

Class 8: PCA mini project

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Today we will do a complete analysis of some breast cancer biopsy data but first let's revisit the main PCA function in R, `prcomp()` and see what `scale=TRUE/FALSE` does.

```
head(mtcars)
```

	mpg	cyl	disp	hp	drat	wt	qsec	vs	am	gear	carb
Mazda RX4	21.0	6	160	110	3.90	2.620	16.46	0	1	4	4
Mazda RX4 Wag	21.0	6	160	110	3.90	2.875	17.02	0	1	4	4
Datsun 710	22.8	4	108	93	3.85	2.320	18.61	1	1	4	1
Hornet 4 Drive	21.4	6	258	110	3.08	3.215	19.44	1	0	3	1
Hornet Sportabout	18.7	8	360	175	3.15	3.440	17.02	0	0	3	2
Valiant	18.1	6	225	105	2.76	3.460	20.22	1	0	3	1

Find the mean and sd value per column of this data set:

```
apply(mtcars, 2, mean)
```

mpg	cyl	disp	hp	drat	wt	qsec
20.090625	6.187500	230.721875	146.687500	3.596563	3.217250	17.848750
vs	am	gear	carb			
0.437500	0.406250	3.687500	2.812500			

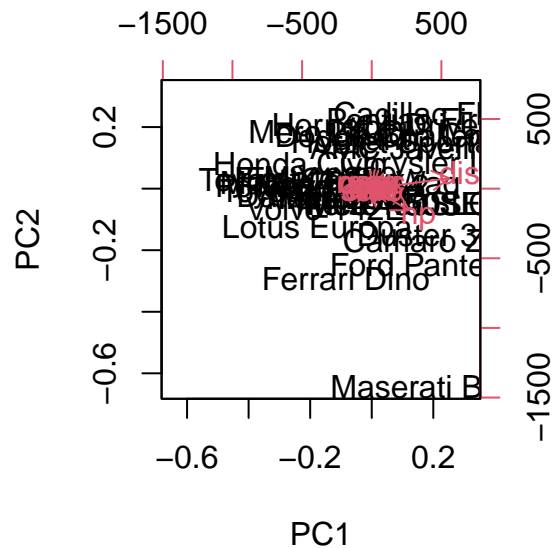
```
apply(mtcars, 2, sd)
```

mpg	cyl	disp	hp	drat	wt
6.0269481	1.7859216	123.9386938	68.5628685	0.5346787	0.9784574
qsec	vs	am	gear	carb	
1.7869432	0.5040161	0.4989909	0.7378041	1.6152000	

It is clear that “disp” and “hp” have the highest mean values and the highest standard deviation. They will likely dominate any analysis I do on this dataset. Let’s see:

```
pc.noscale <- prcomp(mtcars, scale=F)
pc.scale <- prcomp(mtcars, scale=T)
```

```
biplot(pc.noscale)
```



```
pc.noscale$rotation[,1]
```

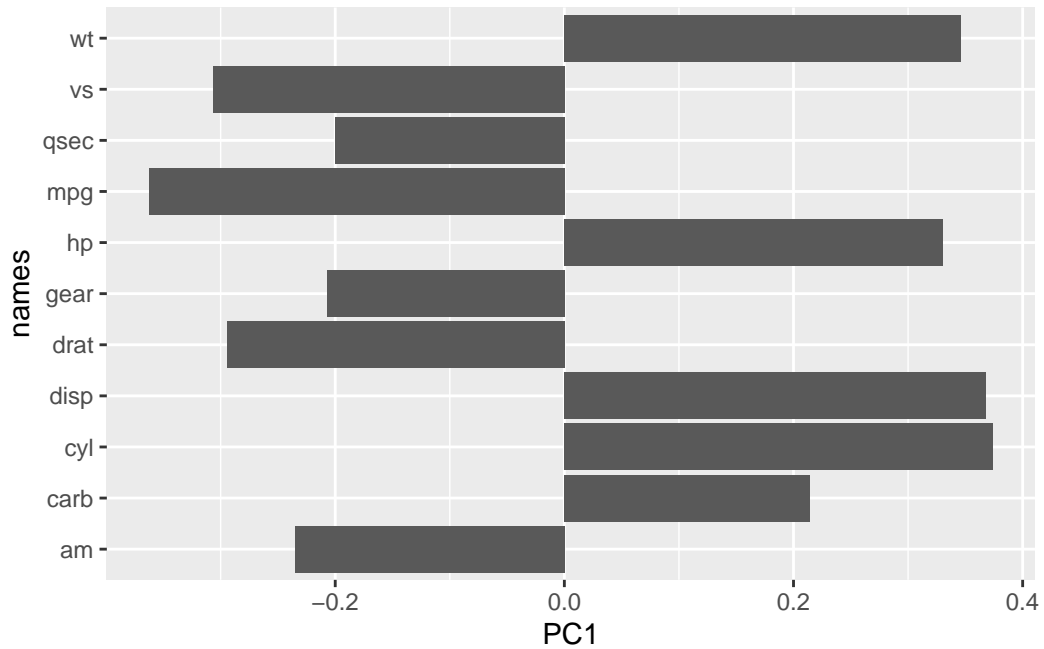
mpg	cyl	disp	hp	drat	wt
-0.038118199	0.012035150	0.899568146	0.434784387	-0.002660077	0.006239405
qsec	vs	am	gear	carb	
-0.006671270	-0.002729474	-0.001962644	-0.002604768	0.005766010	

Plot the loadings:

```
library(ggplot2)

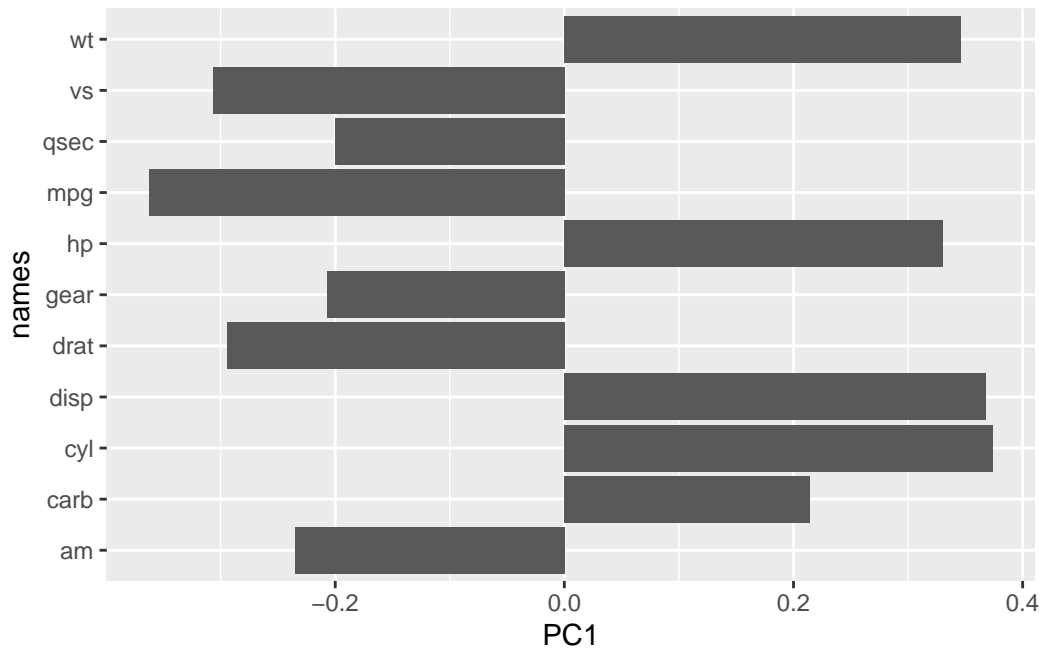
r2 <- as.data.frame(pc.scale$rotation)
r2$names <- rownames(pc.scale$rotation)
```

```
ggplot(r2) +
  aes(PC1, names) +
  geom_col()
```

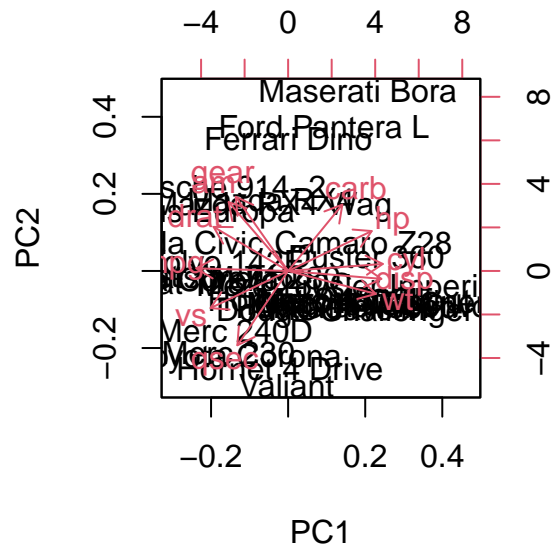


```
r1 <- as.data.frame(pc.scale$rotation)
r1$names <- rownames(pc.scale$rotation)

ggplot(r1) +
  aes(PC1, names) +
  geom_col()
```



```
biplot(pc.scale)
```



Take-Home: Generally we always want to set `scale=TRUE` when we do this type

of analysis to avoid our analysis being dominated by individual variables with the largest variance just due to their unit of measurement.

FNA breast cancer data

Preparing the data

Load the data into R:

```
wisc.df <- read.csv("WisconsinCancer.csv", row.names = 1)
head(wisc.df)
```

	diagnosis	radius_mean	texture_mean	perimeter_mean	area_mean
842302	M	17.99	10.38	122.80	1001.0
842517	M	20.57	17.77	132.90	1326.0
84300903	M	19.69	21.25	130.00	1203.0
84348301	M	11.42	20.38	77.58	386.1
84358402	M	20.29	14.34	135.10	1297.0
843786	M	12.45	15.70	82.57	477.1

	smoothness_mean	compactness_mean	concavity_mean	concave.points_mean
842302	0.11840	0.27760	0.3001	0.14710
842517	0.08474	0.07864	0.0869	0.07017
84300903	0.10960	0.15990	0.1974	0.12790
84348301	0.14250	0.28390	0.2414	0.10520
84358402	0.10030	0.13280	0.1980	0.10430
843786	0.12780	0.17000	0.1578	0.08089

	symmetry_mean	fractal_dimension_mean	radius_se	texture_se	perimeter_se
842302	0.2419	0.07871	1.0950	0.9053	8.589
842517	0.1812	0.05667	0.5435	0.7339	3.398
84300903	0.2069	0.05999	0.7456	0.7869	4.585
84348301	0.2597	0.09744	0.4956	1.1560	3.445
84358402	0.1809	0.05883	0.7572	0.7813	5.438
843786	0.2087	0.07613	0.3345	0.8902	2.217

	area_se	smoothness_se	compactness_se	concavity_se	concave.points_se
842302	153.40	0.006399	0.04904	0.05373	0.01587
842517	74.08	0.005225	0.01308	0.01860	0.01340
84300903	94.03	0.006150	0.04006	0.03832	0.02058
84348301	27.23	0.009110	0.07458	0.05661	0.01867
84358402	94.44	0.011490	0.02461	0.05688	0.01885
843786	27.19	0.007510	0.03345	0.03672	0.01137

	symmetry_se	fractal_dimension_se	radius_worst	texture_worst
--	-------------	----------------------	--------------	---------------

842302	0.03003		0.006193	25.38	17.33
842517	0.01389		0.003532	24.99	23.41
84300903	0.02250		0.004571	23.57	25.53
84348301	0.05963		0.009208	14.91	26.50
84358402	0.01756		0.005115	22.54	16.67
843786	0.02165		0.005082	15.47	23.75
	perimeter_worst	area_worst	smoothness_worst	compactness_worst	
842302	184.60	2019.0	0.1622		0.6656
842517	158.80	1956.0	0.1238		0.1866
84300903	152.50	1709.0	0.1444		0.4245
84348301	98.87	567.7	0.2098		0.8663
84358402	152.20	1575.0	0.1374		0.2050
843786	103.40	741.6	0.1791		0.5249
	concavity_worst	concave.points_worst	symmetry_worst		
842302	0.7119		0.2654		0.4601
842517	0.2416		0.1860		0.2750
84300903	0.4504		0.2430		0.3613
84348301	0.6869		0.2575		0.6638
84358402	0.4000		0.1625		0.2364
843786	0.5355		0.1741		0.3985
	fractal_dimension_worst				
842302		0.11890			
842517		0.08902			
84300903		0.08758			
84348301		0.17300			
84358402		0.07678			
843786		0.12440			

Exploratory data analysis

Q1. How many observations are in this data-set?

```
nrow(wisc.df)
```

```
[1] 569
```

Q2. How many of the observations have a malignant diagnosis?

The `table()` function is super useful here:

```
table(wisc.df$diagnosis)
```

```
  B    M  
357 212
```

or you could do...

```
sum(wisc.df$diagnosis == "M")
```

```
[1] 212
```

Q3. How many variables/features in the data are suffixed with _mean?

First how many columns do we have?

```
ncol(wisc.df)
```

```
[1] 31
```

Now we want to know the names of the columns.

```
colnames(wisc.df)
```

```
[1] "diagnosis"          "radius_mean"  
[3] "texture_mean"       "perimeter_mean"  
[5] "area_mean"          "smoothness_mean"  
[7] "compactness_mean"   "concavity_mean"  
[9] "concave.points_mean" "symmetry_mean"  
[11] "fractal_dimension_mean" "radius_se"  
[13] "texture_se"         "perimeter_se"  
[15] "area_se"            "smoothness_se"  
[17] "compactness_se"     "concavity_se"  
[19] "concave.points_se"  "symmetry_se"  
[21] "fractal_dimension_se" "radius_worst"  
[23] "texture_worst"      "perimeter_worst"  
[25] "area_worst"         "smoothness_worst"  
[27] "compactness_worst"  "concavity_worst"  
[29] "concave.points_worst" "symmetry_worst"  
[31] "fractal_dimension_worst"
```

A useful function for this is the `grep()` function.

```
m <- grep("_mean", colnames(wisc.df) )  
length(m)
```

```
[1] 10
```

Before we go any further, we need to exclude the diagnoses column from any future analysis - this tells us whether a sample is cancer or non-cancer.

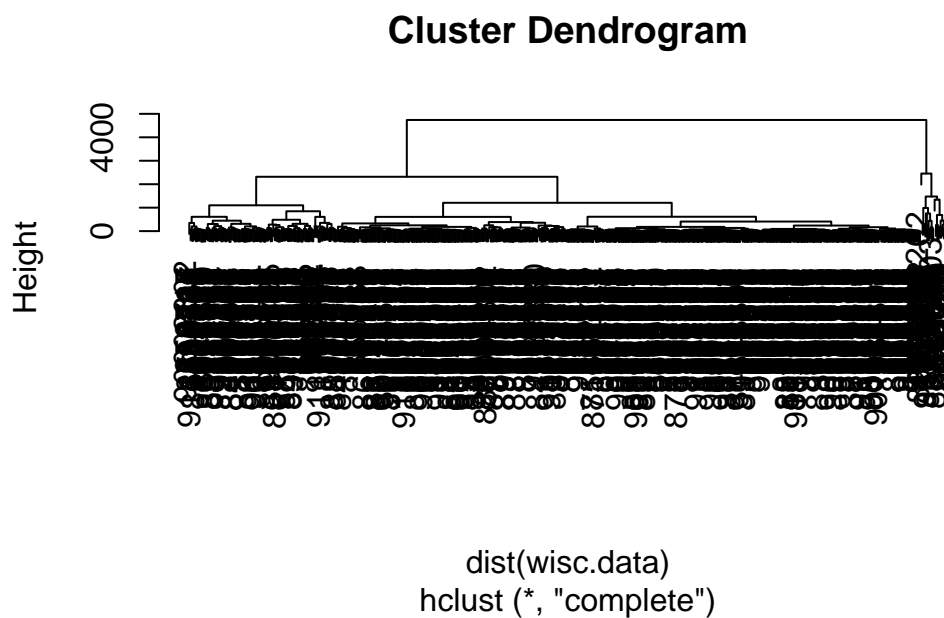
```
diagnosis <- as.factor(wisc.df$diagnosis)  
head(diagnosis)
```

```
[1] M M M M M M  
Levels: B M
```

```
wisc.data <- wisc.df[,-1]
```

Let's see if we can cluster the `wisc.data` to find some structure in the data-set.

```
hc <- hclust( dist(wisc.data) )  
plot(hc)
```



Principal Component Analysis (PCA)

Performing PCA

```
wisc.pr <- prcomp( wisc.data, scale=T)
summary(wisc.pr)
```

Importance of components:

	PC1	PC2	PC3	PC4	PC5	PC6	PC7
Standard deviation	3.6444	2.3857	1.67867	1.40735	1.28403	1.09880	0.82172
Proportion of Variance	0.4427	0.1897	0.09393	0.06602	0.05496	0.04025	0.02251
Cumulative Proportion	0.4427	0.6324	0.72636	0.79239	0.84734	0.88759	0.91010

	PC8	PC9	PC10	PC11	PC12	PC13	PC14
Standard deviation	0.69037	0.6457	0.59219	0.5421	0.51104	0.49128	0.39624
Proportion of Variance	0.01589	0.0139	0.01169	0.0098	0.00871	0.00805	0.00523
Cumulative Proportion	0.92598	0.9399	0.95157	0.9614	0.97007	0.97812	0.98335

	PC15	PC16	PC17	PC18	PC19	PC20	PC21
Standard deviation	0.30681	0.28260	0.24372	0.22939	0.22244	0.17652	0.1731
Proportion of Variance	0.00314	0.00266	0.00198	0.00175	0.00165	0.00104	0.0010
Cumulative Proportion	0.98649	0.98915	0.99113	0.99288	0.99453	0.99557	0.9966

	PC22	PC23	PC24	PC25	PC26	PC27	PC28
Standard deviation	0.16565	0.15602	0.1344	0.12442	0.09043	0.08307	0.03987
Proportion of Variance	0.00091	0.00081	0.0006	0.00052	0.00027	0.00023	0.00005
Cumulative Proportion	0.99749	0.99830	0.9989	0.99942	0.99969	0.99992	0.99997

	PC29	PC30
Standard deviation	0.02736	0.01153
Proportion of Variance	0.00002	0.00000
Cumulative Proportion	1.00000	1.00000

Q4. From your results, what proportion of the original variance is captured by the first principal components (PC1)?

```
pr.var <- wisc.pr$sdev^2
pve <- pr.var/sum(pr.var)
pve[1]
```

```
[1] 0.4427203
```

Q5. How many principal components (PCs) are required to describe at least 70% of the original variance in the data?

```
summary(wisc.pr)
```

Importance of components:

	PC1	PC2	PC3	PC4	PC5	PC6	PC7
Standard deviation	3.6444	2.3857	1.67867	1.40735	1.28403	1.09880	0.82172
Proportion of Variance	0.4427	0.1897	0.09393	0.06602	0.05496	0.04025	0.02251
Cumulative Proportion	0.4427	0.6324	0.72636	0.79239	0.84734	0.88759	0.91010

	PC8	PC9	PC10	PC11	PC12	PC13	PC14
Standard deviation	0.69037	0.6457	0.59219	0.5421	0.51104	0.49128	0.39624
Proportion of Variance	0.01589	0.0139	0.01169	0.0098	0.00871	0.00805	0.00523
Cumulative Proportion	0.92598	0.9399	0.95157	0.9614	0.97007	0.97812	0.98335

	PC15	PC16	PC17	PC18	PC19	PC20	PC21
Standard deviation	0.30681	0.28260	0.24372	0.22939	0.22244	0.17652	0.1731
Proportion of Variance	0.00314	0.00266	0.00198	0.00175	0.00165	0.00104	0.0010
Cumulative Proportion	0.98649	0.98915	0.99113	0.99288	0.99453	0.99557	0.9966

	PC22	PC23	PC24	PC25	PC26	PC27	PC28
Standard deviation	0.16565	0.15602	0.1344	0.12442	0.09043	0.08307	0.03987
Proportion of Variance	0.00091	0.00081	0.0006	0.00052	0.00027	0.00023	0.00005
Cumulative Proportion	0.99749	0.99830	0.9989	0.99942	0.99969	0.99992	0.99997

	PC29	PC30
Standard deviation	0.02736	0.01153
Proportion of Variance	0.00002	0.00000
Cumulative Proportion	1.00000	1.00000

```
which(cumsum(pve) > 0.7)[1]
```

```
[1] 3
```

```
sum(pve[1:3])
```

```
[1] 0.7263637
```

Three PCs are required to describe at least 70% of the original variance in the data.

Q6. How many principal components (PCs) are required to describe at least 90% of the original variance in the data?

```
summary(wisc.pr)
```

Importance of components:

	PC1	PC2	PC3	PC4	PC5	PC6	PC7
Standard deviation	3.6444	2.3857	1.67867	1.40735	1.28403	1.09880	0.82172
Proportion of Variance	0.4427	0.1897	0.09393	0.06602	0.05496	0.04025	0.02251
Cumulative Proportion	0.4427	0.6324	0.72636	0.79239	0.84734	0.88759	0.91010
	PC8	PC9	PC10	PC11	PC12	PC13	PC14
Standard deviation	0.69037	0.6457	0.59219	0.5421	0.51104	0.49128	0.39624
Proportion of Variance	0.01589	0.0139	0.01169	0.0098	0.00871	0.00805	0.00523
Cumulative Proportion	0.92598	0.9399	0.95157	0.9614	0.97007	0.97812	0.98335
	PC15	PC16	PC17	PC18	PC19	PC20	PC21
Standard deviation	0.30681	0.28260	0.24372	0.22939	0.22244	0.17652	0.1731
Proportion of Variance	0.00314	0.00266	0.00198	0.00175	0.00165	0.00104	0.0010
Cumulative Proportion	0.98649	0.98915	0.99113	0.99288	0.99453	0.99557	0.9966
	PC22	PC23	PC24	PC25	PC26	PC27	PC28
Standard deviation	0.16565	0.15602	0.1344	0.12442	0.09043	0.08307	0.03987
Proportion of Variance	0.00091	0.00081	0.0006	0.00052	0.00027	0.00023	0.00005
Cumulative Proportion	0.99749	0.99830	0.9989	0.99942	0.99969	0.99992	0.99997
	PC29	PC30					
Standard deviation	0.02736	0.01153					
Proportion of Variance	0.00002	0.00000					
Cumulative Proportion	1.00000	1.00000					

```
which(cumsum(pve) > 0.9)[1]
```

```
[1] 7
```

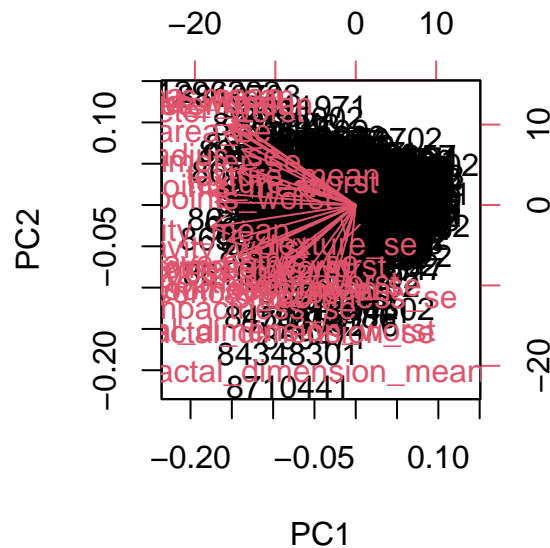
```
sum(pve[1:7])
```

```
[1] 0.9100953
```

Seven PCs are required to describe at least 90% of the original variance in the data

Interpreting PCA results

```
biplot(wisc.pr)
```



Q7. What stands out to you about this plot? Is it easy or difficult to understand? Why?

The overlapping of text and numbers stands out to me about this plot. It is very difficult to understand because of the excessive overlapping of data points.

This biplot sucks! We need to build our own PCA score plot of PC1 vs PC2.

```
attributes(wisc.pr)
```

```
$names
[1] "sdev"      "rotation" "center"    "scale"     "x"

$class
[1] "prcomp"
```

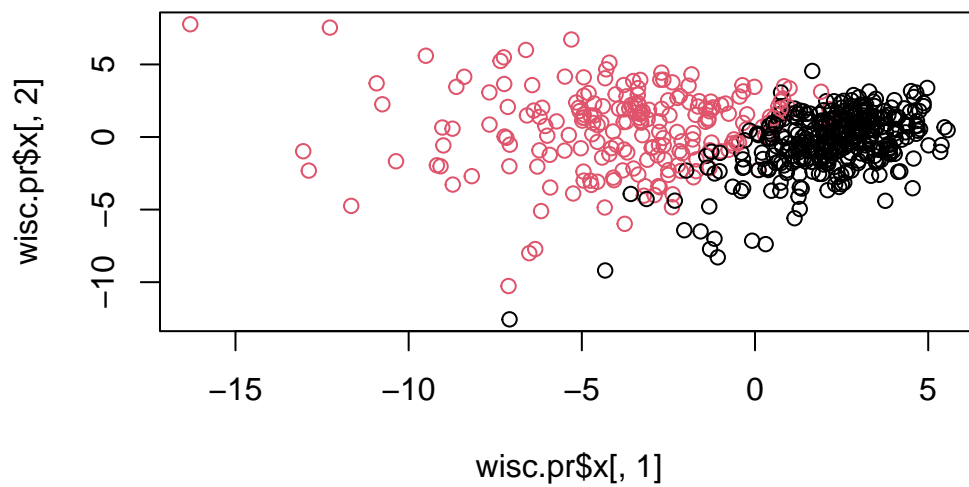
```
head(wisc.pr$x)
```

	PC1	PC2	PC3	PC4	PC5	PC6
842302	-9.184755	-1.946870	-1.1221788	3.6305364	1.1940595	1.41018364
842517	-2.385703	3.764859	-0.5288274	1.1172808	-0.6212284	0.02863116
84300903	-5.728855	1.074229	-0.5512625	0.9112808	0.1769302	0.54097615
84348301	-7.116691	-10.266556	-3.2299475	0.1524129	2.9582754	3.05073750

84358402	-3.931842	1.946359	1.3885450	2.9380542	-0.5462667	-1.22541641						
843786	-2.378155	-3.946456	-2.9322967	0.9402096	1.0551135	-0.45064213						
	PC7	PC8	PC9	PC10	PC11	PC12						
842302	2.15747152	0.39805698	-0.15698023	-0.8766305	-0.2627243	-0.8582593						
842517	0.01334635	-0.24077660	-0.71127897	1.1060218	-0.8124048	0.1577838						
84300903	-0.66757908	-0.09728813	0.02404449	0.4538760	0.6050715	0.1242777						
84348301	1.42865363	-1.05863376	-1.40420412	-1.1159933	1.1505012	1.0104267						
84358402	-0.93538950	-0.63581661	-0.26357355	0.3773724	-0.6507870	-0.1104183						
843786	0.49001396	0.16529843	-0.13335576	-0.5299649	-0.1096698	0.0813699						
	PC13	PC14	PC15	PC16	PC17							
842302	0.10329677	-0.690196797	0.601264078	0.74446075	-0.26523740							
842517	-0.94269981	-0.652900844	-0.008966977	-0.64823831	-0.01719707							
84300903	-0.41026561	0.016665095	-0.482994760	0.32482472	0.19075064							
84348301	-0.93245070	-0.486988399	0.168699395	0.05132509	0.48220960							
84358402	0.38760691	-0.538706543	-0.310046684	-0.15247165	0.13302526							
843786	-0.02625135	0.003133944	-0.178447576	-0.01270566	0.19671335							
	PC18	PC19	PC20	PC21	PC22							
842302	-0.54907956	0.1336499	0.34526111	0.096430045	-0.06878939							
842517	0.31801756	-0.2473470	-0.11403274	-0.077259494	0.09449530							
84300903	-0.08789759	-0.3922812	-0.20435242	0.310793246	0.06025601							
84348301	-0.03584323	-0.0267241	-0.46432511	0.433811661	0.20308706							
84358402	-0.01869779	0.4610302	0.06543782	-0.116442469	0.01763433							
843786	-0.29727706	-0.1297265	-0.07117453	-0.002400178	0.10108043							
	PC23	PC24	PC25	PC26	PC27							
842302	0.08444429	0.175102213	0.150887294	-0.201326305	-0.25236294							
842517	-0.21752666	-0.011280193	0.170360355	-0.041092627	0.18111081							
84300903	-0.07422581	-0.102671419	-0.171007656	0.004731249	0.04952586							
84348301	-0.12399554	-0.153294780	-0.077427574	-0.274982822	0.18330078							
84358402	0.13933105	0.005327110	-0.003059371	0.039219780	0.03213957							
843786	0.03344819	-0.002837749	-0.122282765	-0.030272333	-0.08438081							
	PC28	PC29	PC30									
842302	-0.0338846387	0.045607590	0.0471277407									
842517	0.0325955021	-0.005682424	0.0018662342									
84300903	0.0469844833	0.003143131	-0.0007498749									
84348301	0.0424469831	-0.069233868	0.0199198881									
84358402	-0.0347556386	0.005033481	-0.0211951203									
843786	0.0007296587	-0.019703996	-0.0034564331									

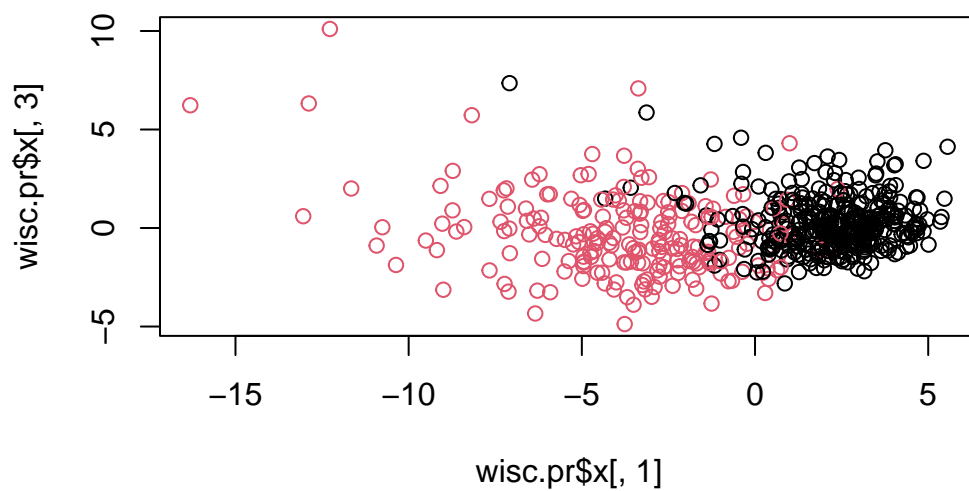
Plot of PC1 vs PC2 the first two columns

```
plot( wisc.pr$x[,1], wisc.pr$x[,2], col=diagnosis )
```



Q8. Generate a similar plot for principal components 1 and 3. What do you notice about these plots?

```
plot( wisc.pr$x[,1], wisc.pr$x[,3], col=diagnosis )
```

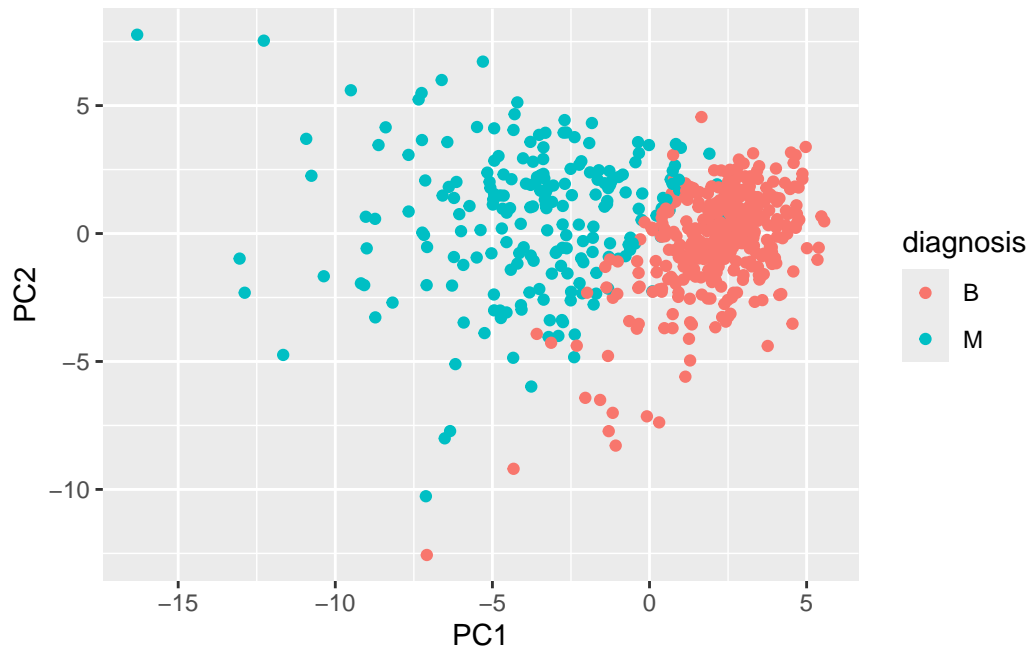


The values on the y-axis of this plot are much lower than the values on the y-axis of the other plot. PC3 is much different from PC2.

Make a ggplot version of the first score plot

```
pc <- as.data.frame(wisc.pr$x)

ggplot(pc) +
  aes(PC1, PC2, col=diagnosis)+
  geom_point()
```



Communicating PCA results

Q9. For the first principal component, what is the component of the loading vector (i.e. `wisc.pr$rotation[,1]`) for the feature `concave.points_mean`?

```
wisc.pr$rotation["concave.points_mean",1]
```

```
[1] -0.2608538
```

Q10. What is the minimum number of principal components required to explain 80% of the variance of the data?

```
which(cumsum(pve) > 0.8)[1]
```

```
[1] 5
```

Hierarchical clustering

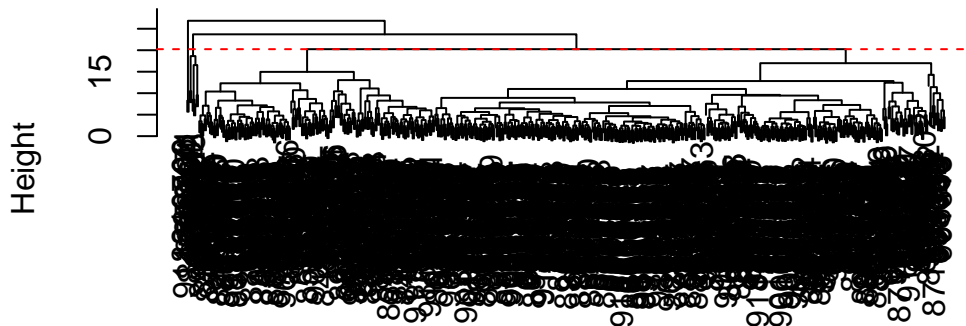
Results of hierarchical clustering

```
data.scaled <- scale(wisc.data)
data.dist <- dist(data.scaled)
wisc.hclust <- hclust(data.dist)
```

Q11. Using the `plot()` and `abline()` functions, what is the height at which the clustering model has 4 clusters?

```
plot(wisc.hclust)
heights <- sort(wisc.hclust$height, decreasing = TRUE)
height_for_4_clusters <- heights[3]
abline(h = height_for_4_clusters, col="red", lty=2)
```

Cluster Dendrogram



```
data.dist
hclust(*, "complete")
```

At height 3, is the height at which the clustering model has 4 clusters.

Selecting number of clusters

```
wisc.hclust.clusters <- cutree(wisc.hclust, k=4)
table(wisc.hclust.clusters, diagnosis)
```

	diagnosis		
wisc.hclust.clusters	B	M	
1	12	165	
2	2	5	
3	343	40	
4	0	2	

Q12. Can you find a better cluster vs diagnoses match by cutting into a different number of clusters between 2 and 10?

```
for (k in 2:10) {
  clusters <- cutree(wisc.hclust, k = k)
  cat("\nNumber of Clusters:", k, "\n")
  print(table(clusters, diagnosis))
}
```

Number of Clusters: 2

	diagnosis		
clusters	B	M	
1	357	210	
2	0	2	

Number of Clusters: 3

	diagnosis		
clusters	B	M	
1	355	205	
2	2	5	
3	0	2	

Number of Clusters: 4

	diagnosis		
clusters	B	M	
1	12	165	
2	2	5	

3	343	40
4	0	2

Number of Clusters: 5

	diagnosis		
clusters	B	M	
1	12	165	
2	0	5	
3	343	40	
4	2	0	
5	0	2	

Number of Clusters: 6

	diagnosis		
clusters	B	M	
1	12	165	
2	0	5	
3	331	39	
4	2	0	
5	12	1	
6	0	2	

Number of Clusters: 7

	diagnosis		
clusters	B	M	
1	12	165	
2	0	3	
3	331	39	
4	2	0	
5	12	1	
6	0	2	
7	0	2	

Number of Clusters: 8

	diagnosis		
clusters	B	M	
1	12	86	
2	0	79	
3	0	3	
4	331	39	
5	2	0	
6	12	1	
7	0	2	

8 0 2

Number of Clusters: 9

	diagnosis	
clusters	B	M
1	12	86
2	0	79
3	0	3
4	331	39
5	2	0
6	12	0
7	0	2
8	0	2
9	0	1

Number of Clusters: 10

	diagnosis	
clusters	B	M
1	12	86
2	0	59
3	0	3
4	331	39
5	0	20
6	2	0
7	12	0
8	0	2
9	0	2
10	0	1

Using different methods

Q13. Which method gives your favorite results for the same data.dist data-set? Explain your reasoning.

Ward.D2 method is my favorite method because it creates compact, evenly-sized clusters. This method also reduces variance within each cluster and aligns better with classifications like, benign vs. malignant tumors in this data set.

OPTIONAL: K-means clustering

K-means clustering and comparing results

```
wisc.km <- kmeans(wisc.data, centers= 2, nstart= 2)
table(wisc.km$cluster, diagnosis)
```

```
      diagnosis
      B      M
1      1 130
2    356   82
```

```
table(wisc.km$cluster, wisc.hclust.clusters)
```

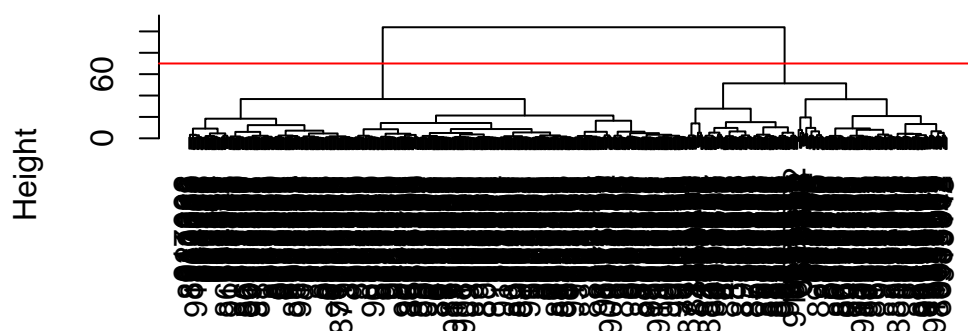
```
      wisc.hclust.clusters
      1      2      3      4
1 109      2     18      2
2   68      5    365      0
```

Combining methods

Clustering in PC space

```
hc <- hclust(dist(wisc.pr$x[,1:2]), method="ward.D2")
plot(hc)
abline(h=70, col="red")
```

Cluster Dendrogram



```
dist(wisc.pr$x[, 1:2])
hclust (*, "ward.D2")
```

Cluster membership vector

```
grps <- cutree(hc, h=70, k=2)
table(grps)
```

```
grps
  1   2
195 374
```

```
table(diagnosis)
```

```
diagnosis
  B   M
357 212
```

Cross-table to see how my clustering groups correspond to the expert diagnosis vector of M and B values.

```
table(grps, diagnosis)
```

```

diagnosis
grps  B  M
1   18 177
2  339  35

```

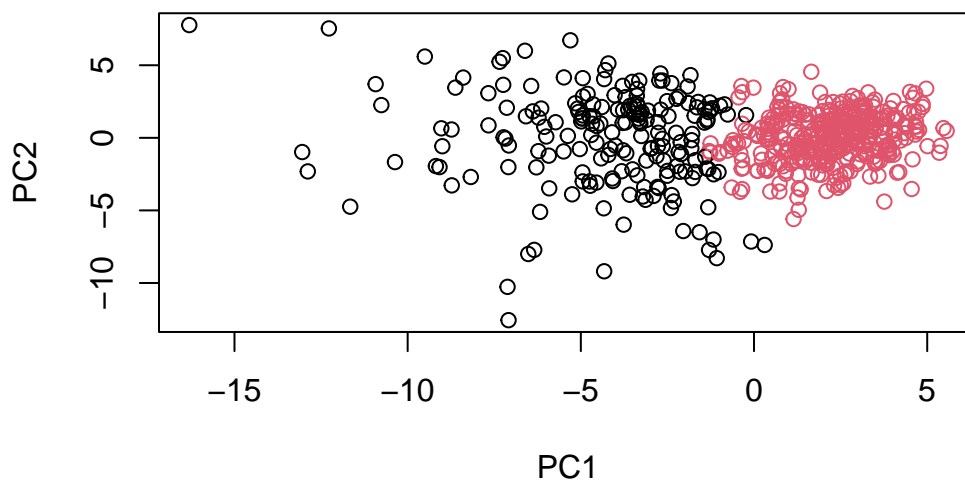
Positive => cancer M Negative => non-cancer B

True = cluster/grp 1 False = grp 2

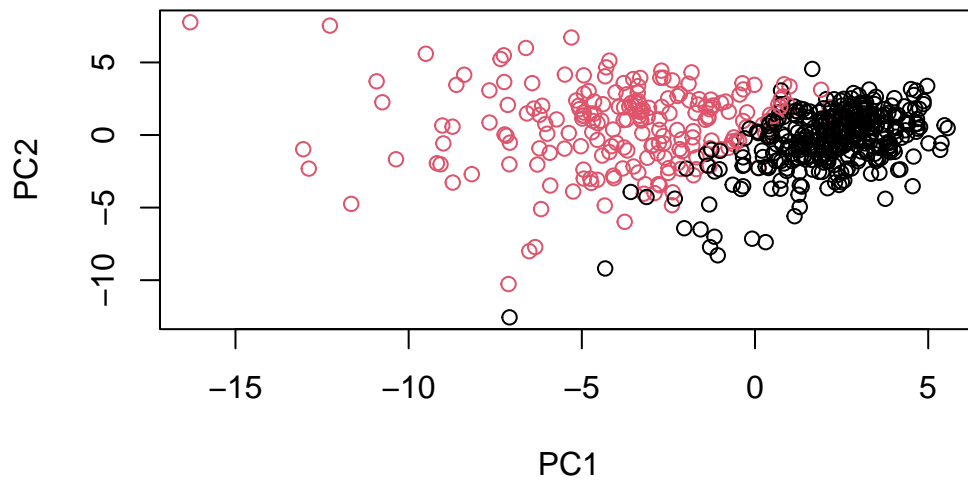
True Positive 177 False Positive 18 True Negative 339 False Negative 35

We can use our PCA results (wisc.pr) to make predications on new unseen data.

```
plot(wisc.pr$x[,1:2], col=grps)
```



```
plot(wisc.pr$x[,1:2], col=diagnosis)
```



Clustering on PCA results

```
wisc.pr.dist <- dist(wisc.pr$x[, 1:7])
wisc.pr.hclust <- hclust(wisc.pr.dist, method="ward.D2")
wisc.pr.hclust.clusters <- cutree(wisc.pr.hclust, k=4)
```

Q15. How well does the newly created model with four clusters separate out the two diagnoses?

```
table(wisc.pr.hclust.clusters, diagnosis)
```

	diagnosis	
wisc.pr.hclust.clusters	B	M
1	0	45
2	2	77
3	26	66
4	329	24

The newly created model with four clusters separates out the two diagnoses shows a different distribution of diagnoses compared to the original model. Cluster 4 contains the majority of

the benign diagnoses, whereas the malignant diagnoses are more spread out between clusters 1, 2, and 3. This suggests the original clustering model is more efficient in separating the diagnoses.

Q16. *How well do the k-means and hierarchical clustering models you created in previous sections (i.e. before PCA) do in terms of separating the diagnoses? Again, use the `table()` function to compare the output of each model (`wisc.km$cluster` and `wisc.hclust.clusters`) with the vector containing the actual diagnoses.*

```
table(wisc.hclust.clusters, diagnosis)
```

	diagnosis	
wisc.hclust.clusters	B	M
1	12	165
2	2	5
3	343	40
4	0	2

```
table(wisc.km$cluster, diagnosis)
```

	diagnosis	
	B	M
1	1	130
2	356	82

The k-means clustering model provides a better separation between the two diagnoses, compared to the hierarchical clustering model. K-means effectively grouped most malignant cases into one cluster and most benign cases into another, despite some mis-classifications. Hierarchical clustering created four clusters, leading to more overlap between benign and malignant diagnoses, and thus a less distinct separation.

Sensitivity/Specificity

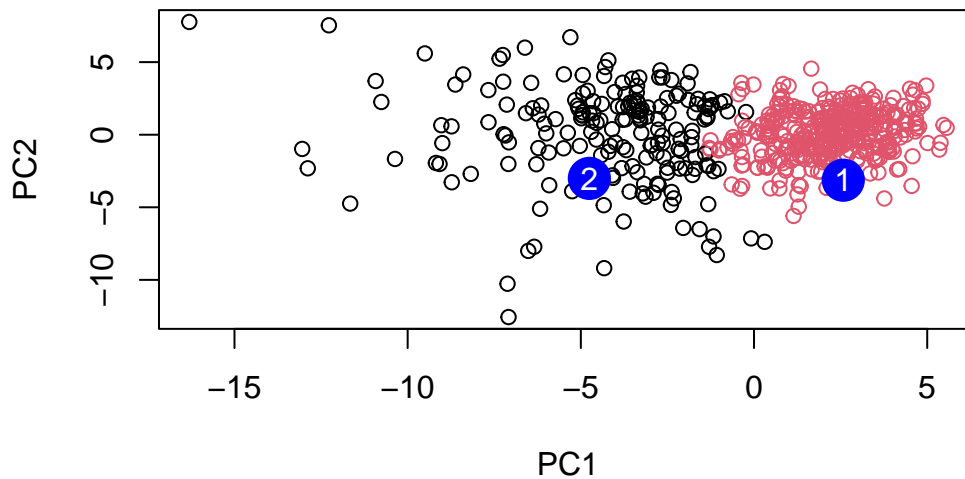
Q17. *Which of your analysis procedures resulted in a clustering model with the best specificity? How about sensitivity?*

The hierarchical clustering model with four clusters had better specificity in detecting benign cases compared to the k-means model. But, the k-means clustering model is better in sensitivity than the hierarchical clustering.

Prediction

```
#url <- "new_samples.csv"
url <- "https://tinyurl.com/new-samples-CSV"
new <- read.csv(url)
npc <- predict(wisc.pr, newdata=new)
```

```
plot(wisc.pr$x[,1:2], col=grps)
points(npc[,1], npc[,2], col="blue", pch=16, cex=3)
text(npc[,1], npc[,2], c(1,2), col="white")
```



Q18. Which of these new patients should we prioritize for follow up based on your results?

Based on these results, patient #2 should be prioritized for a follow up. This is because they are in the group of malignant.