

Environmental heterogeneity patterns plant species richness and turnover in two hyperdiverse floras

Ruan van Mazijk, Michael D. Cramer & G. Anthony Verboom

2019-03-03

Department of Biological Sciences, University of Cape Town, Rondebosch, South Africa

Corresponding author: RVM (ruanvmazijk@gmail.com, +27 21 650 3684)

ORCID nos.: RVM: 0000-0003-2659-6909, MDC: 0000-0003-0989-3266, GAV: 0000-0002-1363-9781

Running title: Environmental heterogeneity and plant species richness

Abstract

Aim: To quantify the explanatory power of environmental heterogeneity in predicting plant species richness and turnover in two mediterranean-type biodiversity hotspots, the southwestern Cape of South Africa and southwest Australia (SWA), due to habitat diversity's stimulating speciation and supporting diverse assemblages. In each region, we compare different forms of environmental heterogeneity, species turnover and the relative importance of different forms of heterogeneity better predict species richness and turnover. We expected the Cape to be more heterogeneous in most environmental axes, and at a finer grain, such that the consequent high levels of species turnover explain the Cape's greater species richness per unit area. We also conjectured that edaphic heterogeneity would be an important factor in predicting species richness patterns in SWA.

Location: The Greater Cape Floristic Region in southwest Africa (the Cape), and the Southwest Australia Floristic Region (SWA)

Taxon: Vascular plants

22 **Methods:** Geospatially explicit floral and environmental data, non-parametric statistics, boosted regression
23 tree modelling

24 **Results:** The Cape is more environmentally heterogeneous and has higher levels of floristic turnover than
25 SWA. We find that environmental heterogeneity is the main predictor of species richness in the Cape, and
26 somewhat less so for SWA. Edaphic conditions are found to be of more biologically important in the Cape,
27 though this is contingent on the quality of the data modelled.

28 **Main conclusions:** The Cape was found to have generally more environmentally heterogenous than SWA,
29 though SWA does possess edaphic heterogeneity as great as that in the Cape at coarse spatial scales, and greater
30 levels of floristic turnover in vascular plant communities between grid-cells than SWA. Vascular plant species
31 richness and turnover in the Cape was associated more strongly with environmental heterogeneity than in SWA,
32 and particularly edaphic heterogeneity, than species richness in SWA. In SWA, patterns of species richness and
33 turnover are more strongly associated with climatic axes. The relative strengths of association of different
34 environmental variables with patterns of species richness and turnover was found to vary with spatial scale.

35 *Keywords:* biodiversity, boosted regression trees, BRTs, environmental heterogeneity, fynbos, Greater Cape
36 Floristic Region, kwongan, macroecology, species richness, species turnover, vascular plants, Southwest
37 Australia Floristic Region

38 **Acknowledgements**

39 This work was funded by the South African Department of Science and Technology (DST) and the National
40 Research Foundation (NRF) under the DST-NRF Freestanding Innovation Honours Scholarship (to RVM), and
41 by the South African Association of Botanists (SAAB) Honours Scholarship (to RVM). Thanks go to the
42 Department of Biological Sciences, University of Cape Town, for providing a 2TB external hard drive for local
43 GIS data storage. Many computations were performed using facilities provided by the University of Cape
44 Town's ICTS High Performance Computing team (<http://hpc.uct.ac.za>).

1 Introduction

Biodiversity represents the variety of species and the ecological and evolutionary processes that bring about those species (Bøhn & Amundsen, 2004). Studying the distribution of biodiversity in space is a major avenue of biological research (Kerr et al., 2001; Kreft & Jetz, 2007). Region-scale geographic patterns in species richness have long been studied, particularly in biodiversity hotspots (Cook et al., 2015). Indeed, “primary geographic patterns” such as these (sensu Juarez-Barrera et al., 2018) are arguably central when studying the distribution of biota across space. The spatial distribution of species richness can be and often is explained in terms of the physical environment. Certain properties of the environment have been suggested to influence species richness in three ways: (i) available resources and energy, which can determine the number of species able to co-exist in an area (Gaston, 2000; Kreft & Jetz, 2007; Mouchet et al., 2015); (ii) environmental stability through time, which enables species’ persistence; and (iii) spatial heterogeneity, which can both stimulate ecological speciation and possible barriers to gene flow and can facilitate greater levels of species’ co-existence (Thuiller et al., 2006; Mouchet et al., 2015; Cramer & Verboom, 2016). The physical environment, then, can be used to explain species richness in both a local-deterministic and historical sense (Ricklefs, 1987).

The maintenance of species richness, particularly the coexistence of high numbers of species in biodiversity hotspots, is often regarded as “paradoxical” (Hart et al., 2017), and is a central problem in macro-ecology and biogeography (Ricklefs, 1987; Kreft & Jetz, 2007; Hart et al., 2017). Species richness is constrained by the ability of habitats to support a variety of species—its ecological carrying capacity (Mateo et al., 2017). This is exemplified in modelling approaches, wherein species richness is a function of environmental predictors in a correlative framework (“macro-ecological models”; Mateo et al., 2017). Macro-ecological models of species richness implicitly assume that communities are saturated, following species-area and species-energy relationships, and at equilibrium with the environment (Mateo et al., 2017).

A solution to the “paradox” of species co-existence is environmental heterogeneity (EH): a more heterogeneous environment exhibits a larger environmental space, thus facilitating co-existence between more species. Heterogeneity in the physical environment is known to be positively associated with species richness (Rensburg et al., 2002; Hart et al., 2017), and has been demonstrated to do so across many taxa—e.g. Canadian butterflies (Kerr et al., 2001), European vertebrates (Mouchet et al., 2015), South African birds (Rensburg et al., 2002), in communities along marine continental margins (Levin et al., 2010), French scarab beetles (Lobo et al., 2004), and for global terrestrial plants (Kreft & Jetz, 2007). The spatial scale of heterogeneity, or “grain” of the environment, is also important to consider (Hart et al., 2017), as spatial scale in absolute environmental

75 conditions has also been explored (Kerr et al., 2001; Baudena et al., 2015; Mouchet et al., 2015). Species
76 co-existence and biodiversity maintenance is indeed suggested to be scale-dependent (Hart et al., 2017).

77 EH is often under-represented in macro-ecological models of species richness, and has recently been found to
78 explain up to ca. 95% of biome level species richness across South Africa (Cramer & Verboom, 2016). Indeed,
79 models that include EH yield better estimates of the richness of the Cape flora (Thuiller et al., 2006; Cramer &
80 Verboom, 2016). Mediterranean-type terrestrial biodiversity hotspots, such as the Cape flora included in the
81 models by Cramer & Verboom (2016), present interesting study systems in which to investigate the relationship
82 between the environment and species richness. These systems exhibit far greater species richness than
83 predicted by their areas, productivities and latitudes (Cowling et al., 1996; Kreft & Jetz, 2007). There are five
84 Mediterranean biodiversity hotspots on Earth: the California Floristic Province, the Mediterranean Basin, the
85 Chilean Winter Rainfall-Valdivian Forests, the Greater Cape Floristic Region, and the Southwest Australia
86 Floristic Region (Cowling et al., 1996; Hopper & Gioia, 2004; Cook et al., 2015). These ecosystems have
87 regular fire-cycles (Cowling et al., 1996), climatic buffering, and long term stability (Kreft & Jetz, 2007),
88 shrubby, sclerophyllous flora (Hopper & Gioia, 2004). Together, they account for ca. 20% of global vascular
89 plant species, yet only ca. 5% of global land surface areas (Cowling et al., 1996). Various hypotheses have
90 been proposed to explain the high levels of plant species richness in these regions (Cook et al., 2015). The
91 species accumulation hypothesis states that the stability of these regions has allowed many species to accrue.
92 The species co-existence hypothesis states that these hotspots may facilitate greater degrees of species
93 co-existence in smaller spatial areas, due to fine-scale heterogeneity in their environments. Indeed, EH has
94 evolutionary implications too, stimulating ecological speciation across sharp environmental gradients.

95 Both the Southwest Australia Floristic Region (SWA) and the Greater Cape Floristic Region (Cape) are
96 Mediterranean-type biodiversity hotspots, particularly in terms of plant species. Where the Cape (with an area
97 of ca. 189,000 km²) is known to contain about 11,400 plant species (about 0.060 species per km²), SWA (area
98 of ca. 270,000 km²) has about 3,700 species (0.014 species per km²) (Manning et al., 2012). So, the Cape has
99 ca. 4.3 times as many species per km² as SWA. The Cape and SWA are appropriately often compared, due to
100 the similarities between their environments (e.g. oligotrophic soils, an oceanically buffered moderate climate)
101 and their plants' ecologies (Hopper & Gioia, 2004). These two regions present unique flora out of the five
102 Mediterranean systems, with high levels of endemism (Cowling et al., 1996), and many obligate fire-adapted
103 species (Cowling et al., 1996). Similarities withstanding, SWA is topographically and edaphically distinct from
104 the Cape. The former is topographically rather uniform (i.e. flat)—uniquely so among the world's five
105 Mediterranean-climate regions (Hopper & Gioia, 2004)). SWA possesses a mesoscale chronosequence dune

106 system (Laliberte et al., 2014; Cook et al., 2015), while the Cape is mountainous, topographically
107 heterogeneous, and therefore associated with a large degree of spatial climatic variability, with a fine-scale
108 mosaic of geologies and soils (Cowling et al., 1996; Cramer et al., 2014; Verboom et al., 2017).

109 Both regions have sources of edaphic heterogeneity, but at different scales. This edaphic variability may aid in
110 explaining the species richness in these regions (Beard et al., 2000; Verboom et al., 2017). EH of many forms
111 will likely be important in macro-ecological models in both regions, as both regions have been relatively
112 environmentally stable over evolutionary time-scales (Wardell-Johnson & Horwitz, 1996; Hopper & Gioia,
113 2004; Lambers et al., 2010; Cramer et al., 2014; Laliberte et al., 2014; Cook et al., 2015). For the Cape, high
114 levels of species richness are thought to result from long term climatic stability, and fine grain variation in
115 geology and soils (Cramer et al., 2014). The question thus arises whether heterogeneity is a significant
116 contributor to SWA species richness. In the absence of topographic variability in SWA, it is proposed that the
117 heterogeneity of that region is due to the juxtaposition of soil types (Laliberte et al., 2014; Cook et al., 2015),
118 creating extreme edaphic variation.

119 Our hypotheses concern the Cape and SWA's environments and floras. Our main hypothesis is that the Cape
120 possesses greater abiotic heterogeneity, and at finer grain, compared to SWA, such as to explain the Cape's
121 greater species richness per unit area, and proposed greater levels of species turnover between areas. We also
122 conjecture that the heterogeneity that predicts species richness in SWA will be more pronounced in terms of
123 edaphic variables. Here we attempt to assess five key predictions of this hypothesis, additionally investigating
124 a seventh prediction to test the conjectured role of edaphic heterogeneity in SWA. Dealing with the two
125 regions' environments, we assess (i) whether the Cape environment is more heterogeneous than that of SWA
126 and (ii) whether the Cape environment has more pronounced heterogeneity at finer scales than that of SWA.
127 Dealing with the distribution of species in the two regions, we assess (iii) whether the Cape exhibits greater
128 levels of species turnover between areas. Relating each regions' environment and flora, we finally assess (iv)
129 whether species richness and species turnover are adequately predicted by EH in both regions and whether (v)
130 species richness and species turnover are better predicted by different forms of EH in either region (e.g. the
131 importance of edaphic heterogeneity in SWA).

2 Materials and methods

2.1 Overview

Our analyses required boundaries for each region and geographically explicit environmental data and vascular plant occurrence records. The environmental variables chosen (Table 1) for this study were intended to cover a reasonable spread of climatic, edaphic, and ecologically relevant environmental axes, and are not intended to be exhaustive. We selected variables describing topography (elevation), productivity (NDVI), soil status and climate and climatic seasonality.

We carried out this investigation at four principal spatial scales: $0.05^\circ \times 0.05^\circ$ squares (the finest common resolution among the environmental data sources used), quarter degree squares (QDS) (Larsen et al., 2009), half degree squares (HDS) (Larsen et al., 2009) and three-quarter degree squares (3QDS). For the Cape, most plant occurrence records are only accurate to QDS level. Thus, analyses involving species occurrence data were necessary limited to scales including and above QDS.

Analyses were performed in R v3.4.0–3.5.1 (R Core Team, 2018). Version-numbers of specific R packages used are presented in the bibliography.

2.2 Environmental data sources

The GCFR was treated as the area occupied by the Succulent Karoo and Fynbos biomes in the current delineation of South Africa's biome boundaries (Mucina & Rutherford, 2006). The SWAFR was treated as the areas occupied by the Southwest Australia savanna, Swan Coastal Plain Scrub and Woodlands, Jarrah-Karri forest and shrublands, Southwest Australia woodlands, Esperance mallee, and Coolgardie woodlands in the World Wildlife Fund Terrestrial Ecoregions dataset (Olson et al., 2001) in order to closely match the currently delineated SWAFR (Gioia & Hopper, 2017, Hopper & Gioia (2004)). For the sake of readability, we shall refer to the GCFR and SWAFR simply as the Cape and SWA from hereon.

Geospatially-explicit raster layers were acquired for a selection of environmental variables (Table 1), for the regions of interest. Raster data were re-projected to a common coordinate reference: WGS84 (NIMA, 2000), using the “rgdal” (Bivand et al., 2017) package in R (R Core Team, 2018). All data were re-sampled to 0.05° resolution using the “resample” function in the R package “raster” (Hijmans, 2016), with the “bilinear” method.

158 An emphasis was made on using satellite-derived environmental data in this work, in order to minimise
159 differences in data quality and methodologies between the Cape and SWA. Additionally, satellite-derived data
160 have been shown to benefit regional-scale species distribution models (Deblauwe et al., 2016), thus motivating
161 their use in this regional-scale study. The environmental data used in this study were derived from NASA's
162 SRTM digital elevation model (Farr et al., 2007), NASA's MODIS/Terra spectroradiometric data for land
163 surface temperature and NDVI, the Climate Hazards Group's CHIRPS rainfall dataset (Funk et al., 2015), and
164 the International Soil Reference and Information Centre's SoilGrids250m edaphic dataset (Hengl et al., 2017)
165 (Table 1). SRTM and MODIS are entirely derived from satellite measurements, whereas CHIRPS is
166 interpolated from weather station data with satellite-derived radiometric measurements. SoilGrids250m is a
167 machine-learning derived product, based on soil measurements as a function of many covariates, including
168 MODIS and STRM sources (see Hengl et al., 2017), using random-forests and other classification-tree-based
169 methods, including gradient-boosting. For the soil data considered here (Table 1), we used depth-interval
170 weighted average values as the value for a particular soil variable in a given place.

171 Climatic and spectral data arise from satellites monitoring properties of the Earth's surface through time. We
172 therefore use the mean annual values for rainfall, surface temperature, and NDVI in each pixel in our analyses.
173 Pronounced seasonality of rainfall is a known feature of mediterranean systems . We describe this seasonality
174 by computing computing the precipitation in the driest quarter (PDQ), using methods based on the "biovars"
175 function in the R package "dismo".

176 **2.3 Plant occurrence data**

177 Geospatially-explicit records of vascular plant occurrences were downloaded from the Global Biodiversity
178 Information Facility (GBIF, Table 1). Queries were made for tracheophyte records from within the borders of
179 the Cape and SWA as treated here (GBIF, 24 July 2017, ???). Only records with defined species and
180 intra-specific ranks were kept. Intra-specific occurrences were treated as simply being representative of their
181 species. This resulted in FIXME unique species names in the Cape, and FIXME in SWA.

182 We cleaned these data using the R package "taxise" (Scott Chamberlain & Eduard Szocs, 2013; Chamberlain et
183 al., 2018) to check that these species names had accepted-status among taxonomic databases. We queried two
184 major taxonomic databases: the Global Name Resolver (GNR), and the Taxonomic Name Resolution Service
185 (TNRS). Should either one of these services return at least one match for a given name, then that name was
186 accepted. Those names for which no full binomial matches were found in either database were excluded from

the final list of species. The number of species names excluded totalled at FIXME and FIXME for the Cape and SWA respectively. Especially for SWA, these numbers may be deemed appreciably high. But, the occurrence records that would be dropped, as a consequence of these names' removals, appeared randomly distributed in geographic space in both regions. As such, any effect of the loss of these records in this analysis is likely uniform within the two regions.

After the unaccepted names were removed, it was important to ensure that a species was not listed under multiple synonyms. Such cases would skew estimates of species richness and turnover in this study. In light of this, the remaining names were queried in the Tropicos and Integrated Taxonomic Information System (ITIS) databases for their known synonyms, again using "taxize". These were collated to produce a nomenclatural "thesaurus" for the Cape and SWA species. This consisted of a list of the accepted species names in a region, each associated with a list of known synonyms. We amended species' names in the GBIF occurrence data, in order ensure species were listed under only one of these synonyms, replacing all appearances of a species' synonyms with the first synonym used in the list.

Lastly, We removed any species from both regions that are invasive aliens or non-indigenous. Alien species lists for plants in South Africa and Australia were acquired from the IUCN's Global Invasive Species Database (<http://www.iucngisd.org/gisd/>).

The final total plant species richness in each region was FIXME and FIXME for the Cape and SWA respectively. These final collections of species occurrence records were converted to raster-layers, wherein pixel-values represented the species richness of vascular plants within that pixel. These rasters were produced at QDS, HDS, and 3QDS resolutions.

2.4 Analyses

2.4.1 Quantifying environmental heterogeneity

In order to assess predictions (i) and (ii), we needed to describe the EH in both regions. Using the R package "raster" (Hijmans, 2016), we used a modified version of the "roughness" index in the "terrain" function. For a three by three neighbourhood N of cells, our index of roughness R is the average square-root of the squared difference between each of the n neighbour cells' values x_i and the central focal cell's value x_{focal} :

$$R(\mathbf{N}) = \frac{1}{n} \sqrt{\sum_{i=1}^n (x_{\text{focal}} - x_i)^2} \quad (1)$$

213 This value, notionally equivalent to the standard deviation of values relative to the focal value, is ascribed to
 214 the focal cell. Note, in order to use as much data from within regions' borders as possible, roughness was
 215 computed if a focal cell had at least one neighbour cell. Using this index, we produced raster layers of each of
 216 our nine environmental variable's heterogeneity. We compared the distributions of "roughness" values in each
 217 variable in each region with non-parametric Mann-Whitney U -tests, as almost all variables were highly
 218 non-normal, and could not be normalised by log-transformations. We also compare the effect size of the Cape
 219 vs SWA using the "common language effect size" ($CLES$), using the R package "canprot". The $CLES$ is the
 220 proportion of all pairwise comparisons between two sample groups' observations where one group's value is
 221 greater than the other's. We calculated the $CLES$ as the proportion of pairs where Cape roughness values
 222 were greater than that of SWA. This allowed us to assess prediction (i). To compare the spatial scales of
 223 heterogeneity (prediction (ii)) between each region, we repeated this analysis at all four spatial scales. This
 224 entailed recalculating the roughness layer for each variable after the original layer (0.05 degrees resolution) had
 225 been rescaled to each of the coarser resolutions.

226 2.4.2 Quantifying species turnover

227 Regarding prediction (iii), we wished to compare the general degree of species turnover in each region. To
 228 compare the extent of species turnover between the Cape and SWA, we determined two metrics of species
 229 turnover. The first, computes the mean species turnover as Jaccard distances (Oksanen et al., 2018) between
 230 each pair of QDS within each HDS (\bar{J}_{QDS} , based on HDS with $2 \leq n \leq 4$ QDS) in both regions. The second
 231 is defined in terms of Whittaker's additive definition of β -diversity, as follows:

$$\gamma = \alpha + \beta \quad (2)$$

232 Here, we treat species richness at the HDS-scale (S_{HDS}) as γ -diversity and at the QDS-scale (\bar{S}_{QDS}) as
 233 α -diversity. Intuitively, the species richness of an area is the result of some combination of the richness of sites
 234 within that area and the difference in species complements between those sites. Thus, we partition γ -diversity
 235 as in Equation (2), such that β -diversity is the difference between γ - and α -diversity. We compare the

distributions of \bar{J}_{QDS} and T_{HDS} using non-parametric Mann-Whitney U -tests, in order to guard against non-normality.

2.4.3 Predicting richness and turnover with environmental heterogeneity

Regarding prediction (iii), we wished to compare the general degree of species turnover in each region. For (iv) and (v) we modelled species richness (S) and turnover as a function of various combinations of environmental and environmental heterogeneity variables in both regions using boosted regression-tree (BRT) modelling techniques. This allowed us to explore which axes of environmental heterogeneity are most influential on vascular plant species richness and turnover, and the differences in the importance of such axes between the Cape and SWA.

BRTs are a flexible machine learning-based model of response variables and do so without involving normal null-hypothesis significance testing (Elith et al., 2008), and have been employed previously to model species richness (Thuiller et al., 2006; see Mouchet et al., 2015; Cramer & Verboom, 2016) as macro-ecological models. BRTs are developed through the iterative generation of non-linear regression trees. BRTs are an ensemble-approach, in which a prediction \hat{y}_i is based on the weighted sum of the predictions of progressively “less important” regression trees (t_k), as opposed to the predictions of one tree (Elith et al., 2008). For $k \rightarrow nt$ number of trees, where each tree is itself a function of the matrix \mathbf{X} of j predictor variables ($t_k = f(x_{ij})$), a BRT-model can be represented as follows:

$$\hat{y}_i = \sum_{k=1}^{nt} w_k t_k \quad (3)$$

BRTs have two major meta-parameters over which users have control (Friedman, 2001): the learning rate (lr , the rate at which iterative trees reduce predictive deviance during model-training, controlling the contribution of each tree to the final model) and tree complexity (tc , the number of nodes on a given regression-tree, i.e. the maximum interaction depth the model is permitted to fit).

BRTs were implemented here to predict both vascular plant species richness and turnover in each HDS, as a function of environmental variables and environmental roughness values in those cells, as Gaussian responses, thus resulting in two BRT-models for each region. We treated richness as S_{HDS} and turnover as \bar{J}_{QDS} . The natural logarithm of species richness was used, in order to satisfy the assumptions of a Gaussian response.

261 Note, this is not strictly because BRTs have any parametric assumptions concerning the distribution of the
262 response variable, but rather to aid in applying the Gaussian-family of BRT algorithms to the richness data
263 available. Additionally, BRTs were implemented to predict vascular plant species richness at the QDS-scale
264 (S_{QDS}), thus resulting in a total of six BRT-models presented here.

265 As recommended by Elith et al. (2008), BRT models were trained on a set of non-collinear predictor variables
266 using “gbm.step” in “dismo” (Hijmans et al., 2017) and “gbm”. Collinear predictor variables can skew the
267 interpretation of results, as the relative influence of mutually collinear variables is reduced. Collinearity
268 among the nine environmental predictor variables and their respective nine roughness-equivalents was assessed
269 using “removeCollinearity” in the R package “virtualspecies” (Leroy et al., 2015) separately for each region,
270 such that variables were no more than 80% collinear (Pearson’s $r \geq 0.80$). When faced with a cluster of
271 collinear variables, one variable was chosen manually therefrom. Where possible, the roughness-equivalent of
272 an environmental variable was included if its absolute-equivalent could also be included. When interpreting the
273 results of BRTs, it is important to consider the effects of the variables included as representative of the effect of
274 the excluded variables with which it was found to be collinear.

275 In order to select ideal lr and tc all models (described below) were trained on the final non-collinear predictor
276 sets iteratively for 25 combinations of a range of tc values (1 to 5) and a range of lr values (0.01, 0.005, 0.001,
277 5×10^{-4} , 1×10^{-4}). The function “gbm.step” optimises the number of trees (nt) using cross-validation during
278 model training (Elith et al., 2008) by halting iteration when predictions begin to overfit. For all models, we
279 used 10 cross-validation folds (i.e. use 10 different randomly selected training data sets), a tolerance-threshold
280 of 0.001, a bagging-fraction of 0.75 (proportion of training data randomly chosen to generate each tree), and
281 trained models starting with 50 trees, with each iterative step adding 50 trees at a time, up to a maximum of
282 10,000 trees. Following this iterative parameter optimisation, Gaussian BRT models were constructed with
283 $tc = 3$ and $lr = 0.001$, along with the other settings described.

284 The optimum configuration of lr and tc for the final model is a trade-off between model fit (e.g. pseudo- R^2 ;
285 Equation (4)) and complexity (nt). A tc of 5 was chosen for the final model. This follows the
286 recommendations of Elith et al. (2008), where lr and tc are advised to be adjusted inversely. This was chosen
287 in order to account for the complex interactions determining species richness. To avoid overfitting, an
288 intermediate lr of 0.001 was chosen.

289 2.4.4 Assessing BRT-predictions' fit

290 BRT-model performance can be described by measuring the variance in a dataset a BRT-model has explained,
291 quantified here by R^2_{pseudo} , which is the proportion of null deviance D_{null} explained by some model i .
292 Formally, it is defined as follows:

$$R^2_{\text{pseudo}} = 1 - \frac{D_i}{D_{\text{null}}} \quad (4)$$

293 The derivation of this metric is not easy to interpret, as it is not immediately clear what model deviance is.
294 Alternatively, comparing expected (i.e. model-predicted) and observed data has more heuristic appeal. We
295 employed this metric of BRT-model performance too. We regressed expected against observed richness and
296 turnover, and calculated the R^2 -value for those regressions (hereafter $R^2_{\text{E-O}}$).

297 The BRT-model fitting algorithm contains intrinsic stochasticity, due to the random partitions made in a dataset
298 during cross-validation. Though this randomness is usually negligible (e.g. variables' contributions vary from
299 run-to-run by a few decimal places), we reran each of the six BRT-models (see above) 1000 times in order to
300 account for this stochasticity. Where indicated, we either present the averages of these replicate-models' results
301 or the results of a representative model from each set of replicates.

302 In order to assess the reliability of the conclusions drawn from these models, we randomly permuted the
303 response data (S_{QDS} , S_{HDS} and \bar{J}_{QDS}) with respect to the environmental and heterogeneity data, and reran all
304 six BRT-models 999 times (with the final non-collinear predictor sets and preconfigurations above). This also
305 allows us to remove any effect of spatial autocorrelation in generating the observed correlations between
306 patterns of species occurrence and environment (Cramer & Verboom, 2016), and to allow us to assess the
307 significance of our results relative to a random null. Notably, as the predictor variables themselves are likely
308 spatially autocorrelated, correlation structure in model residuals is accounted for by the correlation structure in
309 the environmental data. Nonetheless, we wished to demonstrate our results more robustly and thus carried out
310 these permutation tests. For all six models, the majority of the 999 permuted models failed to find associations
311 between the response and predictor variables. The results of those that succeeded to fit a model to completion
312 (usually ca. 200 out of 999) are presented. The replicate and permuted BRT-models were compared using
313 various measures of model performance (above; *nt*, R^2_{pseudo} (Equation (4)), $R^2_{\text{E-O}}$) and the ranks of these
314 values for each replicate BRT-model relative to the 999 permuted models for that region/scope.

3 Results

3.1 Describing environmental heterogeneity across scales

Across all variables considered, the Cape is more environmentally heterogeneous in the majority of pairwise comparisons of grid-cells ($CLES > 0.50$, Mann-Whitney U -test: $P < 0.05$, Figure 1). The degree to which the Cape is more heterogeneous varies between environmental variables. These effects also vary somewhat across spatial scales. In some variables, the differentiation between Cape and SWA heterogeneity lessens at coarser scales (Figure 1b). Indeed, when comparing the overall ranking and medians of Cape vs SWA roughness values for each variable, we only find non-significant differences at the 3QDS scale (Mann-Whitney U tests, $P > 0.05$, Figure 1b).

Most obviously, and as expected, topographic heterogeneity is generally greater in the Cape than in SWA (Figure 1). Though SWA has a slightly wider distribution of elevational roughness values at coarse scales compared to fine scales than the Cape, the relative difference in heterogeneity between the two regions seems invariant with spatial scale ($CLES \approx 0.95$, Figure 1b). This concurs with our expectations, as the Cape is mountainous and known to have steep elevational gradients, while SWA is much more topographically uniform. Elevational roughness differs between the two regions in the manner we expected, giving us more confidence in the environmental reality of our other results here.

Climatic heterogeneity is less differentiated between the Cape and SWA than with topographic heterogeneity (Figure 1a), though the Cape is indeed more climatically heterogeneous (Figure ??b). Notably, the difference between roughness in mean annual rainfall (R MAP) and land surface temperature (R Surface T) in the Cape and SWA is less pronounced when considered at coarse spatial scales (Figure ??b). At all spatial scales considered, roughness in rainfall seasonality (R PDQ), however, is equally more heterogeneous in the Cape than SWA. Biological productivity, as measured by NDVI, is fairly similarly heterogeneous in the Cape and SWA ($CLES < 0.60$, Figure 1). Concerning edaphic variables, the Cape and SWA are similarly heterogeneous at coarser scales, particularly in terms of CEC and Soil C ($CLES \approx 0.50$, Figure 1b).

3.2 Comparing species turnover in the two regions

Following calculations of \bar{J}_{QDS} and T_{HDS} for each HDS-cell in each region, we also used non-parametric Mann-Whitney U -tests to compare the distributions of values in the Cape and SWA. The Cape possesses

generally greater floristic turnover than SWA, for both measures of turnover defined here ($P < 0.0001$, Figure 2a,b). \bar{J}_{QDS} measures the average pairwise Jaccard distance between QDS-cells in each HDS-cell. T_{HDS} , however, represents the inferred β component of γ -diversity. As γ -diversity ($= S_{\text{HDS}}$) in the Cape has a greater β -diversity component ($= T_{\text{HDS}}$) than SWA, the complement is necessarily true: γ -diversity in the Cape has a lesser α -diversity component ($= \bar{S}_{\text{QDS}}$) than SWA.

3.3 Predicting richness and turnover with environmental heterogeneity

We found vascular plant species richness and turnover both to be predicted primarily by environmental heterogeneity in the Cape (Figure 3a–c) and at least in-part by environmental heterogeneity in SWA (Figure 3d–f). Our six BRT-models performed adequately, and detected relationships between patterns of species occurrence and the environment that do not occur in the permuted datasets (Figures 4 and 3, Table 2).

BRT-models of species richness at the QDS-scale in each region generally performed best, as these models had fit greater number of trees (nt , Figure 4a), and possess higher R^2 -values (Figure 4b,c). SWA models of species richness and turnover at the HDS-scale out-performed Cape models. At the QDS-scale, the Cape and SWA models performed equally well (Figure 4, Table 3).

Across our BRT-models of species richness and turnover, the importance of different environmental variables in predictions differed substantially between the Cape and SWA. Additionally, the relative importance of absolute and heterogeneity variables also differs between the Cape and SWA (Figure 3). Most obviously, species richness and turnover in the Cape are predicted mostly by environmental heterogeneity, which is not the case in SWA (Figure 3). Species richness and turnover in the Cape are predicted by a broad suite of environmental variables, with no individual variable contributing more than ca. 20% to any model prediction (Figure 3a–c). The SWA models' predictions, however, are largely determined by MAP (Figure 3d–f).

Species richness at QDS-scales ($= \bar{S}_{\text{QDS}}$), and to a lesser extent at HDS-scales ($= S_{\text{HDS}}$), in the Cape is predicted largely edaphic conditions (Figure 3a,b). Species richness in SWA, at both scales, is mostly predicted by MAP and other climatic variables (Figure 3d,e). Interestingly, topographic heterogeneity did not feature as highly in contributing to Cape predictions as we expected (Figure 3a–c).

It is important to consider variables not included formally in these BRT-models that were found to be collinear with some of the variables included (see SI). Here, we interpret the effects of variables excluded from the analyses as well as those included, as the forms and importances of these relationships are likely similar. In the

370 Cape (concerning clusters of collinear variables relevant to those retained during BRT-model
371 fitting), MAP was included in the BRT-analyses as representative of a cluster of collinear variables consisting
372 of itself, NDVI, surface T and soil C at the. Roughness in soil clay content represented itself, roughness in soil
373 pH and roughness in NDVI. In SWA, MAP was select as representative of itself, NDVI and soil C.

374 Our BRT-models of species richness in both regions rank environmental variables somewhat differently at
375 QDS- and HDS-scales (Figure 3a,b,d,e). These differences in rankings are similar to those between two
376 unrelated lists ($P_{1-2} > 0.05$, Figure 5). This suggests some scale-dependence of different environmental
377 variables' associations with species richness.

378 It is noteworthy that BRT-models of species turnover ($= \bar{J}_{QDS}$, at HDS-scales) (Figure 3c,f) rank variables
379 similarly to models of richness at HDS-scales ($P_{2-3} \leq 0.005$, Figure 5). This is likely due to the fact that
380 proportional floristic turnover covaries with species richness. As such, though the signs of relationships
381 determining turnover may differ from those determining richness, the importances of different variables would
382 be similar.

383 In addition to different variables being more strongly associated with species richness and turnover in the Cape
384 compared to SWA (Figure 3), the forms of those relationships vary (Figure 6). We found MAP, and roughness
385 therein, to relate positively with species richness in both regions at both scales (Figure 6a,b,d,e). As MAP is
386 collinear with NDVI and soil C in both regions (and surface T in the Cape), this can be interpreted as the signal
387 of a biological productivity and resource availability associating with high levels of species richness.

388 The positive association of heterogeneity variables in the Cape as opposed to SWA (Figure 6a,b vs d,e) concurs
389 with their greater importance in BRT-model predictions (Figure 3).

390 The fact that species turnover ($T_{HDS} = \bar{J}_{QDS}$) in the Cape and SWA is largely predicted by the same
391 variables as species richness, but with opposite signs to its relationships (Figure 6c,f), is indicative of the
392 richness-dependence of the measure of floristic turnover used here (Jaccard distances) to quantify turnover at
393 the HDS-scale.

394 **4 Discussion**

395 Here we have provided support for the hypothesis that the difference in plant species richness between the
396 Cape and SWA is accounted for by the greater abiotic heterogeneity in the Cape. As expected, the Cape is

397 shown to possess (i) a quantifiably more heterogeneous environment that is (ii) generally heterogeneous at a
398 finer spatial scale than SWA, with (iii) greater levels of floristic turnover. We have shown that vascular plant
399 species richness (iv) can be explained in terms of environmental conditions including environmental
400 heterogeneity in both the Cape and SWA. Also, (v) the sets of environmental axes that explain plant species
401 richness differ between the Cape and SWA. These findings contribute towards an understanding of the
402 ecological conditions associated with high levels of species co-existence in these two regions, and strengthens
403 the generality of a positive relationship between EH and biodiversity.

404 Cook et al. (2015) summarised the suites of hypotheses for the high levels of plant diversity in
405 mediterranean-type ecosystems. Broadly, these biodiversity hotspots are thought to arise due to greater levels
406 of evolutionary diversification, ecological co-existence and time for species accumulation by either
407 immigration or in situ evolution. Our study concerns the dual role of EH in facilitating ecological speciation
408 along environmental gradients and a diversity of habitats in which greater numbers of species may co-exist. We
409 have correlative evidence for these processes, in that EH was positively associated with high levels of species
410 richness and turnover in both regions. Further research should be aimed at distinguishing the roles of ecological
411 speciation and species co-existence associated with EH in mediterranean-type ecosystems. The role of
412 environmental stability is relevant here (Cook et al., 2015, Pinto-Ledezma et al., 2018), both by allowing more
413 continuous species accumulation when habitats persist through geologic time and by allowing environmental
414 gradients to be stable long enough for ecological speciation to occur (Pinto-Ledezma et al., 2018). In the
415 contexts of the Cape and SWA, this long term environmental stability, and the stability of EH through time, is
416 indeed expected to contribute to these regions biodiversity (Hopper, 1979; Cowling et al., 1996).

417 *Ed: I'd like to include more information on the landscape/geologic history of the regions here, and link this*
418 *to the notion of environmental stability and "species accumulation", sensu Cook et al. (2015) and*
419 *Pinto-Ledezma et al. (2018).*

420 The Cape and SWA regions present differentiable environmental spaces, each with varying degrees of
421 heterogeneity across spatial scales and environmental axes. The clear distinction of the regions' topographic
422 features is as expected. The Cape region has been found previously to have the second highest median
423 topographic heterogeneity of the five Mediterranean-climate regions (Bradshaw & Cowling, 2014). In the
424 models developed by Cramer & Verboom (2016) for South Africa, roughness in topography was largely
425 superseded as an important predictor of species richness by other roughness variables. This is the case with our
426 results too, likely as topographic complexity can be considered a proxy for gradients in other environmental

axes, such as rainfall or temperature. In line with our hypotheses, a combination of absolute and heterogeneity variables are associated with species richness and turnover. These effects are region-specific, particularly in that patterns in the Cape are more strongly determined by edaphic conditions and EH while SWA patterns are more strongly determined by absolute environmental conditions, particularly climate. Kreft & Jetz (2007) modelled global terrestrial vascular plant species richness, which focussed primarily on using absolute environmental variables, underestimated the richness of the Cape flora. Though Kreft & Jetz (2007) did include topographic heterogeneity in their predictor set, topography is often a proxy for more biologically meaningful variables (Cramer & Verboom, 2016). This explains why the inclusion of these variables (e.g. roughness in mean annual precipitation) yields more accurate predictions of species richness. Indeed, Thuiller et al. (2006) also included topographic heterogeneity.

We also detected strongly non-linear relationships between environmental and heterogeneity variables and species richness and turnover. We expected this, as non-linear relationships are common in ecology (Zanne et al. 2018), making our use of machine-learning methods appropriate here.

Comparing our models to those fit by Cramer & Verboom (2016), we did not directly find spatial heterogeneity in NDVI to be an important determinant of species richness and turnover patterns in the Cape. Rather absolute MAP, which is collinear with absolute NDVI, was found to be important. This speaks to the relationship between vegetation productivity and climate, and how suitably productive habitats can foster more species. An additional interpretation of spatial heterogeneity (and indeed absolute conditions) in NDVI is that of changing canopy light-contexts (Cramer & Verboom, 2016). Power et al. (2017) found that floristic turnover between biomes in South Africa to be associated with differences in biomes' leaf area indices. We found NDVI to be more heterogeneous across the Cape than SWA (Figures 1). NDVI is an integrating variable, which captures information about productivity, light availability, and soil nutrients (Power et al., 2017). Absolute NDVI and its correlates can be interpreted as contributing to predicting species richness in the Cape. This demonstrates the role of ecological productivity in facilitating the co-existence diverse species assemblages. EH, then, must be considered alongside resource- and energy-availability axes when interpreting the drivers of species richness. In so much as a diverse environmental space supports more species, the materials and productivity required for biota to thrive are also needed to support species (Gaston, 2000; Kerr et al., 2001; Bøhn & Amundsen, 2004; Kreft & Jetz, 2007). As such, our findings, along with those of previous studies (Rensburg et al., 2002; Thuiller et al., 2006; Kreft & Jetz, 2007; Cramer & Verboom, 2016), suggest that there are ecological and evolutionary consequence to both resource availability and EH.

457 We detected a strong set of links between various edaphic properties and vascular plant species richness and
458 turnover in the Cape. The importance of absolute pH speaks to the association of many Cape communities with
459 nutrient impoverished, acidic soils. We did not, however, find edaphic conditions to be as important as we
460 expected in SWA. This may be due to a real lack of importance of edaphic features in driving assemblage
461 patterns in SWA. Alternatively, there is a risk that the quality of soil data provided to our models, from
462 SoilGrids250m, is insufficient for our purposes here. There are two potential reasons for this. Firstly, the
463 SoilGrids250m data product is acknowledged to be less accurate at the scales we investigate here, such that the
464 true variation in soil properties in SWA is not represented in our dataset. Secondly, it could be that the link
465 between soils and assemblage patterns in SWA exists, but simply at a finer scale than that at which we
466 performed our analyses here. These two reasons are not mutually exclusive, in that SoilGrids250m may be
467 both inaccurate at the scales we consider here and that our analysis considers scales greater than those
468 important in SWA plant community assemblage patterns. This is particularly relevant to our study, as the Cape
469 has a much wider range of scales exhibited in the heterogeneity across its environmental axes. Notably, each
470 region has finer scale heterogeneity in some variables, and coarser scale in others—neither region is more fine
471 or coarse than the other over all environmental axes.

472 *Ed: I'd obviously like to include more information on the landscape/geologic history of the regions here*
473 *too, and with reference to particular soil variables (Mike?).*

474 Another issue related to spatial scales, as with any ecological study, is that of spatially uniform and
475 representative plant species occurrence data. Efforts were made herein to minimise differences in data-quality
476 between SWA and Cape by using remote-sensing derived environmental data. Species occurrence data,
477 however, prove difficult to obtain with uniform methodology. South African biodiversity data is stored only to
478 QDS-level accuracy. As such, our analyses were limited to that as the finest spatial scale, such that our
479 environmental datasets necessarily, then, were also limited to the QDS-scale. This may complicate the
480 interpretation of our analyses, as these data may be too coarse in scale to capture patterns of species richness
481 and turnover and their associations with the environment in reality in empirically heterogeneous environments
482 like the Cape and SWA.

483 In addition to the scale of our data, spatial bias in species occurrence data must also be considered. GBIF data
484 are sourced from both herbaria and plot-monitoring datasets. The relative contribution of each of these to GBIF
485 data may vary between countries and regions—i.e. between the Cape and SWA. As recently noted by Guerin
486 (2018), herbarium records and plot-based inventories describe regional flora with varying accuracy, especially

487 with respect to documenting high levels of floristic turnover. We emphasise here our concerns that there the
488 floral occurrence data and soils information use in this study are especially at risk of spatial bias and/or
489 inaccuracies.

490 ...

491 Jaccard distances, as used here as \bar{J}_{QDS} , are highly richness dependent. Although they represent the
492 proportional floristic turnover between cells, this proportion itself is sensitive to the richness of sites under
493 comparison. For example, the turnover between two low richness sites has is likely to be greater by chance,
494 due to the absence or presence of a few species, than the turnover between two high richness sites, where the
495 absence or presence of a few species does not greatly affect the turnover calculated.

496 ...

497 Our findings here are correlative. There are, however, many proposed mechanisms to explain the correlative
498 signals demonstrated here. Our findings support the hypothesis that mediterranean systems' plant species
499 richness is a function of spatial variability in environmental conditions. This can stimulate diversification, and
500 maintain that diversity by providing a range of habitats for species co-existence. Oligotrophic soils can stimulate
501 an increase in functional diversity, through the evolution of diverse nutrient acquisition strategies (Lambers et
502 al., 2010; Verboom et al., 2017)—e.g. sclerophylly (Cramer et al., 2014; Cook et al., 2015). An aspect of the
503 environment I have neglected to consider is fire, shown to also contribute to predictions here in the Cape
504 (Cramer & Verboom, 2016). Cardillo (2012) have shown the structuring forces behind species co-occurrence
505 patterns, and thus likely species richness, differ between species-pairs with different post-fire responses and
506 those with similar post-fire responses.

507 Following from the understanding that functionally diverse assemblages, which are more likely to be more
508 species rich, are likely to arise and/or occur in areas with diverse ecological pressures (Molina-Venegas et al.,
509 2015), one would expect, then, heterogeneous habitats such as those in Mediterranean-type biodiversity
510 hotspots to exhibit high levels functional beta diversity along steep environmental gradients (Molina-Venegas
511 et al., 2015). If the niches concerning these functions are phylogenetically conserved among those biota, then
512 one would also expect high levels of species and phylogenetic beta diversity along these gradients
513 (Molina-Venegas et al., 2015). This concurs with the notion put forward by Power et al. (2017), wherein
514 megadiverse systems such as these represent the results of “phylogenetic niche conservatism on a
515 heterogeneous landscape”. Thus, species and phylogenetic turnover should covary with environmental

516 heterogeneity in some way. Indeed, endemism, at certain scales, could also follow this pattern. Thuiller et al.
517 (2006) demonstrated that there are phylogenetic and biome-related determinants of species richness. This
518 makes sense in light of the low probability of lineages crossing biome boundaries in Mediterranean systems
519 (Power et al., 2017). NDVI and light availability, and the heterogeneity therein, are associated with high levels
520 of floristic turnover (Power et al., 2017). This may be indicative of ecological specialisation precluding species
521 from crossing these boundaries, thus increasing the level of endemism within a region, while also increasing
522 the level of turnover, and thus likely species richness, along environmental gradients. Although, this may be
523 debated. Beard et al. (2000) state that the high levels of endemism in SWA are function of habitat specialisation
524 to soil mosaics. Cf. Laliberte et al. (2014), who say that this endemism is likely due to environmental filtering
525 along these soil turnover sequences, as opposed to the juxtaposition of specialised species along soil gradients.

526 *Ed: Is it worth expanding this discussion to functional diversity and phylogenetic diversity metrics too?*

527 Summarily, we have demonstrated support for the generality of EH as a meaningful predictor of patterns of
528 species richness and turnover in mediterranean-type ecosystems. In SWA and the Cape, high levels of are also
529 likely the results of long-term landscape and climatic stability (Hopper, 1979). Thus, the roles of environmental
530 variability through space that persists through deep time are two important ways in which the environment
531 relates to biodiversity in these regions. The efficacy of environmental stability through geological time should
532 be explored further, comparing the Cape and SWA.

Table 1: Georeferenced vascular plant species occurrence and environmental data sources used in this study. Data were acquired for the Cape and SWA regions, with the temporal extent of data products used described where applicable. Abbreviations are as follows: MAP, mean annual precipitation; PDQ, precipitation in the driest quarter; CEC, cation exchange capacity.

Variable	Source	Temporal extent	Citation
Plant species occurrences	GBIF	TODO	??, ??
Elevation	SRTM v2.0		??
NDVI	MODIS (MOD13C2)	Feb. 2000 to Apr. 2017	??
Climatic variables			
Surface temperature	MODIS (MOD11C3)	Feb. 2000 to Apr. 2017	??
MAP	CHIRPS v2.0	Jan. 1981 to Feb. 2017	??
PDQ	CHIRPS v2.0	Jan. 1981 to Feb. 2017	??
Soil variables			
CEC	SoilGrids250m (CECSOL M 250m)		??
Clay	SoilGrids250m (CLYPPT M 250m)		
Soil C	SoilGrids250m (OCDENS M 250m)		
pH	SoilGrids250m (PHIKCL M 250m)		

Table 2: Average proportional-ranks for BRT-model performance measures (nt , R^2_{pseudo} (Equation (4)), $R^2_{\text{E-O}}$ (see text)) of the 1000 replicate BRT-models relative to 999 BRT-models fit to permuted datasets. Each of the 1000 replicate BRT-models was ranked against the 999 permuted BRT-models. The average rank of each, as a proportion, is presented.

Model	nt	R^2_{pseudo}	$R^2_{\text{E-O}}$
QDS-richness			
GCFR	1.000	1.000	1.000
SWAFR	1.000	1.000	1.000
HDS-richness			
GCFR	0.987	1.000	0.988
SWAFR	1.000	1.000	1.000
HDS-turnover			
GCFR	0.977	0.992	0.979
SWAFR	0.997	1.000	1.000

Table 3: Estimated differences between replicate Cape and SWA BRT-models' performance measures (nt , R^2_{pseudo} (Equation (4)), $R^2_{\text{E-O}}$ (see text)) following two-sided t -tests. Positive values indicate that the Cape models had greater values. In all cases, the Cape and SWA had highly significantly different values for these quality measures ($P < 0.0001$).

Model	nt	R^2_{pseudo}	$R^2_{\text{E-O}}$
QDS-richness	542.938	0.063	-0.005
HDS-richness	-808.994	-0.064	-0.233
HDS-turnover	-997.045	-0.052	-0.296

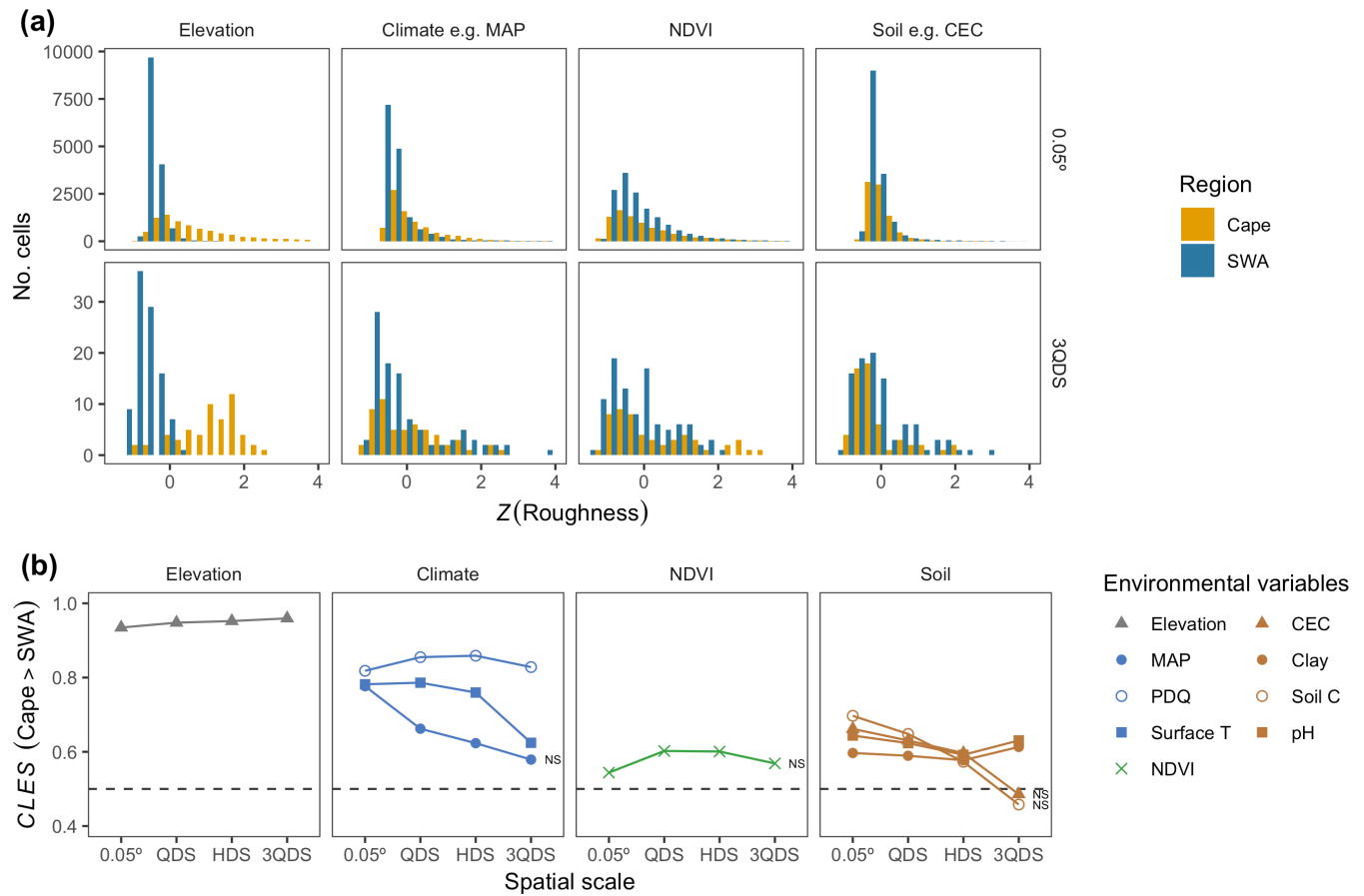


Figure 1: Comparisons of different types of environmental heterogeneity between the the Greater Cape Floristic Region (Cape) and the Southwest Australia Floristic Region (SWA). We present (a) distributions of roughness values (Equation (1)) for example variables from each broad category of the environment concerned. (b) The common language effect size ($CLES$; see text) of Cape versus SWA roughness values is shown for all variables, grouped by broad categories of the environment, describing differences in the distributions of Cape and SWA roughness values. We used Mann-Whitney U -tests to assess differences in these distributions. Non-significant differences ($P_U > 0.05$) are denoted as such (“NS”). Note, U -tests were performed using only a random set of 5000 cells at the 0.05-degree-scale, as the U -test as implemented in R cannot handle more than that many values to compare.

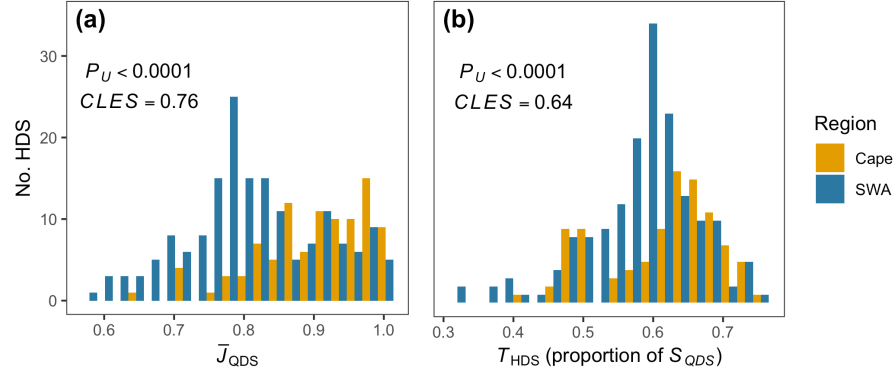


Figure 2: Species turnover, described in two forms ((a) mean Jaccard distance between QDS in each HDS (\bar{J}_{QDS}), (b) additively defined turnover (T_{HDS} , Equation (2)) as a proportion of HDS richness (S_{HDS})), compared between the Cape and SWA. Mann-Whitney U -tests between the Cape and SWA distributions of \bar{J}_{QDS} and T_{HDS} yielded significant differences (see P -values and common language effect sizes ($CLES$) inset).

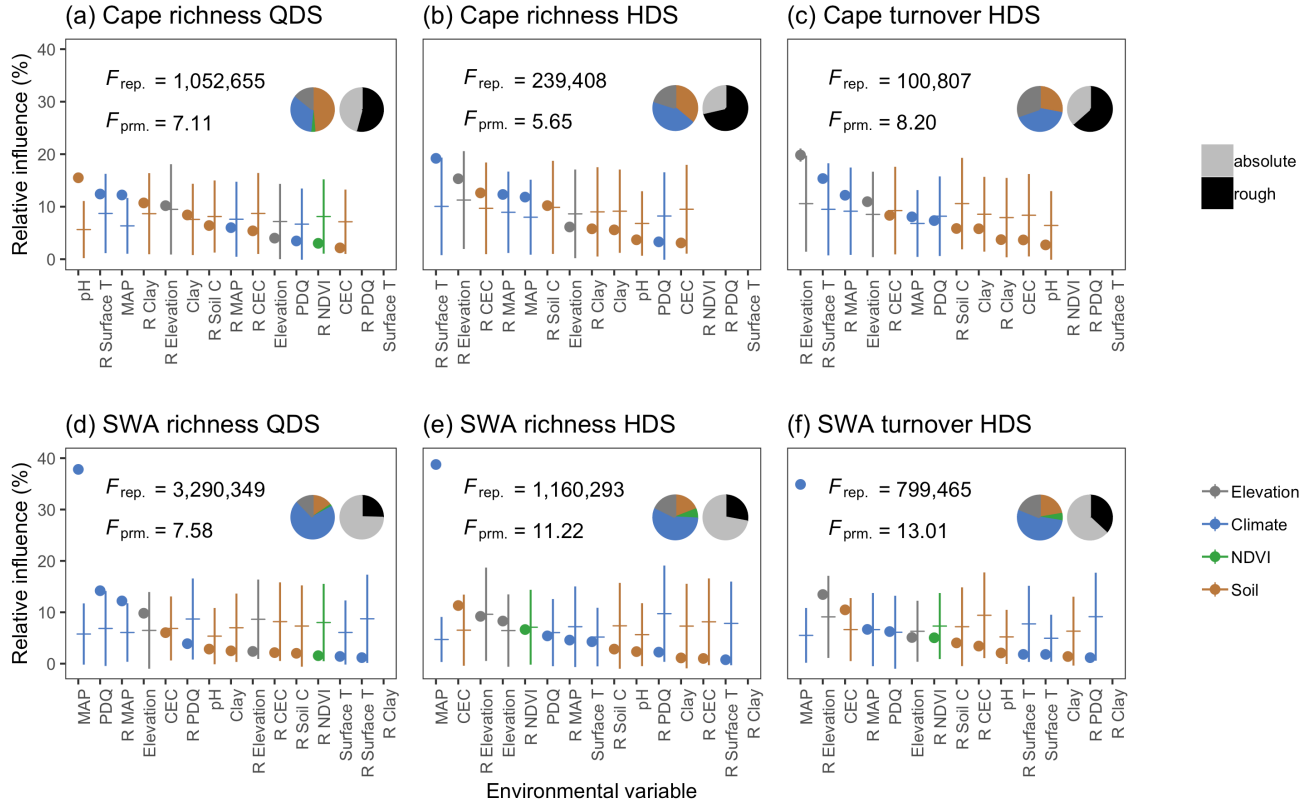


Figure 3: Relative influence of environmental variables (including heterogeneity variables—prefixed with “R”) in boosted regression tree (BRT) model predictions in the Greater Cape Floristic Region (Cape, a–c) and the Southwest Australia Floristic Region (SWA, b–d) of vascular plant species richness at the (b,e) QDS-scale ($= \bar{S}_{QDS}$), (a,d) HDS-scale ($= S_{HDS}$) and (c,f) turnover ($= \bar{J}_{QDS}$). All BRT-models were permitted to fit three-way interactions between environmental variables. Points denote the mean contribution of an environmental variable to model-predictions across the 1000 replicate BRT-models for that region/scope. Horizontal ticks denote the mean for the 999 permuted BRT-models. Standard deviations above and below these means are shown with vertical lines. Note, in the case of the replicate, standard deviations are so small such that the vertical lines are obscured by the points. Colours represent the general category of the environment to which a variable belongs (keyed), as in Figure 1b. Left-most piecharts inset in each panel display the same information. Right-most piecharts group contributions according to whether a variable was absolute or roughness-transformed (keyed). F -statistics inset are for one-way ANOVAs of differences in variables’ relative influences—for both the replicate ($F_{rep.}$) and permuted ($F_{prm.}$) BRT-models.

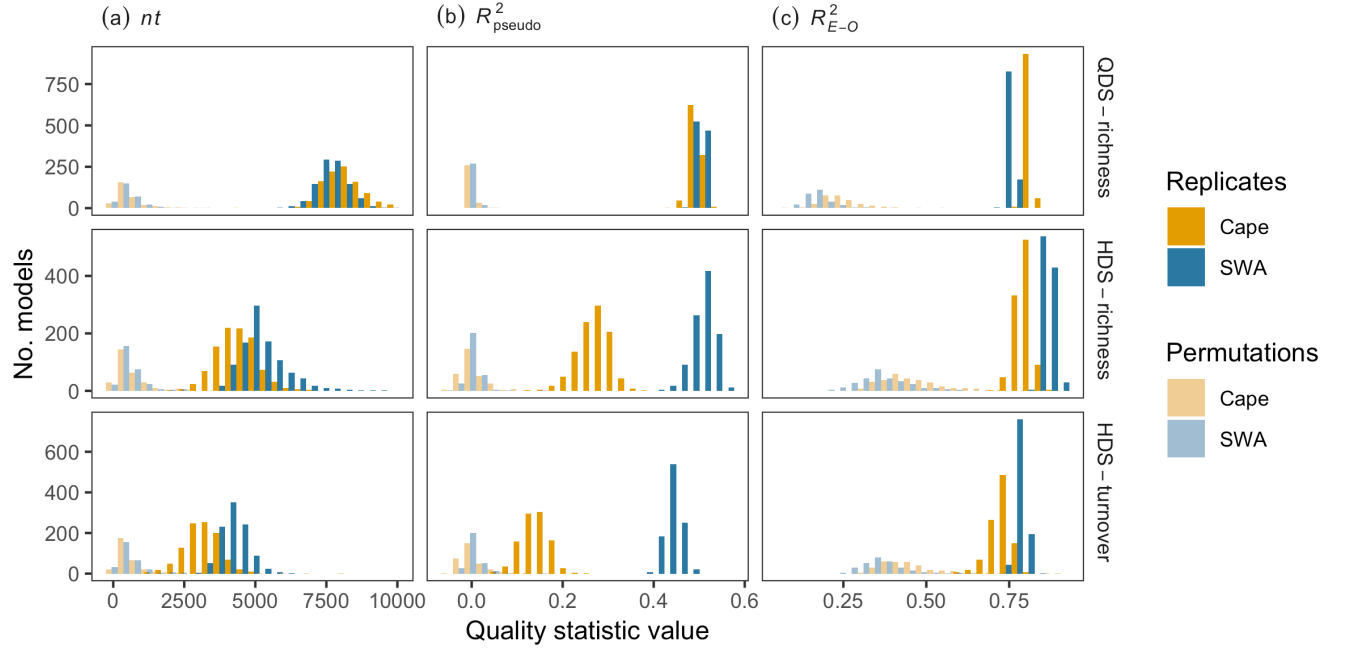


Figure 4: Distributions of three measures of boosted regression tree (BRT) model performance: (a) the number of trees in the model nt , (b) R^2_{pseudo} (Equation (4)), (c) R^2_{E-O} (see text). These measures are presented for the six sets of permuted (pale bars) and six sets of replicate BRT-models (dark bars) as in Figure 3, coloured according to the region of interest as in Figures 1a and 2. In all cases, replicate BRT-models almost entirely out-rank the permuted models in terms of performance (Table 2) and the Greater Cape Floristic Region (Cape) and Southwest Australia Floristic Region (SWA) models had significantly different values for each metric (Table 3). Note, the actual differences between Cape and the SWA models' values is not realistically significant in some cases (e.g. the difference in nt between the Cape and SWA QDS richness models is statistically significant, but are observedly so similar as not to affect interpretation).

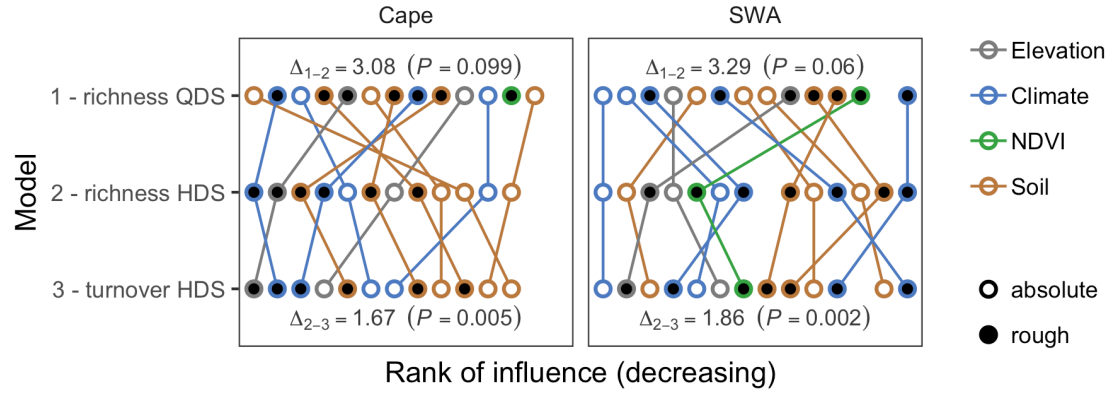


Figure 5: Differences in the rankings of environmental variables' (including heterogeneity variables) relative influences on boosted regression tree (BRT) model predictions of vascular plant species richness and turnover in (a) the Greater Cape Floristic Region (Cape) and (b) Southwest Australia Floristic Region (SWA) (as in Figure 3). Each point represents an environmental variable's rank in BRT-model importance, decreasing in importance from left to right. Rankings used here are the same as that of the average relative influence for variables across replicate BRT-models, presented in Figure 3. Coloured lines connect points representing the same environmental variable. Points' outlines are coloured according to the general category of the environment (keyed) to which a variable belongs, as in Figures 1b and 3, while points' centres are coloured according to whether a variable was roughness-transformed or not. The comparisons of variables' rankings of interest are between QDS- and HDS-scale richness ($= \bar{S}_{QDS}$ and S_{HDS} respectively; rows nos. 1 and 2) and between HDS-scale richness and turnover ($= \bar{J}_{QDS}$) (rows nos. 2 and 3). Statistics (Δ - and P -values) inset at the top and bottom of each panel refer to these comparisons respectively. Δ -values represent the average absolute difference in ranks across variables between two models' rankings. The associate P -value results from ranking the observed Δ -values against 999 Δ -values based on random permutations of variables' rankings (SI1), such that more significant P -values denote rankings more similar than would be expected by chance.

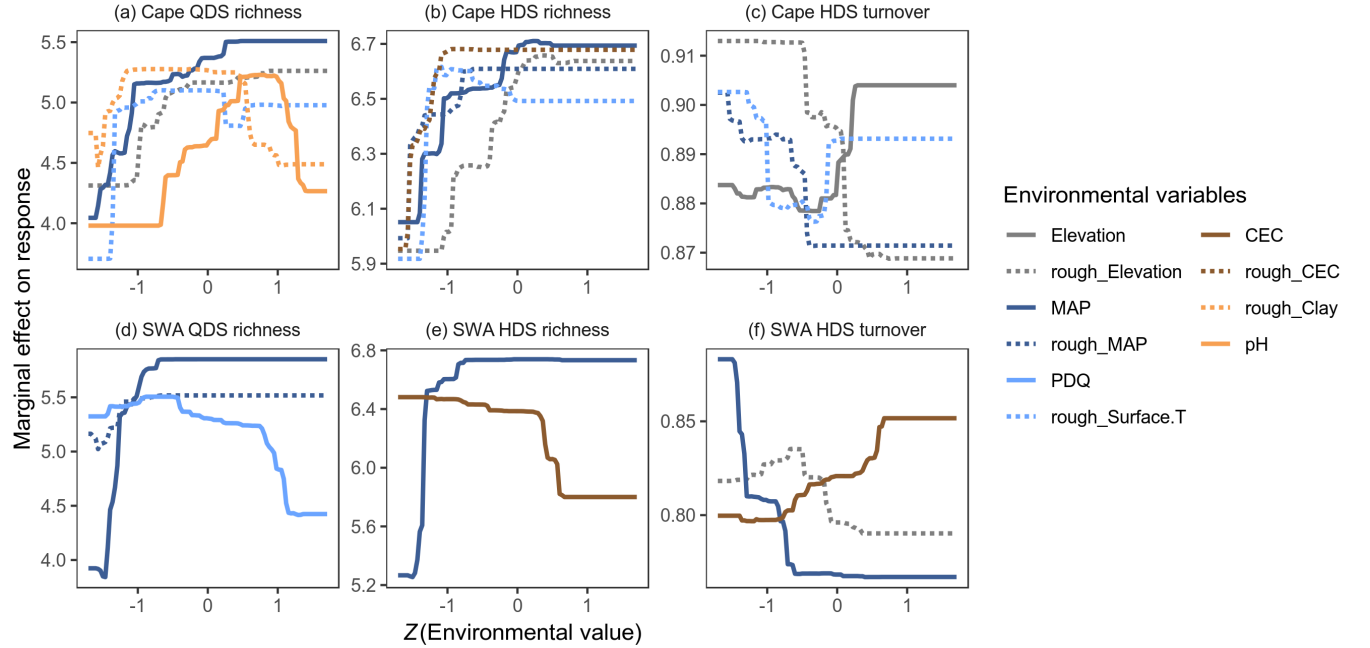


Figure 6: Marginal effects of environmental conditions and heterogeneity on vascular plant species richness at the QDS-scale ($= \bar{S}_{QDS}$; a,d), HDS-scale ($= S_{HDS}$; b,e) and turnover ($= \bar{J}_{QDS}$; c,f) in response variables in the Greater Cape Floristic Region (Cape; a–c) and Southwest Australia Floristic Region (SWA; d–f) following boosted regression tree (BRT) modelling. Marginal effect functions presented are derived from a representative BRT-model from the set of replicate BRT-models (for each of the six modelling cases) (see SI regarding how representative BRT-models were selected). Marginal effects represent the effect of a predictor variable when all other predictors are set at their means. Marginal effect functions are shown for environmental variables that contributed $\geq 10\%$ to a model’s predictions. Functions are coloured as keyed, with solid lines representing absolute environmental variables and dotted representing heterogeneity variables (“rough”). Environmental variables were all rescaled here such as to be centred on zero (i.e. Z -transformed), facilitating comparison of functions’ forms.

535 **Biosketches**

536 **Ruan van Mazijk**

537 **Michael D. Cramer**

538 **G. Anthony Verboom**

539 **Author contributions**

540 MDC and GAV conceived the study question, which RVM investigated under their supervision for his BSc
541 Hons project. The analyses and programming work were largely devised by RVM, with input from the other
542 authors, and was carried out by RVM. RVM wrote the first draft of the manuscript and all authors contributed
543 equally thereafter.

References

- Baudena, M., Sánchez, A., Georg, C.-P., Ruiz-Benito, P., Rodríguez, M.Á., Zavala, M.A., & Rietkerk, M. (2015) Revealing patterns of local species richness along environmental gradients with a novel network tool. *Scientific Reports*, **5**, 11561.
- Beard, J.S., Chapman, A.R., & Gioia, P. (2000) Species richness and endemism in the Western Australian flora. *Journal of Biogeography*, **27**, 1257–1268.
- Bivand, R., Keitt, T., & Rowlingson, B. (2017) rgdal: Bindings for the Geospatial Data Abstraction Library. R package version 1.2-7..
- Bradshaw, P.L. & Cowling, R.M. (2014) Landscapes, rock types, and climate of the Greater Cape Floristic Region. *Fynbos: Ecology, evolution and conservation of a megadiverse region* (ed. by N. Allsopp, J.F. Colville, and G.A. Verboom), pp. 26–46. Oxford University Press, Oxford.
- Bøhn, T. & Amundsen, P.-A. (2004) Ecological Interactions and Evolution: Forgotten Parts of Biodiversity? *BioScience*, **54**, 804.
- Cardillo, M. (2012) The phylogenetic signal of species co-occurrence in high-diversity shrublands: different patterns for fire-killed and fire-resistant species. *BMC Ecology*, **12**, 21.
- Chamberlain, S., Szoecs, E., Foster, Z., Arendsee, Z., Boettiger, C., Ram, K., Bartomeus, I., Baumgartner, J., O'Donnell, J., Oksanen, J., Tzovaras, B.G., Marchand, P., & Tran, V. (2018) *Taxize: Taxonomic information from around the web*.
- Cook, L.G., Hardy, N.B., & Crisp, M.D. (2015) Three explanations for biodiversity hotspots: small range size, geographical overlap and time for species accumulation. An Australian case study. *New Phytologist*, **207**, 390–400.
- Cowling, R.M., Rundel, P.W., Lamont, B.B., Arroyo, M.K., & Arianoutsou, M. (1996) Plant diversity in mediterranean-climate regions. *Trends in Ecology and Evolution*, **11**, 362–366.
- Cramer, M.D. & Verboom, G.A. (2016) Measures of biologically relevant environmental heterogeneity improve prediction of regional plant species richness. *Journal of Biogeography*, 1–13.
- Cramer, M.D., West, A.G., Power, S.C., Skelton, R., & Stock, W.D. (2014) Plant ecophysiological diversity.

570 *Fynbos: Ecology, evolution and conservation of a megadiverse region* pp. 248–272. Oxford University
571 Press, Oxford.

572 Deblauwe, V., Droissart, V., Bose, R., Sonké, B., Blach-Overgaard, A., Svenning, J.C., Wieringa, J.J., Ramesh,
573 B.R., Stévant, T., & Couvreur, T.L.P. (2016) Remotely sensed temperature and precipitation data improve
574 species distribution modelling in the tropics. *Global Ecology and Biogeography*, **25**, 443–454.

575 Elith, J., Leathwick, J.R., & Hastie, T. (2008) A working guide to boosted regression trees. *Journal of Animal*
576 *Ecology*, **77**, 802–813.

577 Farr, T., Rosen, P., Caro, E., Crippen, R., Duren, R., Hensley, S., Kobrick, M., Paller, M., Rodriguez, E., Roth,
578 L., Seal, D., Shaffer, S., Shimada, J., Umland, J., Werner, M., Oskin, M., Burbank, D., & Alsdorf, D. (2007)
579 The shuttle radar topography mission. *Reviews of Geophysics*, **45**, 1–33.

580 Funk, C.C., Peterson, P.J., Landsfeld, M., Pedreros, D.H., Verdin, J., Shukla, S., Husak, G., Rowland, J.D.,
581 Harrison, L., Hoell, A., & Michaelsen, J. (2015) The climate hazards infrared precipitation with stations—a
582 new environmental record for monitoring extremes. *Scientific Data*, **2**, 150066.

583 Gaston, K.J. (2000) Global patterns in biodiversity. *Nature*, **405**, 220–227.

584 GBIF (24 July 2017) GBIF Occurrence Download..

585 GBIF (24 July 2017) GBIF Occurrence Download..

586 Gioia, P. & Hopper, S.D. (2017) A new phytogeographic map for the Southwest Australian Floristic Region
587 after an exceptional decade of collection and discovery. *Botanical Journal of the Linnean Society*, **184**, 1–15.

588 Guerin, G. et al. (2018) When macroecological transitions are a fiction of sampling: Comparing herbarium
589 records to plot-based species inventory data. *Ecography*, 1864–1875.

590 Hart, S.P., Usinowicz, J., & Levine, J.M. (2017) The spatial scales of species coexistence. *Nature Ecology &*
591 *Evolution*, **1**, 1066–1073.

592 Hengl, T., Mendes de Jesus, J., Heuvelink, G.B.M., Ruiperez Gonzalez, M., Kilibarda, M., Blagoti?, A.,
593 Shangguan, W., Wright, M.N., Geng, X., Bauer-Marschallinger, B., Guevara, M.A., Vargas, R., MacMillan,
594 R.A., Batjes, N.H., Leenaars, J.G.B., Ribeiro, E., Wheeler, I., Mantel, S., & Kempen, B. (2017)
595 SoilGrids250m: Global gridded soil information based on machine learning. *PLoS ONE*, **12**, e0169748.

596 Hijmans, R.J. (2016) raster: Geographic Data Analysis and Modeling. R package version 2.5-8..

597 Hijmans, R.J., Phillips, S., Leathwick, J., & Elith, J. (2017) dismo: Species Distribution Modeling. R package
598 version 1.1-4..

599 Hopper, S.D. (1979) Biogeographical Aspects of Speciation in the Southwest Australian Flora. *Annual Review*
600 *of Ecology and Systematics*, **10**, 399–422.

601 Hopper, S.D. & Gioia, P. (2004) The Southwest Australian Floristic Region: Evolution and Conservation of a
602 Global Hot Spot of Biodiversity. *Annual Review of Ecology, Evolution, and Systematics*, **35**, 623–650.

603 Kerr, J.T., Southwood, T.R., & Cihlar, J. (2001) Remotely sensed habitat diversity predicts butterfly species
604 richness and community similarity in Canada. *Proceedings of the National Academy of Sciences of the*
605 *United States of America*, **98**, 11365–11370.

606 Kreft, H. & Jetz, W. (2007) Global patterns and determinants of vascular plant diversity. *Proceedings of the*
607 *National Academy of Sciences*, **104**, 5925–5930.

608 Laliberte, E., Zemunik, G., & Turner, B.L. (2014) Environmental filtering explains variation in plant diversity
609 along resource gradients. *Science*, **345**, 1602–1605.

610 Lambers, H., Brundrett, M.C., Raven, J.A., & Hopper, S.D. (2010) Plant mineral nutrition in ancient
611 landscapes: high plant species diversity on infertile soils is linked to functional diversity for nutritional
612 strategies. *Plant and Soil*, **334**, 11–31.

613 Larsen, R., Holmern, T., Prager, S.D., Maliti, H., & Røskoft, E. (2009) Using the extended quarter degree grid
614 cell system to unify mapping and sharing of biodiversity data. *African Journal of Ecology*, **47**, 382–392.

615 Leroy, B., Meynard, C.N., Bellard, C., & Courchamp, F. (2015) Virtualspecies, an r package to generate virtual
616 species distributions. *Ecography*.

617 Levin, L.A., Sibuet, M., Gooday, A.J., Smith, C.R., & Vanreusel, A. (2010) The roles of habitat heterogeneity
618 in generating and maintaining biodiversity on continental margins: an introduction. *Marine Ecology*, **31**, 1–5.

619 Lobo, J.M., Jay-robert, P., Lumaret, J.-p., Lobo, J.M., Jay-robert, P., & Lumaret, J.-p. (2004) Modelling the
620 Species Richness Distribution for French Aphodiidae (Coleoptera, Scarabaeoidea). *Ecography*, **27**, 145–156.

621 Manning, J., Goldblatt, P., & Others (2012) *Plants of the Greater Cape Floristic Region. 1: The Core Cape*

622 *flora*. South African National Biodiversity Institute,

623 Mateo, R.G., Mokany, K., & Guisan, A. (2017) Biodiversity Models: What If Unsaturation Is the Rule? *Trends*
624 *in Ecology & Evolution*, **32**, 556–566.

625 Molina-Venegas, R., Aparicio, A., Slingsby, J.A., Lavergne, S., & Arroyo, J. (2015) Investigating the
626 evolutionary assembly of a Mediterranean biodiversity hotspot: Deep phylogenetic signal in the distribution of
627 eudicots across elevational belts. *Journal of Biogeography*, **42**, 507–518.

628 Mouchet, M., Levers, C., Zupan, L., Kuemmerle, T., Plutzar, C., Erb, K., Lavorel, S., Thuiller, W., & Haberl,
629 H. (2015) Testing the effectiveness of environmental variables to explain European terrestrial vertebrate
630 species richness across biogeographical scales. *PLoS ONE*, **10**, 1–16.

631 Mucina, L. & Rutherford, M.C. (2006) *The vegetation of South Africa, Lesotho and Swaziland*. South
632 African National Biodiversity Institute,

633 NIMA (2000) Amendment 1. 3 January 2000. Department of Defense World Geodetic System 1984. Its
634 Definition and Relationships with Local Geodetic Systems. 1–3.

635 Oksanen, J., Blanchet, F.G., Friendly, M., Kindt, R., Legendre, P., McGlinn, D., Minchin, P.R., O’Hara, R.B.,
636 Simpson, G.L., Solymos, P., Stevens, M.H.H., Szoecs, E., & Wagner, H. (2018) *Vegan: Community ecology*
637 *package*.

638 Olson, D.M., Dinerstein, E., Wikramanayake, E.D., Burgess, N.D., Powell, G.V.N., Underwood, E.C.,
639 D’amico, J.A., Itoua, I., Strand, H.E., Morrison, J.C., & Others (2001) Terrestrial Ecoregions of the World: A
640 New Map of Life on Earth: A new global map of terrestrial ecoregions provides an innovative tool for
641 conserving biodiversity. *BioScience*, **51**, 933–938.

642 Power, S.C., Verboom, G.A., Bond, W.J., & Cramer, M.D. (2017) Environmental correlates of biome-level
643 floristic turnover in South Africa. *Journal of Biogeography*, **44**, 1745–1757.

644 R Core Team (2018) *R: A Language and Environment for Statistical Computing. Version 3.5.0*. R
645 Foundation for Statistical Computing, Vienna, Austria.

646 Rensburg, B.J. van, Chown, S.L., & Gaston, K.J. (2002) Species Richness, Environmental Correlates, and
647 Spatial Scale: A Test Using South African Birds. *The American Naturalist*, **159**, 566–577.

648 Ricklefs, R.E. (1987) Community diversity: relative roles of local and regional processes. *Science, New*
649 *Series*, **235**, 167–171.

650 Scott Chamberlain & Eduard Szocs (2013) Taxize - taxonomic search and retrieval in r. *FI000Research*,.

651 Thuiller, W., Midgley, G.F., Rouget, M., Cowling, R.M., F. Midgley, G., Rougeti, M., & M. Cowling, R. (2006)
652 Predicting patterns of plant species richness in megadiverse South Africa. *Ecography*, **29**, 733–744.

653 Verboom, G.A., Stock, W.D., & Cramer, M.D. (2017) Specialization to extremely low-nutrient soils limits the
654 nutritional adaptability of plant lineages. *The American Naturalist*, **In press**,.

655 Wardell-Johnson, G. & Horwitz, P. (1996) Conserving biodiversity and the recognition of heterogeneity in
656 ancient landscapes: a case study from south-western Australia. *Forest Ecology and Management*, **85**,
657 219–238.