

# Using RAMPAGE to identify and annotate promoters in insect genomes

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**Abstract.** Application of Transcription Start Site (TSS) profiling technologies, coupled with large-scale next-generation sequencing (NGS) has yielded valuable insights into the location, structure and activity of promoters across diverse metazoan model systems. In insects, TSS profiling has been used to characterize the promoter architecture of *Drosophila melanogaster* [1], and, shortly thereafter, to reveal widespread transposon-driven alternative promoter usage in *D. melanogaster* [2].

In this chapter we highlight the utility of one TSS profiling method, RAMPAGE (RNA annotation and mapping of promoters for analysis of gene expression), for the precise, quantitative identification of promoters in insect genomes. We demonstrate this using our tools GoRAMPAGE [3] and TSRchitect [4], providing details instructions with the aim of taking the user from raw reads to processed results.

**Keywords:** *cis*-regulatory regions, promoter architecture, transcription initiation, transcription start sites (TSSs)

## 1 Introduction

### 1.1 TSS Profiling Identifies Promoters at Genome-Scale

The promoter, defined in eukaryotes as the genomic region bound by RNA Polymerase II immediately prior to transcription initiation [5], is the site where regulatory signals unite to direct gene expression. The identification of promoter regions is a valuable step for understanding the *cis*-regulatory signals that are present in an organism, and is also important for genome annotation. However, despite the rapid accumulation of genome sequences across metazoan and arthropod diversity, accurate annotation of promoter regions remains sparse. This is because—absent empirically-defined information—precisely identifying

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sequence motifs that demarcate the promoter is unreliable. In contrast with current *in silico* approaches, direct mapping of TSSs identifies the location of the core promoter. Cap Analysis of Gene Expression (CAGE) [6], one of the first methods devised to identify 5'-ends of mRNAs at large-scale, involves selective capture of 5'-capped transcripts, first-strand reverse-transcription and ligation of a short oligonucleotide (CAGE tag). CAGE was initially utilized by the FANTOM (Functional Annotation of the Mammalian Genome) consortium to identify promoter architecture in human and mouse [7], providing the first glimpse of the global landscape of transcription initiation. At the onset of the NGS era, CAGE was coupled with massively-parallel sequencing to generate 5'-ends of mRNAs at substantially higher scale. This advance provided more extensive coverage of the expressed transcriptome, and provided increased sensitivity for quantitative measurements *i.e.* measurement of promoter activity.

## 1.2 Promoter Architecture of *Drosophila melanogaster*

Hoskins and colleagues [1] performed CAGE in *D. melanogaster* as part of the modENCODE consortium, identifying promoters at large-scale and characterizing the promoter architecture of an insect genome for the first time. Hoskins [1] indicated that TSS distributions at *Drosophila* promoters exhibit a range of shapes that can be generally grouped into two major classifications: *peaked* and *broad*. Peaked promoters have a single, major TSS position occupying a narrow genomic region, whereas broad promoters lack a single, major TSS and contain TSSs across a wider region [8, 9]. The authors also showed a strong association between promoter class and motif composition (consistent with previous findings [8, 10]). Peaked promoters were associated with positionally-enriched *cis*-regulatory motifs including TATA, Initiator (Inr) and DPE, while broad promoters contained an enrichment of less-well characterized motifs, including *Ohler6* and *Ohler7* [11]. The existence of two promoter classes appears to be conserved among metazoans, and has been reported (using TSS profiling methodologies) in insects, cladocerans [12], fish [13] and mammals [14, 9].

## 1.3 Promoter Structure of Insects

Beyond *D. melanogaster*, few investigations have utilized TSS profiling in insect genomes. As a consequence, what is known about promoter architecture in insects is largely restricted to the *Drosophila* genus. As part of the modENCODE effort, CAGE was performed in multiple tissues and developmental stages of the *Drosophila pseudoobscura*. TSSs were found to be highly similar between species: more than 80% of TSSs (81%) of aligned, CAGE-identified TSSs from *D. pseudoobscura* were positioned within 20nt of their counterparts in *D. melanogaster*. An enrichment of the CA dinucleotide was detected at the TSS ( $[-1, +1]$ ), and the motifs corresponding to TATA, Inr and DPE were positioned at the same locations relative to the TSS in both species. The one other insect species for

which TSS profiling has been applied is the Tsetse fly (*Glossina morsitans morsitans*) [15]. Using TSS-seq (specifically Oligo-capping; for details see [16]), the authors identified 3134 mapping to 1424 genes. The authors found a preference for CA and AA dinucleotides at the TSS, and observe the major core promoter elements observed in *Drosophila*: TATA, Inr, DPE, in addition to MTE (Motif Ten Element). As in *D. melanogaster*, peaked promoters were more likely to contain TATA and Inr than broad promoters. While the taxonomic sampling of species for TSS profiling has been limited, the existing studies are sufficient to provide a general picture of insect promoter architecture. A major demarcation between the promoter architecture of insects and mammals appears to be the large fraction of mammalian promoters found in CpG islands [15]. CpG island promoters (CPIs) form the largest class of promoter in mammals [17]; by contrast, CPIs are not known to exist as a class in invertebrates.

#### 1.4 Paired-end TSS Profiling with RAMPAGE

The most recent major methodological advance in TSS Profiling is RAMPAGE (RNA Annotation and Mapping of Promoters for the Analysis of Gene Expression) [2, 18]. RAMPAGE is a protocol for 5'-cDNA sequencing that combines cap trapping and template-switching with paired-end sequence information. A key advantage of generating paired-end sequence is transcript connectivity, which provides a direct link between a given 5'-end and its associated mRNA molecule [2]. Because short or spurious RNAs are found within the transcriptome, transcript connectivity allows the TSSs (and thus promoters) of full-length mRNAs to be unambiguously identified, which benefits genome annotation and improves interpretation of transcript species. Batut and colleagues [2] generated libraries from total RNA isolated from 36 stages across the life cycle of *D. melanogaster* providing a comprehensive gene expression and promoter atlas for fruit fly and in the process demonstrating the utility of RAMPAGE. RAMPAGE is currently being applied as part of the latest iteration of ENCODE to identify promoters in human, but as of this writing it has not been applied to any non-*Drosophila* insect model system. In anticipation of the future application of TSS profiling into other insect model systems here we provide a documented protocol for the computational processing RAMPAGE data, using selected libraries from Batut *et al.* [2]. This method will consist of two parts: first, we will process, filter and align the sequenced RAMPAGE libraries to the *D. melanogaster* genome. Second, we will identify TSSs and promoters from the aligned sequences and associate them with coding regions. In closing, we will consider further applications of this data and discuss the utility of reproducible workflows in bioinformatic analysis.

## 2 Materials

The analyses described herein require a workstation capable of doing modern bioinformatics, including a reasonably-appointed laptop. An intermediate understanding of the Linux/Unix command line will be extremely useful, although

we make efforts to explain the procedures with clarity. In addition, it will likely be necessary for the participant to have superuser privileges on the machine. If you do not have a machine (or have access to one) that meets these requirements, it is recommended that you consider cloud-based cyberinfrastructure, including Amazon Web Services (AWS; <https://aws.amazon.com/>) or CyVerse (<http://www.cyverse.org/>) [19]. The former is a well-known pay-per-use solution, while the latter is an NSF-funded resource that makes compute allocations freely available to the public.

## 2.1 Hardware

1. x86-64 compatible processors
2. At least 8GB RAM
3. 30GB+ hard disk space

## 2.2 Operating System

- 64 bit Linux (preferred) or Mac OS X (with Command Line Tools from XCode)

## 2.3 Software

Below is a list of the software packages required for this demonstration (*see Note 1*).

### Sequence retrieval

1. SRA Toolkit [20] (<https://www.ncbi.nlm.nih.gov/sra/docs/toolkitsoft/>)

### GoRAMPAGE

1. GoRAMPAGE [3] (<https://github.com/brendelGroup/GoRAMPAGE>)
2. fastq-multx [21] (<https://github.com/brwnj/fastq-multx/blob/master/README.md>)
3. FASTX-Toolkit [22] ([http://hannonlab.cshl.edu/fastx\\_toolkit/Index.html](http://hannonlab.cshl.edu/fastx_toolkit/Index.html))
4. TagDust2 [23] (<https://sourceforge.net/projects/tagdust/>)
5. Samtools [24] (<http://www.htslib.org/doc/samtools.html>)
6. STAR [25] (<https://github.com/alexdobin/STAR>)

### TSRchitect

1. R (v. 3.4 and up) [26] (<https://www.r-project.org/>)
2. Bioconductor (v. 3.5 and up) [27] (<http://bioconductor.org/>)
3. TSRchitect [4] (<http://bioconductor.org/packages/release/bioc/html/TSRchitect.html>)
4. Various R package dependencies (*see Methods*)

## 2.4 Online Appendix

We created an online appendix to serve as a companion to this chapter, which contains both scripts and select files to assist you in completing this tutorial. Please find the repository at [https://github.com/rtraborn/MMB\\_appendix](https://github.com/rtraborn/MMB_appendix) (*see Note 2*).

## 130 2.5 Installation of R packages

131 For installation of the software listed above, please follow the instructions pro-  
 132 vided by each respective package. Part of our analysis will require the use of R  
 133 packages found in the Bioconductor suite [27]. To install Bioconductor, please  
 134 type the following from an R console:

```
135 source("https://bioconductor.org/biocLite.R")
136 biocLite()
```

137 We will use the R package *TSRchitect* to identify promoters from aligned RAM-  
 138 PAGE libraries. Prior to running the analysis, it will be necessary to install a  
 139 series of prerequisite packages to *TSRchitect* from Bioconductor. Please install  
 140 these packages as follows (as before, from an R console):

```
141 source("https://bioconductor.org/biocLite.R")
142 biocLite(c("AnnotationHub", "BiocGenerics", "BiocParallel",
143 "ENCODEExplorer", "GenomicAlignments", "GenomeInfoDb",
144 "GenomicRanges", "IRanges", "methods",
145 "Rsamtools", "rtracklayer", "S4Vectors",
146 "SummarizedExperiment"))
```

147 To install *TSRchitect*, please type the following from an R console:

```
148 source("https://bioconductor.org/biocLite.R")
149 biocLite("TSRchitect")
```

150 Finally, please confirm that *TSRchitect* has been installed correctly by loading  
 151 it from your R console as follows:

```
152 library(TSRchitect) #installing TSRchitect
```

## 153 3 Methods

### 154 3.1 Retrieving the RAMPAGE sequence data from NCBI

155 To begin our analysis, we must download the RAMPAGE data to our worksta-  
 156 tion. We will utilize tools provided by the SRA Toolkit, which should already  
 157 be installed on your machine (see **Materials**). The command *fastq-dump* al-  
 158 lows one to directly retrieve data from the GEO database using the appropriate  
 159 identifier(s). While there are 36 RAMPAGE libraries in the Batut *et al.* pa-  
 160 per, we will select a subset of these to analyze here. We will compare samples  
 161 from selected embryonic (E01h-E03h) and larval (L1-L3) tissues, representing  
 162 the beginning and end of embryonic development. For more information about  
 163 the experiment and the available RAMPAGE libraries, please see the following  
 164 link: <https://www.ncbi.nlm.nih.gov/Traces/study/?acc=SRP011193>.

165  
 166 First, let's proceed with downloading the libraries from early embryonic tis-  
 167 sues (see **See Note 3**). We will make a new folder (entitled "*fastq\_files/*")  
 168 to house these files.

```

169 mkdir fastq_files
170 cd fastq_files
171
172 fastq-dump --split-files SRR424683
173 fastq-dump --split-files SRR424684
174 fastq-dump --split-files SRR424685

```

175 We continue by downloading the data from late larval tissues.

```

176
177 fastq-dump --split-files SRR424707
178 fastq-dump --split-files SRR424708
179 fastq-dump --split-files SRR424709
180
181
182

```

183 Once the download of the aforementioned files are complete, you should see a  
 184 total of 12 (6  $\times$  2) separate fastq files in your current working directory:

```

185 ls -l *.fastq | wc -l
186 cd ..

```

### 187 3.2 Creating symlinks to the files

188 Our workflow expects fastq files that have the format “\*.R1/R2.clipped.fq”.  
 189 Rather than rename them, we can simply create brand new symbolic links (sym-  
 190 links) to the files, as follows:

```

191 cd ..
192 mkdir -p output/reads/clipped
193 cd output/reads/clipped
194
195 #embryonic libraries
196 ln -s ../../../../fastq-files/SRR424683_1.fastq E01h.R1.clipped.fq
197 ln -s ../../../../fastq-files/SRR424683_2.fastq E01h.R2.clipped.fq
198 ln -s ../../../../fastq-files/SRR424684_1.fastq E02h.R1.clipped.fq
199 ln -s ../../../../fastq-files/SRR424684_2.fastq E02h.R2.clipped.fq
200 ln -s ../../../../fastq-files/SRR424685_1.fastq E03h.R1.clipped.fq
201 ln -s ../../../../fastq-files/SRR424685_2.fastq E03h.R2.clipped.fq
202
203 #larval libraries
204 ln -s ../../../../fastq-files/SRR424707_1.fastq L1.R1.clipped.fq
205 ln -s ../../../../fastq-files/SRR424707_2.fastq L1.R2.clipped.fq
206 ln -s ../../../../fastq-files/SRR424708_1.fastq L2.R1.clipped.fq
207 ln -s ../../../../fastq-files/SRR424708_2.fastq L2.R2.clipped.fq
208 ln -s ../../../../fastq-files/SRR424709_1.fastq L3.R1.clipped.fq

```

```

209 ln -s ../../../../fastq-files/SRR424709_2.fastq L3.R2.clipped.fq
210
211 cd ../../.. #returning to the output directory

```

### 212 3.3 Downloading genomic data from *D. melanogaster*

213 Now that we have the fastq files from the RAMPAGE libraries downloaded and  
 214 named appropriately, we now must retrieve the genome assembly and rRNA  
 215 sequences from *D. melanogaster*. The genome assembly is required for aligning  
 216 the RAMPAGE reads, and the rRNA sequences are required to filter out match-  
 217 ing reads in the sequenced RAMPAGE libraries, since our sample is intended  
 218 to contain only capped RNA transcripts. Please download the rRNA sequences  
 219 from the link we provide below. These sequences were retrieved separately from  
 220 Genbank at the NCBI database.

221  
 222 To retrieve the genome assembly from the ENSEMBL database, please do the  
 223 following:

```

224 mkdir genome
225 cd genome
226 wget ftp://ftp.ensembl.org/pub/release-78/fasta/drosophila_melanogaster/dna/Drosophila_m
227 #uncompressing the file
228 gzip -d Drosophila_melanogaster.BDGP5.dna.toplevel.fa.gz
229 cd ..

```

230 Please navigate to the rRNA file "Dmel\_rRNA.fasta" found in the Appendix.

```

231 head -n 3
232 >ref|NR_133562.1| Drosophila melanogaster 28S ribosomal RNA (28SrRNA:CR45844), rRNA
233 TTATATACAACCTCAACTCATATGGGACTACCCCTGAATTGAAGCATATTAATTAGGGGAGGAAAAGAA
234 ACTAACAAGGATTTTCTTAGTAGCGGCGAGCGAAAAGAAAACAGTTCAGCACTAAGTCACTTTGTCTATA

```

### 235 3.4 Filtering and alignment of RAMPAGE reads using 236 GoRAMPAGE

237 At this stage we are ready to commence with the rRNA filtering and alignment  
 238 of the RAMPAGE libraries. We will use GoRAMPAGE, a tool we developed,  
 239 to perform these tasks in a concerted workflow. GoRAMPAGE runs TagDust  
 240 [23] to remove rRNA and low-complexity reads, and uses STAR [25] to align  
 241 RAMPAGE (or other paired-end) reads to a given genome assembly.

242 **Setting up the GoRAMPAGE job.** Please refer to the script "GoRAMPAGE\_script\_MMB.sh"  
 243 and (using a text editor) provide the appropriate paths to the genome assembly,  
 244 output directory (see above) and rRNA sequences (see **Note 4**). GoRAMPAGE  
 245 jobs can optionally be run in parallel (see **Note 5**). The script can be executed  
 246 as follows:

```

247 #vi GoRAMPAGE_script_MMB.sh #updating with a text editor
248 ./GoRAMPAGE_script_MMB.sh

```

249 If everything is working correctly you should start to see the results of the job  
 250 being written to the file "errScript". You can inspect the progress during the  
 251 run using the *less* command.

```

252 less -S errScript

```

253 Should the run fail before completion, any associated error messages will be  
 254 printed to the errScript file. Once the job is complete, you should see the message  
 255 "GoRAMPAGE job is complete!" appear on the command-line terminal.

256 **Inspecting the rRNA filtering results.** To evaluate the results from Step  
 257 3 (rRNA filtering), please navigate to the top level of the "output" directory  
 258 and open the file "LOGFILES". You'll see the recorded progress of the program  
 259 Tagdust and a record of the results. We notice that (for the L3h library) 1046448  
 260 of reads (78.1%) were "extracted", meaning that slightly more than 20% of  
 261 reads were removed because of matches with ribosomal sequences. The removed  
 262 reads from all libraries are found in the "dusted\_discard" directory, and the  
 263 extracted reads are found in the current directory. Due to their sheer abundance  
 264 within cells, ribosomal RNA sequences are an inevitable contaminant within TSS  
 265 profiling libraries. For analysis purposes, it is important that these sequences be  
 266 removed, which is what has been completed here.  
 267 Since this step was conducted appropriately, we can proceed to the next step.

268 **Evaluating the alignments.** The folder "alignments/" in your GoRAMPAGE  
 269 output folder will now contain 6 .bam files, each representing the distinct RAM-  
 270 PAGE libraries selected for our analysis. Typing "ls -l" from the command line  
 271 will show that these files are symlinks to the original alignment files found  
 272 in the "STARoutput/" directory. "STARoutput/", as its name suggests, con-  
 273 tains the output from the STAR alignment, and this includes the alignment files  
 274 "\*.sortedByCoord.out.bam", and four additional log files. The files with the suf-  
 275 fix "\*.STAR.Log.final.out" each contain a summary of the alignment, such as  
 276 the number of input reads, the percentage of uniquely-mapped reads and the  
 277 percentage of unmapped reads. An inspection of these log files indicates that  
 278 the alignments have similar mapping rates ( 70-80%), a reasonable outcome for  
 279 our purposes.

280  
 281 Now that our RAMPAGE libraries are filtered and aligned, we can commence  
 282 with the second half of our analysis.

### 283 3.5 Promoter identification from aligned RAMPAGE libraries

284 We can now use the prepared alignment files to identify TSSs and promoters from  
 285 the selected RAMPAGE libraries. There are currently several tools available



for this purpose. *CAGEr*, developed by Haberle [28], was utilized to perform TSS identification as part of the FANTOM5 efforts. We will use *TSRchitect* in this demonstration, since it was specifically designed to analyze paired-end TSS profiling datasets, and also because it is more flexible with respect to model system (*i.e.* it does not require a corresponding *BSTGenome* package). The latter feature will be helpful when analyzing the non-*D. melanogaster* TSS profiling datasets that we expect to be generated in the near future.

**Setting up the Analysis.** *TSRchitect*, the package we'll use for this analysis, is an R package available in the Bioconductor suite of genomics tools [27]. It makes use of existing packages and data structures within this environment, where available, to identify promoters from sequence alignments. Since you have already installed *TSRchitect* and its dependencies (see section 2.3), we are set to proceed.

There are two general ways one can choose to run *TSRchitect*. The first is interactively *i.e.* typing the instructions directly into an R console. While this is a perfectly acceptable way to run analyses using package, for larger jobs it will likely be more efficient (and likely more reproducible) to run a dedicated R script. We have provided a sample script "`MMB_chapter_TSRchitect.R`" to make it easier for you to set up an R script. In the section to follow, we will go through the output of the analysis. For further details on how to use *TSRchitect*, please see its documentation at its Bioconductor page found here: <https://www.bioconductor.org/packages/release/bioc/html/TSRchitect.html>.

**Running the Analysis.** To run *TSRchitect* using the batch script, provide full paths for the variables "`BAMDIR`" and "`DmAnnot`" in the script provided (see **Note 6**). *BAMDIR* should be a path to the subdirectory "`alignments/`" in RAMPAGE output directory you specified earlier, and *DmAnnot* should be a full path to the *D. melanogaster* gene annotation listed above.

Once this is complete, we can run the batch script from the Linux command-line as follows:

```
R CMD BATCH MMB_chapter_TSRchitect.R
#assumes variables BAMDIR and DmAnnot have already been set
bg #puts this job in the background
```

Once the job is underway, you can monitor its progress by looking at the contents of the `.Rout` file (in this case, "`MMB_chapter_TSRchitect.Rout`"). The job should complete within an hour on most systems.

**Reviewing the *TSRchitect* script.** Before we evaluate the results (which will have been written to your working directory after running the batch script), there are some important aspects of the analysis to review. We discuss these for

326 informational purposes only; it will not necessary to perform these commands  
 327 separate from the batch script provided. First, we must initialize the *tssObject*  
 328 (which stores the information about the experiment) appropriately (*see Note 7*).

329

330 The inputs in this case are BAM files (*inputType*="bam"); *TSRchitect* also ac-  
 331 cepts input in BED format.

```
332 DmRAMPAGE <- loadTSSobj(experimentTitle = "RAMPAGE Tutorial", \
333   inputDir=BAMDIR, inputType="bam", isPairedEnd=TRUE, \
334   sampleNames=c("E1h", "E2h", "E3h", "L1", "L2", "L3"), \
335   replicateIDs=c(1,1,1,2,2,2))
```

336 A critical step in our analysis is identifying TSRs from the aligned TSS data;  
 337 to do this we use the function *determineTSR*. We have selected the job to run  
 338 on 4 cores in this example (*n.cores*=4). Please enter the number of cores ap-  
 339 propriate for your system. Because we want to identify TSRs from every one  
 340 of the selected RAMPAGE libraries, we specify *tssSet*="all". The parameter  
 341 *tagCountThreshold* was set to 25, meaning that only TSSs supported by 25 or  
 342 more 5' RAMPAGE reads will be included within a TSR. Setting *writeTable* to  
 343 "TRUE" means that the identified TSRs from each set will be written to the  
 344 working directory.

```
345 DmRAMPAGE <- determineTSR(experimentName=DmRAMPAGE, n.cores=4, \
346   tsrSetType="replicates", tssSet="all", tagCountThreshold=25, \
347   clustDist=20, writeTable=TRUE)
```

348 *TSRchitect* can incorporate the tag abundances from each of the samples  
 349 and append them to the list of identified TSRs. This is useful for downstream  
 350 analysis of differential expression.

```
351 DmRAMPAGE <- addTagCountsToTSR(experimentName=DmRAMPAGE, \
352   tsrSetType="replicates", tsrSet=1, tagCountThreshold=10, \
353   writeTable=TRUE)
```

354 We can use *TSRchitect* to import an annotation file (or, alternatively, use an  
 355 existing one from *AnnotationHub*) and use it to associate our set of identified  
 356 TSRs with coding genes. We can specify the maximum distances (both up-  
 357 and downstream) between the TSR and the annotation using the arguments  
 358 *upstreamDist* and *downstreamDist*.

```
359 DmRAMPAGE <- importAnnotationExternal(experimentName=DmRAMPAGE, \
360   fileType="gff3", annotFile=DmAnnot)
361
362 DmRAMPAGE <- addAnnotationToTSR(experimentName=DmRAMPAGE, \
363   tsrSetType="replicates", tsrSet=1, \
364   upstreamDist=1000, downstreamDist=200, feature="gene", \
365   featureColumnID="ID", writeTable=TRUE)
```

Now we have generated a set of identified TSSs, TSRs from all 6 RAMPAGE libraries, and have associated the identified TSRs with annotated genes. Next, we will merge the libraries into two samples according to condition: early embryonic (E1h, E2h, E3h) and late larval (L1, L2, L3) using the information we provided when we initialized the *tssObject* at the start of this section. After merging, we identify promoters i) within the merged samples and ii) within the entire dataset combined, and associate with the *D. melanogaster* gene annotation as described previously (not shown).

```
#merging the sample data into two groups
DmRAMPAGE <- mergeSampleData(DmRAMPAGE)

# ... identifying TSRs from the merged samples:
DmRAMPAGE <- determineTSR(experimentName=DmRAMPAGE, \
  n.cores=4, tsrSetType="merged", \
  tssSet="all", tagCountThreshold=40, \
  clustDist=20, writeTable=TRUE)
```

**Evaluating the results** Our analysis using *TSRchitect* is now complete. Your working directory should now contain the following:

- TSSs from each sample *e.g.* TSSset-1.txt: (6)
- TSRs from each sample (in both .txt and .tab formats): (12)
- TSRs from each merged group (in both .txt and .tab formats): *e.g.* TSRsetMerged-1.txt: (4)
- TSRs from the combined set of TSSs: TSRsetCombined.tab: (1)

Let's briefly review the files. We can quickly obtain the counts on the command line, as follows:

```
wc -l *.tab
8377 TSRset-1.tab
6159 TSRset-2.tab
4814 TSRset-3.tab
17924 TSRset-4.tab
11851 TSRset-5.tab
3242 TSRset-6.tab
13986 TSRsetCombined.tab
7344 TSRsetMerged-1.tab
12126 TSRsetMerged-2.tab
85823 total
```

We will see that we have identified between roughly 3,200 and 18,000 TSRs within the individual RAMPAGE samples, which is attributable to the differences in library sizes. We detect 7,344 TSRs within the early embryonic samples ("TSRsetMerged-1.tab") and 12,126 TSRs in the late larval samples ("TSRsetMerged-2.tab"). Within the combined samples ("TSRsetCombined.tab")

we find 13,986 TSRs, which is similar to the number reported by Hoskins *et. al.* [1].

In addition to identifying the position of a given TSRs, *TSRchitect* records other useful information about its properties. The *width* of a TSR refers the span of the genomic region it occupies (in bp), and the *Shape Index* (SI) is measure of the relative peakedness of the TSR. We can see an example of this in the file "TSRsetMerged-1.txt".

seq	start	end	strand	nTSSs	tsrWidth	shapeIndex	featureID
2L.67043.67044.+			2L	67043	67044 +	270 2	1 NA
2L.74089.74115.+			2L	74089	74115 +	341 27	0.13 NA
2L.94739.94752.+			2L	94739	94752 +	1650 14	0.55 FBgn0031
2L.102386.102386.+			2L	102386	102386 +	284 1	2 FBgn0031

### 3.6 Summary

The workflow provided here is intended to serve as a useful entry point for the analysis of TSS profiling data in insects. On the computational side, we have provided an open source set of tools so that the uninitiated genome scientist can begin to analyze RAMPAGE (or other forms of TSS profiling data) quickly. While the analysis centered on *D. melanogaster* via the use of public datasets, it is anticipated that this will assist groups who may be interested in performing TSS profiling in their preferred insect model system.

The application of TSS profiling technology across a more representative sample of insect diversity will improve our understanding of the positions and general structure *cis*-regulatory regions in this phylum.

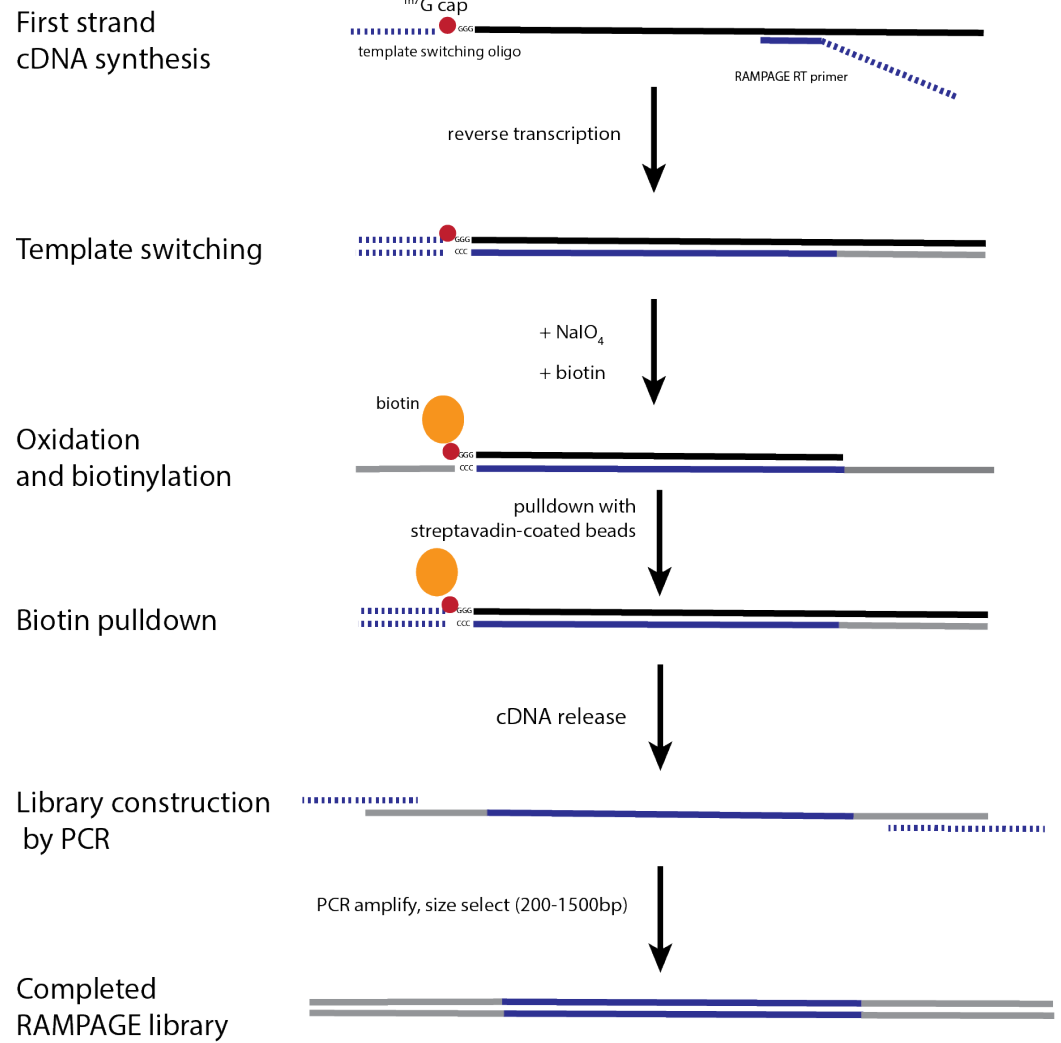
### 3.7 Figures

## 4 Notes

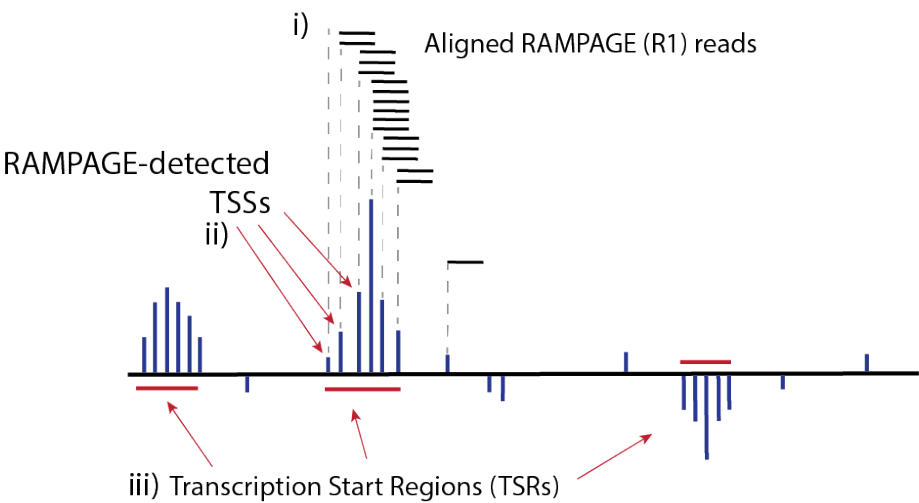
1. Please consult the GoRAMPAGE documentation found here:  
<https://github.com/BrendelGroup/GoRAMPAGE>. Installation instructions for the prerequisites of GoRAMPAGE (which includes some of the items listed) are found at the following link:  
<https://github.com/BrendelGroup/GoRAMPAGE/tree/master/src>.
2. You can clone this appendix to your workspace on the command line using git, as follows:

```
git clone https://github.com/rtraborn/MMB_appendix.git
```

The "scripts/" folder in the Appendix contains code for you to run the two major workflows described in this chapter. The "additional\_files/" folder contains the following files which are necessary for the analysis: i) a fasta file containing ribosomal RNA sequences for *D. melanogaster* (*Dmel\_rRNA.fasta*) and ii) a gene annotation for *D. melanogaster* (*Drosophila\_melanogaster.BDGP5.78.gff*).



**Fig. 1.** Test caption for figure 1



**Fig. 2.** Test caption for figure 2

3. Since these fastq files are paired-end, we use the argument `-split-files` to  
generate separate files for each read pair.
4. If you are running this on a cluster with a job scheduler you'll need to add  
the necessary headers to the top of the script and submit the job in the  
appropriate manner.
5. For parallel execution, GoRAMPAGE uses the Linux package *GNU parallel*  
[29]. Please see the GoRAMPAGE documentation for more information.
6. To do this, please edit the batch script `TSRchitect_script_MMB.R` with a  
text editor of your choice.
7. Because the samples provided derive from related developmental stages, we  
will merge them for annotation purposes using the argument `replicateIDs`,  
(though it must be emphaized that they are not replicates).

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**Disclosure Declaration**

The authors declare that they have no competing interests.

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## 6 Checklist of Items to be Sent to Volume Editors

Here is a checklist of everything the volume editor requires from you:

- ☐ The final L<sup>A</sup>T<sub>E</sub>X source files
- ☐ A final PDF file
- ☐ A copyright form, signed by one author on behalf of all of the authors of the paper.
- ☐ A readme giving the name and email address of the corresponding author.