

# Environmental heterogeneity patterns plant species richness and turnover in two hyperdiverse floras

**Running title:** Environmental heterogeneity and plant species richness

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## Abstract

**Aim:** To quantify the explanatory power of heterogeneity in predicting plant species richness and turnover here in the Greater Cape Floristic Region and in the Southwest Australia Floristic Region. We compare the environmental heterogeneity in each region, how species richness and turnover interact in each region to produce the observed patterns of richness, and what different forms of environmental heterogeneity better predict richness in each region. We expect the Cape to be more heterogeneous in most environmental axes, and at a finer grain, such that the consequent high levels of species turnover explain the Cape's greater species richness per unit area. We also conjecture that edaphic heterogeneity will be an important factor in predicting richness in SW Australia.

**Location:** The Greater Cape Floristic Region (GCFR) in southwest Africa, and the Southwest Australia Floristic Region (SWAFR)

**Taxon:** Vascular plants

**Methods:** Geospatially explicit floral and environmental data, non-parametric statistics, boosted regression tree modelling

**Results:** The Cape is more environmentally heterogeneous and has higher levels of vascular plant species turnover than SWA. We find that environmental heterogeneity is the main predictor of species richness in the Cape, and somewhat less so for SWA. Species turnover is best predicted by environmental heterogeneity in both regions.

**Main conclusions:**

*Keywords:* biodiversity, environmental heterogeneity, fynbos, Greater Cape Floristic Region, kwongan, macroecology, species richness, species turnover, vascular plants, Southwest Australia Floristic Region

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## 1 Introduction

Biodiversity represents the variety of living things, and the variety of ecological and evolutionary processes responsible for it (Bøhn & Amundsen, 2004). Studying the distribution of biodiversity in space is a major avenue of biological research (???, Kreft & Jetz, 2007). Regional-scale geographic patterns in species richness have long been studied, particularly in biodiversity hotspots (Cook et al., 2015). The spatial distribution of species richness can be explained in terms of the physical environment. Properties of the environment have been suggested to influence species richness in three ways: (i) productivity, water, and energy to enable organismal growth, and resources (i.e. niche space) to support a wider range of species (Gaston, 2000; Kreft & Jetz, 2007; Mouchet et al., 2015); (ii) stability, which enables species' persistence; and (iii) heterogeneity, which enables ecological speciation and possible barriers to gene flow, and with a wider variety of environments to facilitate species' co-existence (Thuiller et al., 2006; Mouchet et al., 2015; Cramer & Verboom, 2016). The physical environment, then, can be used to explain species richness in a

41 local-deterministic sense, and in a historical context (Ricklefs, 1987).

42 The maintenance of species richness, particularly the coexistence of high numbers of species in biodiversity  
43 hotspots, is often regarded as “paradoxical” (Hart et al., 2017), and is a central problem in ecology (Ricklefs,  
44 1987; Kreft & Jetz, 2007; Hart et al., 2017). Species richness is constrained by the ability of habitats to support  
45 a variety of species—its ecological carrying capacity (Mateo et al., 2017). This is exemplified in approaches to  
46 modelling species richness as a function of environmental predictors in a correlative framework  
47 (“macro-ecological models”; Mateo et al., 2017). Macroecological models of species richness implicitly  
48 assume that communities are saturated, following species-area and species-energy relationships, and at  
49 equilibrium with the environment (Mateo et al., 2017).

50 A solution to the paradox of species coexistence is environmental heterogeneity (EH): a more heterogeneous  
51 environment has a larger environmental space, and can thus facilitate co-existence between species at the scale  
52 of that heterogeneity. EH can also stimulate ecological speciation, should the region be environmentally stable  
53 over evolutionary time-scales. Heterogeneity in the physical environment is known to be positively associated  
54 with species richness (Rensburg et al., 2002; Hart et al., 2017), and has been demonstrated to do so across  
55 many taxa—e.g. Canadian butterflies (??), European vertebrates (Mouchet et al., 2015), South African birds  
56 (Rensburg et al., 2002), in communities along marine continental margins (Levin et al., 2010), French scarab  
57 beetles (Lobo et al., 2004), and for global terrestrial plants (Kreft & Jetz, 2007). The spatial scale of  
58 heterogeneity, or “grain” of the environment, is important to consider (Hart et al., 2017), in the same way that  
59 the spatial of absolute environmental conditions has also been considered (??; Baudena et al., 2015; Mouchet  
60 et al., 2015). Species co-existence and biodiversity maintenance is indeed suggested to be scale-dependent  
61 (Hart et al., 2017).

62 EH is often under-represented in macro-ecological models of species richness, and has recently been found to  
63 explain up to ca. 95% of biome level species richness across South Africa (Cramer & Verboom, 2016). Models  
64 that include EH yield better estimates of the richness of the Cape flora, as they account for the role  
65 heterogeneous environments such as those in the Cape facilitate species coexistence (Thuiller et al., 2006;  
66 Cramer & Verboom, 2016). Mediterranean-type terrestrial biodiversity hotspots, such as the Cape flora  
67 included in the models by Cramer & Verboom (2016), present interesting study systems in which to investigate  
68 the relationship between the environment and species richness. These systems exhibit far greater species  
69 richness than predicted by their areas, productivities and latitudes (Cowling et al., 1996; Kreft & Jetz, 2007).  
70 There are five Mediterranean biodiversity hotspots on Earth: the California Floristic Province, the

71 Mediterranean Basin, the Chilean Winter Rainfall-Valdivian Forests, the Greater Cape Floristic Region, and the  
72 Southwest Australia Floristic Region (Cowling et al., 1996; Hopper & Gioia, 2004; Cook et al., 2015). These  
73 ecosystems have regular fire-cycles (Cowling et al., 1996), climatic buffering, and long term stability (Kreft &  
74 Jetz, 2007), shrubby, sclerophyllous flora (Hopper & Gioia, 2004). Together, they account for ca. 20% of  
75 global vascular plant species, yet only ca. 5% of global land surface areas (Cowling et al., 1996). Various  
76 hypotheses have been proposed to explain the high levels of plant species richness in these regions (Cook et al.,  
77 2015). The species accumulation hypothesis states that the stability of these regions has allowed many species  
78 to accrue. The species co-existence hypothesis states that these hotspots may facilitate greater degrees of  
79 species co-existence in smaller spatial areas, due to fine-scale heterogeneity in their environments. Indeed, EH  
80 has evolutionary implications too, stimulating ecological speciation across sharp environmental gradients.

81 Both the Southwest Australia Floristic Region (SWA) and the Greater Cape Floristic Region (Cape) are  
82 Mediterranean-type biodiversity hotspots, particularly in terms of plant species. Where the Cape (with an area  
83 of ca. 189,000 km<sup>2</sup>) is known to contain about 11,400 plant species (about 0.060 species per km<sup>2</sup>), SWA (area  
84 of ca. 270,000 km<sup>2</sup>) has about 3,700 species (0.014 species per km<sup>2</sup>) (???). So, the Cape has ca. 4.3 times as  
85 many species per km<sup>2</sup> as SWA. The Cape and SWA are appropriately often compared, due to the similarities  
86 between their environments (e.g. oligotrophic soils, an oceanically buffered moderate climate) and their plants'  
87 ecologies (Hopper & Gioia, 2004). These two regions present unique flora out of the five Mediterranean  
88 systems, with high levels of endemism (Cowling et al., 1996), and many obligate fire-adapted species (Cowling  
89 et al., 1996). Similarities withstanding, SWA is topographically and edaphically distinct from the Cape. The  
90 former is topographically rather uniform (i.e. flat)—uniquely so among the world's five Mediterranean-climate  
91 regions (Hopper & Gioia, 2004)). SWA possesses a mesoscale chronosequence dune system (Laliberte et al.,  
92 2014; Cook et al., 2015), while the Cape is mountainous, topographically heterogeneous, and therefore  
93 associated with a large degree of spatial climatic variability, with a fine-scale mosaic of geologies and soils  
94 (Cowling et al., 1996; Cramer et al., 2014; Verboom et al., 2017).

95 Both regions have sources of edaphic heterogeneity, but at different scales. This edaphic variability may aid in  
96 explaining the species richness in these regions (Beard et al., 2000; Verboom et al., 2017). EH can stimulate  
97 ecological speciation, should the region be stable over evolutionary time-scales, as is likely the case in both the  
98 Cape and SWA (Wardell-Johnson & Horwitz, 1996; Hopper & Gioia, 2004; Lambers et al., 2010; Cramer et al.,  
99 2014; Laliberte et al., 2014; Cook et al., 2015). For the Cape, this richness is largely known to result from long  
100 term climatic stability, and fine grain variation in geology and soils (Cramer et al., 2014). The question thus  
101 arises whether heterogeneity is a significant contributor to SWA species richness as is likely the case in the

102 Cape. In the absence of topographic variability in SWA, it is proposed that the heterogeneity of that region is  
103 due to the juxtaposition of soil types (Laliberte et al., 2014; Cook et al., 2015), creating extreme edaphic  
104 variation.

## 105 **1.1 Hypothesis-v1**

106 Our main hypothesis is that the greater abiotic heterogeneity in the Cape, and the finer grain of that  
107 heterogeneity, compared to that of the SWA, accounts for the Cape's greater species richness per unit area. We  
108 expect the relationships between EH, species richness, and species turnover in these two regions to demonstrate  
109 this. As stated above, heterogeneous environments can (i) support diverse species assemblages, and (ii)  
110 stimulate ecological speciation. Thus, we expect species richness to covary with heterogeneity. Additionally, as  
111 one moves across a heterogeneous landscape, we expect to find greater turnover in community composition, as  
112 different environments support different species. Thus, areas of greater turnover should also be more rich, due  
113 to potential complementarity between neighbouring communities increasing total richness. Consequently, we  
114 expect that EH positively influences species richness and species turnover, and that species turnover itself  
115 positively influences species richness.

## 116 **1.2 Hypothesis-v2**

117 Aim: This study investigates the role EH plays in explaining vascular plant species richness in the Cape and  
118 SWA. We compare the relative importance of heterogeneity between the two regions, as heterogeneity has the  
119 evolutionary role of facilitating speciation, and the ecological role of supporting diverse species assemblages.  
120 Spatial scale of that heterogeneity is also considered, as the heterogeneity-richness relationship can vary with  
121 habitat grain-size.

122 Our hypotheses concern the Cape and SWA's environments and floras. Our main hypothesis is that the Cape  
123 possesses greater abiotic heterogeneity, and at finer grain, compared to SWA, such as to explain the Cape's  
124 greater species richness per unit area, and proposed greater levels of species turnover between areas. We also  
125 conjecture that the heterogeneity that predicts species richness in SWA will be more pronounced in terms of  
126 edaphic variables. Here we attempt to assess six key predictions of this hypothesis, additionally investigating a  
127 seventh prediction to test the conjectured role of edaphic heterogeneity in SWA. Dealing with the two regions'  
128 environments, we assess (i) whether the Cape environment is more heterogeneous than that of SWA and (ii)

129 whether the Cape environment has more pronounced heterogeneity at finer scales than that of SWA. Dealing  
130 with the distribution of species in the two regions, we assess (iii) whether the Cape exhibits greater levels of  
131 species turnover between areas. Relating each regions' environment and flora, we finally assess (iv) whether  
132 species richness and species turnover are adequately predicted by EH in both regions and whether (v) Species  
133 richness and species turnover are better predicted by different forms of EH in either region (e.g. the importance  
134 of edaphic heterogeneity in SWA).

135 ...

136 We employ classical statistical methods to analyse publicly available geospatial and species occurrence  
137 datasets.

138 ...

139 Species distribution models (SDMs), or environmental niche models, are sets of empirical methods that relate  
140 observed species presences (or similar data) to environmental and spatial variables, often correlatively (Guisan  
141 & Thuiller, 2005). As SDMs rely chiefly on correlating observed species ranges with the conditions thereof,  
142 they provide only a model of the realised niche of a species (Raes, 2012), which can cause issues when  
143 attempting to predict responses of species to changing climate. Other assumptions of typical SDMs include that  
144 the range of species considered is in equilibrium with the environment (Altwegg et al., 2014; Guisan & Thuiller,  
145 2005; Hannah et al., 2005), thus limiting the efficacy of these models on dynamically ranged or highly vagile  
146 species (Hannah et al., 2007). Regardless, given the dynamic nature of biotic ranges under climate change,  
147 SDMs are a valuable tool in identifying the contemporary risks posed to global and regional biodiversity.

148 The GCFR is a megadiverse terrestrial biogeographic region, with high levels of endemism. Midgley et al.  
149 (2003) investigated the responses the now defunct Cape Floristic Region (CFR) flora to climate change. Using  
150 bioclimatic envelope models (a form of SDM), they modelled the Fynbos Biome's distribution as a whole, and  
151 select Cape-endemic Proteaceae species' distributions, under current and future climate (climate scenario  
152 HadCM2), and again with the impact of land use change. Their Fynbos model was an indicator of regional  
153 priority for species level modelling efforts, showing a general southwards contraction of the biome. Their  
154 specific Proteaceae models yielded various results: complete extinctions for some species, range contractions  
155 for most, improbable range shifts in some, and range expansions in few. The range shifts predicted therein were  
156 acknowledged to be improbable, due to the unmodelled limitations of plant dispersal and edaphic dependence.  
157 Midgley et al. (2003) concluded that climate change is likely to have severely negative for the CFR flora.

158 However, as will be outlined below, their methods may be overpredicting losses due to climate change.

159 ...

160 The last 20 years have seen much ecological research interest in and development of SDMs, using many  
161 statistical and machine-learning-based methodologies (Altwegg et al., 2014; Elith et al., 2008, 2011; Guisan &  
162 Thuiller, 2005). Machine-learning-based methods in SDMs include MaxEnt (Elith et al., 2011), genetic  
163 algorithms, and adaptive neural networks (Hannah et al., 2005). The use of more advanced statistics  
164 (e.g. Bayesian frameworks, ordination methods (Hannah et al., 2005)) is also seen. These two avenues of  
165 research have intersected in the development of boosted regression trees (BRTs) (originally “gradient boosting  
166 machine”; Friedman, 1999) a system of recursively generated, non-linear regression trees, as outlined by Elith  
167 et al. (2008). BRTs have been used as SDMs in southern Africa before (e.g. Thuiller et al., 2006), sometimes  
168 for conservation purposes (e.g. Coetzee et al., 2009), but BRTs have yet to be used specifically to assess the  
169 responses of the regional flora to climate change. BRTs have more flexibility in their predictions than more  
170 traditional methods (e.g. GAMs), as they are non-linear and machine-learning-based (Elith et al., 2008).

171 ...

## 172 **2 Materials and methods**

### 173 **2.1 Overview**

174 Our analyses required definitions of the boundaries of the two regions, environmental data for each, and  
175 geospatially-explicit vascular plant occurrence records, all based on publicly available data. The environmental  
176 variables chosen (Table 1) for this study were intended to cover a reasonable spread of climatic, edaphic, and  
177 ecologically relevant environmental axes, and are not intended to be exhaustive. We selected variables  
178 describing topography (elevation), productivity (NDVI), soil status, and climate and climatic seasonality.

179 We carried out this investigation at four principal spatial scales: 0.05° x 0.05° squares (the finest common  
180 resolution among the environmental data sources used), quarter degree squares (QDS) (Larsen et al., 2009),  
181 half degree squares (HDS) (Larsen et al., 2009) and three-quarter degree squares (3QDS). For the Cape, most  
182 plant occurrence records are only accurate to QDS level. Thus, any analysis involving species data was  
183 necessary limited to scales above and including QDS.

Analyses were performed in R v3.4.0–3.5.1 (R Core Team, 2018). Version-numbers of specific R packages used are presented in the bibliography.

## 2.2 Environmental data sources

Geospatially-explicit raster layers were acquired for a selection of environmental variables (Table 1), for the regions of interest. Here, the GCFR was treated as the areas occupied by the Succulent Karoo and Fynbos biomes in the current delineation of South Africa’s biome boundaries (Mucina & Rutherford, 2006). The SWAFR was treated as the areas occupied by the Southwest Australia savanna, Swan Coastal Plain Scrub and Woodlands, Jarrah-Karri forest and shrublands, Southwest Australia woodlands, Esperance mallee, and Coolgardie woodlands in the World Wildlife Fund Terrestrial Ecoregions dataset (Olson et al., 2001) in order to closely match the currently delineated SWAFR (Gioia & Hopper, 2017, Hopper & Gioia (2004)). For the sake of readability, we shall refer to the GCFR and SWAFR simply as the Cape and SWA from hereon.

Raster data were re-projected to a common coordinate reference: WGS84 (NIMA, 2000), using the “rgdal” (???) package in R (R Core Team, 2018). All data were re-sampled to 0.05° resolution using the “resample” function in the R package “raster” (???), with the “bilinear” method.

An emphasis was made on using satellite-derived environmental data in this work, in order to minimise differences in data quality and methodologies between the Cape and SWA. Additionally, satellite-derived data have been shown to benefit regional-scale species distribution models (Deblauwe et al., 2016), thus motivating their use in this regional-scale study. The environmental data used in this study were derived from NASA’s SRTM digital elevation model (Farr et al., 2007), NASA’s MODIS/Terra spectroradiometric data for land surface temperature and NDVI, the Climate Hazards Group’s CHIRPS rainfall dataset (Funk et al., 2015), and the International Soil Reference and Information Centre’s SoilGrids250m edaphic dataset (Hengl et al., 2017) (Table 1). SRTM and MODIS are entirely derived from satellite measurements, whereas CHIRPS is interpolated from weather station data with satellite-derived radiometric measurements. SoilGrids250m is a machine-learning derived product, based on soil measurements as a function of many covariates, including MODIS and STRM sources (see Hengl et al., 2017), using random-forests and other classification-tree-based methods, including gradient-boosting. For the soil data considered here (Table 1), we used depth-interval weighted average values as the value for a particular soil variable in a given place.

Climatic and spectral data arise from satellites monitoring properties of the Earth’s surface through time. We



212 therefore use the mean annual values for rainfall, surface temperature, and NDVI in each pixel in our analyses.  
213 Pronounced seasonality of rainfall is a known feature of Mediterranean systems (???). We describe this  
214 seasonality by computing computing the precipitation in the driest quarter (PDQ), using code from within the  
215 “biovars” function in the R package “dismo”.

## 216 **2.3 Plant occurrence data**

217 Geospatially-explicit records of vascular plant occurrences were downloaded from the Global Biodiversity  
218 Information Facility (GBIF, Table 1). Queries were made for tracheophyte records from within the borders of  
219 the Cape and SWA as treated here (GBIF, 24 July 2017, GBIF (24 July 2017)). Only records with defined  
220 species and intra-specific ranks were kept. Intra-specific occurrences were treated as simply being  
221 representative of their species. This resulted in FIXME unique species names in the Cape, and FIXME in SWA.

222 We cleaned these data using the R package “taxise” (???, (???)) to check that these species names had  
223 accepted-status among taxonomic databases. I queried two major taxonomic databases: the Global Name  
224 Resolver (GNR), and the Taxonomic Name Resolution Service (TNRS). Should one of either service return at  
225 least one match for a given name, then that name was deemed accepted. Those names for which no full  
226 binomial matches were found in either database were excluded from the final list of species. The number of  
227 species names excluded totalled at FIXME and FIXME for the Cape and SWA respectively. Especially for  
228 SWA, these numbers may be deemed appreciably high. But, the occurrence records that would be dropped, as a  
229 consequence of these names’ removals, seemed to be distributed randomly in geographic space in both regions.  
230 As such, any effect of the loss of these records in this analysis is uniform within the two regions.

231 After the unaccepted names were removed, it was important to ensure that a species was not listed under  
232 multiple synonyms. Such cases would skew the species richness data used in this study. In light of this, the  
233 remaining names were queried in the Tropicos and Integrated Taxonomic Information System (ITIS) databases  
234 for their known synonyms, using “taxize”. These were collated to produce a nomenclatural “thesaurus” for the  
235 Cape and SWA species. This consisted of a list of the accepted species names in a region, each associated with  
236 a list of known synonyms. I amended species’ names in the GBIF occurrence data, in order ensure species were  
237 listed under only one of these synonyms, as follows: For each entry in the thesaurus, for each synonym of that  
238 entry, if that synonym appeared in the GBIF species list, I replaced all appearances of that synonym in the  
239 species list with the original name from the thesaurus-entry that that synonym came from.

240 Lastly, I removed any species from both regions that are invasive aliens or non-indigenous. Alien species lists  
241 for plants in South Africa and Australia were acquired from the IUCN’s Global Invasive Species Database  
242 (<http://www.iucngisd.org/gisd/>).

243 The final total plant species richness in each region was FIXME and FIXME for the Cape and SWA  
244 respectively. These final collections of species occurrence records were converted to raster-layers, wherein  
245 pixel-values represented the species, genus and family richness of vascular plants within that pixel. These  
246 rasters were produced at QDS, HDS, and 3QDS resolutions.

## 247 **2.4 Analyses**

### 248 **2.4.1 Quantifying environmental heterogeneity**

249 First, in order to assess predictions (i) and (ii), we needed to describe the EH in both regions. Using the R  
250 package “raster” (??), we used a modified version of the “roughness” index in the “terrain” function. For a  
251 three by three neighbourhood  $N$  of cells, our index of roughness is the average square-root of the squared  
252 difference between each of the  $n$  neighbour cells’ values  $x_i$  and the central focal cell’s value  $x_{focal}$ :

$$Roughness(N) = \frac{1}{n} \sqrt{\sum_{i=1}^n (x_{focal} - x_i)^2}, \quad (1)$$

253 This value, notionally the standard deviation of values relative to the focal value, is ascribed to the focal cell.  
254 Note, in order to use as much data from within regions’ borders as possible, roughness was computed if a focal  
255 cell had at least one neighbour cell—that is, roughness is defined where  $n_{x_{focal}} \geq 1$ . Using this index of  
256 heterogeneity, we produced raster layers of each of our nine environmental variable’s heterogeneity. We  
257 compared the distributions of roughness values in each variable in each region with non-parametric  
258 Mann-Whitney  $U$ -tests, as almost all variables could not be normalised by log-transformations. We also  
259 compare the effect size of the Cape vs SWA using the “common language effect size” ( $CLES$ ), using the R  
260 package “canprot”. The  $CLES$  is the proportion of all pairwise comparisons between two sample groups’  
261 observations where one group’s value is greater than the other. In our case, of all pairwise comparisons of a  
262 variable’s roughness values between the Cape and SWA, we calculated the  $CLES$  as the proportion of pairs  
263 where Cape roughness values were greater than that of SWA. This allowed us to assess prediction (i). To  
264 compare the spatial scales of heterogeneity (prediction (ii)) between each region, we repeated this analysis at

all four spatial scales. Once again, this entailed recalculating the roughness layer for each variable after the original layer (0.05 degrees resolution) had been rescaled to each of the coarser resolutions.

#### 2.4.2 Quantifying species turnover

Regarding prediction (iii), we wished to compare the general degree of species turnover in each region. To compare the extent of species turnover between the Cape and SWA, we determined two metrics of species turnover. The first, computes the mean species turnover as Jaccard distance (???) between each pair of QDS within each HDS ( $\bar{J}_{QDS}$ , based on HDS with  $2 \leq n \leq 4$  QDS) in both regions. The second is defined in terms of Whittaker's additive definition of  $\beta$ -diversity (???), where

$$\gamma = \alpha + \beta \quad (2)$$

Here, we treat species richness at the HDS-scale ( $S_{HDS}$ ) as  $\gamma$  and at the QDS-scale as  $\alpha$ . Intuitively, the species richness of an area is the result of the richness of sites within that area and the difference in species complements between those sites. Thus, we use here an additively defined residual turnover ( $T_{HDS}$ ; i.e.  $\beta = \alpha - \gamma$ ) as the proportion of  $S_{HDS}$  unexplained by the mean species richness of HDS' constituent QDS ( $\bar{S}_{QDS}$ ). We compare the distributions of  $\bar{J}_{QDS}$  and  $T_{HDS}$  using non-parametric Mann-Whitney  $U$ -tests, in order to guard against non-normality.

#### 2.4.3 Predicting richness and turnover with environmental heterogeneity

Regarding prediction (iii), we wished to compare the general degree of species turnover in each region. For (iv) and (v) we modelled species richness ( $S$ ) and turnover as a function of various combinations of environmental and environmental heterogeneity variables in both regions using boosted regression-tree (BRT) modelling techniques. Allowing us to explore which axes of environmental heterogeneity have most influence on vascular plant species richness and turnover, and the differences in the importance of such axes between the Cape and SWA.

BRTs are a flexible machine learning-based model of response variables and do so without involving normal null-hypothesis significance testing (Elith et al., 2008), and have been employed previously to model species richness (Thuiller et al., 2006; see Mouchet et al., 2015; Cramer & Verboom, 2016) as macro-ecological

models. BRTs are developed through the iterative generation of non-linear regression trees. BRTs are an ensemble-approach, in which a prediction  $\hat{y}_i$  is based on the weighted sum of the predictions of progressively “less important” regression trees ( $t_k$ ), as opposed to the predictions of one tree (Elith et al., 2008). For  $k \rightarrow nt$  number of trees, where each tree is itself a function of the matrix  $X$  of  $j$  predictor variables ( $t_k = f(x_{ij})$ ),

$$\hat{y}_i = \sum_{k=1}^{nt} w_k t_k. \quad (3)$$

BRTs have two major metaparameters that users have control over (???): the learning rate ( $lr$ , the rate at which iterative trees reduce predictive deviance during model-training, conceptually a “shrinkage factor” controlling the contribution of each tree to the final model) and tree complexity ( $tc$ , the number of nodes on a given regression-tree, i.e. the maximum interaction depth the model is permitted to fit).

BRTs were implemented here to predict both vascular plant species richness and turnover in each HDS, as a function of environmental variables and environmental roughness values in those cells, as Gaussian responses, thus resulting in two BRT-models for each region. We treated richness as  $S_{HDS}$  and turnover as  $\bar{J}_{QDS}$ . The natural logarithm of species richness was used, in order to satisfy the assumptions of a Gaussian response. Note, this is not strictly because BRTs have any parametric assumptions concerning the distribution of the response variable, but rather to aid in applying the Gaussian-family of BRT algorithms to the richness data available.

As recommended by Elith et al. (2008), BRT models were trained on a set of non-collinear predictor variables using “gbm.step” in “dismo” (???) and “gbm” (???). Collinear predictor variables can skew the interpretation of results, as the relative influence of mutually collinear variables is reduced. Collinearity among the 9 environmental predictor variables and their respective 9 roughness-equivalents was assessed using “removeCollinearity” in the R package “virtualspecies” (???) separately for each region, such that variables were no more than 80% collinear (Pearson’s  $r \geq 0.80$ ). When faced with a cluster of collinear variables, one variable was chosen manually therefrom. Where possible, the roughness-equivalent of an environmental variable was included if its absolute-equivalent could also be included. When interpreting the results of BRTs, it is important to consider the effects of the variables included as representative of the effect of the excluded variables with which it is collinear.

In order to select ideal  $lr$  and  $tc$  all models (described below) were trained on the final non-collinear predictor sets iteratively for 25 combinations of a range of  $tc$  values (1 to 5) and a range of  $lr$  values (0.01, 0.005, 0.001,  $5 \times 10^{-4}$ ,  $1 \times 10^{-4}$ ). The function “gbm.step” optimises the number of trees ( $nt$ ) using cross-validation during

model training (Elith et al., 2008) by halting iteration when predictions begin to overfit. For all models, we used 10 cross-validation folds (i.e. use 10 different randomly selected training data sets), a tolerance-threshold of 0.001, a bagging-fraction of 0.75 (proportion of training data randomly chosen to generate each tree), and trained models starting with 50 trees, with each iterative step adding 50 trees at a time, up to a maximum of 10,000 trees.

Following this iterative parameter optimisation, Gaussian BRT models were constructed with  $tc = 3$  and  $lr = 0.001$ , along with the other settings described. The models were developed with all variables (those retained, above) and then simplified using the protocol suggested by Elith et al. (2008) to retain the minimum number of variables contributing to the models, using “gbm.simplify” in “dismo”.

BRT-model performance can be described by measuring the variance a model has explained and by comparing observed and model-expected species richness (and turnover). For the former, we use  $R_{pseudo}^2$ , which is the proportion of null deviance  $D_{null}$  explained by a model  $i$ :

$$R_{pseudo}^2 = 1 - \frac{D_i}{D_{null}} \quad (4)$$

For the latter, we regressed expected against observed richness and turnover, and calculated the  $R^2$ -value for those regressions (hereafter  $R_{O-E}^2$ ).

**TODO:** In order to assess the reliability of the conclusions drawn from these models, we randomly permuted the response data ( $S_{HDS}$  and  $\bar{J}_{QDS}$ ) with respect to the environmental and heterogeneity data, and refit all 4 BRT-models 1000 times (with the final non-collinear predictor sets and preconfigurations above). This also allows us to remove any effect of spatial autocorrelation in generating the observed correlations between patterns of species occurrence and environment (???)

## 3 Results

### 3.1 Describing environmental heterogeneity across scales

Across all variables considered, the Cape is more environmentally heterogeneous in the majority of pairwise comparisons of grid-cells ( $CLES > 0.50$ , Mann-Whitney  $U$ -test:  $P < 0.05$ , Figure 1). The Cape is thus more environmentally heterogeneous than SWA overall, but the degree to which it is more heterogeneous

varies between environmental variables. These effects also vary somewhat with the spatial scale concerned. In some variables, the differentiation between Cape and SWA heterogeneity lessens at coarser scales (Figure 1b). Indeed, when comparing the overall ranking and medians of Cape vs SWA roughness values for each variable, we only find non-significant differences at the coarser 3QDS scale (Mann-Whitney  $U$  tests,  $P > 0.05$ , Figure 1b).

Most obviously, and as expected, topographic heterogeneity is greatest in the Cape (Figure 1). Though SWA has a slightly wider distribution of elevational roughness values at coarse scales (e.g. 3QDS) compared to fine scales ( $0.05^\circ$ ), so does the Cape. As such, the relative difference between the two regions seems invariant with spatial scale ( $CLES \approx 0.95$ , Figure 1b). This concurs with our expectations, as the Cape is mountainous and known to have steep elevational gradients (??), while SWA is much more topographically uniform.

Climatic heterogeneity presents less differentiation between the Cape and SWA than elevational roughness (Figure 1a), though still the Cape predominates (Figure ??b). Notably, the difference between Cape and SWA mean annual rainfall and land surface temperature heterogeneity lessens when considered at coarse spatial scales (3QDS scale, Figure ??b). Rainfall seasonality (PDQ), however, is similarly more heterogeneous in the Cape across all spatial scales considered.

Biological productivity, as measured by NDVI, varies spatially to a similar extent in the Cape and SWA (i.e. is more similarly heterogeneous,  $CLES < 0.60$ , Figure 1).

Concerning edaphic variables, the Cape and SWA are similarly heterogeneous at coarser scales, particularly in terms of CEC and Soil C (Figure 1b).

### 3.2 Comparing species turnover in the two regions

Following calculations of  $\bar{J}_{QDS}$  and  $T_{HDS}$  for each HDS-cell in each region, we used non-parametric Mann-Whitney  $U$ -tests to compare the distributions of values in the Cape and SWA. The Cape possesses generally greater floristic turnover than SWA, no matter how turnover is defined ( $P < 0.0001$ , Figure 2a,b).

### 3.3 Predicting richness and turnover with environmental heterogeneity

Vascular plant species richness and turnover are both predicted at least in-part, if not primarily, by environmental heterogeneity in both regions (Figure 3). BRT-models of species richness in each region

366 performed better than the turnover models (e.g. Cape richness vs turnover:  $R^2_{pseudo} = 0.30$  vs 0.19,  
367  $R^2_{O-E} = 0.74$  vs 0.71, respectively; Figure 3).

368 BRT-models found climatic variables to be of most importance when predicting species richness and turnover  
369 (Figure 3). Elevational heterogeneity features prominently in all models too, ranking at worst the third most  
370 important predictor.

371 Concerning BRT-models of vascular plant species richness, the sets of environmental variables important to  
372 predicting species richness differ substantially between the Cape and SWA (Figure 3a,b). While species  
373 richness in the Cape is more strongly predicted by elevational, climatic and edaphic heterogeneity (Figure 3a),  
374 richness in SWA is mostly predicted by absolute MAP (Figure 3b). Indeed, Cape species richness is mostly  
375 predicted by environmental heterogeneity variables (Figure 3a inset), while SWA species richness is mostly  
376 predicted by absolute environmental variables (Figure 3b inset).

377 Concerning BRT-models of vascular plant species turnover, elevational heterogeneity is the most important  
378 predictor in both the Cape and SWA (Figure 3c,d). Also in both regions land surface temperature heterogeneity  
379 is an important predictor, especially in SWA (Figure 3c,d). Moreover, species turnover in SWA was found to  
380 only depend on two heterogeneity variables (Figure 3d).

## 381 **4 Discussion**

382 Conclusion: The Cape is more generally environmentally heterogeneous than the SWA, as expected (see  
383 prediction (i)). Though, there are cases where the SWA is arguably at-least-as-heterogeneous as the Cape, and  
384 we can observe here extreme regions of high edaphic heterogeneity, at fine scales, in SWA. These surpass the  
385 edaphic heterogeneity of the Cape, supporting our seventh prediction/conjecture.

386 We also have support for prediction (ii), as seen in Fig. ??.

387 I have provided support for the hypothesis that the difference in plant species richness between the GCFR and  
388 SWAFR is accounted for by the fact that the GCFR is more abiotically heterogeneous than the SWAFR. As  
389 expected, the GCFR is shown to possess (i) a quantifiably more heterogeneous environment, and (ii) is  
390 heterogeneous at a finer spatial scale than the SWAFR. I have shown that vascular plant species richness (iii)  
391 can be explained in terms of environmental conditions, including environmental heterogeneity, in both the  
392 GCFR and SWAFR. Also, I have shown that (iv) the set of environmental axes that explain plant species

393 richness, both absolute and as heterogeneity, differs between the GCFR and SWAFR. These findings contribute  
394 towards an understanding of the ecological conditions that facilitate species coexistence (and likely stimulate  
395 ecological speciation) in these two regions.

396 These two regions present differentiable environmental spaces, each with heterogeneity varying across spatial  
397 scales. The clear separation of the regions' topographic features is as expected (Figures ??A, ??). Indeed,  
398 topography seems to be the most striking distinction between the regions. The Cape region has been found  
399 previously to have the second highest median topographic heterogeneity of the five Mediterranean-climate  
400 regions (Bradshaw & Cowling, 2014). The GCFR has a much wider range of scales exhibited in the  
401 heterogeneity across its environmental axes. Notably, each region has finer scale heterogeneity in some  
402 variables, and coarser scale in others—neither region is necessarily more fine or coarse than the other, as it  
403 depends on the variable concerned. BRT-models of species richness in both regions reveal species richness to  
404 depend on those environmental axes that differentiate the two regions (Figures ??, ??). The importance of  
405 variables is also shown to vary with spatial scale (Figure ??), as previously suggested may be the case when  
406 modelling geographic patterns of biodiversity (Baudena et al., 2015). Indeed, as Cowling et al. (1996)  
407 describes differing patterns of species richness across spatial scales, so do the predictors of those patterns vary  
408 with scale (Hart et al., 2017).

409 The fact that a combination of absolute and roughness variables is also as predicted by the hypothesis in this  
410 study. In the models developed by Cramer & Verboom (2016) for South Africa, roughness in topography was  
411 largely superseded as an important predictor of species richness by other roughness variables. My models,  
412 however, did not show this. Similar to the study by Rensburg et al. (2002), my models revealed roughness in  
413 topography and other variables to be important. Although, Rensburg et al. (2002) considered differences  
414 within pixels, as opposed to this study, which considered differences between pixels. My models, those of  
415 Cramer & Verboom (2016), and those of Rensburg et al. (2002), do not all concur as to the role of roughness in  
416 elevation vs. more biologically meaningful variables in explaining species richness. The source of these  
417 discrepancies is unclear, though no doubt complex. The complements of environmental variables and  
418 methodologies used in these studies do differ, limiting extensive comparison between these analyses.

419 The determinants of vascular plant species are shown to be region specific (Figures ??, ??, ??). The importance  
420 of MAP and roughness in rainfall seasonality (PCV) in predicting richness in the SWAFR (Figure ??I, ??J),  
421 aligns with the steep climatic gradients observed there (Cook et al., 2015). The soil variables that determine  
422 plant species richness in the model for the SWAFR (Figures ??K, ??L) differ to those that determine richness in



the GCFR (Figures ??G, ??H), further highlighting the edaphic differences between these two regions. Although both are nutrient leached systems, the SWAFR is flat, with soil-chronosequences (Laliberte et al., 2014; Cook et al., 2015), while the GCFR is mountainous (Cowling et al., 1996; Cramer et al., 2014; Verboom et al., 2017). The importance of roughness in soil density, and absolute texture, in the SWAFR (Figures ??K, ??L) highlights the changes in soil that are associable with age of the substrate ( e.g. particle size) as being biologically relevant to species richness. The positive effect of soil clay content on species richness in the SWAFR aligns with the findings of Laliberte et al. (2014) that richness in the SWAFR increases with soil age.

NDVI is more heterogeneous across the GCFR than the SWAFR (Figures ??A). The fact that thermal variables tend to be more rough in the GCFR (Figure ??A) is likely due to possible covariance of the MODIS/Terra products with topography, as MODIS data used here describes land surface temperature. As the GCFR is topographically rugged, the roughness of NDVI may arise from this. Despite this, NDVI is an integrating variable, which captures information about productivity, light availability, and soil nutrients (Power et al., 2017). The fact that absolute NDVI contributes to predicting species richness in the GCFR, especially at finer spatial scales (Figure ??E) demonstrates the role of ecological productivity in facilitating the coexistence diverse species assemblages. Environmental heterogeneity, then, is integral to explaining patterns of species richness, but must be considered along with resource- and energy-availability axes. In so much as a diverse environmental space supports more species, the materials and productivity required for biota to thrive are also needed to support species (???; Gaston, 2000; Bohn & Amundsen, 2004; Kreft & Jetz, 2007). As such, my findings, along with those of previous studies (Rensburg et al., 2002; Thuiller et al., 2006; Kreft & Jetz, 2007; Cramer & Verboom, 2016), suggest that there is ecological and evolutionary consequence to resource availability *and* environmental heterogeneity, in that they tend to be positively associated with species richness.

The combined BRT-model of species richness for both regions reveals soil clay content as an important predictor, at coarse spatial scales, despite this variable not being particularly important within each region separately (Figure ??). Though this model does not strictly consider the regions as separate, this finding may indicate that the relationship between clay content and species richness differs between the regions. So far as clay content can be used to predict species richness, it matters more to those predictions when applied to large sections ( i.e. coarse scales) of each regions.

Kreft & Jetz (2007) modelled global terrestrial vascular plant species richness, which focussed on primarily absolute environmental values, underestimated the richness of the Cape flora. Though Kreft & Jetz (2007) did include topographic heterogeneity in their predictor set, topography is often a proxy for more biologically

453 meaningful variables (Cramer & Verboom, 2016). This explains why the inclusion of these variables (  
454 e.g. roughness in mean annual precipitation) yields more accurate predictions of species richness. Indeed,  
455 Thuiller et al. (2006) also included topographic heterogeneity. Cramer & Verboom (2016) described 68% of  
456 species richness at the QDS scale across South Africa. Regarding the GCFR, depending on whether one  
457 consults pseudo- $R^2$  (Table 3), the ratio of mean predicted to observed richness per grid-cell (Table 5), or the  
458 distributions of predicted vs. observed richness values per grid-cell (Figure ??), I have achieved a similarly  
459 suitable level of predictive accuracy. There is, though, still unexplained species richness in light of my models.  
460 As Cramer & Verboom (2016), Rensburg et al. (2002), Thuiller et al. (2006), and Mouchet et al. (2015) have  
461 done, these macro-ecological models are a-historical. Evolutionary considerations of species richness in  
462 geographic space are worthwhile, especially in regions with environments stable over evolutionary time.

463 The findings here are correlative. There are, however, many proposed mechanisms to explain the correlative  
464 signals demonstrated here. My findings support the hypothesis that Mediterranean systems' plant species  
465 richness is a function of spatial variability in environmental conditions. This can stimulate diversification, and  
466 maintain that diversity by providing a range of habitats for species co-existence. Oligotrophic soils can stimulate  
467 an increase in functional diversity, through the evolution of diverse nutrient acquisition strategies (Lambers et  
468 al., 2010; Verboom et al., 2017) ( e.g. sclerophylly (Cramer et al., 2014; Cook et al., 2015)). An aspect of the  
469 environment I have neglected to consider is fire, shown to also contribute to predictions here in the GCFR  
470 (Cramer & Verboom, 2016). Cardillo (2012) have shown the structuring forces behind species co-occurrence  
471 patterns, and thus likely species richness, differ between species-pairs with different post-fire responses and  
472 those with similar post-fire responses.

473 Though the GCFR was correctly predicted to have, on average, more species per grid-cell at HDS and 3QDS  
474 scales than the SWAFR, this was not the case for QDS grid-cells (Table 5). This demonstrates that the GCFR is  
475 indeed overall more rich in plant species than the SWAFR, but a given HDS in the SWAFR contains fewer  
476 species than a given GCFR HDS. Thus, the greater richness in the GCFR is a product of greater turnover in  
477 species at spatial scales no more coarse than the HDS. Species turnover is an interesting aspect to species  
478 richness studies, as it species turnover is implicit to species-area and co-existence-area relationships (Hart et  
479 al., 2017). One could expect patterns of endemism and species turnover to concur with patterns in  
480 environmental heterogeneity to some degree.

481 Following from the understanding that functionally diverse assemblages, which are more likely to be more  
482 species rich, are likely to arise and/or occur in areas with diverse ecological pressures (Molina-Venegas et al.,

2015), one would expect, then, heterogeneous habitats such as those in Mediterranean-type biodiversity hotspots to exhibit high levels functional beta diversity along steep environmental gradients (Molina-Venegas et al., 2015). If the niches concerning these functions are phylogenetically conserved among those biota, then one would also expect high levels of species and phylogenetic beta diversity along these gradients (Molina-Venegas et al., 2015). This concurs with the notion put forward by Power et al. (2017), wherein megadiverse systems such as these represent the results of “phylogenetic niche conservatism on a heterogeneous landscape”. Thus, species and phylogenetic turnover should covary with environmental heterogeneity in some way. Indeed, endemism, at certain scales, could also follow this pattern. Thuiller et al. (2006) demonstrated that there is phylogenetic and biome related determinants of species richness. This makes sense, in light of the difficulty of crossing biome boundaries in Mediterranean systems (Power et al., 2017). NDVI and light availability, and the heterogeneity therein, are associated with high levels of floristic turnover (Power et al., 2017). This may be indicative of ecological specialisation precluding species from crossing these boundaries, thus increasing the level of endemism within a region, while also increasing the level of turnover, and thus likely species richness, along environmental gradients. Although, this may be debated. Beard et al. (2000) state that the high levels of endemism in SWAFR are function of habitat specialisation to soil mosaics. Cf. Laliberte et al. (2014), who say that this endemism is likely due to environmental filtering along these soil turnover sequences, as opposed to the juxtaposition of specialised species along soil gradients.

I have demonstrated support for the idea that environmental heterogeneity is positively associated with species richness, particularly Mediterranean systems. In the SWAFR and the GCFR, high levels of endemism and biodiversity are also likely the results of long-term landscape and climatic stability (Hopper, 1979). Thus, the roles of environmental variability through space, and stability through time, are the two main ways in which the environment relates to biodiversity in these regions.

## Table captions

Captions are also repeated alongside their respective tables for readability.

Table 1: Georeferenced vascular plant species occurrence and environmental data sources used in this study. Data were acquired for the Cape and SWA regions, with the temporal extent of data products used described where applicable. Abbreviations are as follows: MAP, mean annual precipitation; PDQ, precipitation in the driest quarter; CEC, cation exchange capacity.

## Figure captions

Captions are also repeated alongside their respective figures for readability.

Figure 1: Types of environmental heterogeneity, compared between the Cape and SWA—namely for (a) elevation, (b) climatic variables, (c) NDVI and (d) soil variables—in each panel consisting of three sub-panels per variable type. The upper row of panels shows example distributions of roughness values (Equation (1)), showing the different extremes in environmental heterogeneity observed in each region when compared at fine ( $0.05^\circ$ ) and coarse (3QDS) scales. Each distribution has under it an area of one. Histograms were constructed using 20 breaks. In the lower row of panels, these distributions of roughness values were compared between the Cape and SWA at each of the four spatial scales, not just  $0.05^\circ$  and 3QDS, using non-parametric Mann-Whitney  $U$ -tests to test for differences. The “common language effect size” ( $CLES$ , see text) describes these differences (b).  $U$ -tests for almost all environmental variables yielded significant differences ( $P < 0.05$ ) between Cape and SWA values (NS, non-significant differences).  $CLES$  for  $0.05^\circ$  res. is for 5000 random cells in each region, as the Mann-Whitney  $U$ -test cannot handle more than a few thousand values per sample when comparing.

Figure 2: Species turnover, described in two forms ((a) mean Jaccard distance between QDS in each HDS ( $\bar{J}_{QDS}$ ), (b) additively defined turnover ( $T_{HDS}$ , see Equation (2)) as a proportion of HDS richness ( $S_{HDS}$ )), compared between the Cape and SWA. Mann-Whitney  $U$ -tests between the Cape and SWA distributions of  $\bar{J}_{QDS}$  and  $T_{HDS}$  yielded significant differences.

Figure 3: Contributions of environmental variables (including heterogeneity variables) to boosted regression tree (BRT) model predictions for the final four models’ response variables in Greater Cape Floristic Region (Cape) and Southwest Australia Floristic Region (SWA): vascular plant species (a,b) richness and (c,d) turnover. All BRT-models were permitted to fit three-way interactions between environmental variables. Coloured bars denote the general category of the environment (keyed) to which a variable belongs, as in Figure 1. Inset piecharts display the same information, but grouped according to whether a variable was absolute or roughness-transformed. Summary statistics for each BRT-models are also inset:  $R_{pseudo}^2$  (Equation (4));  $R_{O-E}^2$ , the  $R^2$  of the regression of expected values as a function of observed values;  $nt$ , the number of regressions trees used in a BRT-model.

## References

- Baudena, M., Sánchez, A., Georg, C.-P., Ruiz-Benito, P., Rodríguez, M.Á., Zavala, M.A., & Rietkerk, M. (2015) Revealing patterns of local species richness along environmental gradients with a novel network tool. *Scientific Reports*, **5**, 11561.
- Beard, J.S., Chapman, A.R., & Gioia, P. (2000) Species richness and endemism in the Western Australian flora. *Journal of Biogeography*, **27**, 1257–1268.
- Bradshaw, P.L. & Cowling, R.M. (2014) Landscapes, rock types, and climate of the Greater Cape Floristic Region. *Fynbos: Ecology, evolution and conservation of a megadiverse region* (ed. by N. Allsopp, J.F. Colville, and G.A. Verboom), pp. 26–46. Oxford University Press, Oxford.
- Böhn, T. & Amundsen, P.-A. (2004) Ecological Interactions and Evolution: Forgotten Parts of Biodiversity? *BioScience*, **54**, 804.
- Cardillo, M. (2012) The phylogenetic signal of species co-occurrence in high-diversity shrublands: different patterns for fire-killed and fire-resistant species. *BMC Ecology*, **12**, 21.
- Cook, L.G., Hardy, N.B., & Crisp, M.D. (2015) Three explanations for biodiversity hotspots: small range size, geographical overlap and time for species accumulation. An Australian case study. *New Phytologist*, **207**, 390–400.
- Cowling, R.M., Rundel, P.W., Lamont, B.B., Arroyo, M.K., & Arianoutsou, M. (1996) Plant diversity in mediterranean-climate regions. *Trends in Ecology and Evolution*, **11**, 362–366.
- Cramer, M.D. & Verboom, G.A. (2016) Measures of biologically relevant environmental heterogeneity improve prediction of regional plant species richness. *Journal of Biogeography*, 1–13.
- Cramer, M.D., West, A.G., Power, S.C., Skelton, R., & Stock, W.D. (2014) Plant ecophysiological diversity. *Fynbos: Ecology, evolution and conservation of a megadiverse region* pp. 248–272. Oxford University Press, Oxford.
- Deblauwe, V., Droissart, V., Bose, R., Sonké, B., Blach-Overgaard, A., Svenning, J.C., Wieringa, J.J., Ramesh, B.R., Stévant, T., & Couvreur, T.L.P. (2016) Remotely sensed temperature and precipitation data improve species distribution modelling in the tropics. *Global Ecology and Biogeography*, **25**, 443–454.
- Elith, J., Leathwick, J.R., & Hastie, T. (2008) A working guide to boosted regression trees. *Journal of Animal Ecology*, **77**, 802–813.
- Farr, T., Rosen, P., Caro, E., Crippen, R., Duren, R., Hensley, S., Kobrick, M., Paller, M., Rodriguez, E., Roth, L., Seal, D., Shaffer, S., Shimada, J., Umland, J., Werner, M., Oskin, M., Burbank, D., & Alsdorf, D. (2007) The shuttle radar topography mission. *Reviews of Geophysics*, **45**, 1–33.
- Funk, C.C., Peterson, P.J., Landsfeld, M., Pedreros, D.H., Verdin, J., Shukla, S., Husak, G., Rowland, J.D., Harrison, L., Hoell, A., & Michaelsen, J. (2015) The climate hazards infrared precipitation with stations—a new environmental record for monitoring extremes. *Scientific Data*, **2**, 150066.
- Gaston, K.J. (2000) Global patterns in biodiversity. *Nature*, **405**, 220–227.

568 GBIF (24 July 2017) GBIF Occurrence Download..

569 GBIF (24 July 2017) GBIF Occurrence Download..

570 Gioia, P. & Hopper, S.D. (2017) A new phytogeographic map for the Southwest Australian Floristic Region after an exceptional decade  
571 of collection and discovery. *Botanical Journal of the Linnean Society*, **184**, 1–15.

572 Hart, S.P., Usinowicz, J., & Levine, J.M. (2017) The spatial scales of species coexistence. *Nature Ecology & Evolution*, **1**, 1066–1073.

573 Hengl, T., Mendes de Jesus, J., Heuvelink, G.B.M., Ruiperez Gonzalez, M., Kilibarda, M., Blagoti?, A., Shangguan, W., Wright, M.N.,  
574 Geng, X., Bauer-Marschallinger, B., Guevara, M.A., Vargas, R., MacMillan, R.A., Batjes, N.H., Leenaars, J.G.B., Ribeiro, E.,  
575 Wheeler, I., Mantel, S., & Kempen, B. (2017) SoilGrids250m: Global gridded soil information based on machine learning.  
576 *PLoS ONE*, **12**, e0169748.

577 Hopper, S.D. (1979) Biogeographical Aspects of Speciation in the Southwest Australian Flora. *Annual Review of Ecology and*  
578 *Systematics*, **10**, 399–422.

579 Hopper, S.D. & Gioia, P. (2004) The Southwest Australian Floristic Region: Evolution and Conservation of a Global Hot Spot of  
580 Biodiversity. *Annual Review of Ecology, Evolution, and Systematics*, **35**, 623–650.

581 Kreft, H. & Jetz, W. (2007) Global patterns and determinants of vascular plant diversity. *Proceedings of the National Academy of*  
582 *Sciences*, **104**, 5925–5930.

583 Laliberte, E., Zemunik, G., & Turner, B.L. (2014) Environmental filtering explains variation in plant diversity along resource gradients.  
584 *Science*, **345**, 1602–1605.

585 Lambers, H., Brundrett, M.C., Raven, J.A., & Hopper, S.D. (2010) Plant mineral nutrition in ancient landscapes: high plant species  
586 diversity on infertile soils is linked to functional diversity for nutritional strategies. *Plant and Soil*, **334**, 11–31.

587 Larsen, R., Holmern, T., Prager, S.D., Maliti, H., & Røskoft, E. (2009) Using the extended quarter degree grid cell system to unify  
588 mapping and sharing of biodiversity data. *African Journal of Ecology*, **47**, 382–392.

589 Levin, L.A., Sibuet, M., Gooday, A.J., Smith, C.R., & Vanreusel, A. (2010) The roles of habitat heterogeneity in generating and  
590 maintaining biodiversity on continental margins: an introduction. *Marine Ecology*, **31**, 1–5.

591 Lobo, J.M., Jay-robert, P., Lumaret, J.-p., Lobo, J.M., Jay-robert, P., & Lumaret, J.-p. (2004) Modelling the Species Richness  
592 Distribution for French Aphodiidae (Coleoptera, Scarabaeoidea). *Ecography*, **27**, 145–156.

593 Mateo, R.G., Mokany, K., & Guisan, A. (2017) Biodiversity Models: What If Unsaturation Is the Rule? *Trends in Ecology &*  
594 *Evolution*, **32**, 556–566.

595 Molina-Venegas, R., Aparicio, A., Slingsby, J.A., Lavergne, S., & Arroyo, J. (2015) Investigating the evolutionary assembly of a  
596 Mediterranean biodiversity hotspot: Deep phylogenetic signal in the distribution of eudicots across elevational belts. *Journal*  
597 *of Biogeography*, **42**, 507–518.

598 Mouchet, M., Levers, C., Zupan, L., Kuemmerle, T., Plutzer, C., Erb, K., Lavorel, S., Thuiller, W., & Haberl, H. (2015) Testing the  
599 effectiveness of environmental variables to explain European terrestrial vertebrate species richness across biogeographical  
600 scales. *PLoS ONE*, **10**, 1–16.

601 Mucina, L. & Rutherford, M.C. (2006) *The vegetation of South Africa, Lesotho and Swaziland*. South African National Biodiversity  
602 Institute,

603 NIMA (2000) Amendment 1. 3 January 2000. Department of Defense World Geodetic System 1984. Its Definition and Relationships  
604 with Local Geodetic Systems. 1–3.

605 Olson, D.M., Dinerstein, E., Wikramanayake, E.D., Burgess, N.D., Powell, G.V.N., Underwood, E.C., D’Amico, J.A., Itoua, I., Strand,  
606 H.E., Morrison, J.C., & Others (2001) Terrestrial Ecoregions of the World: A New Map of Life on Earth: A new global map of  
607 terrestrial ecoregions provides an innovative tool for conserving biodiversity. *BioScience*, **51**, 933–938.

608 Power, S.C., Verboom, G.A., Bond, W.J., & Cramer, M.D. (2017) Environmental correlates of biome-level floristic turnover in South  
609 Africa. *Journal of Biogeography*, **44**, 1745–1757.

610 R Core Team (2018) *R: A Language and Environment for Statistical Computing. Version 3.5.0*. R Foundation for Statistical  
611 Computing, Vienna, Austria.

612 Rensburg, B.J. van, Chown, S.L., & Gaston, K.J. (2002) Species Richness, Environmental Correlates, and Spatial Scale: A Test Using  
613 South African Birds. *The American Naturalist*, **159**, 566–577.

614 Ricklefs, R.E. (1987) Community diversity: relative roles of local and regional processes. *Science, New Series*, **235**, 167–171.

615 Thuiller, W., Midgley, G.F., Rouget, M., Cowling, R.M., F. Midgley, G., Rougeti, M., & M. Cowling, R. (2006) Predicting patterns of  
616 plant species richness in megadiverse South Africa. *Ecography*, **29**, 733–744.

617 Verboom, G.A., Stock, W.D., & Cramer, M.D. (2017) Specialization to extremely low-nutrient soils limits the nutritional adaptability  
618 of plant lineages. *The American Naturalist*, **In press**.

619 Wardell-Johnson, G. & Horwitz, P. (1996) Conserving biodiversity and the recognition of heterogeneity in ancient landscapes: a case  
620 study from south-western Australia. *Forest Ecology and Management*, **85**, 219–238.

## 621 **Biosketches**

622 **Ruan van Maziik** is currently a Masters student at the University of Cape Town, interested in phylogenetic  
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625 **G. Anthony Verboom**

## 626 **Author contributions**

627 MDC and GAV conceived the study question, which RVM investigated under their supervision for his BSc  
628 Hons project. The analyses and programming work were largely devised by RVM, with input from the other  
629 authors, and was carried out by RVM. RVM wrote the first draft of the manuscript and all authors contributed  
630 equally thereafter.



Table 1: Georeferenced vascular plant species occurrence and environmental data sources used in this study. Data were acquired for the Cape and SWA regions, with the temporal extent of data products used described where applicable. Abbreviations are as follows: MAP, mean annual precipitation; PDQ, precipitation in the driest quarter; CEC, cation exchange capacity.

Variable	Source	Temporal extent	Citation
Plant species occurrences	GBIF	TODO	??, ??
Elevation	SRTM v2.0		??
NDVI	MODIS (MOD13C2)	Feb. 2000 to Apr. 2017	??
<b>Climatic variables</b>			
Surface temperature	MODIS (MOD11C3)	Feb. 2000 to Apr. 2017	??
MAP	CHIRPS v2.0	Jan. 1981 to Feb. 2017	??
PDQ	CHIRPS v2.0	Jan. 1981 to Feb. 2017	??
<b>Soil variables</b>			
CEC	SoilGrids250m (CECSOL M 250m)		??
Clay	SoilGrids250m (CLYPPT M 250m)		
Soil C	SoilGrids250m (OCDENS M 250m)		
pH	SoilGrids250m (PHIKCL M 250m)		

Table 2: (ref:quality-rank-data)

Model	$nt$	$R^2_{pseudo}$	$R^2_{E-O}$
<b>QDS-richness</b>			
GCFR	1.000	1.000	1.000
SWAFR	1.000	1.000	1.000
<b>HDS-richness</b>			
GCFR	0.987	1.000	0.988
SWAFR	1.000	1.000	1.000
<b>HDS-turnover</b>			
GCFR	0.977	0.992	0.979
SWAFR	0.997	1.000	1.000

Table 3: (ref:quality-region-data)

Model	$nt$	$R^2_{pseudo}$	$R^2_{E-O}$
QDS-richness	542.938	0.063	-0.005
HDS-richness	-808.994	-0.064	-0.233
HDS-turnover	-997.045	-0.052	-0.296

## 632 Figures

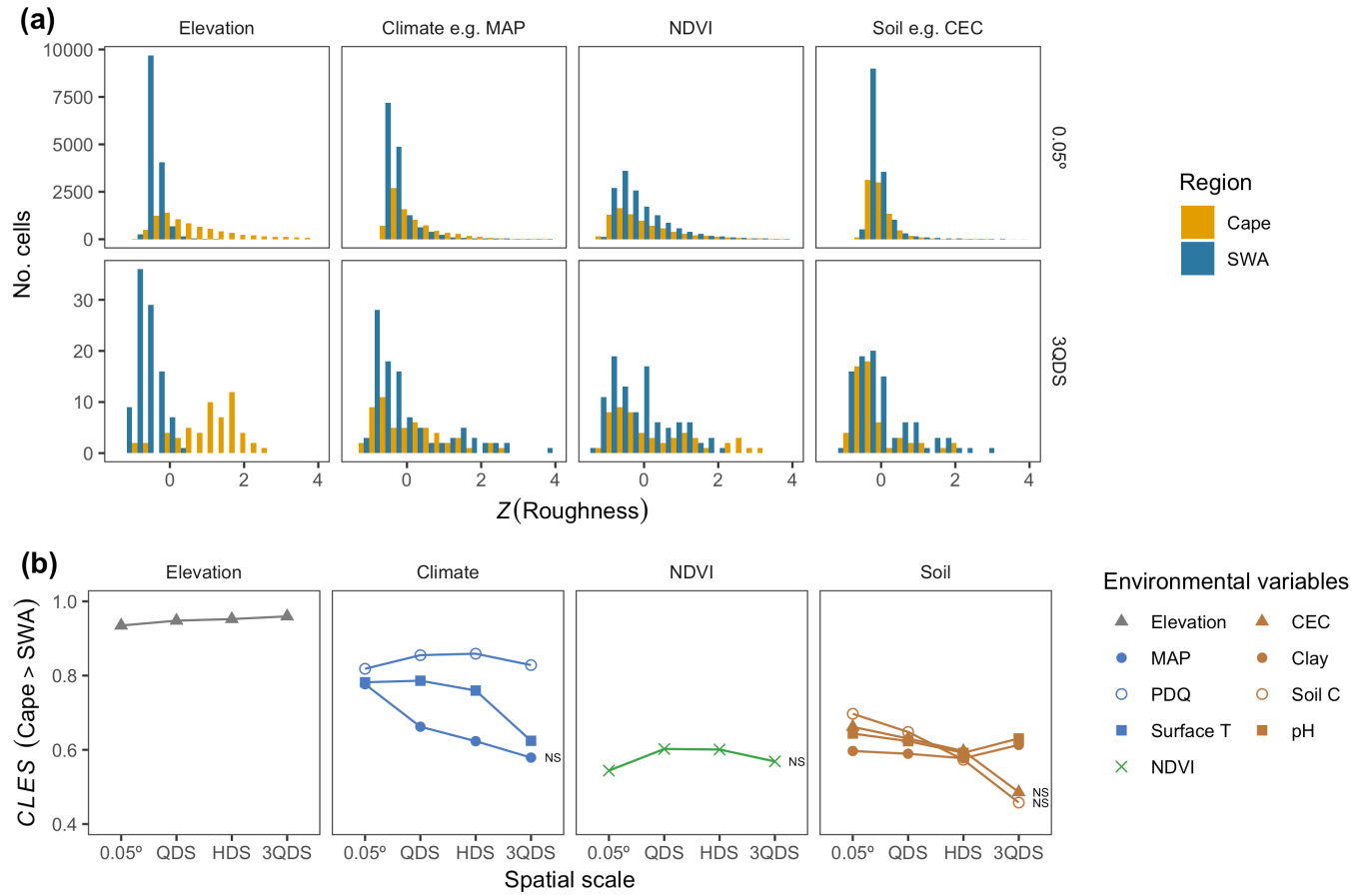


Figure 1: Types of environmental heterogeneity, compared between the the Cape and SWA—namely for (a) elevation, (b) climatic variables, (c) NDVI and (d) soil variables—in each panel consisting of three sub-panels per variable type. The upper row of panels shows example distributions of roughness values (Equation (1)), showing the different extremes in environmental heterogeneity observed in each region when compared at fine (0.05°) and coarse (3QDS) scales. Each distribution has under it an area of one. Histograms were constructed using 20 breaks. In the lower row of panels, these distributions of roughness values were compared between the Cape and SWA at each of the four spatial scales, not just 0.05° and 3QDS, using non-parametric Mann-Whitney  $U$ -tests to test for differences. The “common language effect size” ( $CLES$ , see text) describes these differences (b).  $U$ -tests for almost all environmental variables yielded significant differences ( $P < 0.05$ ) between Cape and SWA values (NS, non-significant differences).  $CLES$  for 0.05 res. is for 5000 random cells in each region, as the Mann-Whitney  $U$ -test cannot handle more than a few thousand values per sample when comparing.

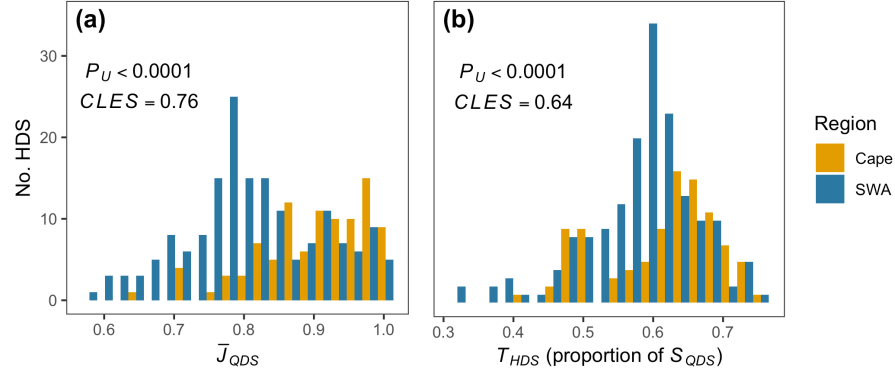


Figure 2: Species turnover, described in two forms ((a) mean Jaccard distance between QDS in each HDS ( $\bar{J}_{QDS}$ ), (b) additively defined turnover ( $T_{HDS}$ , see Equation (2)) as a proportion of HDS richness ( $S_{HDS}$ )), compared between the Cape and SWA. Mann-Whitney  $U$ -tests between the Cape and SWA distributions of  $\bar{J}_{QDS}$  and  $T_{HDS}$  yielded significant differences.

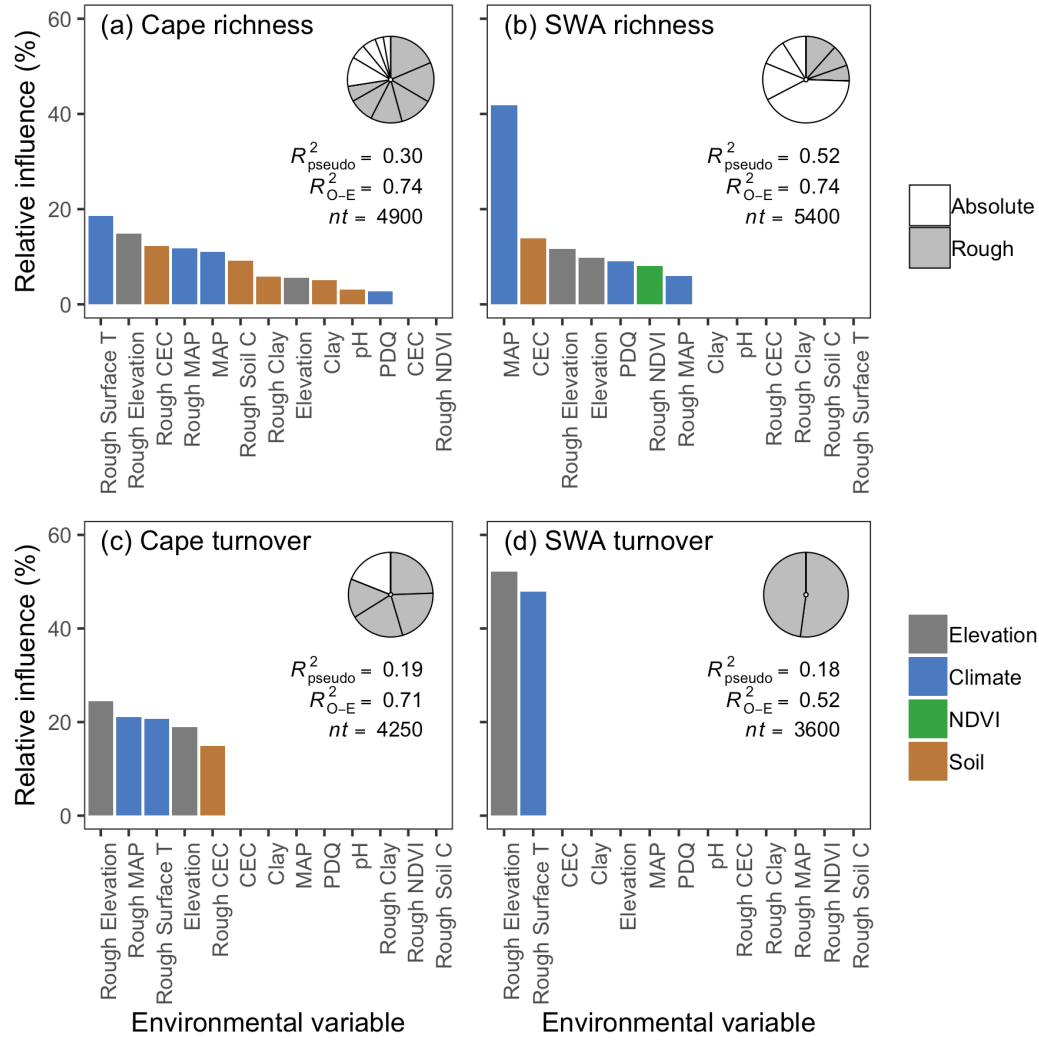


Figure 3: Contributions of environmental variables (including heterogeneity variables) to boosted regression tree (BRT) model predictions for the final four models' response variables in Greater Cape Floristic Region (Cape) and Southwest Australia Floristic Region (SWA): vascular plant species (a,b) richness and (c,d) turnover. All BRT-models were permitted to fit three-way interactions between environmental variables. Coloured bars denote the general category of the environment (keyed) to which a variable belongs, as in Figure 1. Inset piecharts display the same information, but grouped according to whether a variable was absolute or roughness-transformed. Summary statistics for each BRT-models are also inset:  $R^2_{pseudo}$  (Equation (4));  $R^2_{O-E}$ , the  $R^2$  of the regression of expected values as a function of observed values;  $nt$ , the number of regressions trees used in a BRT-model.

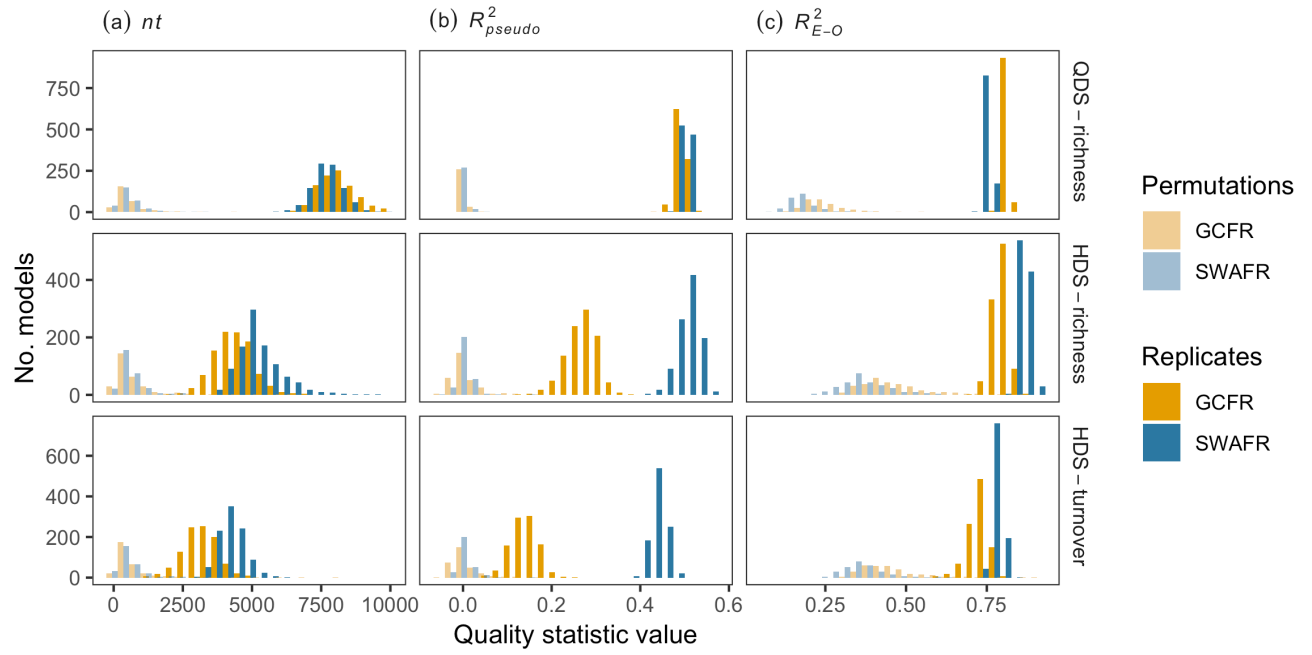


Figure 4: (ref:BRT-quality-plot)