

# Predicting Protein Organelle Localization with Transfer Learning

Ruge Zhao,¹ Siyi Tang²

<sup>1</sup> Department of Statistics, Stanford University <sup>2</sup> Department of Electrical Engineering, Stanford University

First conv layer: 4 input channels

Pretrained RGB weights

& assigned pre-trained R weights to Y

ResNet50 pretrained

on ImageNet

Fig. 2: Overview of preprocessing and transfer learning

## Stanford

### Motivation

- Advances in high-throughput microscopy images aid understanding of complex protein functions in human cells
- Accurate image classifications required for largescale cell images provided by Human Protein Atlas (HPA) [1]
- Machine learning algorithms proven to be effective in automating the classification [2-4]

### Goal

- Predict protein organelle localization labels in human cell samples
- Explore effectiveness of transfer learning

#### **Data**

- 31072 samples, 28 classes
- Each sample represented by 4 channels (RGBY)
- Each sample labeled with ≥ 1 protein organelle locations
- Imbalanced classes (e.g. 12885 vs 11)
- Train/validation/test sets: 24858/3107/3107

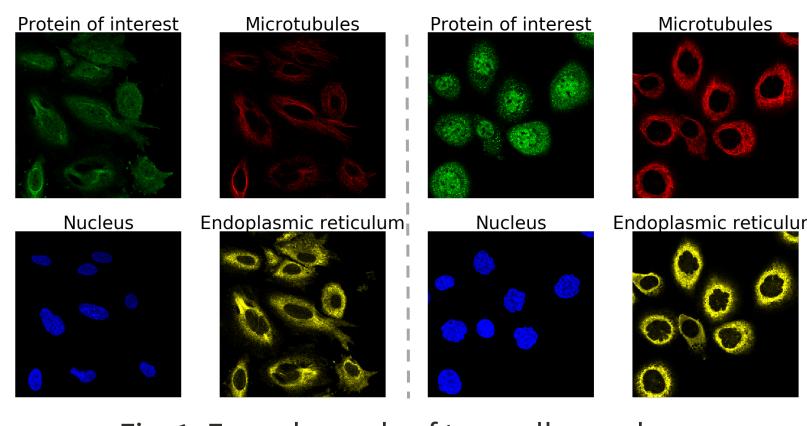


Fig. 1: Four channels of two cell samples

## **Preprocessing**

- Standardized with respect to train set
- Data augmentation: random rotate, flip, lighting

### Baseline

- 28 one-vs-all classifiers
- Logistic regression (LR) & random forest (RF)

## Transfer learning of ResNet50

**Preprocessed images** 

ResNet50 pre-trained on ImageNet dataset

#### Initializing network weights

Standardize, resize

Data augmentation

- RGB: initialized with pre-trained weights
- Y: initialized with pre-trained R weights

#### Loss functions

**Input images** 

- Binary cross-entropy (BCE) loss  $L_{BCE}(p_t) = -\log(p_t)$
- Focal loss [5]

$$L_{FL}(p_t) = -(1 - p_t)^{\gamma} \log(p_t)$$

where  $p_t = p$  if y = 1, else  $p_t = (1 - p)$ ;  $y \in \{0,1\}$ : true label;  $p \in [0,1]$ : predicted prob.

- Focal loss equivalent to BCE loss when  $\gamma=0$ Training of the network
- Freeze all layers except last, train for 1 epoch
- Unfreeze all layers and train until overfit

### Choosing optimal learning rate

 Picked learning rate where loss is low but still clearly decreasing

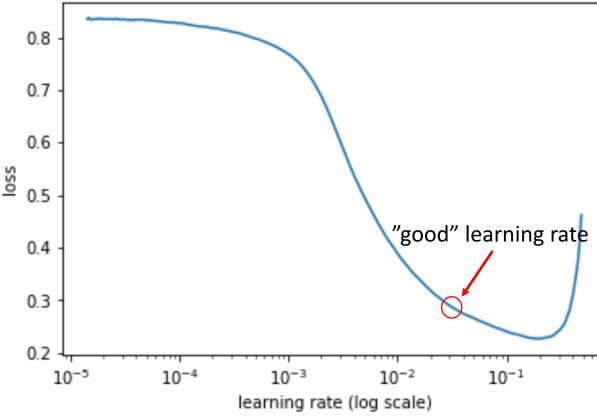


Fig. 3: Learning rate vs training loss

#### Differential learning rates

1. Train with all layers

2. Unfreeze all layers

except last freezed

& train until overfit

 Later layers need more fine-tuning to capture dataset-specific features

Sigmoid

28 classes

multi-label

Nucleoplasm

Nuclear

membrane

Cytoplasmic

bodies

**Rods & rings** 

Higher learning rate for later layers

## Training with stochastic gradient descent with restarts (SGDR) [6]

- Learning rate annealing during each cycle, and restarts learning rate at next cycle
- Encourages to find "smooth" rather than "spiky" local minimum

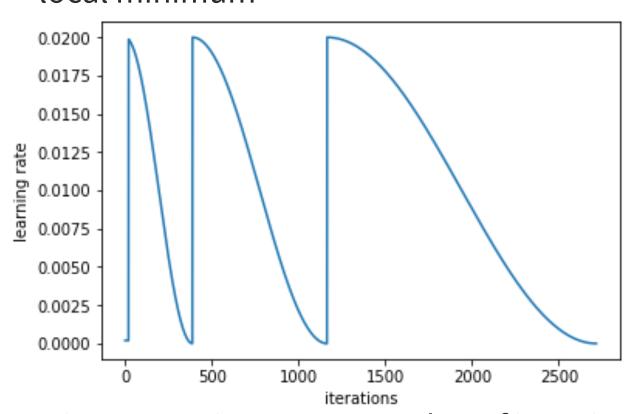


Fig. 4: Learning rate vs number of iterations during training with SGDR

#### Results

#### Baseline

- LR performed better than RF
- Best baseline model: macro F1 = 0.12

### **Transfer learning models**

- Best model without SGDR: Focal loss with  $\gamma$ =0.2
- With SGDR:
  - SGDR improved macro F1
  - Focal loss and BCE loss had comparable performance.

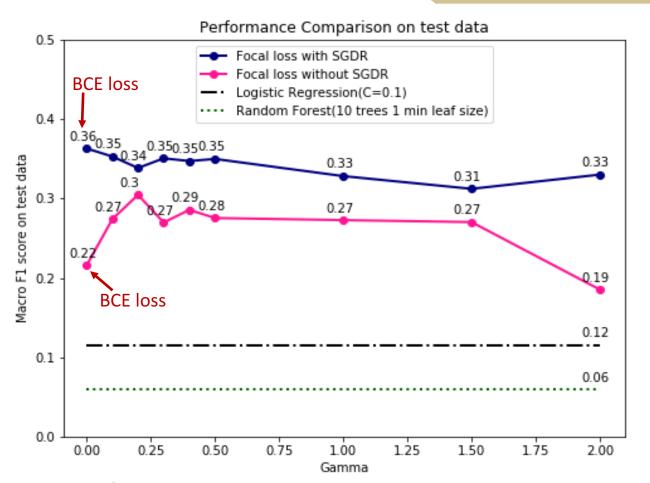


Fig. 5: Performance comparison on test data: macro F1

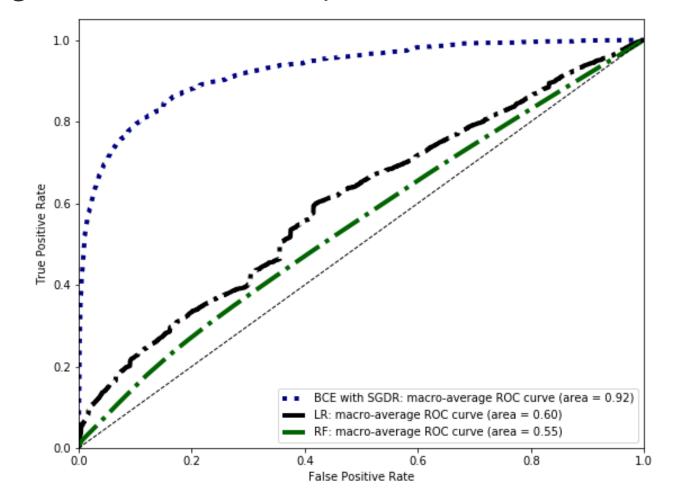


Fig. 6: Performance comparison on test data: ROC AUC

### **Conclusion and future work**

- ResNet pre-trained on ImageNet data has learned transferable features applicable to the HPA dataset
- Transfer learning model outperformed traditional machine learning models

#### **Future work**

- Experiment with highest resolution for all models
- Reweigh / resample majority and minority classes

## References

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