

mini project

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2/14/2022

```
#Import file first
fna.data <- 'WisconsinCancer.csv'
wisc.df <- read.csv(fna.data, row.names=1)

#head(wisc.df)
```

Omit the first column (diagnosis) because it's unnecessary for analysis for now. However, we will store the vector for use as a factor later.

```
# We can use -1 here to remove the first column
wisc.data <- wisc.df[,-1]

#Store as a vector
diagnosis <- wisc.df$diagnosis
```

Question 1

There are 569 observations in this dataset.

```
#Number of observations = number of rows in the dataframe
nrow(wisc.df)
```

```
## [1] 569
```

Question 2

There are 212 malignant diagnoses.

```
#Find out how many patients had a malignant diagnosis using grep for 'M' within character vector diagnosis
#grep() returns the positions in the character vector; find the length of this list to get number
length(grep('M', diagnosis))
```

```
## [1] 212
```

Question 3

10 variables in the data are suffixed with '_mean'.

```
length(grep('_mean', colnames(wisc.df)))
```

```
## [1] 10
```

PCA

```
# Check column means and standard deviations
colMeans(wisc.data)
```

##	radius_mean	texture_mean	perimeter_mean
##	1.412729e+01	1.928965e+01	9.196903e+01
##	area_mean	smoothness_mean	compactness_mean
##	6.548891e+02	9.636028e-02	1.043410e-01
##	concavity_mean	concave.points_mean	symmetry_mean
##	8.879932e-02	4.891915e-02	1.811619e-01
##	fractal_dimension_mean	radius_se	texture_se
##	6.279761e-02	4.051721e-01	1.216853e+00
##	perimeter_se	area_se	smoothness_se
##	2.866059e+00	4.033708e+01	7.040979e-03
##	compactness_se	concavity_se	concave.points_se
##	2.547814e-02	3.189372e-02	1.179614e-02
##	symmetry_se	fractal_dimension_se	radius_worst
##	2.054230e-02	3.794904e-03	1.626919e+01
##	texture_worst	perimeter_worst	area_worst
##	2.567722e+01	1.072612e+02	8.805831e+02
##	smoothness_worst	compactness_worst	concavity_worst
##	1.323686e-01	2.542650e-01	2.721885e-01
##	concave.points_worst	symmetry_worst	fractal_dimension_worst
##	1.146062e-01	2.900756e-01	8.394582e-02

```
apply(wisc.data,2,sd)
```

##	radius_mean	texture_mean	perimeter_mean
##	3.524049e+00	4.301036e+00	2.429898e+01
##	area_mean	smoothness_mean	compactness_mean
##	3.519141e+02	1.406413e-02	5.281276e-02
##	concavity_mean	concave.points_mean	symmetry_mean
##	7.971981e-02	3.880284e-02	2.741428e-02
##	fractal_dimension_mean	radius_se	texture_se
##	7.060363e-03	2.773127e-01	5.516484e-01
##	perimeter_se	area_se	smoothness_se
##	2.021855e+00	4.549101e+01	3.002518e-03
##	compactness_se	concavity_se	concave.points_se
##	1.790818e-02	3.018606e-02	6.170285e-03
##	symmetry_se	fractal_dimension_se	radius_worst
##	8.266372e-03	2.646071e-03	4.833242e+00
##	texture_worst	perimeter_worst	area_worst
##	6.146258e+00	3.360254e+01	5.693570e+02
##	smoothness_worst	compactness_worst	concavity_worst

```
##          2.283243e-02          1.573365e-01          2.086243e-01
## concave.points_worst symmetry_worst fractal_dimension_worst
##          6.573234e-02          6.186747e-02          1.806127e-02
```

```
#Run PCA on data!
wisc.pr <- prcomp(wisc.data, scale=TRUE)

summary(wisc.pr)
```

```
## Importance of components:
##          PC1      PC2      PC3      PC4      PC5      PC6      PC7
## Standard deviation  3.6444 2.3857 1.67867 1.40735 1.28403 1.09880 0.82172
## Proportion of Variance 0.4427 0.1897 0.09393 0.06602 0.05496 0.04025 0.02251
## Cumulative Proportion 0.4427 0.6324 0.72636 0.79239 0.84734 0.88759 0.91010
##          PC8      PC9      PC10     PC11     PC12     PC13     PC14
## Standard deviation  0.69037 0.6457 0.59219 0.5421 0.51104 0.49128 0.39624
## Proportion of Variance 0.01589 0.0139 0.01169 0.0098 0.00871 0.00805 0.00523
## Cumulative Proportion 0.92598 0.9399 0.95157 0.9614 0.97007 0.97812 0.98335
##          PC15     PC16     PC17     PC18     PC19     PC20     PC21
## Standard deviation  0.30681 0.28260 0.24372 0.22939 0.22244 0.17652 0.1731
## Proportion of Variance 0.00314 0.00266 0.00198 0.00175 0.00165 0.00104 0.0010
## Cumulative Proportion 0.98649 0.98915 0.99113 0.99288 0.99453 0.99557 0.9966
##          PC22     PC23     PC24     PC25     PC26     PC27     PC28
## Standard deviation  0.16565 0.15602 0.1344 0.12442 0.09043 0.08307 0.03987
## Proportion of Variance 0.00091 0.00081 0.0006 0.00052 0.00027 0.00023 0.00005
## Cumulative Proportion 0.99749 0.99830 0.9989 0.99942 0.99969 0.99992 0.99997
##          PC29     PC30
## Standard deviation  0.02736 0.01153
## Proportion of Variance 0.00002 0.00000
## Cumulative Proportion 1.00000 1.00000
```

Question 4

44.27% of the original variance is captured by first principal component (PC1).

Question 5

3 PCs are required to describe at least 70% of the variance in the data.

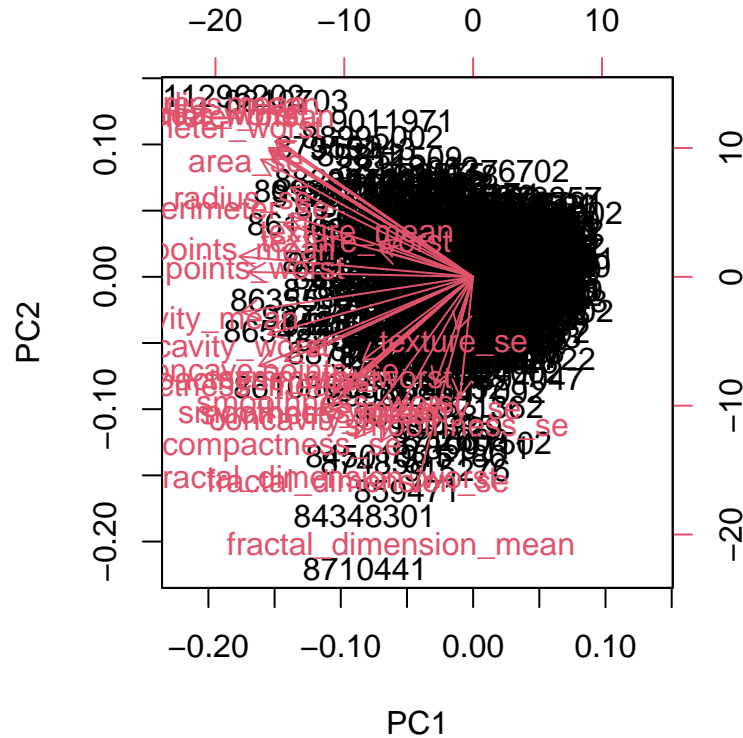
Question 6

7 PCs are required to describe at least 90% of the variance in the data.

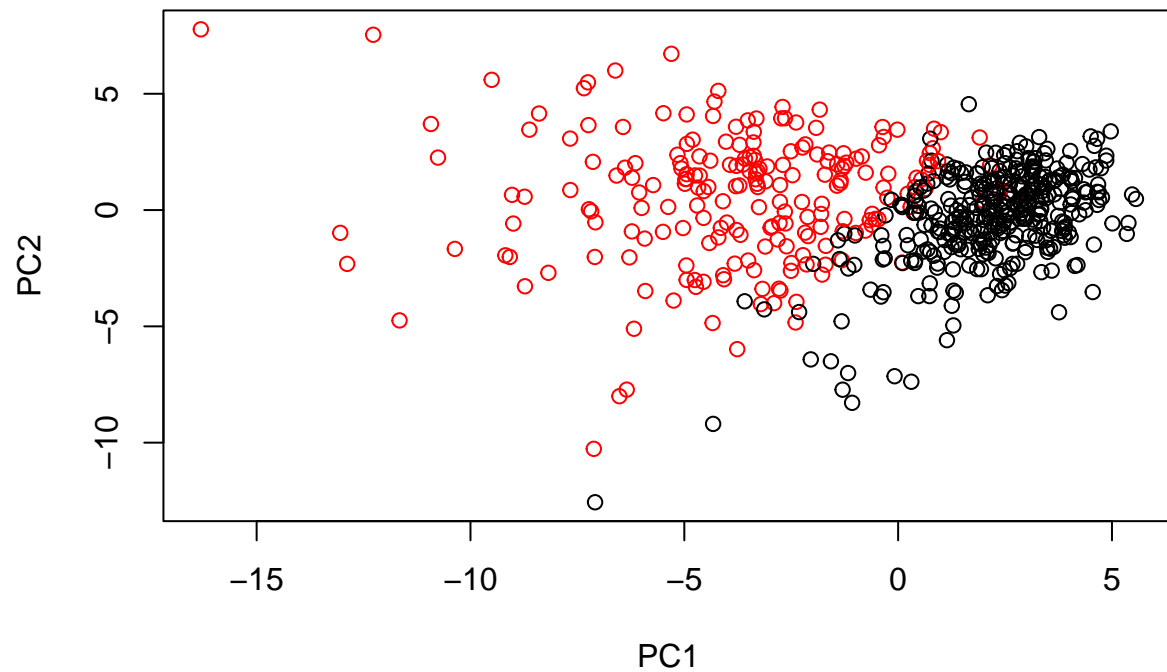
Question 7

This biplot is much too cluttered because here are too many variables to account for per observation and the dataset is also much too large for this visualization to be effective.

```
biplot(wisc.pr)
```

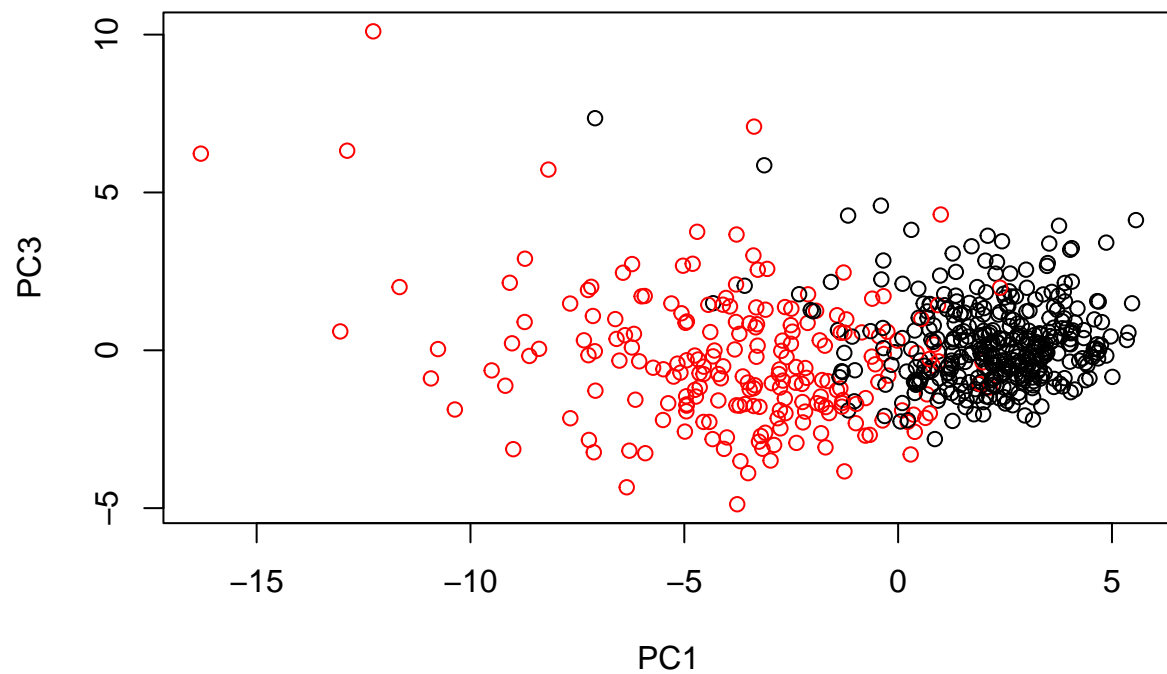


```
# Scatter plot observations by components 1 and 2
#Want each point colored based on diagnosis vector using ifelse()
#ifelse(conditional, if yes, if not)
plot(wisc.pr$x[,1], wisc.pr$x[,2], col = ifelse(diagnosis == 'M','red','black'),
      xlab = "PC1", ylab = "PC2")
```



Question 8 The plot of PCs 1 and 2 seems to generate 2 slightly more separate clusters than the plot of PCs 1 and 3. This may be because PC2 explains a greater proportion of variance than PC3.

```
# Repeat for components 1 and 3
plot(wisc.pr$x[,1], wisc.pr$x[,3], col = ifelse(diagnosis == 'M', 'red', 'black'),
      xlab = "PC1", ylab = "PC3")
```

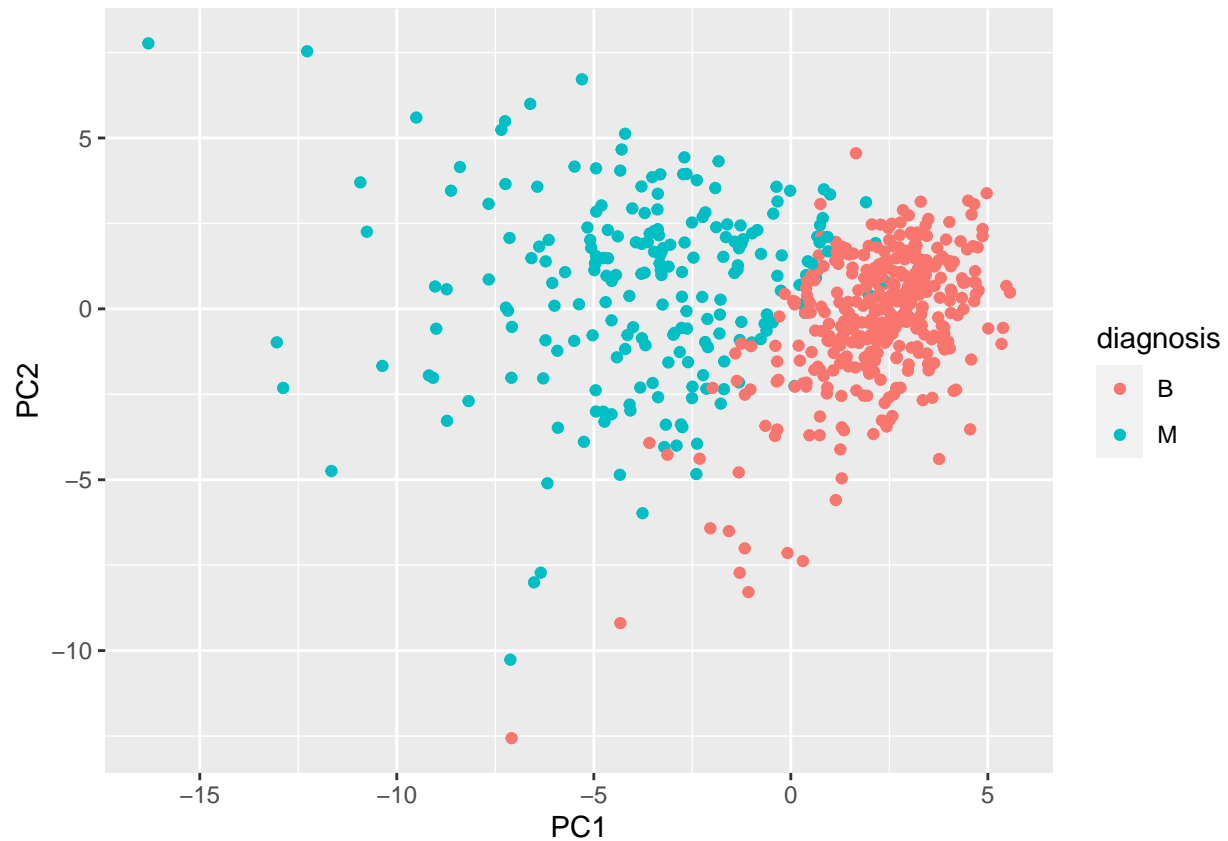


Now plot in ggplot2!

```
# Create a data.frame for ggplot
df <- as.data.frame(wisc.pr$x)
df$diagnosis <- diagnosis

# Load the ggplot2 package
library(ggplot2)

# Make a scatter plot colored by diagnosis
ggplot(df) +
  aes(PC1, PC2, col=diagnosis) +
  geom_point()
```

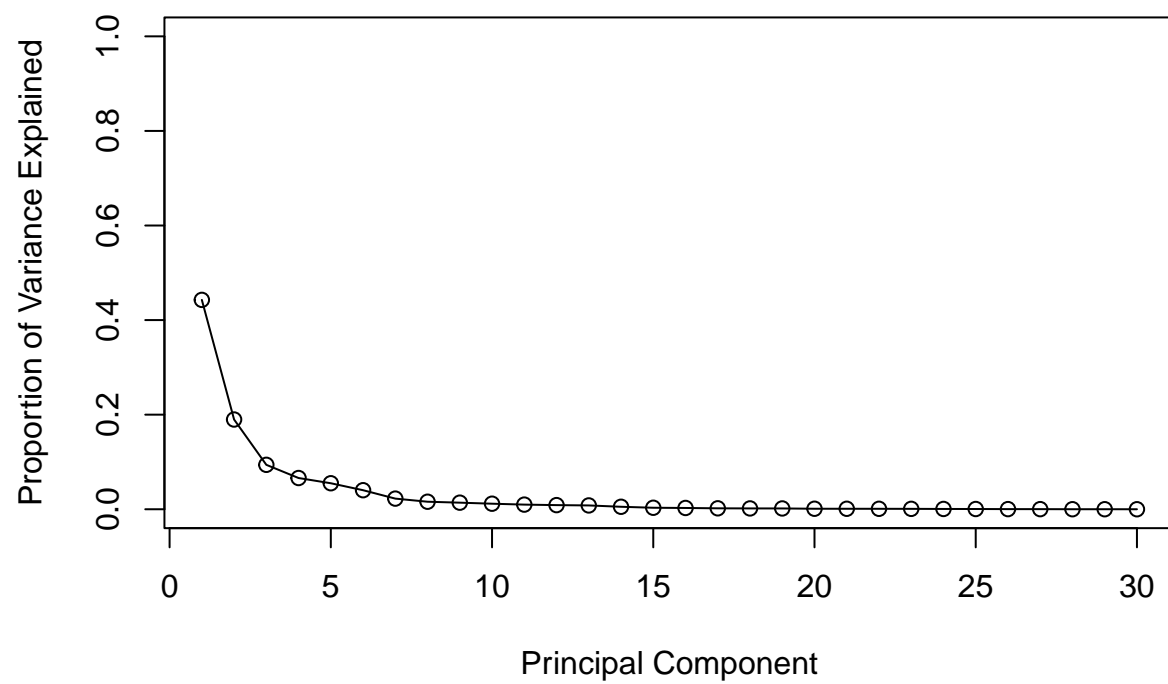


```
# Calculate variance of each component
pr.var <- wisc.pr$sdev^2
head(pr.var)
```

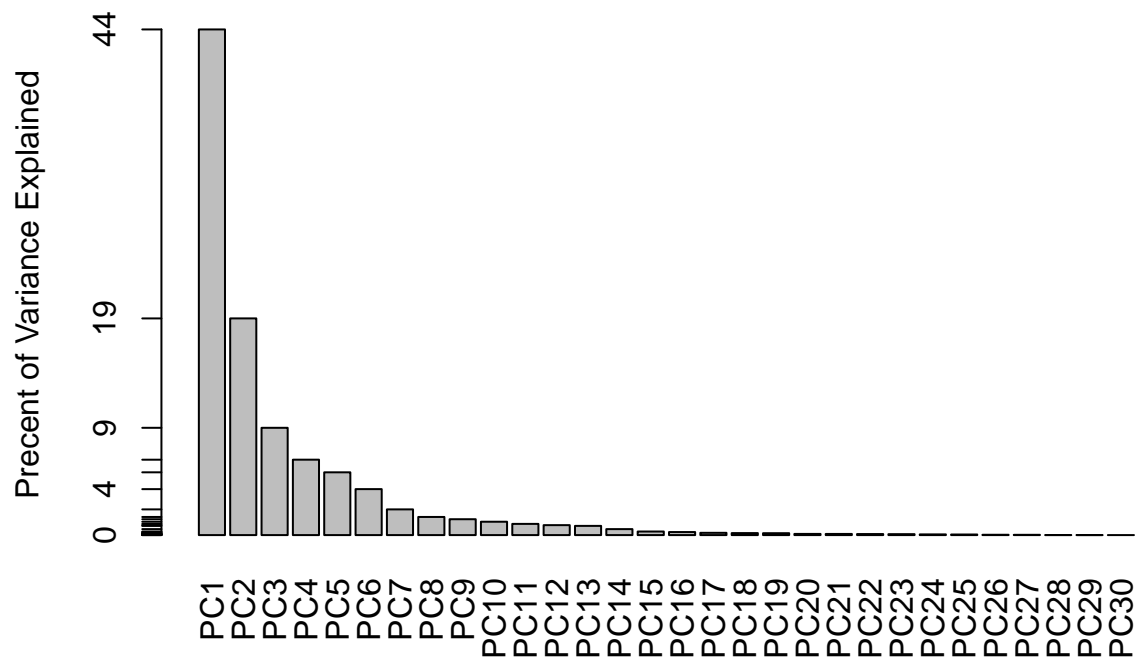
```
## [1] 13.281608  5.691355  2.817949  1.980640  1.648731  1.207357
```

```
# Variance explained by each principal component: pve
pve <- wisc.pr$sdev^2/sum(wisc.pr$sdev^2)

# Plot variance explained for each principal component
plot(pve, xlab = "Principal Component",
     ylab = "Proportion of Variance Explained",
     ylim = c(0, 1), type = "o")
```



```
# Alternative scree plot of the same data, note data driven y-axis
barplot(pve, ylab = "Precent of Variance Explained",
        names.arg=paste0("PC",1:length(pve)), las=2, axes = FALSE)
axis(2, at=pve, labels=round(pve,2)*100 )
```

Question 9

Loading scores show the influence of each variable on each principle component. The loading for `concave.points_mean` for the first principal component is -0.26085376.

```
wisc.pr$rotation[,1]
```

```
##          radius_mean          texture_mean          perimeter_mean
##          -0.21890244          -0.10372458          -0.22753729
##          area_mean          smoothness_mean          compactness_mean
##          -0.22099499          -0.14258969          -0.23928535
##          concavity_mean          concave.points_mean          symmetry_mean
##          -0.25840048          -0.26085376          -0.13816696
##          fractal_dimension_mean          radius_se          texture_se
##          -0.06436335          -0.20597878          -0.01742803
##          perimeter_se          area_se          smoothness_se
##          -0.21132592          -0.20286964          -0.01453145
##          compactness_se          concavity_se          concave.points_se
##          -0.17039345          -0.15358979          -0.18341740
##          symmetry_se          fractal_dimension_se          radius_worst
##          -0.04249842          -0.10256832          -0.22799663
##          texture_worst          perimeter_worst          area_worst
##          -0.10446933          -0.23663968          -0.22487053
##          smoothness_worst          compactness_worst          concavity_worst
```

```
##           -0.12795256           -0.21009588           -0.22876753
## concave.points_worst symmetry_worst fractal_dimension_worst
##           -0.25088597           -0.12290456           -0.13178394
```

Question 10

A minimum of 5 PCs are required to explain 80% of the variance in the data.

Hierarchical Clustering

```
# Scale the wisc.data data using the "scale()" function
data.scaled <- scale(wisc.data)

#Calculate Euclidean distances
data.dist <- dist(data.scaled)

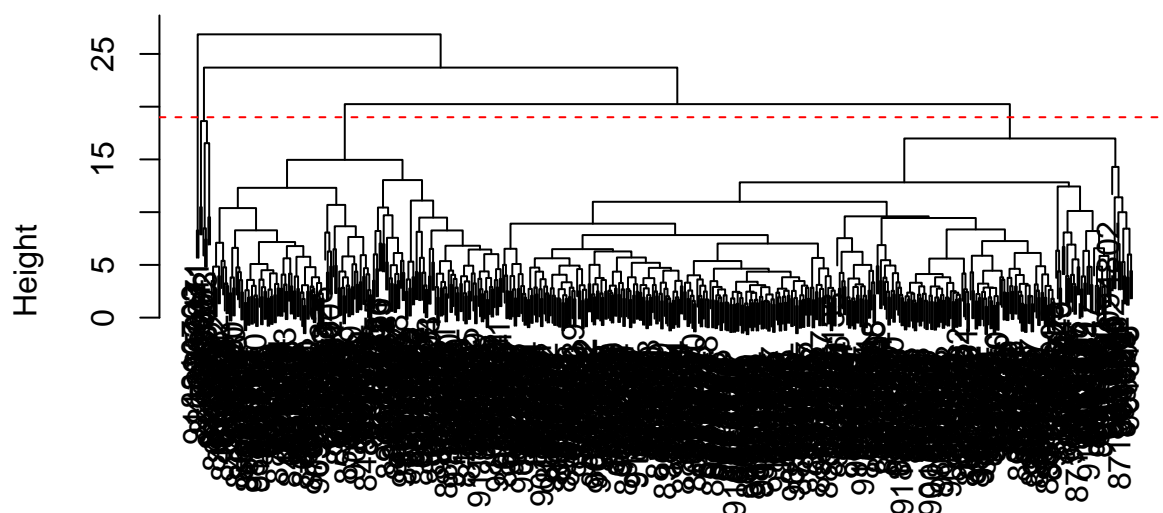
#Perform clustering using complete linkage
wisc.hclust <- hclust(data.dist, method='complete')
```

Question 11

The height at which the clustering model has 4 clusters is h=19.

```
plot(wisc.hclust)
abline(wisc.hclust, h=19, col="red", lty=2)
```

Cluster Dendrogram



```
data.dist
hclust (*, "complete")
```

Cut the tree so there are only 4 clusters (change argument k)

```
wisc.hclust.clusters <- cutree(wisc.hclust, k=4)
```

```
#Compare cluster membership to diagnoses
table(wisc.hclust.clusters, diagnosis)
```

```
##              diagnosis
## wisc.hclust.clusters  B  M
##              1  12 165
##              2   2   5
##              3 343  40
##              4   0   2
```

Question 12

The cluster vs. diagnoses match is only slightly better when the tree is cut into higher numbers of clusters, with certain clusters being able to completely match to a benign diagnosis (there isn't much of a change in the matching to malignant diagnoses).

```
wisc.hclust.clusters3 <- cutree(wisc.hclust, k=2)
table(wisc.hclust.clusters3, diagnosis)
```

```
##              diagnosis
## wisc.hclust.clusters3  B  M
```

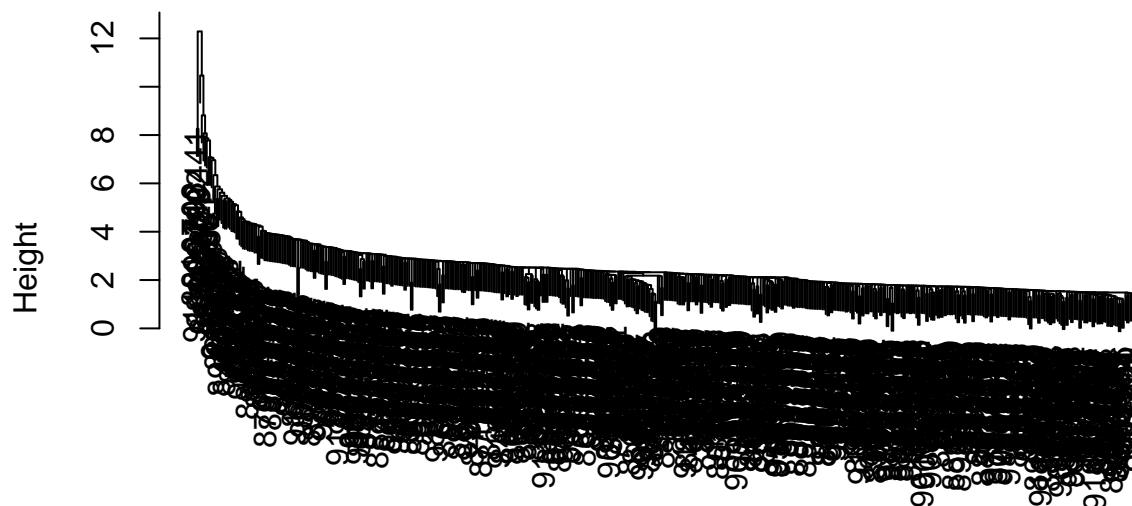
```
##          1 357 210
##          2   0   2
```

Question 13

Ward's method gives my favorite results for this dataset because for the number of clusters that I chose (k=4), it provided greater separation between clusters than any of the other methods. Each cluster more clearly mapped to a particular diagnosis more clearly than with the other methods.

```
#Perform clustering using single linkage
wisc.hclust_single <- hclust(data.dist, method='single')
plot(wisc.hclust_single)
```

Cluster Dendrogram



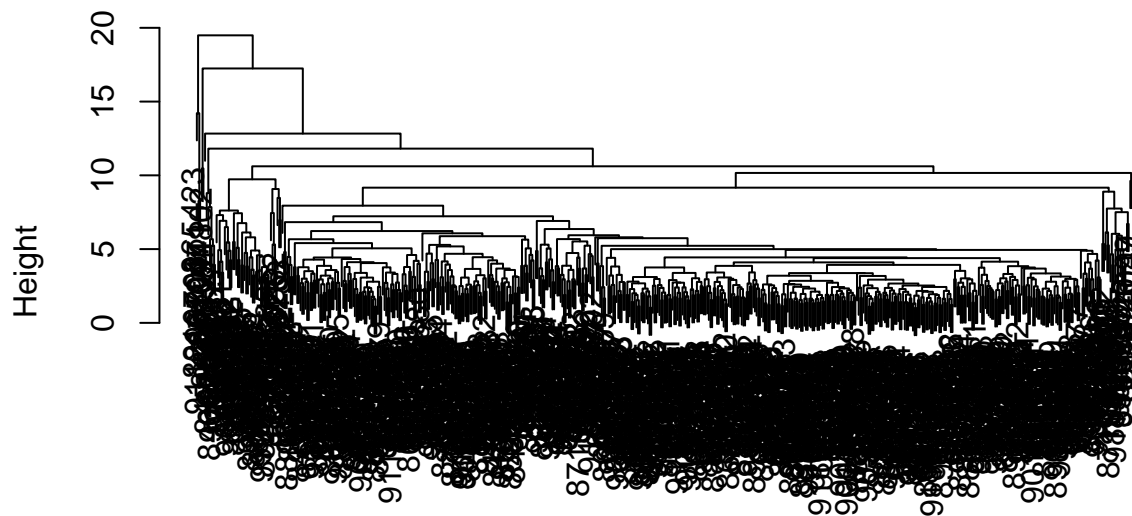
```
data.dist
hclust (*, "single")
```

```
wisc.hclust.clusters_s <- cutree(wisc.hclust_single, k=4)
table(wisc.hclust.clusters_s, diagnosis)
```

```
##          diagnosis
## wisc.hclust.clusters_s  B  M
##          1 356 209
##          2   1   0
##          3   0   2
##          4   0   1
```

```
#Perform clustering using average linkage
wisc.hclust_av <- hclust(data.dist, method='average')
plot(wisc.hclust_av)
```

Cluster Dendrogram



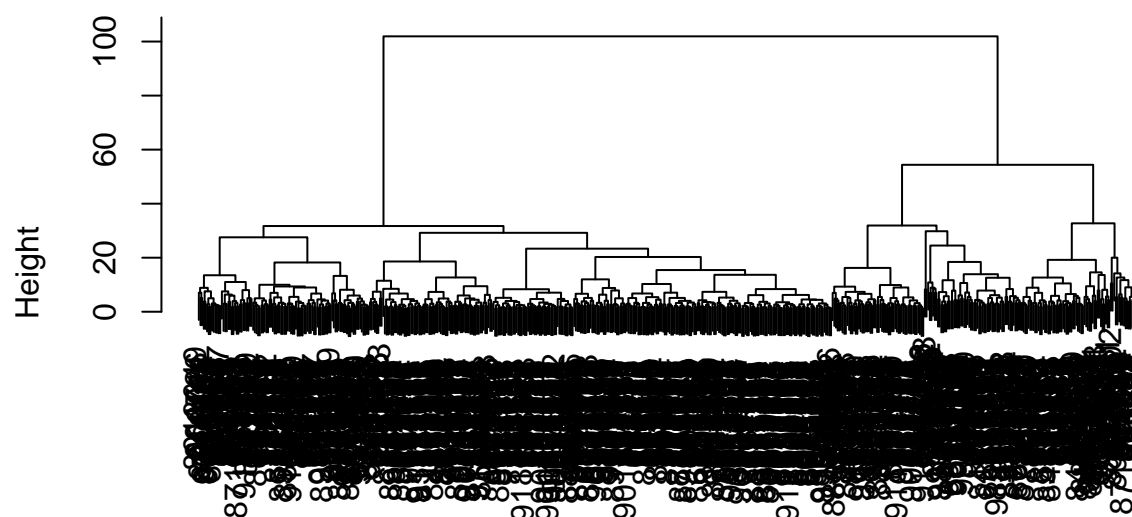
data.dist
hclust (*, "average")

```
wisc.hclust.clusters_av <- cutree(wisc.hclust_av, k=4)
table(wisc.hclust.clusters_av, diagnosis)
```

```
##              diagnosis
## wisc.hclust.clusters_av  B  M
##              1 355 209
##              2   2   0
##              3   0   1
##              4   0   2
```

```
#Perform clustering using Ward's method
wisc.hclust_ward <- hclust(data.dist, method='ward.D2')
plot(wisc.hclust_ward)
```

Cluster Dendrogram



```
data.dist
hclust (*, "ward.D2")
```

```
wisc.hclust.clusters_w <- cutree(wisc.hclust_ward, k=4)
table(wisc.hclust.clusters_w, diagnosis)
```

```
##           diagnosis
## wisc.hclust.clusters_w  B  M
##           1    0 115
##           2    6  48
##           3   337  48
##           4    14   1
```

K-Means Clustering

Question 14

K-means clustering separates out diagnosis clusters more robustly than does the hierarchical clustering methods we used above for the same # of clusters k ($k=2$). In the k-means clustering, cluster 2 is very clearly the malignant diagnosis cluster and cluster 1 much more clearly skews towards diagnoses.

```
wisc.km <- kmeans(wisc.data, centers= 2, nstart= 20)
table(wisc.km$cluster, diagnosis)
```

```
##      diagnosis
##      B      M
```

```
## 1 356 82
## 2 1 130
```

```
#Compare to hierarchical clustering
table(wisc.hclust.clusters3, diagnosis)
```

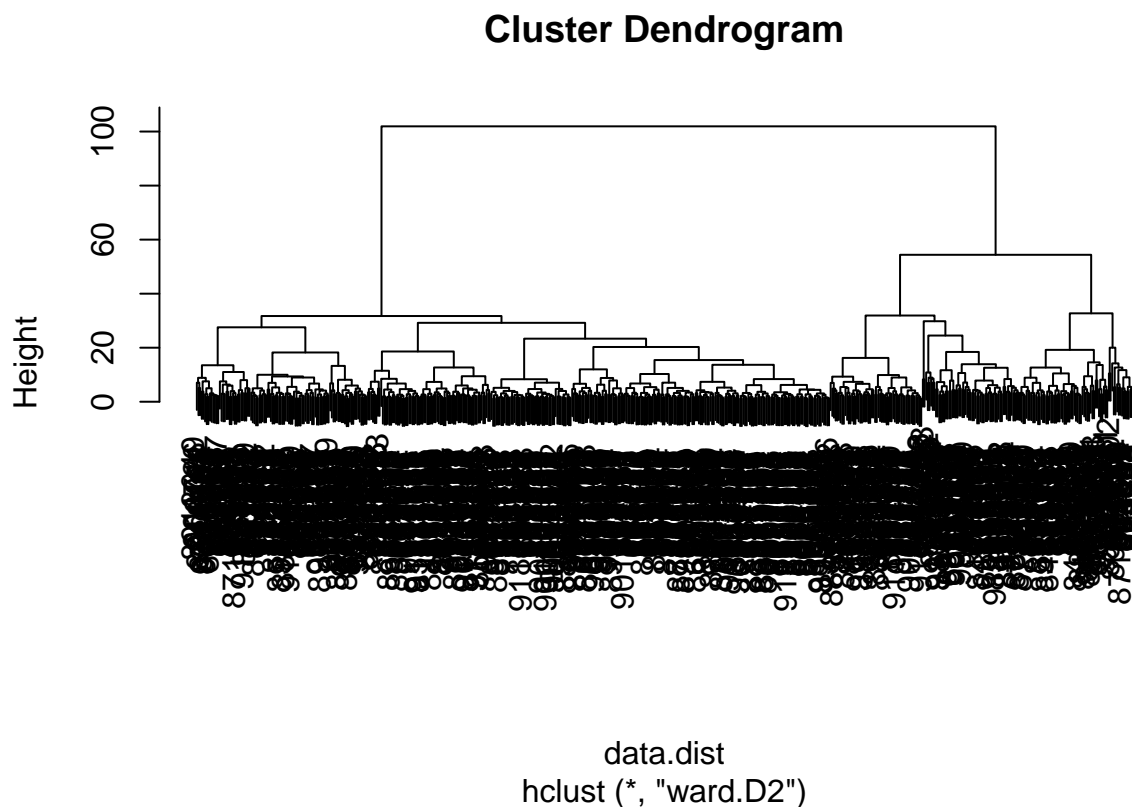
```
##              diagnosis
## wisc.hclust.clusters3  B  M
##                   1 357 210
##                   2  0  2
```

Combining Methods

```
wisc.pr.hclust <- hclust(data.dist, method='ward.D2')
grps <- cutree(wisc.pr.hclust, k=2)
table(grps)
```

```
## grps
## 1 2
## 184 385
```

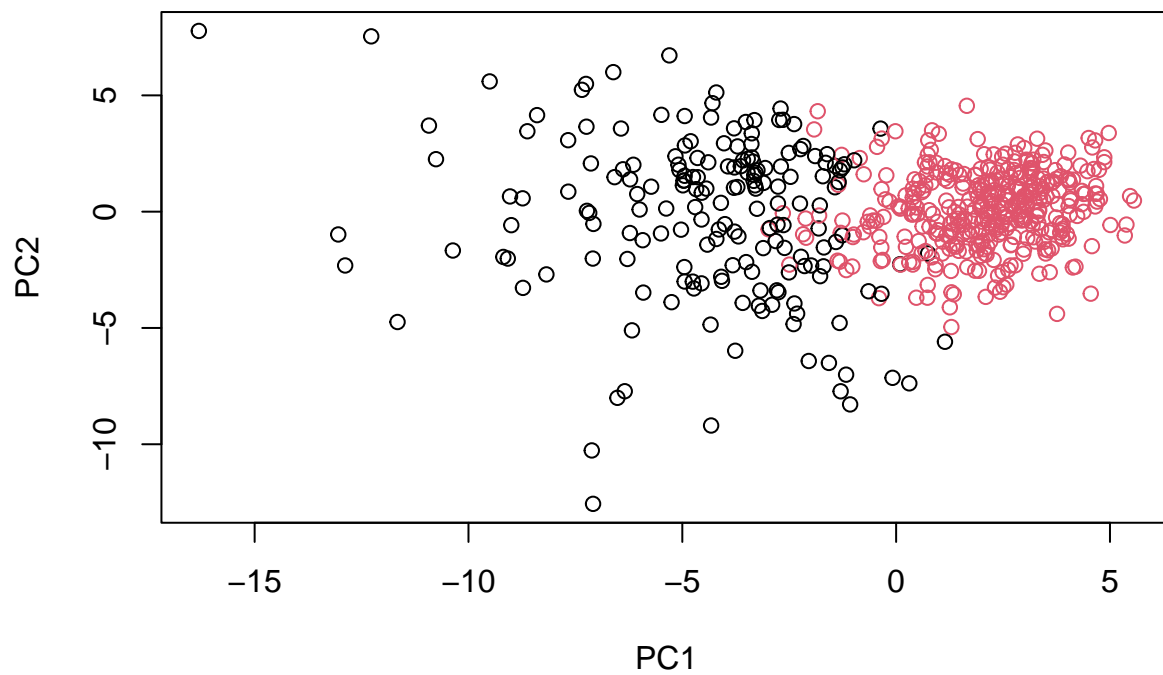
```
plot(wisc.pr.hclust)
```



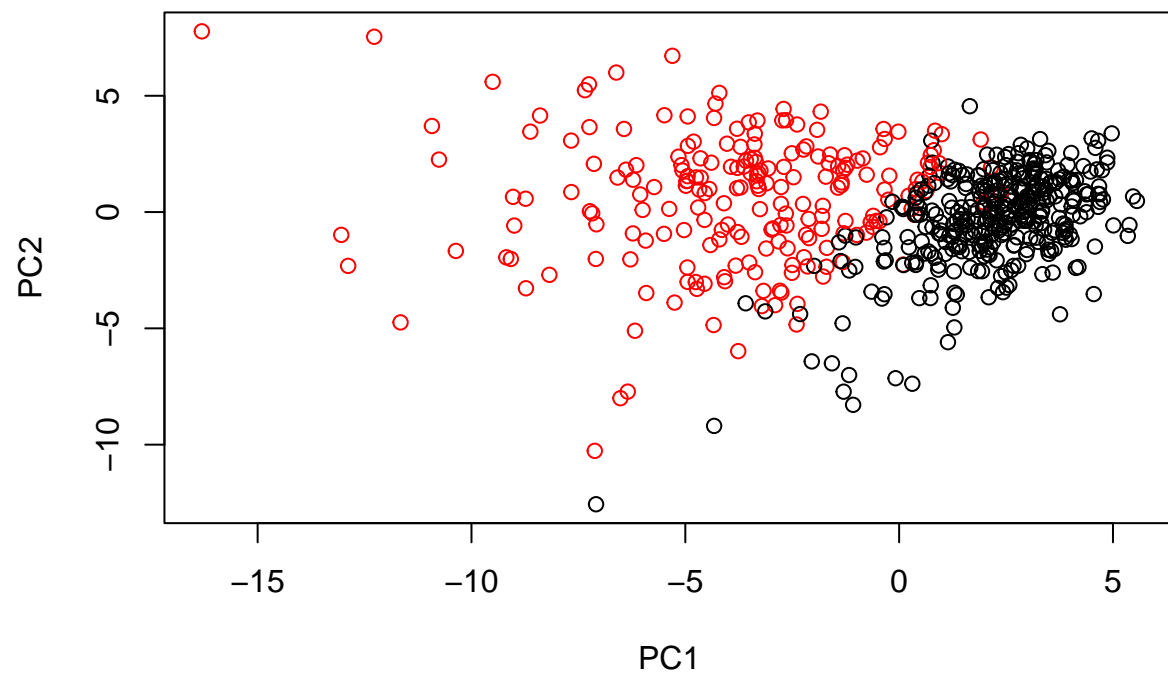
```
table(grps, diagnosis)
```

```
##      diagnosis  
## grps   B    M  
##    1  20 164  
##    2 337  48
```

```
plot(wisc.pr$x[,1:2], col=grps)
```



```
#Now the colors are based on diagnosis rather than the cluster they belong to  
plot(wisc.pr$x[,1:2], col=ifelse(diagnosis == 'M','red','black'))
```

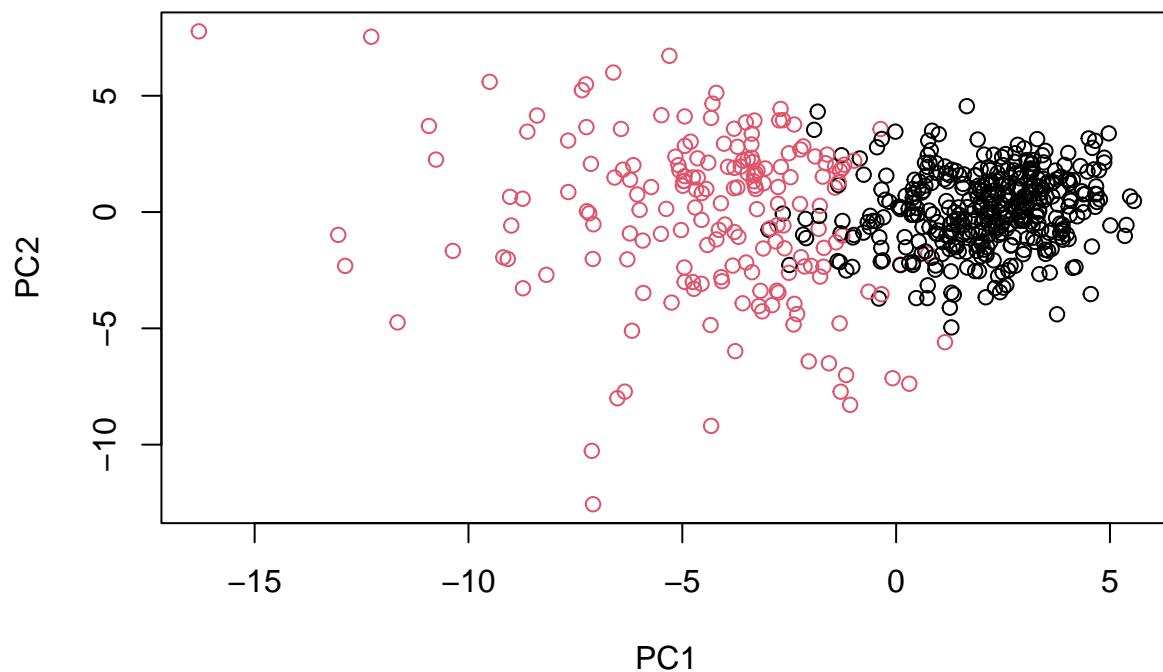
```
g <- as.factor(grps)
levels(g)
```

```
## [1] "1" "2"
```

```
g <- relevel(g,2)
levels(g)
```

```
## [1] "2" "1"
```

```
# Plot using our re-ordered factor
plot(wisc.pr$x[,1:2], col=g)
```



Question 15

with 4 clusters, the model using just the first 7 PCs does not separate out the clusters very well.

```
#Use the distance along the first 7 PCs for clustering
dist_first7 <- dist(wisc.pr$x[,1:7])
wisc.pr.hclust <- hclust(dist_first7, method="ward.D2")

#Separate into 2 clusters
wisc.pr.hclust.clusters <- cutree(wisc.pr.hclust, k=2)
table(wisc.pr.hclust.clusters, diagnosis)
```

```
##                diagnosis
## wisc.pr.hclust.clusters  B  M
##                1  28 188
##                2 329  24
```

```
#When separated into 4 clusters:
wisc.pr.hclust.clusters4 <- cutree(wisc.pr.hclust, k=4)
table(wisc.pr.hclust.clusters4, diagnosis)
```

```
##                diagnosis
## wisc.pr.hclust.clusters4  B  M
##                1    0  45
```

```
##           2   2  77
##           3  26  66
##           4 329  24
```

```
#Original
table(wisc.hclust.clusters, diagnosis)
```

```
##           diagnosis
## wisc.hclust.clusters  B   M
##           1  12 165
##           2   2   5
##           3 343  40
##           4   0   2
```