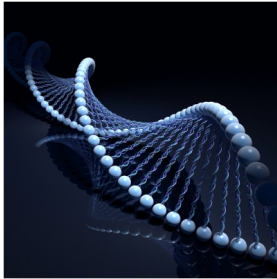


LGBIO2010: Large scale gene expression analysis

Pierre Dupont



UCL – ICTEAM

P. Dupont (UCL)

LGBIO2010

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Outline

- 1 Introduction
- 2 Preprocessing
- 3 Unsupervised gene selection
- 4 Supervised gene selection
 - Filters
 - Non-specific filtering
 - Fold changes
 - t-Test
 - Mutual information
 - Multivariate filters
 - Wrappers
 - Embedded Methods
 - Classification

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- 2 Preprocessing
- 3 Unsupervised gene selection
- 4 Supervised gene selection

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Introduction

Gene expression

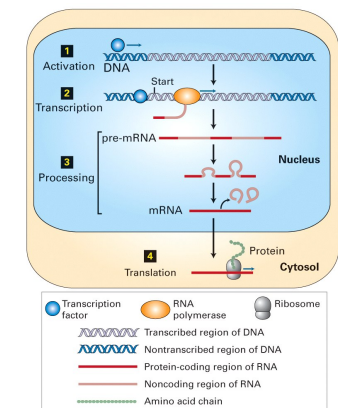
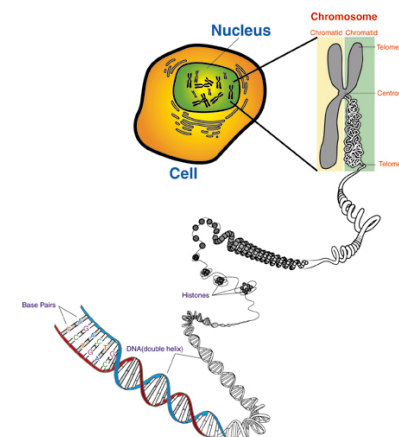


Illustration from Molecular Cell Biology, 5e (© WHFreeman 2004).

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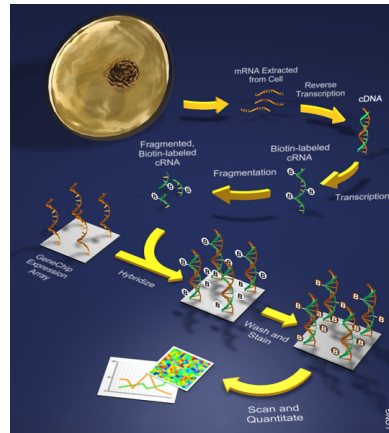
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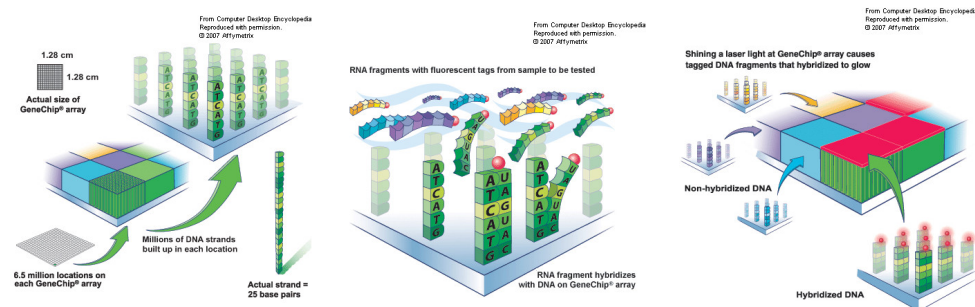
DNA Microarrays

DNA Microarrays measure the level of expression of **all genes** in a **single experiment**

- 1 Data measurements
- 2 Preprocessing and sample normalization
- 3 Gene selection and sample classification
- 4 Diagnosis, prognosis or prediction of the response to a treatment



Affymetrix® technology



Alternative measurement technologies

- Other companies sell DNA chips (Agilent®, ...)
- Multiplex qPCR (Applied Biosystems®, ...)
 - ▶ larger dynamic range than microarrays
 - ▶ limited to ≈ 100 genes
- RNAseq (Illumina®, Ion Torrent®, ...)
 - ▶ fastly evolving
 - ▶ scaling effects influence per sample cost

Example: diagnosis

Biomarkers for an **early diagnosis** of rheumatoid infections

Prediction problem: multi-class feature selection

- Rheumatoid arthritis
- Lupus
- Psoriatic rheumatism
- Microcrystalline arthritis
- Inflammatory osteoarthritis



RHEUMAGENE research project with Prof. Lauwerys (UCL/IREC/RUMA)

Example: prognosis

Biomarkers to predict the risk of allergies of newborns



- More than **30%** of children are allergic in industrial countries
- Predicting who is more likely to become allergic is a path to prevention and possible treatment

CRISTALL research project with Profs. Sokal and Smets (UCL/IREC/PEDI)

Example: response to treatment prediction

Gene profiling for cancer treatment

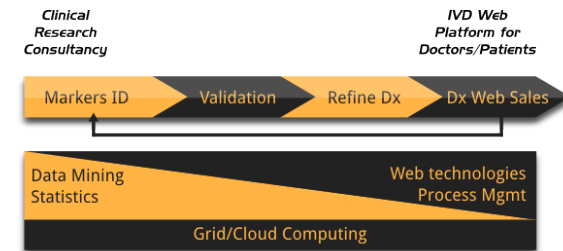
Objective

Identify biomarkers for predicting patient response to MAGE-A3 immuno-therapy against melanoma **before** treatment



In collaboration with GSK Biologicals - WO/2010/029174 (patent).

A UCL spin-off



www.dnalytics.com

Supervised selection

	gene 1	gene 2	...	gene p	response
sample 1	$x_{1,1}$	$x_{1,2}$...	$x_{1,p}$	y_1
...
sample n	$x_{n,1}$	$x_{n,2}$...	$x_{n,p}$	y_n
test sample	x_1	x_2	...	x_p	?

- The number p of **input dimensions** (probes, probesets or genes) may be very large ($10^4 \dots 10^6$)
- The number n of **samples** is typically much smaller ($\approx 50 \dots 100$)
- Each sample is characterized by a vector \mathbf{x} of p measurements
- Each **training sample** has a known response: class label y ($y \in \{-1, 1\}$ or $y \in \mathbb{N}$) or $y \in \mathbb{R}$

Gene selection

Find a small subset of **genes**, (a.k.a **features**, **attributes** or **input variables**), to predict the response or class y of new samples

Gene selection

Objectives

- Insight into the data and the predictive model
- Link between data analysis and medical expert
- Biological validation on a few genes rather than thousand ones
- Reduction of the financial cost of a diagnosis/prognosis kit (technological constraints)

Difficulties

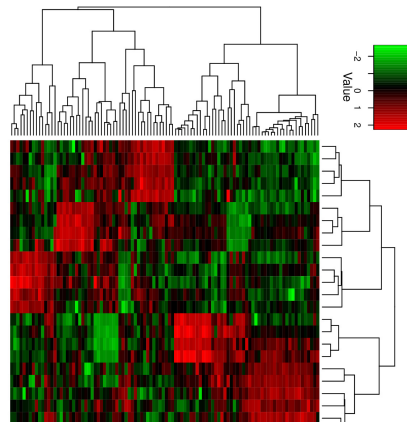
- Measurements are noisy
- Gene expression varies due to many factors (gender, cell type, growth of the organism, chemical environment of the cell, ...) often not related to the response to be predicted
- Financial cost: 500 ... 1,000 €/experiment
- Small n (e.g. 50), large p (e.g. 50,000) problems

Unsupervised selection

	gene 1	gene 2	...	gene p	cluster
sample 1	$x_{1,1}$	$x_{1,2}$...	$x_{1,p}$?
...
sample n	$x_{n,1}$	$x_{n,2}$...	$x_{n,p}$?
cluster	?	?	...	?	

Objective

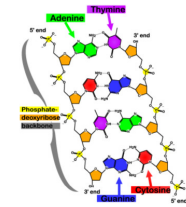
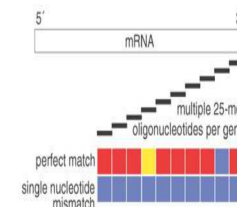
Find **clusters** of genes and/or samples that share a **similar profile**: up or down regulated genes across the same samples



Outline

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Summarization



- Define a single probeset expression level from the various probe intensities
- Popular techniques: MAS 5.0, RMA, **GC-RMA**
 - 1 background adjustment: optical noise correction, probe affinity adjustment (influenced by the GC content), RMA ignores the MM probes
 - 2 sample normalization: quantiles should be stable across samples, after conversion to **log** intensities for (GC-)RMA
 - 3 summarization: median polish

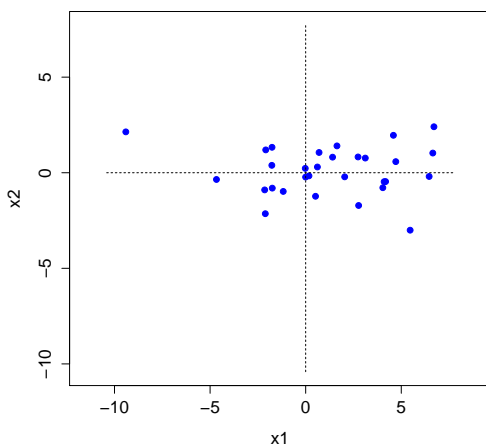
Feature normalization

	gene 1	gene 2	...	gene p
sample 1	$x_{1,1}$	$x_{1,2}$...	$x_{1,p}$
...
sample n	$x_{n,1}$	$x_{n,2}$...	$x_{n,p}$

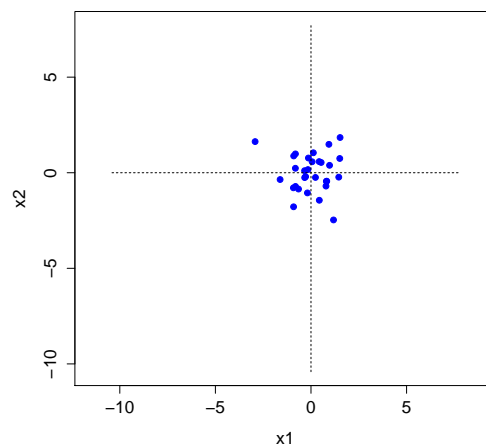
- Make sure that each gene (probeset) has roughly the same expression range across all samples
- Z-score normalization**
Replace $x_{i,j}$ by $\frac{x_{i,j} - \mu_j}{s_j}$ with μ_j the mean level of expression of probeset j over the training samples and s_j its standard deviation

Feature normalization example

Before



After



Distance between expression values

	gene 1	gene 2	...	gene p
sample 1	$x_{1,1}$	$x_{1,2}$...	$x_{1,p}$
...
sample n	$x_{n,1}$	$x_{n,2}$...	$x_{n,p}$

Euclidean distance

$$d(\mathbf{x}_1, \mathbf{x}_2) = \|\mathbf{x}_1 - \mathbf{x}_2\| = \sqrt{\sum_{i=1}^n (x_{i,1} - x_{i,2})^2}$$

Correlation based distance

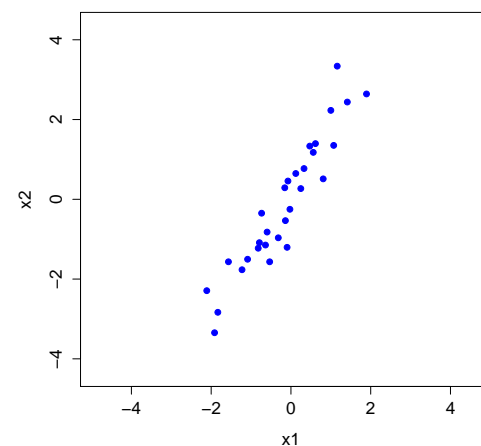
$$d(\mathbf{x}_1, \mathbf{x}_2) = 1 - \frac{1}{2}(1 + \text{corr}(\mathbf{x}_1, \mathbf{x}_2))$$

Correlation between expression values

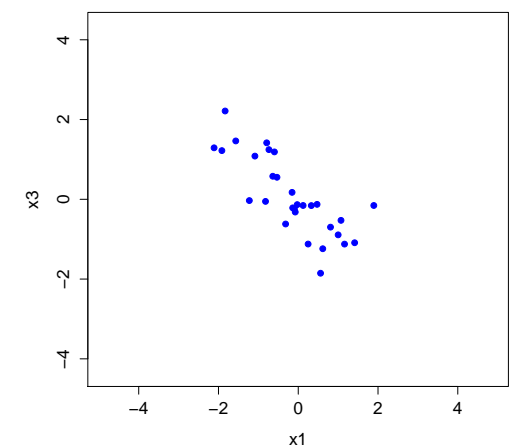
Are both genes over/under expressed on the same samples?

Is one gene over-expressed when the other is under-expressed?

Positive Correlation



Negative Correlation



Pearson correlation

For two random vectors (e.g. gene expression values) $\mathbf{x}_1, \mathbf{x}_2$ measured over n samples

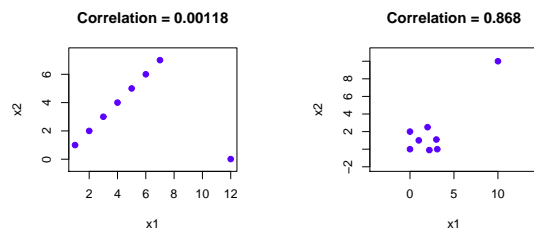
$$\text{corr}(\mathbf{x}_1, \mathbf{x}_2) = \frac{\sum_{i=1}^n (x_{i,1} - \bar{x}_1)(x_{i,2} - \bar{x}_2)}{\sqrt{\sum_{i=1}^n (x_{i,1} - \bar{x}_1)^2 \sum_{i=1}^n (x_{i,2} - \bar{x}_2)^2}}$$

- $\text{corr}(\mathbf{x}_1, \mathbf{x}_2) = \pm 1$ if \mathbf{x}_1 and \mathbf{x}_2 are perfectly linearly correlated
- $\text{corr}(\mathbf{x}_1, \mathbf{x}_2) = 0$ if they are not linearly correlated
- whenever \mathbf{x}_1 and \mathbf{x}_2 are normalized to zero mean and unit variance:

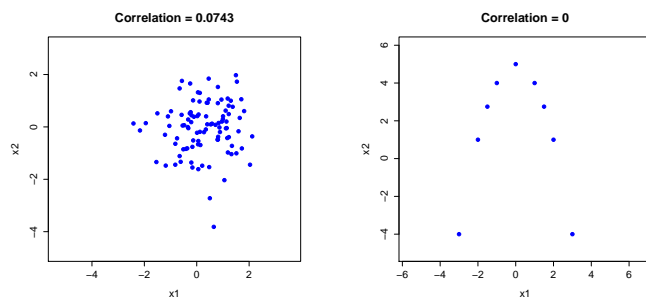
$$\text{corr}(\mathbf{x}_1, \mathbf{x}_2) = \sum_{i=1}^n x_{i,1} x_{i,2} = \mathbf{x}_1^\top \mathbf{x}_2$$

Pitfalls with correlation measures

- Correlation is very sensitive to outliers

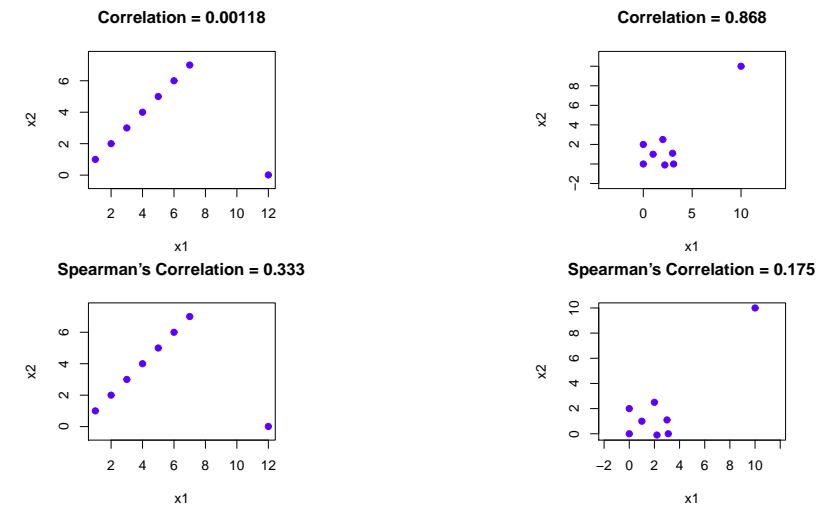


- Correlation measures linear dependence

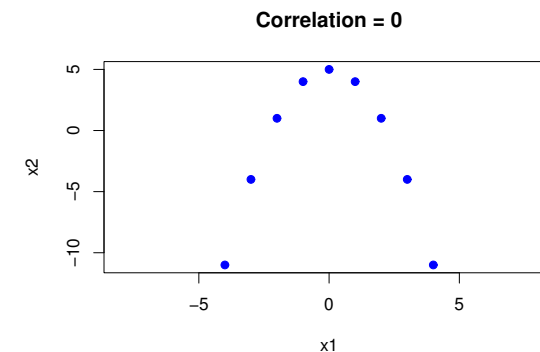


Spearman's rank correlation: less sensitive to outliers

- 1 Replace feature value by feature value rank across observations
- 2 Compute Pearson correlation between rank vectors



Uncorrelated features are not necessarily independent



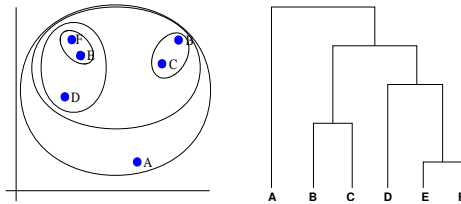
- $\text{corr}(\mathbf{x}_1, \mathbf{x}_2) = 0$ (both Pearson and Spearman correlations)
- $P(\mathbf{x}_2 | \mathbf{x}_1) \neq P(\mathbf{x}_2)$

Outline

- 1 Introduction
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Agglomerative Hierarchical clustering

Each observation represents
either a **sample** across genes
or a **gene** across samples



Algorithm AGGLOMERATIVEHIERARCHICALCLUSTERING

Input: D a set of observations $\vec{x}_1, \dots, \vec{x}_m$; $d(\vec{x}, \vec{y})$ a distance measure between observations

Output: A tree T of subsets of D

// Initialize a set \mathcal{D} of clusters D_1, \dots, D_m

$\mathcal{D} \leftarrow \{\{\vec{x}_1\}, \dots, \{\vec{x}_m\}\}$ // Initial clusters are tree leaves

$T \leftarrow$ a partial tree whose leaves are the \vec{x}_i 's

while $|\mathcal{D}| > 1$ **do**

 Choose pair of clusters (D_i, D_j) in \mathcal{D} such that $\text{Distance}(D_i, D_j, d)$ is minimal

 Define a new cluster $D_k = D_i \cup D_j$

$\mathcal{D} \leftarrow \mathcal{D} \cup D_k - \{D_i, D_j\}$

 Add D_k as parent node of D_i and D_j in the tree T

return T

Distance measure between clusters

Single-link or nearest neighbor rule

$$\text{Distance}(D_i, D_j, d) = \min_{\vec{x} \in D_i, \vec{y} \in D_j} d(\vec{x}, \vec{y})$$

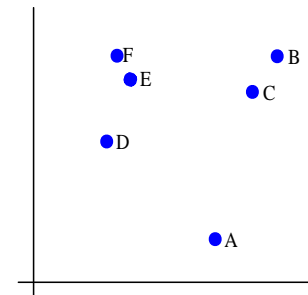
Complete-link or farthest neighbor rule

$$\text{Distance}(D_i, D_j, d) = \max_{\vec{x} \in D_i, \vec{y} \in D_j} d(\vec{x}, \vec{y})$$

Average-link rule

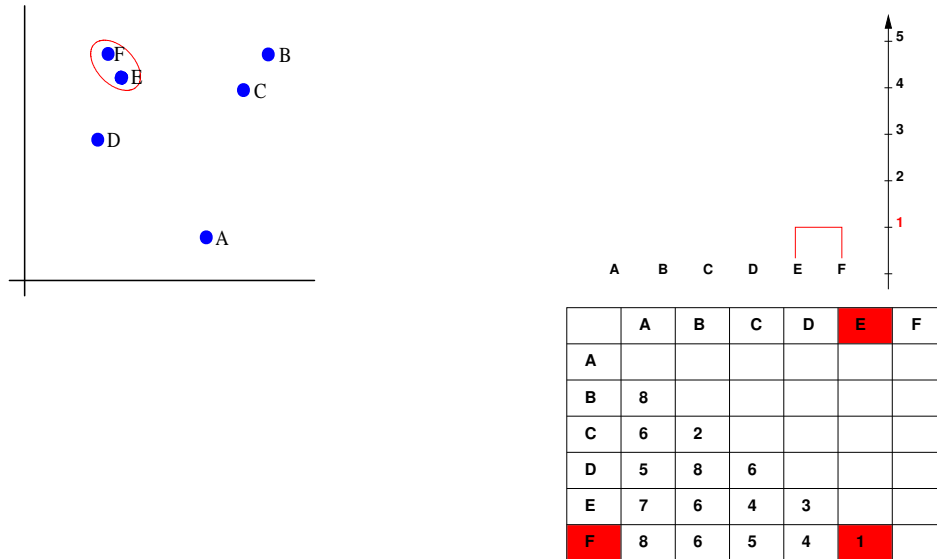
$$\text{Distance}(D_i, D_j, d) = \frac{1}{|D_i| \cdot |D_j|} \sum_{\vec{x} \in D_i, \vec{y} \in D_j} d(\vec{x}, \vec{y})$$

Hierarchical clustering example

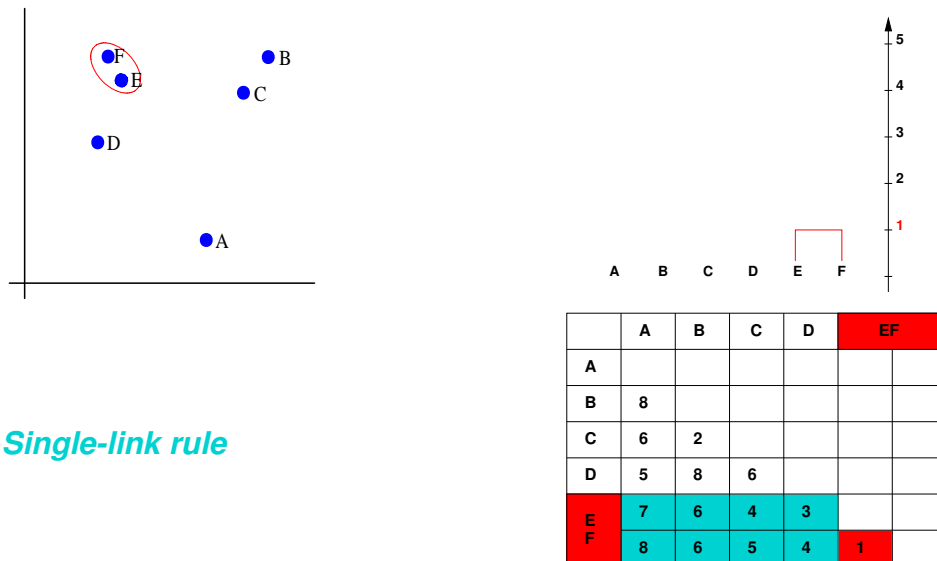


	A	B	C	D	E	F
A						
B	8					
C	6	2				
D	5	8	6			
E	7	6	4	3		
F	8	6	5	4	1	

Hierarchical clustering example

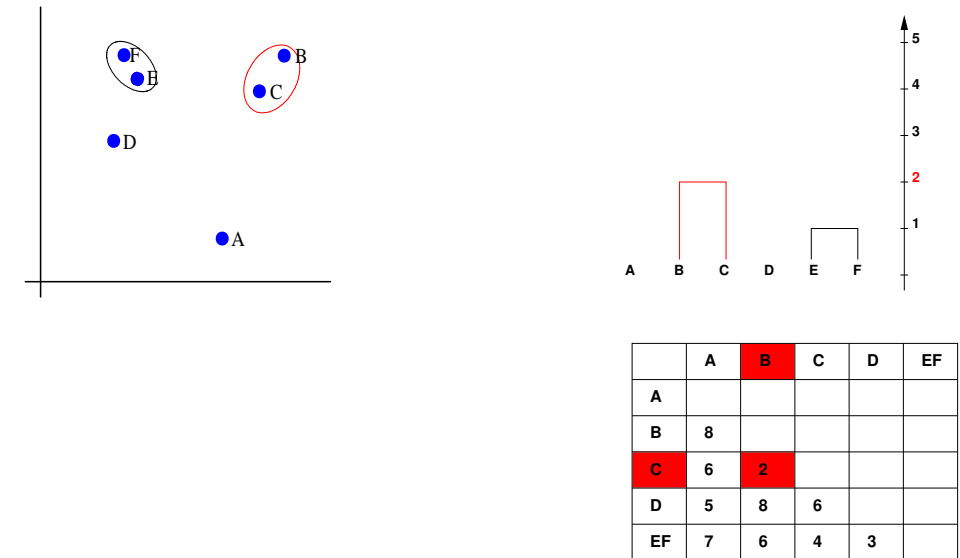


Hierarchical clustering example

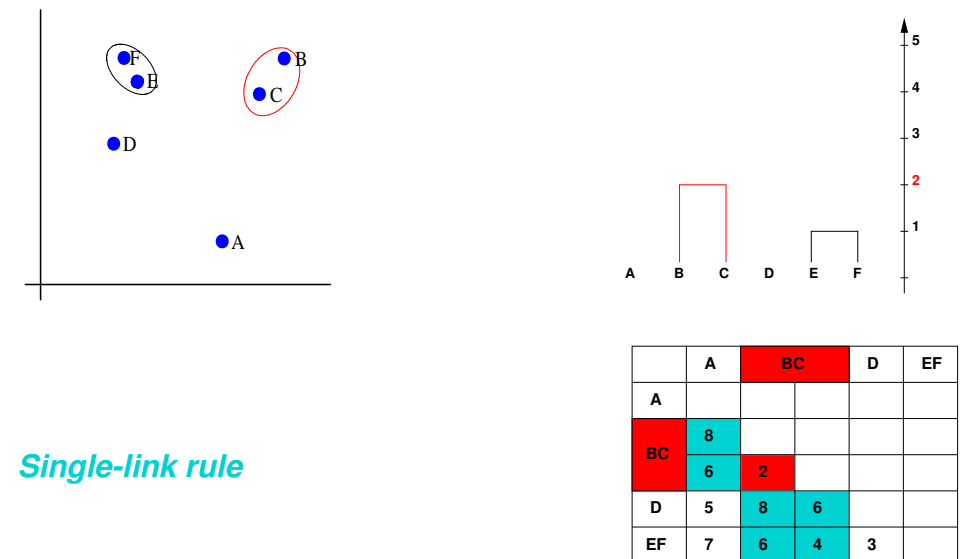


Single-link rule

Hierarchical clustering example

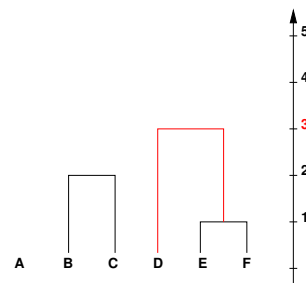
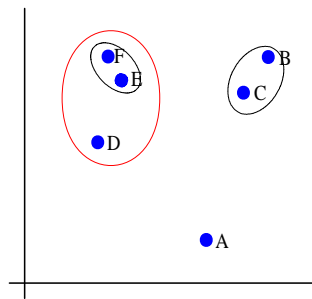


Hierarchical clustering example



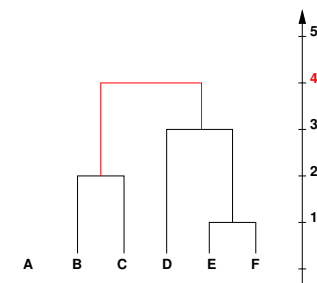
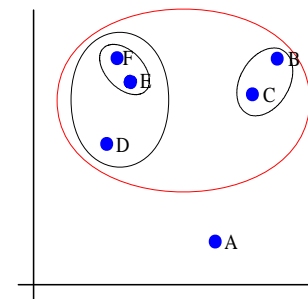
Single-link rule

Hierarchical clustering example



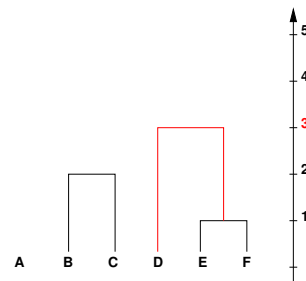
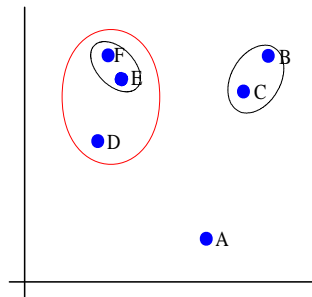
	A	BC	D	EF
A				
BC	6			
D	5	6		
EF	7	4	3	

Hierarchical clustering example



	A	BC	DEF
A			
BC	6		
DEF	5	4	

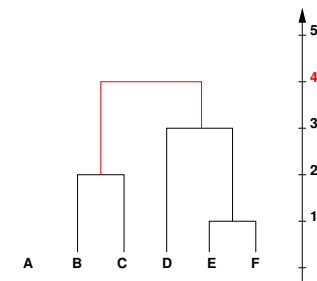
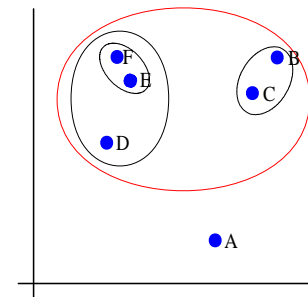
Hierarchical clustering example



	A	BC	DEF
A			
BC	6		
DEF	5	6	
	7	4	3

Single-link rule

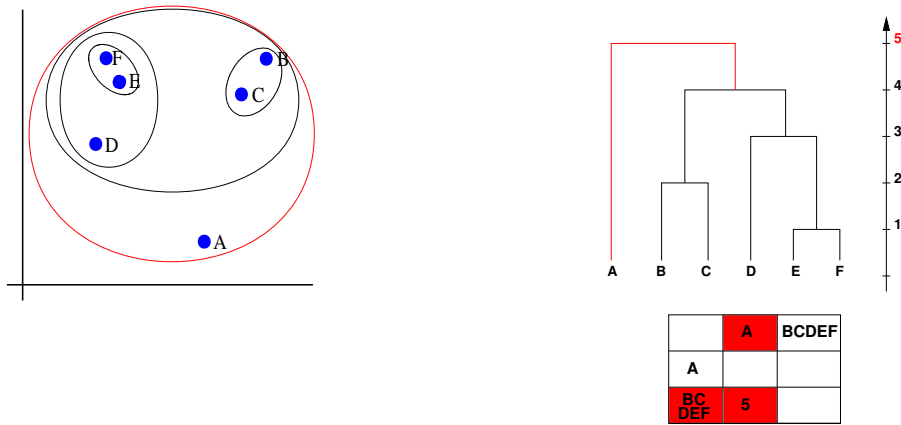
Hierarchical clustering example



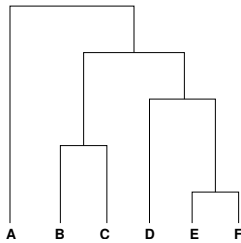
	A	BCDEF
A		
BC	6	
DEF	5	4

Single-link rule

Hierarchical clustering example



A note on phylogeny



This hierarchical clustering algorithm, with the average-link rule, is known as the UPGMA algorithm used in phylogeny

- The observations are (fragments) of sequences representative of some species, called **taxa**
- The pairwise distance measure is based on **alignment scores**, generally corrected according to an evolutionary model (e.g. Kimura)
- The final tree is interpreted as a **phylogenetic tree** and the branch length as representative of **time**

Outline

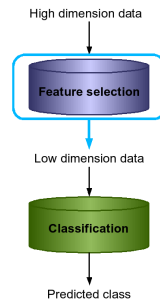
- 1 Introduction
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 - Wrappers
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Supervised gene selection

	gene 1	gene 2	...	gene p	class
sample 1	$x_{1,1}$	$x_{1,2}$...	$x_{1,p}$	+
sample 2	+
...
sample n-1	-
sample n	$x_{n,1}$	$x_{n,2}$...	$x_{n,p}$	-
test sample	x_1	x_2	...	x_p	?

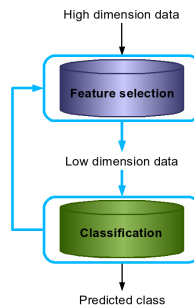
- we discuss binary classification first:
e.g. responders (+ or class 1) vs non-responders (- or class 2)
- samples can be indexed by their class label
 - means and variances can be computed on samples of a given class
- find a subset of **most discriminating genes** for the **prediction** of the class of any **new sample**

Feature selection: filters



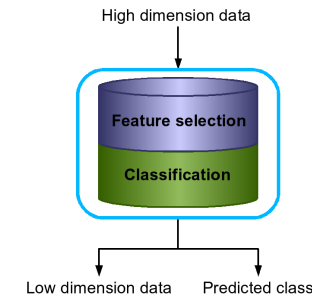
- Use only the training data + class labels during the feature selection step
- Standard techniques: **fold changes**, **t-Test**, **mutual information**, ...
- Train a single classifier taking the selected features as inputs
- The simplest and less computing intensive approach

Feature selection: wrappers



- Train a classifier on several subsets of all possible features
 - ▶ Exhaustive evaluation of all possible subsets is unfeasible
Note: there are $\mathcal{O}(2^p)$ subsets with $p \geq 10,000$
 - ▶ Typical solutions: use **feature ranking** or **forward/backward selection**
- Select the feature set that optimizes the performance of the trained classifier

Feature selection: embedded approaches

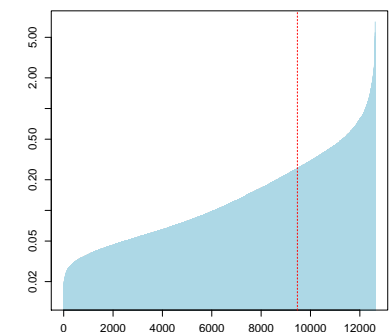


- Define the feature selection and the classifier estimation as a **combined optimization process**
- Include classifier optimization in the feature selection process
- More elegant/relevant but also more computing intensive than a filter

Non-specific filtering

	gene 1	gene 2	...	gene p	class
sample 1	$X_{1,1}$	$X_{1,2}$...	$X_{1,p}$	+
sample 2	+
...
sample n-1	-
sample n	$X_{n,1}$	$X_{n,2}$...	$X_{n,p}$	-

- genes with a small variance across all training samples are unlikely to be discriminating between classes
- keep only those **genes** (e.g. 25 %) with the **larger variances**
 - ▶ before normalization to unit variance!



Fold changes

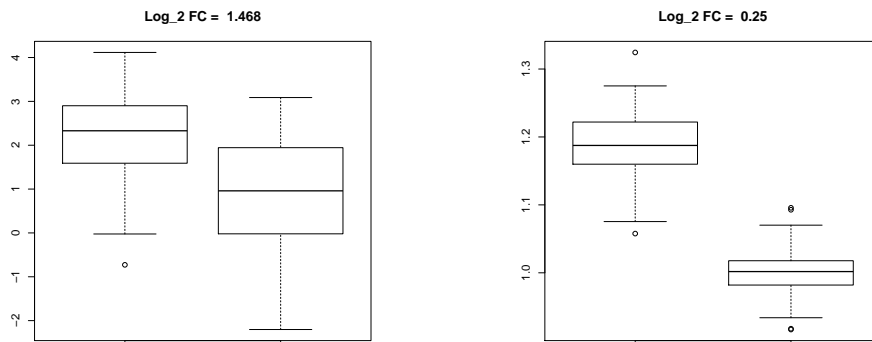
	gene 1	gene 2	...	gene p	class
sample 1	$x_{1,1}$	$x_{1,2}$...	$x_{1,p}$	+
sample 2	+
...
sample n-1	-
sample n	$x_{n,1}$	$x_{n,2}$...	$x_{n,p}$	-

Select genes with the **larger fold changes** between both conditions

$$\frac{\bar{x}_1}{\bar{x}_2} \text{ or } \log \frac{\bar{x}_1}{\bar{x}_2} = \log \bar{x}_1 - \log \bar{x}_2 \text{ or } \bar{x}_1 - \bar{x}_2$$

Comments on fold changes

- whenever $\bar{x}_1 < \bar{x}_2$, one considers a small value as important
 - ▶ $\log_2 \frac{\bar{x}_1}{\bar{x}_2}$ should be ≥ 1 or ≤ -1
- is a two-fold change significant?
 - ▶ dependence on the measurement technology
 - ▶ dependence on the class conditional variance



t-Test relevance index

- A **feature relevance** $J(x)$ can be defined according to the distance between the average feature value in each class
- The larger the distance the better, relatively to standard deviations

t-Test statistic (actually Welch t-statistics)

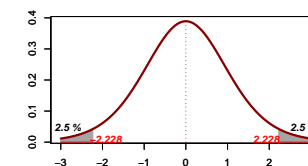
$$J(x) = \frac{\bar{x}_1 - \bar{x}_2}{\sqrt{S_1^2/n_1 + S_2^2/n_2}}$$

with n_1 (resp. n_2) the number of examples labeled as + (resp. -) and the estimated variances in each class $S_i^2 = \frac{1}{n_i-1} \sum_{j=1}^{n_i} (x_{ij} - \bar{x}_i)^2$

Confidence measure

- The Welch statistics follows a t -distribution with a number of degrees of freedom equal to:

$$\frac{(S_1^2/n_1 + S_2^2/n_2)^2}{(S_1^2/n_1)(n_1-1) + (S_2^2/n_2)(n_2-1)}$$
- p -values assess the significance of the difference between the two class means



- A feature is selected if its associated p -value is below a prescribed threshold (e.g. 5% $\Rightarrow |J(x)| \geq 2.228$ when d.f. = 10)

The R Project for Statistical Computing

An efficient way of computing p -values, and **many** other useful things...

<http://www.r-project.org/>

```
> t.test(x1,x2)
Welch Two Sample t-test
data:  x1 and x2
t = 0.9183, df = 7.002, p-value = 0.389
alternative hypothesis: true difference in means is not equal to 0
95 percent confidence interval:
 -0.908216  2.061549
sample estimates:
mean of x mean of y
 1.866667  1.290000
```

where x_1 (resp. x_2) is the vector of expression values of a given gene from samples labeled as class **1** (resp. class **2**)

p -value $> 0.05 \Rightarrow$ the difference between the 2 class means is not considered significant for this feature \Rightarrow discard the feature

Alternatives to a simple t-Test

- **Mann-Whitney** rank test is an alternative *non-parametric* test
- **ANOVA** offers a generalization of the t-Test in a multi-class (> 2) setting
- Pairwise t-tests between one class and the others is a common alternative
- **Kruskal-Wallis** is a generalization of Mann-Whitney to multi-class

The multiple test problem

A microarray experiment to distinguish between patients with a positive or a negative diagnosis

Among 50,000 gene expression values measured in each experiment, only those genes that are differently expressed, with a p -value $\leq 0.05 = \alpha$, are selected

The probability of type I error of a statistical test

Conclude that the mean expression values among the 2 classes are significantly different for a given gene while they are not \Rightarrow the feature is falsely selected with probability α

Test multiplicity

- The test will be performed for each gene \Rightarrow 50,000 times from the **same** experiment
- If $\alpha = 0.05$, we are expecting to select wrongly $50,000 \times .05 = 2,500$ genes !!

Multiple test correction

Bonferroni correction

Divide the critical value (e.g. $\alpha = .05$) by the number of tests n_t performed

Example: $\frac{\alpha}{n_t} = \frac{.05}{50,000} = 10^{-6}$

\Rightarrow only genes with associated p -values $\leq 10^{-6}$ are selected

Very conservative, often leads to select no feature

False Discovery Rate correction

Benjamini-Hochberg correction

- 1 Select a confidence level α (e.g. 0.05)
- 2 Rank the p -values (one for each feature) in **increasing** order
 $p_1 \leq p_2 \leq \dots \leq p_{n_t}$
- 3 Iterate over the n_t features
 ($n_t = p$ with data in \mathbb{R}^p , not to be confused with p -values)
 - Find the **maximal** index i such that $\frac{p_i \times n_t}{i} < \alpha$
- 4 Keep all features up to index i_{max}

Notes:

- If $p_{n_t} < 0.05$ FDR correction leads to select all features
- FDR correction is equivalent to Bonferroni correction whenever a single feature is selected: $p_1 \times n_t < \alpha \Leftrightarrow p_1 < \frac{\alpha}{n_t}$
- Those corrections do not change the relative ranking of features, just the selection threshold
- See R function `p.adjust`

Feature ranking with mutual information

$$I(X; Y) = - \sum_{ij} P(x_i, y_j) \log_2 \frac{P(x_i, y_j)}{P(x_i)P(y_j)}$$

$$= - \sum_{ij} P(x_i, y_j) \log_2 \frac{P(y_j|x_i)}{P(y_j)}$$

- A feature X is **more relevant** if its mutual information with the class value is **higher**
- If X tends to bring no information to predict Y then
 $P(y_j|x_i) \approx P(y_j)$ and $I(X; Y) \rightarrow 0$
- $I(X; Y) = 0$ **if and only if** X and Y are independent
- $I(X; Y)$ is **invariant under rescaling** of the variables X and Y
 (often rescaled to unit variance)

Univariate versus multivariate filters

- Correlation measures, t-Test (ANOVA), and $I(X; Y)$ are **univariate** filters
- Mutual information can be used to select **several variables** at a time $I(X_1, \dots, X_k; Y)$ **but** MI depends on the distributions $P(X_1, \dots, X_k, Y)$, $P(X_1, \dots, X_k)$ and $P(Y)$, which need to be reliably estimated
 - replace the joint problem by an approximation with a greedy selection of features

Maximum relevance minimum redundancy

[Peng et al., 05]

- 1 Select the feature with maximum mutual information with the response
 - $\hat{X} = \operatorname{argmax}_X I(X; Y)$
 - $\Phi = \{\hat{X}\}$ // Initialize the set of selected features
 - $F = \{X_1, \dots, X_p\} \setminus \{\hat{X}\}$ // The remaining set of features
- 2 Repeat

$$\hat{X} = \operatorname{argmax}_{X \in F} \left[\underbrace{I(X; Y)}_{\text{maximize relevance}} \right]$$

$$- \underbrace{\frac{1}{|\Phi|} \sum_{X_j \in \Phi} I(X; X_j)}_{\text{minimize redundancy}}$$

$\Phi \leftarrow \Phi \cup \{\hat{X}\}; F \leftarrow F \setminus \{\hat{X}\}$
until an appropriate number of features are selected

Filters in a nutshell



- Use only the training data + class labels during selection
- Filters offer interesting **baselines** which are **fast to compute**
- Popular **univariate** filters are based on a **t-Test** (with multiple test correction) or mutual information
 - ▶ they ignore the interactions between genes!
- Maximum relevance minimum redundancy is a popular **multivariate** extension

Are filters independent from a predictive model?



- The two step approach is sometimes considered as a benefit since the features are claimed to be selected **independently from** the subsequent classifier/regression **model**
 - ▶ Is it really better? (see embedded methods)
 - ▶ Is it really the case?

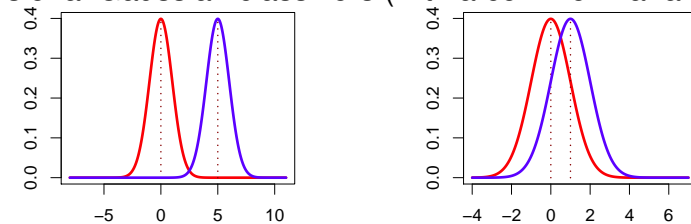
t-Test revisited

t-Test statistic

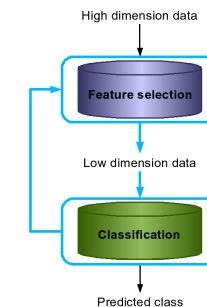
A feature x is selected whenever the difference between the class means is significant (after correction for multiplicity)

$$J(x) = \frac{\bar{x}_1 - \bar{x}_2}{\sqrt{S_1^2/n_1 + S_2^2/n_2}}$$

Equivalently, one easily discriminates between the classes using 2 uni-dimensional Gaussian classifiers (with a common variance)

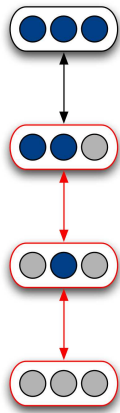


Wrapper principle

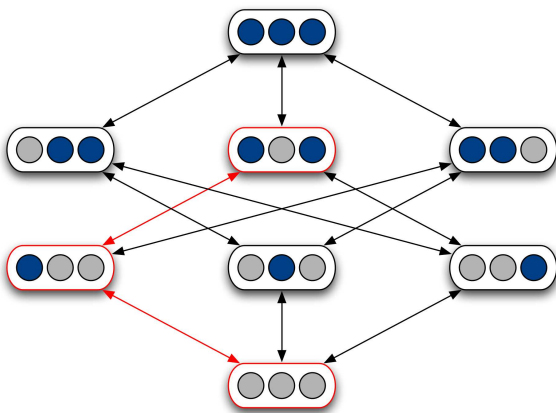


- **Estimate a classifier** from a given subset of all possible features
- Select the feature subset that **optimizes the performance** of the classifier (usually on an independent validation set)
 - ▶ Feature selection depends on the evaluation protocol of the classifier
 - ▶ There are $O(2^p)$ possible subsets

Univariate feature ranking



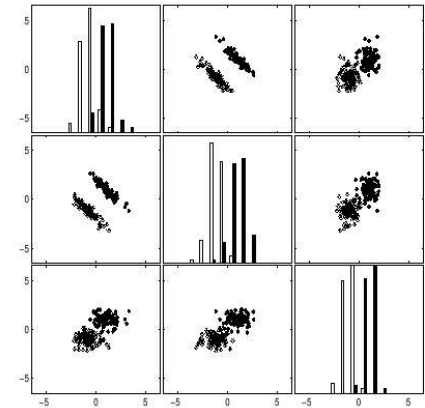
Multivariate Forward/Backward selection



- Forward selection goes bottom-up
- Backward selection goes top-down

Search order matters

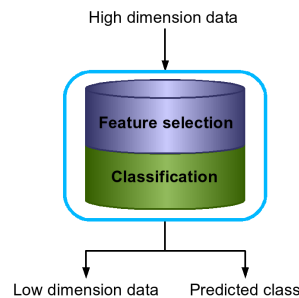
- x_3 alone is better than x_1 or x_2 alone, but x_1 together with x_2 offer the best discrimination
- **2 best features:**
 - ▶ Univariate feature ranking selects (x_3, x_2)
 - ▶ Forward selection selects (x_3, x_1)
 - ▶ Backward selection selects (x_1, x_2)
- **Single best feature:**
 - ▶ Forward selection or univariate feature ranking selects x_3
 - ▶ Backward selection selects x_2



Wrappers in a nutshell

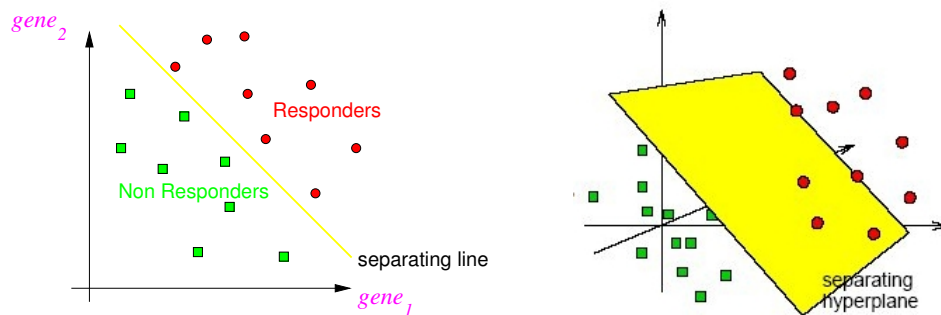
- A wrapper with **univariate feature ranking** offers a **good baseline**
 - ▶ The t-Test statistics can be used to rank features only (no need for multiple test correction nor fixing a confidence measure)
 - ▶ Classifier performance is used to decide how many features to keep
 - ▶ This can outperform a pure filter approach while not increasing much the computational cost
- A **backward selection** may be preferable over a forward selection, but should not be used to select just a few features (what “a few” means depends on the data...)
- More sophisticated search strategies are possible (backward + forward, randomized search, ...)
- If one can afford the computational cost of a multivariate selection, one should probably consider an **embedded approach**

Embedded Methods



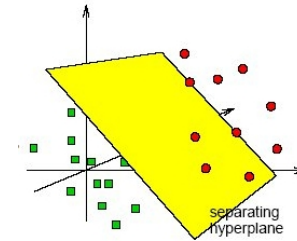
- Define the feature selection and the classifier estimation as a **combined optimization process**
- The features are selected as a by-product of the estimated classifier and its parameters

Linear Discriminants



- The actual number of dimensions may be $\approx 10,000$ for microarray classification
- The linear discriminant is a **hyperplane** in $\mathbb{R}^{\geq 10,000}$
- Decision rule: $\text{sign}(\sum_{j=1}^p w_j x_j + w_0) = \text{sign}(\mathbf{w}^T \mathbf{x})$ (with $x_0 \triangleq 1$)
 $\Rightarrow |w_j|$ is a measure of the importance of the j^{th} feature

Linear Separability

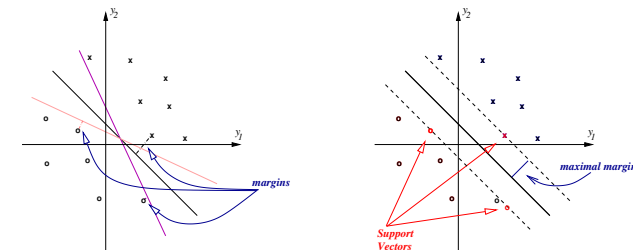


Facts

- The data is **linearly separable** if the two classes can be perfectly separated by a hyperplane
- A hyperplane in $\mathbb{R}^{10,000}$ can **separate perfectly** at least **10,001** (unaligned) points, given any possible 2 class labeling
- There is no problem to find a perfect linear separator of less than **100** points in $\mathbb{R}^{\geq 10,000}$ (e.g. for microarray data)
- The problem is that there are many *apparently perfect* models

Linear Support Vector Machines in a nutshell

- When the data is linearly separable the separating hyperplane is not unique but the **maximal margin hyperplane** separates the data with the largest margin
- For each separating hyperplane, there is an associated set of **support vectors**



Recursive Feature Elimination [Guyon *et al.* , 02]

Embedded Backward Selection

- 1 Estimate a SVM on a given set of dimensions
(initially p dimensions)
 - ▶ Decision rule: $\text{sign}(\sum_{j=1}^p w_j x_j + w_0)$
- 2 Consider $|w_j|$ as the relevance of the j^{th} dimension
- 3 Remove the least relevant dimension(s)
- 4 Iterate 1 to 3 on a reduced feature set

General Summary

- Feature selection aims at reducing the dimensionality of the data while preserving the interpretation of the original features
- **Filters** methods use only the data + class labels:
 - ▶ simple, fast, generally univariate (often an implicit use of a classifier)
- **Wrappers** take the **performance** of the classifier into account
 - ▶ Multivariate as soon as the classifier is multivariate
 - ▶ Often computing intensive
- **Embedded methods** take the **structure** of the classifier into account
 - ▶ More elegant and often faster than wrappers, not always better in terms of performance
 - ▶ A way to get an insight into a black-box classifier



Further information

- LINGI2262 Machine Learning: classification and evaluation (Semester 2)
- LELEC2870 Machine Learning: regression, dimensionality reduction and data visualization (Semester 1)
- LINGI2369 Artificial Intelligence and Machine Learning Seminar (Semester 1)

Further Reading I

-  Guyon, I., Gunn, S., Nikarvesh, M. and Zadeh, L.A. (editors) (2006). *Feature Extraction: Foundations and Applications*. Springer.
-  Hastie, T., Tibshirani, R., and Friedman, J. (2009). *The Elements of Statistical Learning: Data Mining, Inference, and Prediction*. (2nd edition), Springer.
-  Abeel, T., Helleputte, T., Dupont, P. and Saeys, Y. (2010) *Robust biomarker identification for cancer diagnosis with ensemble feature selection methods* *Bioinformatics*, Vol. 26 (3), pp. 392-398.
-  Bolstad, B.M., Irizarry, R.A., Astrand, M. and Speed, T.P. (2003) *A comparison of normalization methods for high density oligonucleotide array data based on variance and bias* *Bioinformatics*, Vol. 19 (2), pp. 185-193.

Further Reading II

-  Guyon, I., Weston, J., Barnhill, S., and Vapnik, V. (2002).
Gene selection for cancer classification using support vector machines.
Machine learning, **46**, 389–422.
-  Peng, H., Long, F., and Ding, C. (2005).
Feature selection based on mutual information criteria of max-dependency, max-relevance, and min-redundancy.
IEEE Transactions on Pattern Analysis and Machine Intelligence, Vol. 27, N° 8, pp. 1226-1238