

# Near Infra-Red Spectroscopy Predicts Crude Protein in Hemp Grain

Ryan Crawford<sup>1</sup>, Jamie Crawford<sup>1</sup>, Lawrence B. Smart<sup>2</sup>, Virginia Moore<sup>3</sup>

<sup>1</sup>Cornell University, Ithaca, NY,

<sup>2</sup>Cornell AgriTech, Geneva, NY,

<sup>3</sup>Cornell University, Ithaca, NY,

---

Corresponding author: Ryan Crawford, [rvc3@cornell.edu](mailto:rvc3@cornell.edu)

**Abstract**

Lorem ipsum dolor sit amet, consectetur adipiscing elit, sed do eiusmod tempor incididunt ut labore et dolore magna aliqua. Ut enim ad minim veniam, quis nostrud exercitation ullamco laboris nisi ut aliquip ex ea commodo consequat. Duis aute irure dolor in reprehenderit in voluptate velit esse cillum dolore eu fugiat nulla pariatur. Excepteur sint occaecat cupidatat non proident, sunt in culpa qui officia deserunt mollit anim id est laborum.

**Plain Language Summary**

Earthquake data for the island of La Palma from the September 2021 eruption is found ...

**incomplete: may contain errors, run-ons, half-thoughts, etc.**

**0.1 INTRODUCTION**

Hemp (*Cannabis sativa* L.) is an annual crop with potential uses as a source of food or feed from grain, and bast fiber or hurd from the stalk. Hemp cultivars are commonly grown for one or both purposes and a cultivar may be referred to as a grain, fiber, or dual-purpose type. Because of protein's nutritional importance, the protein content of a grain crop is an prime consideration for researchers, producers, and consumers. Whole hemp grain typically contains approximately 20-30% protein (Bárta et al., 2024; Ely & Fike, 2022; **callaway2004?**). Crude protein (CP) is often used as a proxy for the direct measurement of protein concentration and consists of the multiplication of nitrogen concentration by a conversion factor because measuring nitrogen concentration is relatively easy and cheap via laboratory assay (Hayes, 2020).

Near-infrared spectroscopy (NIRS) technology is rapid, non-destructive, and cheap, and consists of the measurement of NIR radiation reflected from a sample (Roberts et al., 2004). NIR spectra from many samples are related to laboratory values for components such as moisture, protein, fat, or fiber (Roberts et al., 2004). NIRS technology has been used since the 1970's to assess forage CP (Reeves, 2012; Williams, 1975). A NIRS calibration set often consists of samples from one species grown in many environments encompassing the range of expected values from the analyte or analytes (Chadalavada et al., 2022). Partial least squares regression (PLSR) is a typical method used in the agricultural and food sciences to relate spectra to analyte (Roberts et al., 2004). PLSR calculates principal components (PCs) which relate to the dependent variable and summarize the spectra and uses a subset of PCs in order to fit the regression model. PLSR is commonly used in spectroscopy because it tends to work well with highly-correlated spectral data. Typically the number of principal components is chosen via cross-validation to avoid overfitting. **CITES FOR ALL OF THIS**

A NIRS-scanned sample of undamaged grain may used for other purposes or it may planted as a seed. In wheat and corn, grain protein content has been shown to be heritable (Geyer et al., 2022; Giancaspro et al., 2019). This suggests (at least potentially) that NIRS technology could serve as resource to more rapidly identify high CP hemp germplasm, enabling the delivery of higher CP hemp grain cultivars faster.

For this study, a benchtop NIR spectrometer was used to develop a model to predict CP content based on a data set of hemp grain representing multiple years, locations, and cultivars from grain and dual-purpose hemp types using PLSR.

**0.2 MATERIALS AND METHODS**

Source: [Article Notebook](#)

Source: [Article Notebook](#)

### 0.2.1 Hemp Grain Sample Background

Spectral data were obtained from whole (unground) hemp grain samples, harvested at maturity, collected from 2017 - 2021 from 18 cultivar trials in New York (NY) (NA samples). Grain samples were obtained by hand sampling or mechanical harvest and were cleaned of chaff and dried at 30 C for six days in a forced-air dryer. In total, 38 cultivars were represented in the data set. Cultivars were grain or dual-purpose types and included both commercially available and experimental material.

All cultivar trials were planted in randomized complete block design with each cultivar replicated four times. The 2017 data were comprised of samples from the same thirteen cultivars sampled from six NY locations. For those trials, grain was harvested from each plot individually and aggregated by cultivar within each trial. Four subsamples were drawn from each aggregated sample and scanned separately. These spectra were averaged at each 2 nm increment. All remaining samples from 2018-2021 were collected on a per-plot basis. All possible cultivars and possible locations were represented in 2017, but only a selected subset of cultivars and locations were represented in 2018-2021.

### 0.2.2 Spectral Data Collection and Preprocessing

A benchtop NIR spectrometer (FOSS/ NIR FOSS/ NIR Systems model 5000) was used to obtain the spectra (FOSS North America, Eden Prairie, MN, USA). Spectra were collected every 2 nm from 1100-2498 nm and the logarithm of reciprocal reflectance was recorded.

WINISI software version 1.02A (Infrasoft International, Port Matilda, PA, USA) was used to average the spectra in 2017, as well as to select samples for laboratory assay. Samples were selected according to their spectral distance from their nearest neighbor within the calibration data set with a cutoff of a distance of 0.6 H, where H is approximately equal to the squared Mahalanobis distance divided by the number of principal components used in the calculation (Garrido-Varo et al., 2019). Prior to selection selection, spectra were preprocessed using SNV-detrend with settings 1,4,4,1 for the derivative, gap, smooth, and smooth 2 settings respectively.

### 0.2.3 Laboratory Validation

Laboratory assays were performed by Dairy One Forage Laboratory (Ithaca, NY). For those assays, 1mm ground samples were analyzed by combustion using a CN628 or CN928 Carbon/Nitrogen Determinator. Samples from 2017 were aggregated as described above, but the remaining samples were not aggregated.

### 0.2.4 Model Development

Calibration and validation sets were created by dividing the laboratory CP values into tertiles according to their percent CP in order to ensure that the range of CP values was present in both calibration and validation sets. Within each tertile, 75% of the samples were randomly assigned to the calibration set and the remaining 25% were assigned to the validation set. For each calibration set, models were developed in caret using PLSR. In fitting the model, the number of principal components was optimized over a grid search from 1-20. Model performance was evaluated with 25 iterations of bootstrapping and minimized root mean squared error (RMSE) in selecting the number of principal components in the final model .

Source: [Article Notebook](#)

Initially a number of common spectral preprocessing methods were tested by creating 100 calibration and validation sets as described above. Spectral data from those data sets were transformed by each of the following methods: 1) first derivative, 2) Savitzky-Golay (SG), 3) gap-segment derivative, 4) standard normal variate (SNV), 4) standard normal variate following Savitzky-Golay (SNV-SG), 5) SNV-detrend, and 6) multiplicative scatter correction. For each of these preprocessing methods,

models were fit and predictions were made on the corresponding validation set (since there were 8 preprocessing methods, 8 separate models were fit for each of the 100 sets. The relationship between the predicted and actual values of the validation set were calculated via RMSE,  $R^2$  and Ratio of Performance to InterQuartile distance (RPIQ), three common model assessment metrics. Larger  $R^2$  and RPIQ, and smaller RMSE values are superior. Analyses of variance (ANOVA) were performed for each of these metrics in order to compare the preprocessing methods. For each ANOVA, each data set was considered as a subject and allowing different variances for each preprocessing method.

Once the most promising preprocessing method was identified, 1000 more data sets were created and analyzed via that method and performance on the validation sets was summarized with RMSE,  $R^2$ , and RPIQ.

### 0.2.5 Additional software used

We used R version 4.3.3 (R Core Team, 2024) and the following R packages: caret v. 6.0.94 (Kuhn & Max, 2008), data.table v. 1.15.2 (Barrett et al., 2024), emmeans v. 1.10.0 (Lenth, 2024), nlme v. 3.1.163 (J. Pinheiro et al., 2023; J. C. Pinheiro & Bates, 2000), pls v. 2.8.3 (Liland et al., 2023), prospectr v. 0.2.7 (Stevens & Ramirez-Lopez, 2024), skimr v. 2.1.5 (Waring et al., 2022), tidymodels v. 1.1.1 (Kuhn & Wickham, 2020), tidyverse v. 2.0.0 (Wickham et al., 2019).

Source: [Article Notebook](#)

## 0.3 RESULTS AND DISCUSSION

### 0.3.1 Laboratory assay CP values

Laboratory assay percent CP values are summarized in the following table. These are similar to the range of CP values observed in the literature, indicating an reasonable basis for a chemometric model. The CP values are left-skewed and two thirds of the samples contained more than 25% CP.

Table 1: Summary of Laboratory Assayed CP Values (Percent Dry Matter)

Mean	Sd	Minimum	First Quartile	Median	Third Quartile	Maximum
26.1	2.5	20.8	23.9	26.4	28.2	30.8

Source: [Article Notebook](#)

### 0.3.2 Preprocessing methods comparison

Source: [Article Notebook](#)

Source: [Article Notebook](#)

Source: [Article Notebook](#)

Source: [Article Notebook](#)

Source: [Article Notebook](#)

Source: [Article Notebook](#)

All preprocessing methods outperformed raw spectral data Table 2. Averaged together, all preprocessed spectra were superior to raw spectra, with lower RMSE, and higher  $R^2$  and RPIQ values (significant at  $\alpha$  level  $<0.001$ ). Preprocessing methods had -11.6 % lower RMSE, and had 3.1% higher  $R^2$  7.4% higher RPIQ than unprocessed spectra.

The SNV-SG method had the lowest RMSE, highest  $R^2$ , and highest RPIQ averaging over all iterations. SNV-SG RMSE averaged 1.4% lower, while  $R^2$  and RPIQ av-

eraged 0.4% and 2.4% higher respectively than the next best preprocessing method (SG in both cases), but the difference between the best and second best method by metric were only statistically significant at  $\alpha < 0.05$  for RPIQ. RPIQ was devised to accurately reflect the spread of data in skewed populations (**bellon-maurel2010?**) and thus offers a robust metric for model assessment in this context, where the CP data are skewed. Therefore the superiority of SNV-SG as measured via RPIQ made it the best choice for the final model.

Table 2: Evaluation of Preprocessing Methods by Metric  $\pm$  Standard Error

Preprocessing Method	RMSE	$R^2$	RPIQ
Standard Normal Variate following	1.02 $\pm$	0.84 $\pm$	3.97 $\pm$
Savitzky-Golay	0.012	0.004	0.076
Savitzky-Golay	1.03 $\pm$	0.83 $\pm$	3.88 $\pm$
	0.012	0.004	0.072
First Derivative	1.07 $\pm$	0.82 $\pm$	3.77 $\pm$
	0.013	0.004	0.075
Standard Normal Variate	1.12 $\pm$	0.80 $\pm$	3.61 $\pm$
	0.016	0.005	0.081
Gap-segment Derivative	1.12 $\pm$	0.81 $\pm$	3.60 $\pm$
	0.018	0.006	0.086
Standard Normal Variate-Detrend	1.13 $\pm$	0.80 $\pm$	3.55 $\pm$
	0.015	0.005	0.079
Multiplicative Scatter Correction	1.17 $\pm$	0.79 $\pm$	3.47 $\pm$
	0.016	0.006	0.080
Raw Spectra	1.22 $\pm$	0.79 $\pm$	3.42 $\pm$
	0.044	0.009	0.105

Source: [Article Notebook](#)

The preprocessing methods examined represent a portion of those available. As well, preprocessing methods tend to have a number of user-adjustable parameters whose various permutations were not tested. This subset of preprocessing methods and parameters nonetheless contained substantial variations in model quality, demonstrating the importance of the selection of an appropriate preprocessing method.

### 0.3.3 Final model development and summary

Source: [Article Notebook](#)

The model improved most rapidly as the number of principal components increased from 1 to 7, with the inclusion of each additional PC being associated with a decrease in RMSE of 5-12% . From 8 to 12 PCs, model performance continued to improve, although gains were more modest (decrease in RMSE of 0.7-3%). With 13 or more PCs, performance gains were minimal and the relative ranks of the models tended to be stable Figure 1.

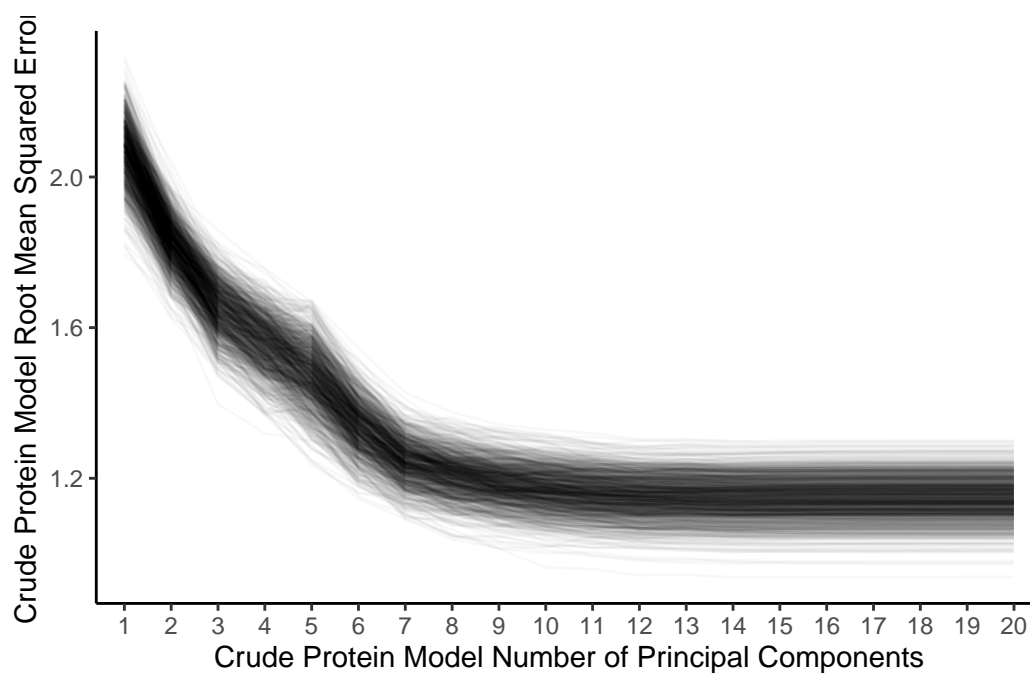


Figure 1: Decreasing RMSE with increasing number of PCs

169 Source: [Article Notebook](#)

170 Source: [Article Notebook](#)

171 Final model performance was similar, but not identical to, that obtained during  
 172 the initial comparison of preprocessing methods. The final models' mean RMSE  
 173 was 1.03,  $R^2$  was 0.83, and RPIQ was 3.89 (all calculated on the test sets). Despite  
 174 the generally good model performance, a subset of poor models can be seen. For  
 175 example, Figure 2 shows twenty-one models with  $R^2$  below 0.7. **more comment on**  
 176 **poor models?**

177 Source: [Article Notebook](#)

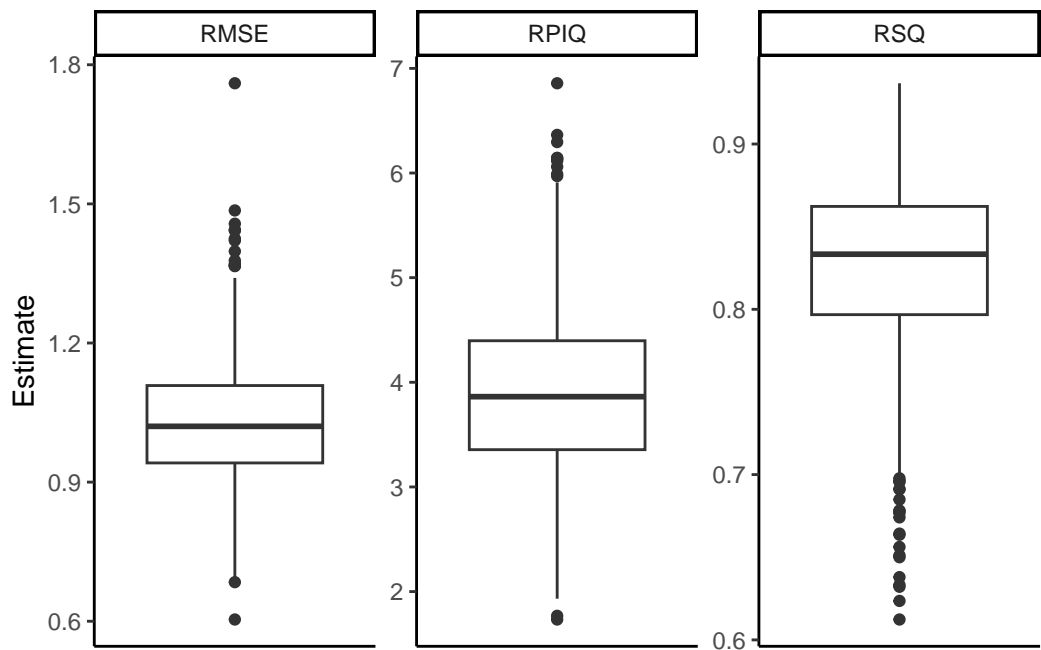


Figure 2: Final model validation set performance (1000 iterations)

178 Source: [Article Notebook](#)

179 Source: [Article Notebook](#)

180 Source: [Article Notebook](#)

```
181
182 Call:
183 lm(formula = difference ~ adj_cp, data = temp_dat)
184
185 Residuals:
186      Min       1Q   Median       3Q      Max
187 -2.51794 -0.58132  0.06936  0.50754  2.74745
188
189 Coefficients:
190             Estimate Std. Error t value Pr(>|t|)
191 (Intercept)  0.80412    0.17827   4.511 1.31e-05 ***
192 adj_cp      -0.15334    0.03051  -5.026 1.44e-06 ***
193 ---
194 Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
195
196 Residual standard error: 0.9138 on 147 degrees of freedom
197 Multiple R-squared:  0.1466,    Adjusted R-squared:  0.1408
198 F-statistic: 25.26 on 1 and 147 DF,  p-value: 1.438e-06
```

199 Source: [Article Notebook](#)

200 Finally, the pattern of errors was examined on a per-sample basis. Figure 3

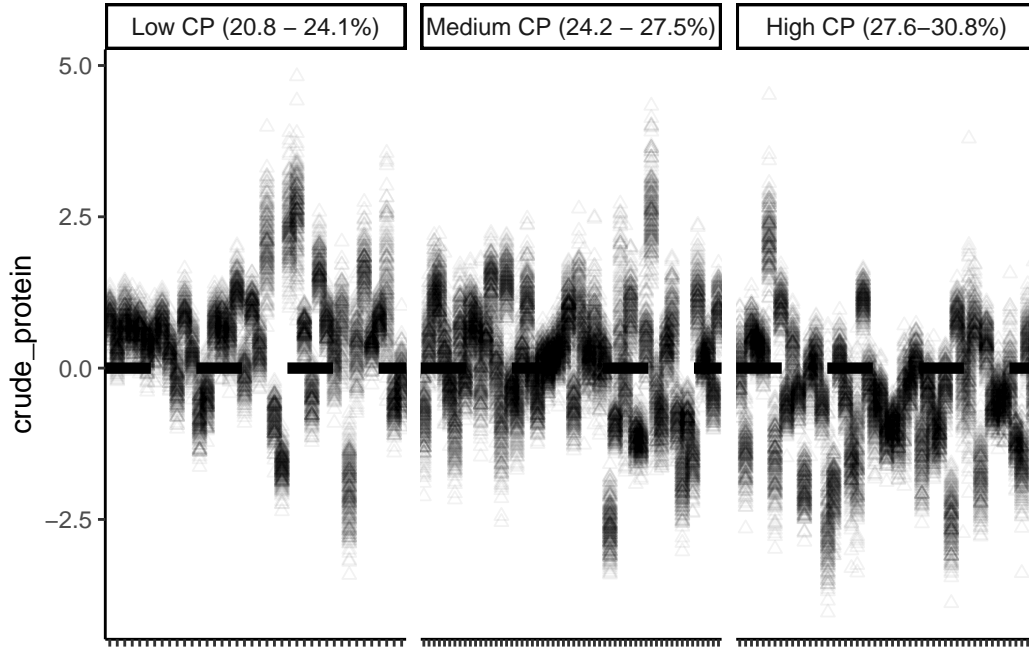


Figure 3: Test set prediction errors on a per-sample basis. Actual sample value set to 0, and samples ranked from least to greatest.

Source: [Article Notebook](#)

Errors tend to be lower at higher levels of actual CP

This study is limited in that it represents the creation of one model based upon spectra collected from one machine. NIRS calibrations can be unique to a particular machine, even if the machines compared are of the same model (**Reeves\_2012?**) . As well, the calibration and validation sets are relatively small.

This research showed the promise of the use of NIRS in order to make predictions concerning %CP in hemp grain using PLS. Promising preprocessing methods were identified and a model was validated. Further research could refine the model by including more samples or by examining other predictive methods.

#### 0.4 ACKNOWLEDGMENTS

#### 0.5 SUPPLEMENTAL MATERIAL

Source: [Article Notebook](#)

Table 3: Tally of hemp cultivars and locations. Private cultivars are labeled “cultivar1”, “cultivar2”, etc.

cultivar2	chazy	freeville	geneva	ithaca	willsboro	Total
altair				1		1
anka		1	3	5	2	11
bialobrzeskie		1	3	4	1	9
canda		1	1	1		3
cfx-1		1	2	5		8
cfx-2		1	2	4		7
crs-1	1	1	2	5		9



Table 3: Tally of hemp cultivars and locations. Private cultivars are labeled “cultivar1”, “cultivar2”, etc.

cultivar2	chazy	freeville	geneva	ithaca	willsboro	Total
cultivar1		1				1
cultivar2				1		1
cultivar3				1		1
cultivar4				1		1
earlina 8			1			1
experimental1				1		1
experimental2				1		1
felina 32		1	2	3		6
futura 75		1	3	4		8
grandi		3	3	4		10
h-51			1	2		3
han-fn-h				1		1
han-nw				1		1
helena		1				1
henola				2		2
hlesia				3		3
hliana			1	1		2
joey		1	1	1		3
katani		2	3	4		9
nebraska (feral)	1			1		2
pewter river		1				1
pico		1	2	5		8
portugal			1			1
rocky hemp			1			1
sterling gold			1			1
swift	1	1		1		3
tygra		1	3	4		8
uso-31	2	1	2	4		9
wojko		1	3	4		8
x-59		2		1		3
Total	5	24	41	76	3	149

214 Source: [Article Notebook](#)

215 **0.6 OPTIONAL SECTIONS**

216 **0.7 REFERENCES**

217 **0.8 FIGURES AND TABLES**

218 Source: [Article Notebook](#)

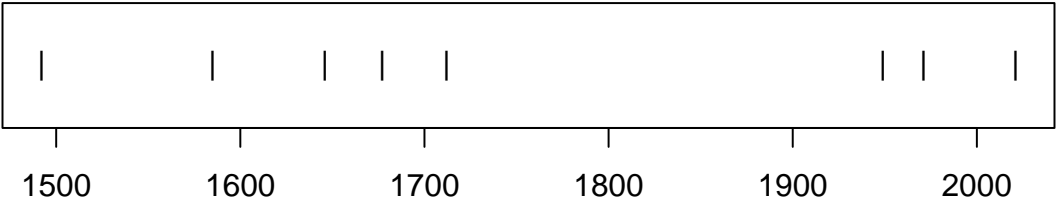


Figure 4: Timeline of recent earthquakes on La Palma

219 Source: [Article Notebook](#)

220 Source: [Article Notebook](#)

221 Based on data up to and including 1971, eruptions on La Palma happen every 79.8  
222 years on average.

223 Studies of the magma systems feeding the volcano, such as ([marrero2019?](#)), have  
224 proposed that there are two main magma reservoirs feeding the Cumbre Vieja vol-  
225 cano; one in the mantle (30-40km depth) which charges and in turn feeds a shallower  
226 crustal reservoir (10-20km depth).

227 Eight eruptions have been recorded since the late 1400s (Figure [4](#)).

228 Data and methods are discussed in Section [0.9](#).

229 Let  $x$  denote the number of eruptions in a year. Then,  $x$  can be modeled by a Pois-  
230 son distribution

$$p(x) = \frac{e^{-\lambda} \lambda^x}{x!} \quad (1)$$

231 where  $\lambda$  is the rate of eruptions per year. Using Equation [1](#), the probability of an  
232 eruption in the next  $t$  years can be calculated.

Table 4: Recent historic eruptions on La Palma

Name	Year
Current	2021
Teneguía	1971
Nambroque	1949
El Charco	1712
Volcán San Antonio	1677
Volcán San Martin	1646
Tajuya near El Paso	1585
Montaña Quemada	1492

233 Table [4](#) summarises the eruptions recorded since the colonization of the islands by  
234 Europeans in the late 1400s.

235 La Palma is one of the west most islands in the Volcanic Archipelago of the Canary  
236 Islands (Figure [5](#)).

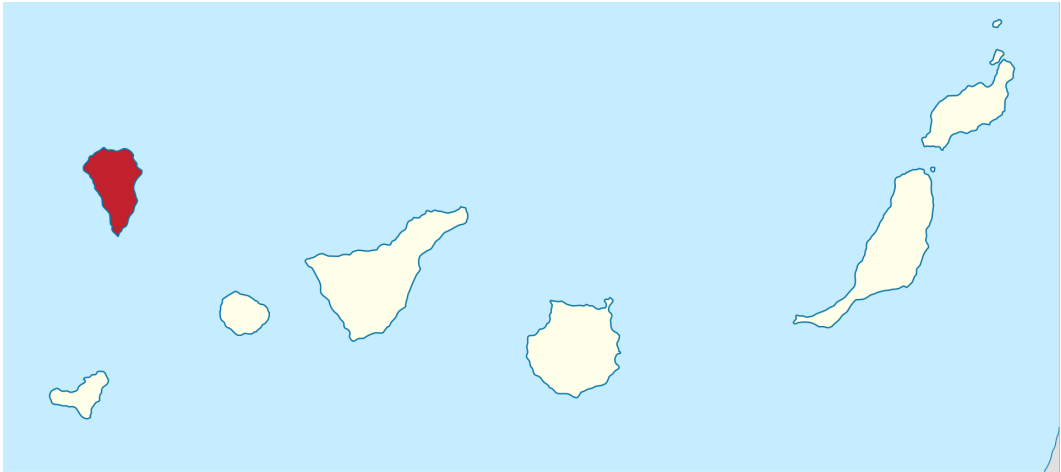


Figure 5: Map of La Palma

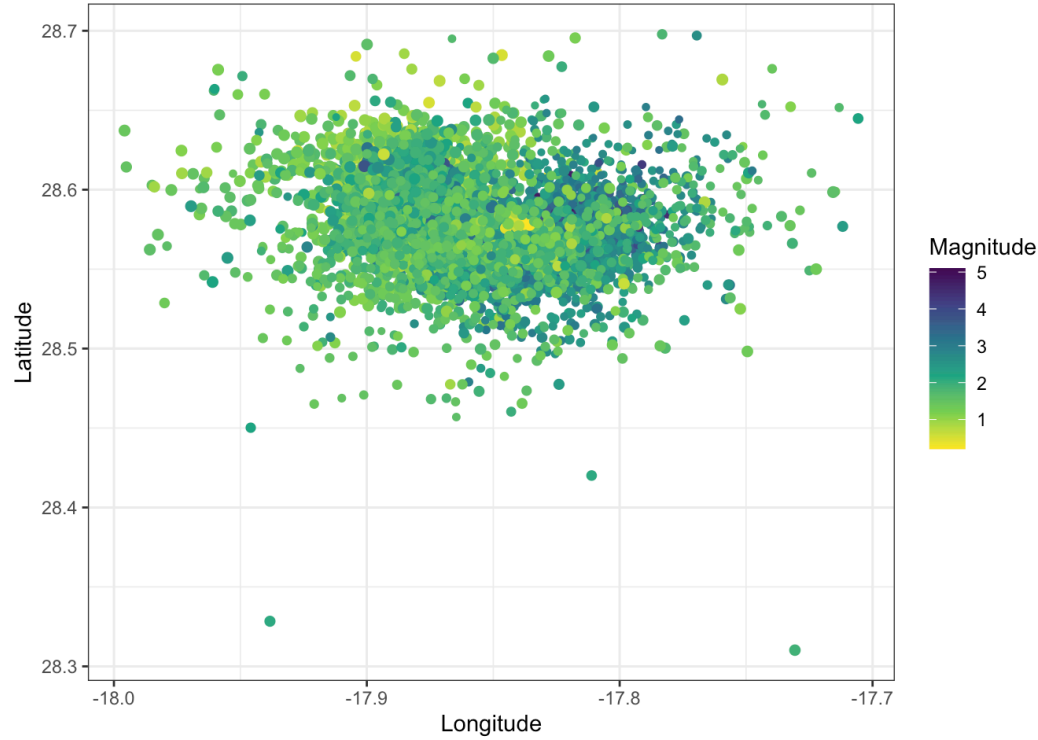


Figure 6: Locations of earthquakes on La Palma since 2017

237 Source: [Explore Earthquakes](#)

238 Figure 6 shows the location of recent Earthquakes on La Palma.

## 0.9 Data & Methods

## 0.10 Conclusion

## References

- Barrett, T., Dowle, M., Srinivasan, A., Gorecki, J., Chirico, M., & Hocking, T. (2024). *data.table: Extension of “data.frame”*. <https://CRAN.R-project.org/package=data.table>
- Bárta, J., Roudnický, P., Jarošová, M., Zdráhal, Z., Stupková, A., Bártová, V., Krejčová, Z., Kyselka, J., Filip, V., Říha, V., Lorenc, F., Bedrníček, J., & Smetana, P. (2024). Proteomic Profiles of Whole Seeds, Hulls, and Dehulled Seeds of Two Industrial Hemp (*Cannabis sativa* L.) Cultivars. *Plants*, 13(1), 111. <https://doi.org/10.3390/plants13010111>
- Chadalavada, K., Anbazhagan, K., Ndour, A., Choudhary, S., Palmer, W., Flynn, J. R., Mallayee, S., Pothu, S., Prasad, K. V. S. V., Varijakshapanikar, P., Jones, C. S., & Kholová, J. (2022). NIR Instruments and Prediction Methods for Rapid Access to Grain Protein Content in Multiple Cereals. *Sensors (Basel, Switzerland)*, 22(10). <https://doi.org/10.3390/s22103710>
- Ely, K., & Fike, J. (2022). Industrial Hemp and Hemp Byproducts as Sustainable Feedstuffs in Livestock Diets. In D. C. Agrawal, R. Kumar, & M. Dhanasekaran (Eds.), *Cannabis/Hemp for Sustainable Agriculture and Materials* (pp. 145–162). Springer. [https://doi.org/10.1007/978-981-16-8778-5\\_6](https://doi.org/10.1007/978-981-16-8778-5_6)
- Garrido-Varo, A., Garcia-Olmo, J., & Fearn, T. (2019). A note on Mahalanobis and related distance measures in WinISI and The Unscrambler. *Journal of Near Infrared Spectroscopy*, 27(4), 253–258. <https://doi.org/10.1177/0967033519848296>
- Geyer, M., Mohler, V., & Hartl, L. (2022). Genetics of the Inverse Relationship between Grain Yield and Grain Protein Content in Common Wheat. *Plants*, 11(16), 2146. <https://doi.org/10.3390/plants11162146>
- Giancaspro, A., Giove, S. L., Blanco, A., & Gadaleta, A. (2019). Genetic Variation for Protein Content and Yield-Related Traits in a Durum Population Derived From an Inter-Specific Cross Between Hexaploid and Tetraploid Wheat Cultivars. *Frontiers in Plant Science*, 10. <https://doi.org/10.3389/fpls.2019.01509>
- Hayes, M. (2020). Measuring Protein Content in Food: An Overview of Methods. *Foods*, 9(10), 1340. <https://doi.org/10.3390/foods9101340>
- Kuhn, M., & Wickham, H. (2020). *Tidymodels: A collection of packages for modeling and machine learning using tidyverse principles*. <https://www.tidymodels.org>
- Kuhn, & Max. (2008). Building predictive models in r using the caret package. *Journal of Statistical Software*, 28(5), 1–26. <https://doi.org/10.18637/jss.v028.i05>
- Lenth, R. V. (2024). *emmeans: Estimated marginal means, aka least-squares means*. <https://CRAN.R-project.org/package=emmeans>
- Liland, K. H., Mevik, B.-H., & Wehrens, R. (2023). *pls: Partial least squares and principal component regression*. <https://CRAN.R-project.org/package=pls>
- Pinheiro, J. C., & Bates, D. M. (2000). *Mixed-effects models in s and s-PLUS*. Springer. <https://doi.org/10.1007/b98882>
- Pinheiro, J., Bates, D., & R Core Team. (2023). *nlme: Linear and nonlinear mixed effects models*. <https://CRAN.R-project.org/package=nlme>
- R Core Team. (2024). *R: A language and environment for statistical computing*. R Foundation for Statistical Computing. <https://www.R-project.org/>
- Reeves, J. B. (2012). Potential of Near- and Mid-infrared Spectroscopy in Biofuel Production. *Communications in Soil Science and Plant Analysis*, 43(1-2), 478–495. <https://doi.org/10.1080/00103624.2012.641844>
- Roberts, C. A., Workman, J., & Reeves, J. B. (2004). *Near-infrared spectroscopy in agriculture*. American Society of Agronomy.
- Stevens, A., & Ramirez-Lopez, L. (2024). *An introduction to the prospectr package*.

- 294 Waring, E., Quinn, M., McNamara, A., Arino de la Rubia, E., Zhu, H., & Ellis,  
295 S. (2022). *skimr: Compact and flexible summaries of data*. [https://CRAN.R](https://CRAN.R-project.org/package=skimr)  
296 [-project.org/package=skimr](https://CRAN.R-project.org/package=skimr)
- 297 Wickham, H., Averick, M., Bryan, J., Chang, W., McGowan, L. D., François, R.,  
298 Grolemund, G., Hayes, A., Henry, L., Hester, J., Kuhn, M., Pedersen, T. L.,  
299 Miller, E., Bache, S. M., Müller, K., Ooms, J., Robinson, D., Seidel, D. P., Spinu,  
300 V., ... Yutani, H. (2019). Welcome to the tidyverse. *Journal of Open Source*  
301 *Software*, 4(43), 1686. <https://doi.org/10.21105/joss.01686>
- 302 Williams, P. C. (1975). Application of near infrared reflectance spectroscopy to anal-  
303 ysis of cereal grains and oilseeds. *Cereal Chemistry*, 52(4 p.561-576), 576–561.