene Dataset	Maker annotation
One Exon	171 5090	
Two Exons	1055	4769
Three Exons	1584	4769
Four Exons	1604	3739
Five +	6871	11817

#### Number of exons/gene





J. F. Denton, J. Lugo-Martinez, A. E. Tucker, D. R. Schrider, W. C. Warren, and M. W. Hahn, "Extensive Error in the Number of Genes Inferred from Draft Genome Assemblies," PLoS Comp Biol, vol. 10, no. 12, p. e1003998, Dec. 2014.

Get single exonic genes from the gff3 file: R package GenomicFeatures (exons by 'gene')



Bedtools getfasta : fasta sequence



**BLASTx** 



Check for the single exonic gene in Maker gff3 : Bedtools intersect

## Example:

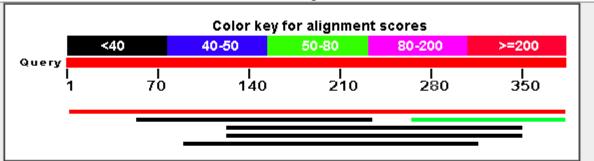
- Gene9711
- Single exonic gene
- On scaffold\_184 from 186739 to 187119
- SNAP derived gene

• One of the many single exonic genes with no evidence.

#### No putative conserved domains have been detected

#### Distribution of 6 Blast Hits on the Query Sequence





#### escriptions

#### Sequences producing significant alignments:

Select: All None Selected:0

AT	Alignments Download GenPept Graphics						
	Description			Query cover		Ident	Accession
	hypothetical protein DAPPUDRAFT 263274 [Daphnia pulex]	229	229	99%	2e-74	100%	gi 321455240 EFX66378.1
	hypothetical protein DAPPUDRAFT 251165 [Daphnia pulex]	57.0	57.0	30%	8e-08	64%	gi 321463868 EFX74880.1
	protein of hypothetical function UPF0052 and CofD [Weissella koreensis]	37.4	37.4	59%	1.7	31%	gi 493899121 WP 006844948.1
	hypothetical protein [Weissella koreensis]	36.2	36.2	59%	4.7	29%	gi 503755542 WP_013989618.1

### **CEGMA**

- CEGMA <- output.cegma.gff</li>
- The assembly was 96.37 % complete
- Bedtools to convert gff3 to bed format

- KOG1078.2 missing from Frozen gene dataset
- KOG1078, Vesicle coat complex COPI, gamma subunit [Intracellular trafficking, secretion, and vesicular transport]

# Questions?