Remote Floating-Gate Field-effect Transistor with 2-Dimensional Reduced Graphene Oxide Sensing Layer for Reliable Detection of SARS-CoV-2 Spike Proteins

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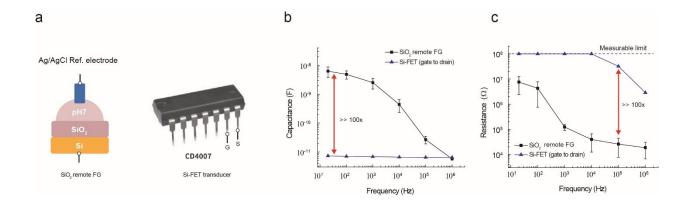
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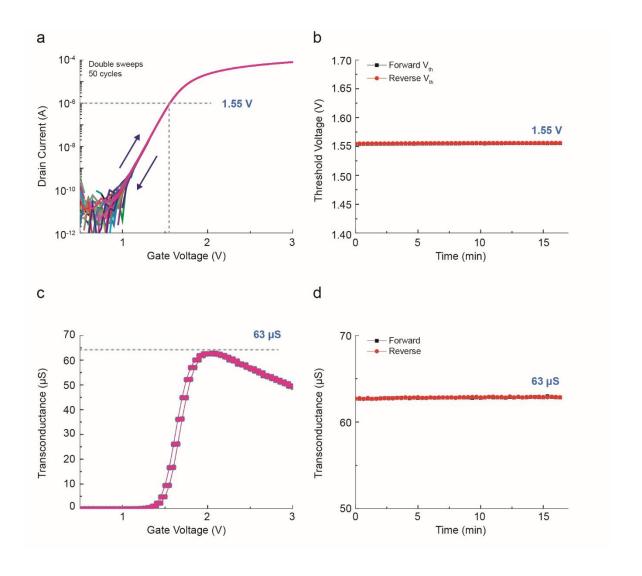
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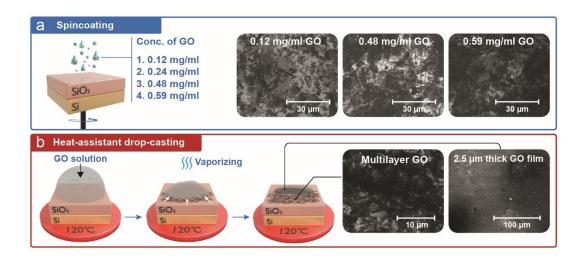
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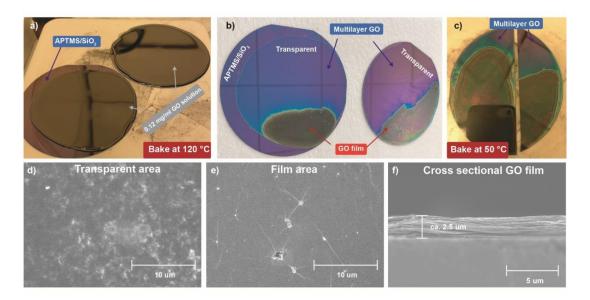
**Figure S1.** (a) Schematic images of device structure for impedance measurement of remote FG and commercial FET (CD4007). Frequency vs (b) capacitance and (c) resistance of a 300-nm-thick SiO<sub>2</sub> and gate-drain of Si-FET chip. For measurement of remote FG, 50 μL pH7 solution was added on the top of SiO<sub>2</sub> surface. AC frequency was applied through the Ag/AgCl reference electrode and the bottom silicon electrode of each FG. The level of voltage set at 1 V during capacitance and resistance measurement. The input impedance of Si-FET is much higher than those of remote SiO<sub>2</sub> FG, meaning that the effect of impedance of FGs on total impedance of remote FGFET is insignificant.



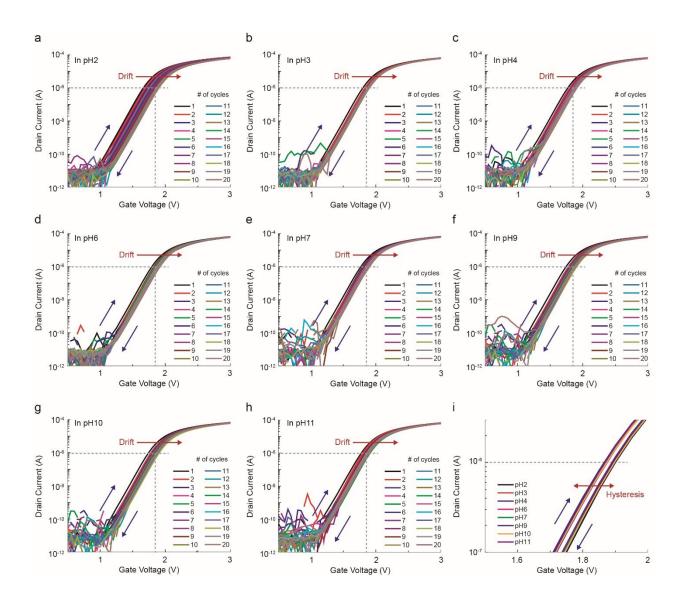
**Figure S2**. (a) Zoomed-in representative transfer curves of Si-FET with the double-sweeping mode the gate voltage  $(V_G)$  ranging from 0 to 5 V for 50 cycles of transfer curves. (b)  $V_{th}$  of the MOSFET measured by each forward- and reverse-sweeping mode over time. (c) Representative transconductance  $(G_m)$  curves of Si-FET with the double-sweeping mode at the  $V_G$  ranging from 0 to 5 V for 50 cycles of transconductance curves. (d)  $G_m$  of the MOSFET measured by each forward- and reverse-sweeping mode over time.



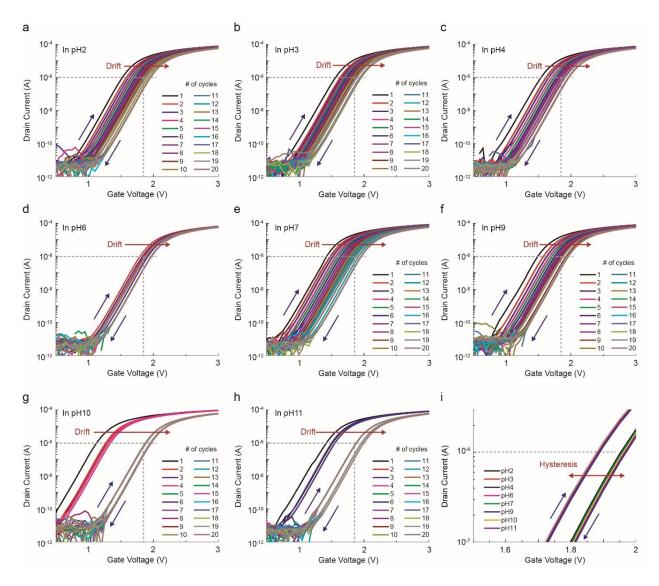
**Figure S3**. SEM images of GO layers (a) spin-coated with 0.12, 0.48, and 0.59 mg/ml GO solutions and (b) made by a heat-assisted drop-casting method.



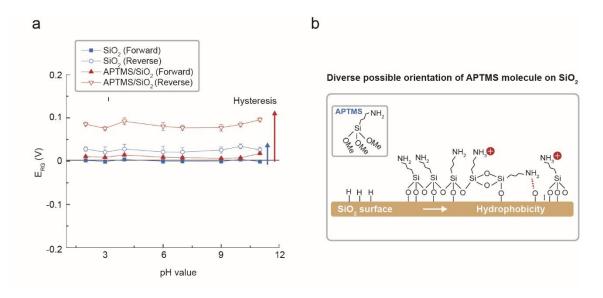
**Figure S4**. Images of GO films made using the heat-assistant drop-casting method: 0.12 mg/ml GO solution dropped on the APTMS/SiO<sub>2</sub> substrate (a) before and (b) after baking at 120 °C for 1 hour. The lower baking temperature at 50 °C allowed the GO film (dark area) to develop on more areas of the wafer (**Figure S4c**) due to the low evaporation rate of the GO solvent. For the dried wafers, transparent and dark areas appeared on the same APTMS/SiO<sub>2</sub> surface from multilayer GO (**Figure S4d**) and continuous thick GO films (**Figure S4e**), respectively. The thickness of the GO film was measured to be about 2.5 μm for cross-sectional SEM images of the GO film (**Figure S4f**).



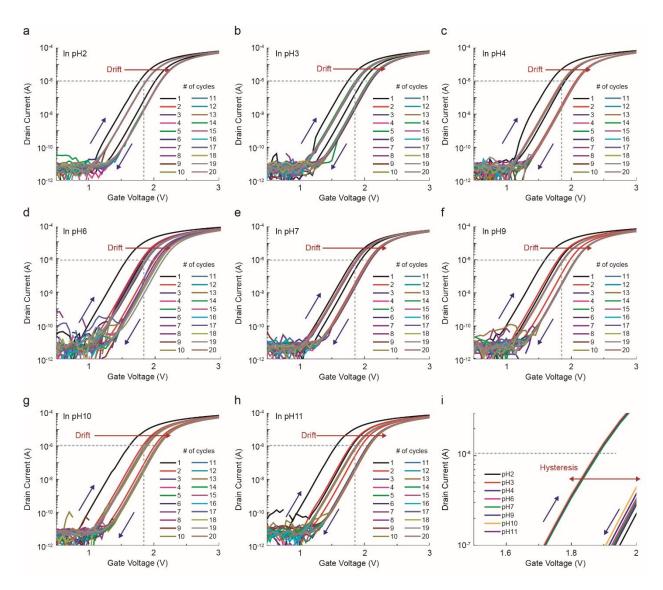
**Figure S5.** Representative transfer curves of the SiO<sub>2</sub> remote FGFET measured in (a) pH2, (b) pH3, (c) pH4, (d) pH6, (e) pH7, (f) pH9, (g) pH10, and (h) pH11 solution. Transfer curves were measured 20 times for each pH solution under the double-sweep mode with a gate voltage ranging from 0 to 5 V. The drain voltage sets at 50 mV over all measurements. Each initial transfer curve for the respective pH value drifted to the direction on the right with a saturation point. (i) Transfer curves at saturation for each pH solution. The transfer curves mostly overlapped in quasi-equilibrium regardless of pH changes.



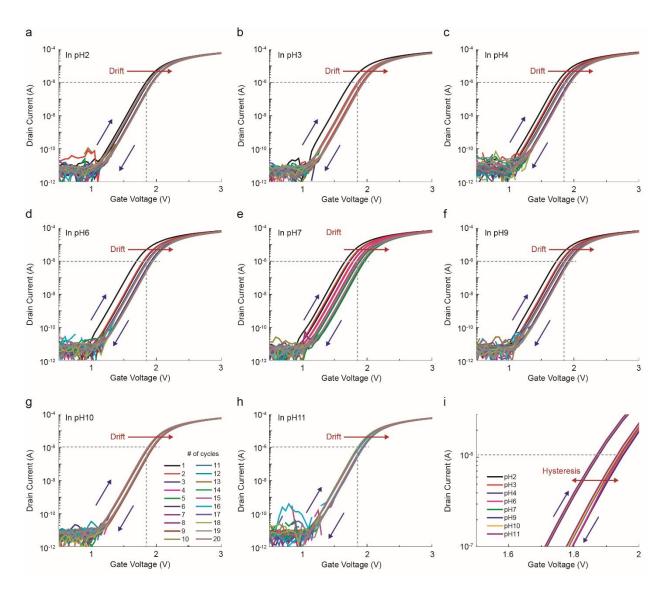
**Figure S6.** Representative transfer curves of APTMS/SiO<sub>2</sub> remote FGFET measured in (a) pH2, (b) pH3, (c) pH4, (d) pH6, (e) pH7, (f) pH9, (g) pH10, and (h) pH11 solution. Transfer curves were measured 20 times for each pH solution under the double-sweep mode with a gate voltage ranging from 0 to 5 V. The drain voltage sets at 50 mV over all measurements. Each initial transfer curve for the respective pH value drifted to the direction on the right with a saturation point. (i) Transfer curves at saturation for each pH solution. The transfer curves mostly overlapped in quasi-equilibrium regardless of pH changes.



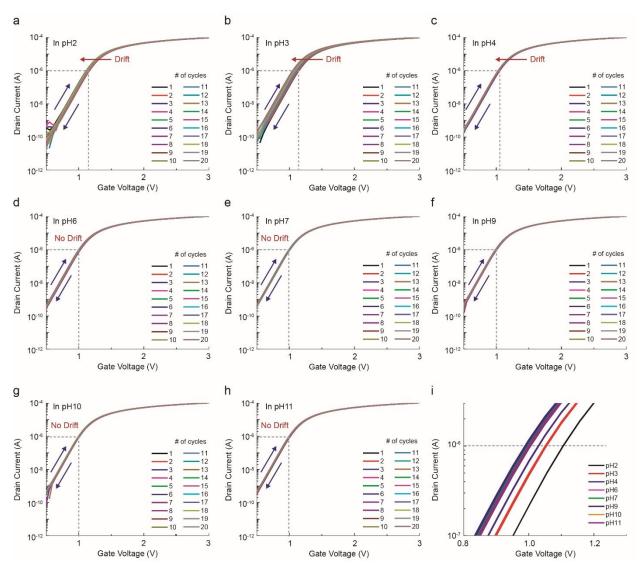
**Figure S7.** (a) The  $E_{RG}^{For}$  and  $E_{RG}^{Rev}$  of SiO<sub>2</sub> and APTMS/SiO<sub>2</sub> in quasi-equilibrium vs. pH value. (b) Schematic images of possible APTMS orientations enabling more hydrophobic surfaces than SiO<sub>2</sub>.



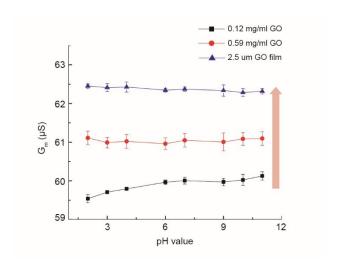
**Figure S8.** Representative transfer curves of the remote FGFET with a FG surface of spin-coated GO (0.12 mg/ml GO) without post-annealing, which was measured in (a) pH2, (b) pH3, (c) pH4, (d) pH6, (e) pH7, (f) pH9, (g) pH10, and (h) pH11 solution. Transfer curves were measured 20 times for each pH solution under the double-sweep mode with a gate voltage ranging from 0 to 5 V. The drain voltage sets at 50 mV over all measurements. Each initial transfer curve for the respective pH value drifted to the direction on the right with a saturation point. (i) Transfer curves at saturation for each pH solution. The transfer curves mostly overlapped in quasi-equilibrium regardless of pH changes.



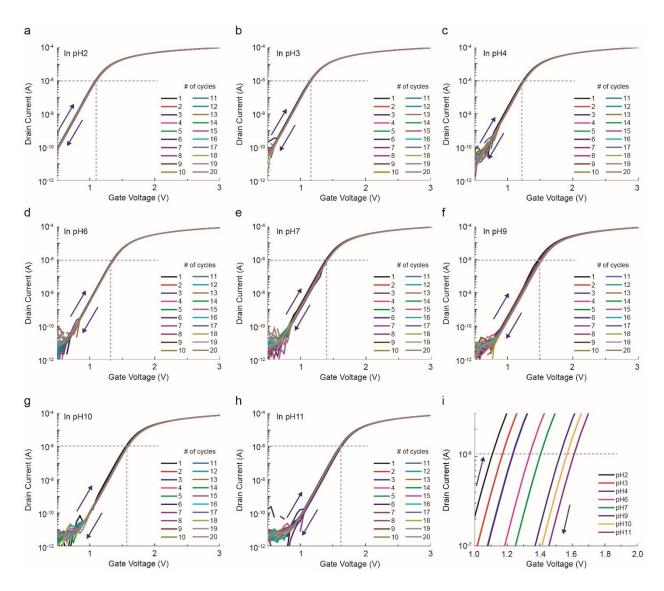
**Figure S9.** Representative transfer curves of the remote FGFET with a FG surface of spin-coated GO (0.59 mg/ml GO) without post-annealing, which was measured in (a) pH2, (b) pH3, (c) pH4, (d) pH6, (e) pH7, (f) pH9, (g) pH10, and (h) pH11 solution. Transfer curves were measured 20 times for each pH solution under the double-sweep mode with a gate voltage ranging from 0 to 5 V. The drain voltage sets at 50 mV over all measurements. Each initial transfer curve for the respective pH value drifted to the direction on the right with a saturation point. (i) Transfer curves at saturation for each pH solution. The transfer curves mostly overlapped in quasi-equilibrium regardless of pH changes.



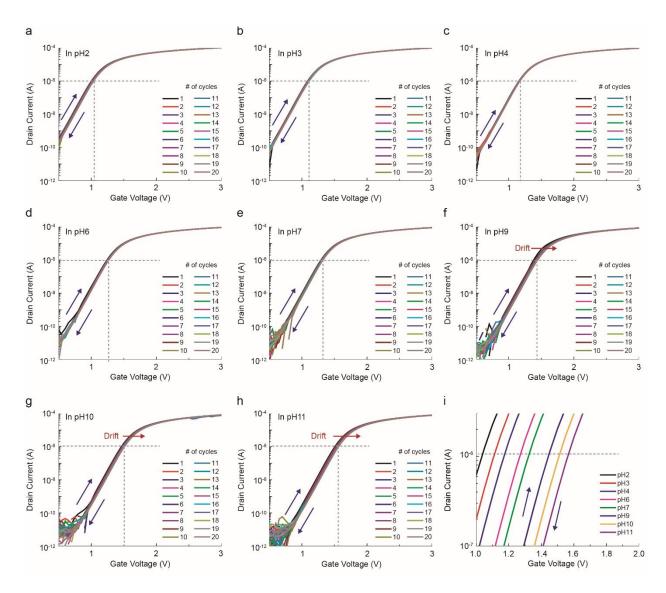
**Figure S10.** Representative transfer curves of the remote FGFET with a FG surface of drop-casted 2.5 μm GO without post-annealing, which was measured in (a) pH2, (b) pH3, (c) pH4, (d) pH6, (e) pH7, (f) pH9, (g) pH10, and (h) pH11 solution. Transfer curves were measured 20 times for each pH solution under the double-sweep mode with a gate voltage ranging from 0 to 5 V. The drain voltage sets at 50 mV over all measurements. Each initial transfer curve for the respective pH value drifted to the direction on the right with a saturation point. (i) Transfer curves at saturation for each pH solution. The transfer curves mostly overlapped in quasi-equilibrium regardless of pH changes.



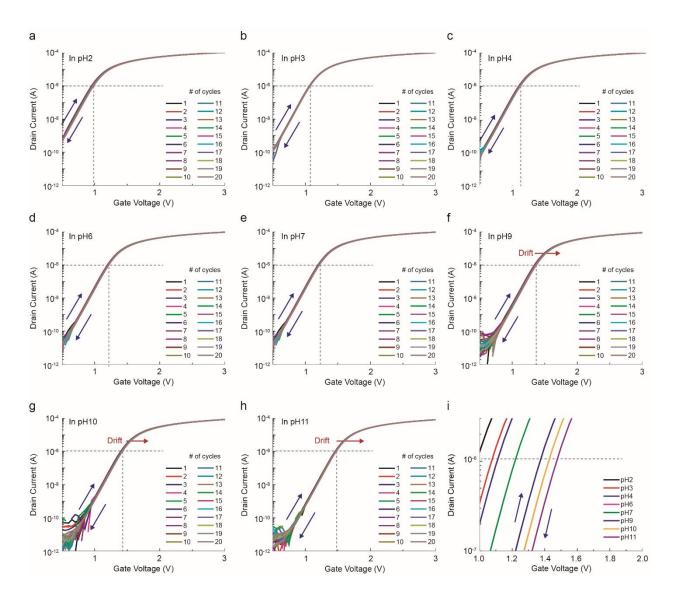
**Figure S11.**  $G_m^{For}$  distributions of 0.12 mg/ml GO films, 0.59 mg/ml GO films, and drop-casted 2.5  $\mu$ m GO films without reduction process over at least four samples as a function of increasing pH.



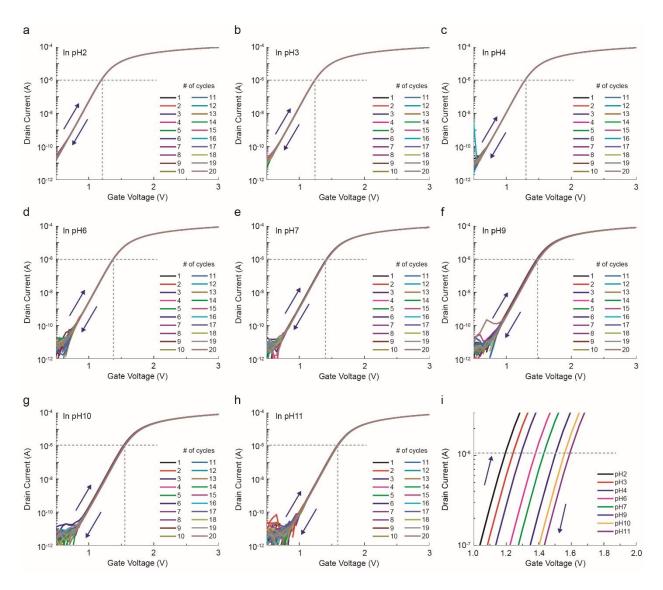
**Figure S12.** Representative transfer curves of the remote FGFET with a 0.59 mg/ml GO FG surface reduced at 200 °C, which was measured in (a) pH2, (b) pH3, (c) pH4, (d) pH6, (e) pH7, (f) pH9, (g) pH10, and (h) pH11 solution. Transfer curves were measured 20 times for each pH solution under the double-sweep mode with a gate voltage ranging from 0 to 5 V. The drain voltage sets at 50 mV over all measurements. Each initial transfer curve for the respective pH value drifted to the direction on the right with a saturation point. (i) Transfer curves at saturation for each pH solution. The transfer curves mostly overlapped in quasi-equilibrium regardless of pH changes.



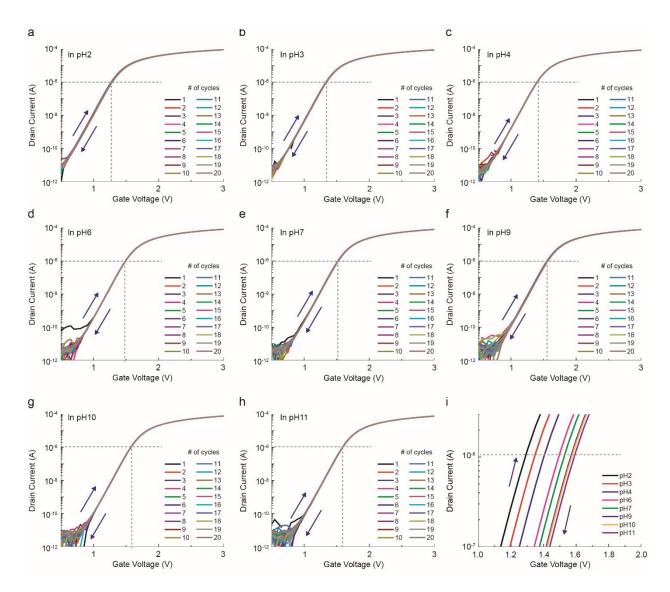
**Figure S13.** Representative transfer curves of the remote FGFET with a 0.59 mg/ml GO FG surface reduced at 300 °C, which was measured in (a) pH2, (b) pH3, (c) pH4, (d) pH6, (e) pH7, (f) pH9, (g) pH10, and (h) pH11 solution. Transfer curves were measured 20 times for each pH solution under the double-sweep mode with a gate voltage ranging from 0 to 5 V. The drain voltage sets at 50 mV over all measurements. Each initial transfer curve for the respective pH value drifted to the direction on the right with a saturation point. (i) Transfer curves at saturation for each pH solution. The transfer curves mostly overlapped in quasi-equilibrium regardless of pH changes.



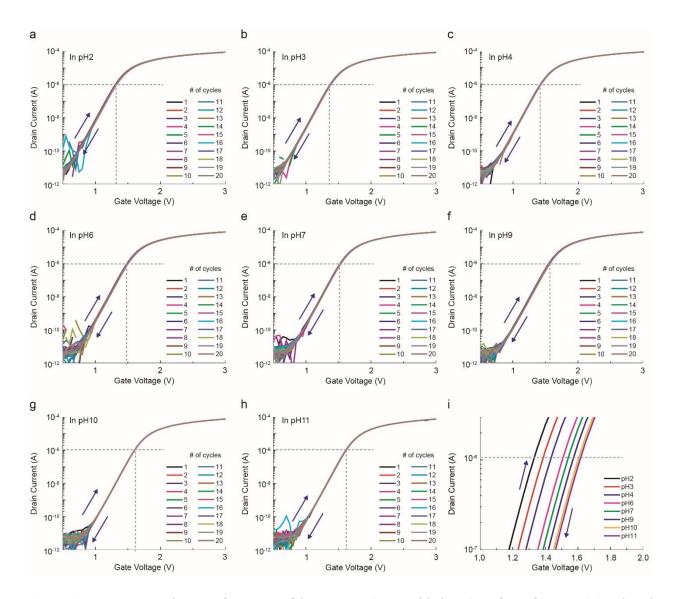
**Figure S14.** Representative transfer curves of the remote FGFET with a 0.59 mg/ml GO FG surface reduced at 400 °C, which was measured in (a) pH2, (b) pH3, (c) pH4, (d) pH6, (e) pH7, (f) pH9, (g) pH10, and (h) pH11 solution. Transfer curves were measured 20 times for each pH solution under the double-sweep mode with a gate voltage ranging from 0 to 5 V. The drain voltage sets at 50 mV over all measurements. Each initial transfer curve for the respective pH value drifted to the direction on the right with a saturation point. (i) Transfer curves at saturation for each pH solution. The transfer curves mostly overlapped in quasi-equilibrium regardless of pH changes.



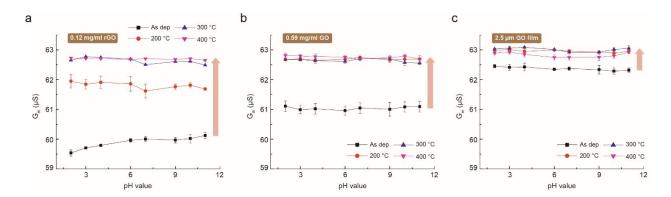
**Figure S15.** Representative transfer curves of the remote FGFET with the FG surface of 2.5 μm GO reduced at 200 °C, which was measured in (a) pH2, (b) pH3, (c) pH4, (d) pH6, (e) pH7, (f) pH9, (g) pH10, and (h) pH11 solution. Transfer curves were measured 20 times for each pH solution under the double-sweep mode with a gate voltage ranging from 0 to 5 V. The drain voltage sets at 50 mV over all measurements. Each initial transfer curve for the respective pH value drifted to the direction on the right with a saturation point. (i) Transfer curves at saturation for each pH solution. The transfer curves mostly overlapped in quasi-equilibrium regardless of pH changes.



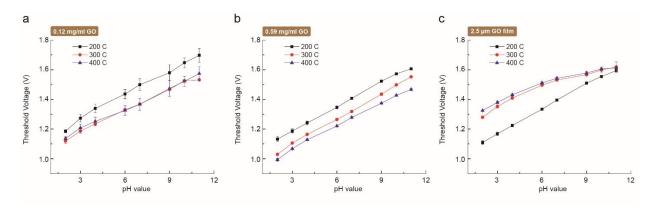
**Figure S16.** Representative transfer curves of the remote FGFET with the FG surface of 2.5 μm GO reduced at 300 °C, which was measured in (a) pH2, (b) pH3, (c) pH4, (d) pH6, (e) pH7, (f) pH9, (g) pH10, and (h) pH11 solution. Transfer curves were measured 20 times for each pH solution under the double-sweep mode with a gate voltage ranging from 0 to 5 V. The drain voltage sets at 50 mV over all measurements. Each initial transfer curve for the respective pH value drifted to the direction on the right with a saturation point. (i) Transfer curves at saturation for each pH solution. The transfer curves mostly overlapped in quasi-equilibrium regardless of pH changes.



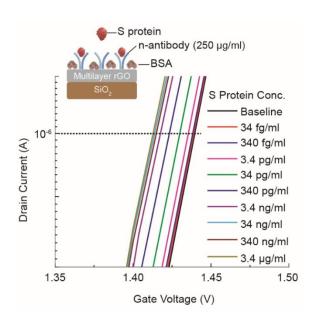
**Figure S17.** Representative transfer curves of the remote FGFET with the FG surface of 2.5 μm GO reduced at 400 °C, which was measured in (a) pH2, (b) pH3, (c) pH4, (d) pH6, (e) pH7, (f) pH9, (g) pH10, and (h) pH11 solution. Transfer curves were measured 20 times for each pH solution under the double-sweep mode with a gate voltage ranging from 0 to 5 V. The drain voltage sets at 50 mV over all measurements. Each initial transfer curve for the respective pH value drifted to the direction on the right with a saturation point. (i) Transfer curves at saturation for each pH solution. The transfer curves mostly overlapped in quasi-equilibrium regardless of pH changes.



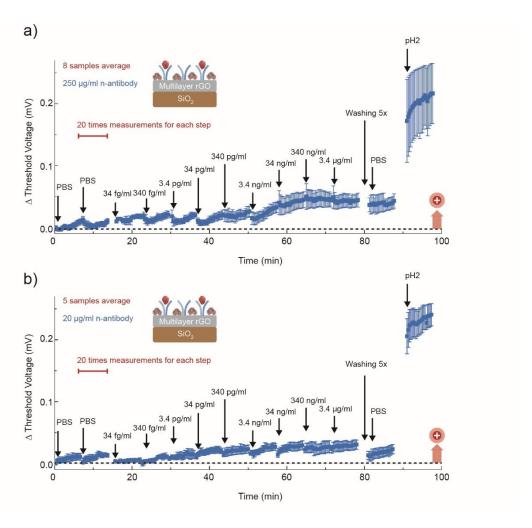
**Figure S18.**  $G_m^{For}$  distributions of (a) 0.12 mg/ml GO films, (b) 0.59 mg/ml GO films, and (c) drop-casted 2.5 µm GO films over at least four samples as a function of increasing pH and reduction temperatures ranging from 200 to 400 °C.



**Figure S19.**  $V_{th}$  distributions from (a) an FG surface with 0.12 mg/ml GO films, (b) 0.59 mg/ml GO films, and (c) drop-casted 2.5  $\mu$ m GO films over at least three samples as a function of increasing pH and reduction temperatures ranging from 200 to 400 °C.



**Figure S20.** Responses of transfer curves of antibody-functionalized rGO (250  $\mu$ g/ml antibody) with increasing concentrations of S protein in a range from 34 fg/ml to 3.4  $\mu$ g/ml in 0.05 × PBS.



**Figure S21.**  $\Delta V_{th}$  distributions of rGO layers functionalized with (a) 250 and (b) 20 μg/ml n-antibodies for increasing S proteins from 34 fg/ml to 3.4 μg/ml over time; each  $\Delta V_{th}$  in the same measurement condition was averaged from 8 and 5 samples of rGO in a different batch, respectively. At the end of the S protein detection, the remote FG surfaces were washed five times with a pure PBS solution and then sequentially measured in a pure PBS media again to confirm the remaining signals between the antibody and the spike protein. After measuring the remote FG surfaces in a PBS solution, the pH2 solution was measured using the same FG surface. A sharp shift at pH2 resulted from changes in surface potentials on rGO in responding to pH changes, indicating the presence of rGO layers even after all the biosensing experiments.

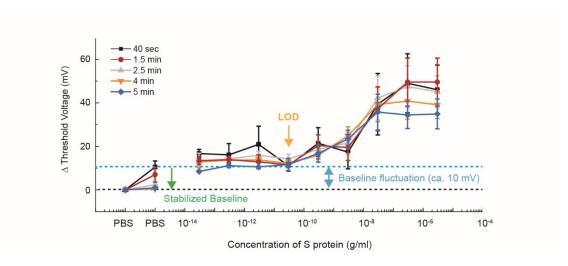
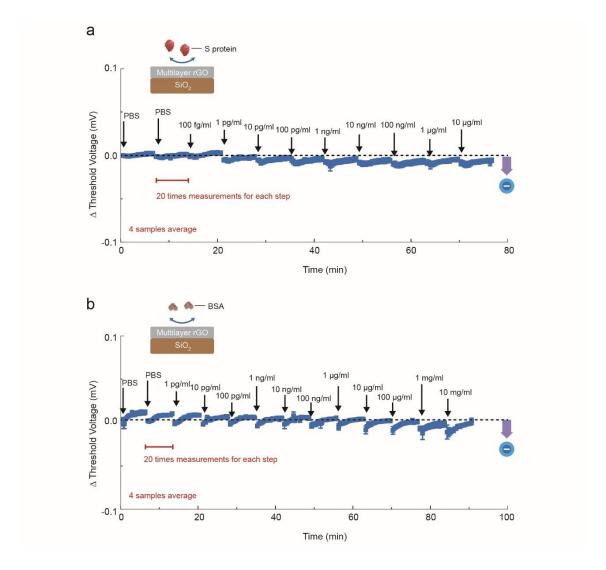
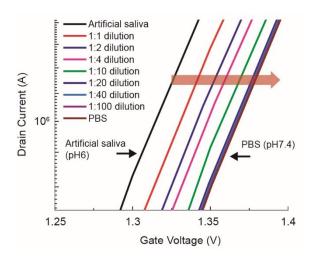


Figure S22.  $\Delta V_{th}$  levels of 250 µg/ml antibody-functionalized rGO in response to S protein concentrations but obtained from different intervals of measurement time in a range from 40 sec to 5 min. Stabilized baseline and  $\Delta V_{th}$  sensing curves began to show  $\Delta V_{th}$  taken after at least 2.5 min measurements. A large difference in baseline, however, was observed from  $\Delta V_{th}$  taken after the measurements with less than 2.5 min. This suggests that for the early stage of measurements, drift effects in  $V_{th}$  and signal fluctuation significantly surged but soon stabilized after some measurement time around reaching quasi-equilibrium. Therefore, longer measurement time of 5 min was chosen to obtain stable  $\Delta V_{th}$  levels in quasi-equilibrium, which could minimize the impact of measurement time and drift on sensitivity of devices. It is noted that the maximum fluctuation in baselines that could occur for any measurement conditions is measured to be ca. 10 mV as shown in Figure S22. Thus, conservative LODs could be determined at the concentration of S protein showing  $\Delta V_{th}$  above 10 mV.



**Figure S23.**  $\Delta V_{th}$  distributions of bare rGO layers for increasing (a) S proteins in a range from 100 fg/ml to 10  $\mu$ g/ml over time and (b) BSA concentrations in a range from 1 pg/ml to 10 mg/ml.



**Figure S24.** Transfer curves of a bare rGO surface measured in a pure artificial saliva, artificial saliva mixtures with different dilution ratios between artificial saliva and  $0.05 \times PBS$ , and a pure  $0.05 \times PBS$ . For artificial solution mixtures, the pure artificial saliva at pH 6 was diluted in a  $0.05 \times PBS$  solution at pH7.4 with a different ratio of the PBS solution. Transfer curve of the artificial solution mixture diluted in a ratio of 1:20 (artificial saliva:PBS) is mostly overlapping with that of a pure  $0.05 \times PBS$ , implying that the pH value of both the artificial solution mixture and the  $0.05 \times PBS$  was controlled at 7.4.