

Final 'Find a Gene' Project

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Question 1

Beginning my search, I knew I wanted to limit my organism to some kind of fungus, because they are very understudied and I think their diversity and versatility are fascinating!

I decided to narrow my search to those proteins that help the fungus *Ophiocordyceps Unilateralis* “zombify” a host insect by taking over the hosts’ neurological systems, eventually killing the host.

protein name: serine/threonine-protein kinase MAK, partial

species: *Ophiocordyceps Unilateralis*

accession number: ADI72911.1

function: The role of MAK-like kinases in this species is to induce behavioral changes in the host by interfering with Mitogen- Activated Protein Kinase signaling pathways. (ChatGPT)



Figure 1: *Ophiocordyceps Unilateralis*

Question 2

Attempting to find a homologous protein:

blast method: NCBI tblastn

database: est

limits/restrictions: none

My BLAST results were as followed:

Job Title **gb|ADI72911.1|**

RID [17SCCHF016](#) Search expires on 05-03 05:03 am [Download All](#) ▼

Program TBLASTN [Citation](#) ▼

Database est [See details](#) ▼

Query ID [ADI72911.1](#)

Description serine/threonine-protein kinase MAK, partial [Ophiocordyc ...

Molecule type amino acid

Query Length 139

Other reports [?](#)

Filter Results

Organism only top 20 will appear ☐ exclude

Type common name, binomial, taxid or group name

[+ Add organism](#)

Percent Identity to **E value** to **Query Coverage** to

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Descriptions Graphic Summary Alignments Taxonomy

Sequences producing significant alignments Download ▼ Select columns ▼ Show 100 ▼ [?](#)

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	Description	Scientific Name	Max Score	Total Score	Query Cover	E value	Per. Ident	Acc. Len	Accession
<input checked="" type="checkbox"/>	MgA0137f MgA Library Zymoseptoria tritici cDNA clone MgA0137 5' mRNA sequence	Zymoseptoria tritici	156	156	91%	3e-46	59.84%	589	AW180074.1
<input checked="" type="checkbox"/>	AculSEQ16326 Aureobasidium pullulans pBluescript (EcoRI-XhoI) Aureobasidium pullulans cDNA clone AculSEQ...	Aureobasidium p...	124	124	91%	2e-33	56.59%	726	DY862280.1
<input checked="" type="checkbox"/>	CATN5183 fwd CATN Nectria haematococca mpVI Sporulation (PDB) 77-13-4 Mycelia 24 hour culture [Nectria] ha...	Fusarium variettenii	100	100	91%	1e-23	48.03%	721	GE229289.1
<input checked="" type="checkbox"/>	Asn_08688 Aspergillus niger cBluescript (EcoRI-XhoI) Aspergillus niger cDNA clone Asn_08688 mRNA sequence	Aspergillus niger	99.0	99.0	74%	2e-23	54.29%	670	DR707892.1
<input checked="" type="checkbox"/>	FQ083057 Botrytis tulipae Bt9901 Botrytis tulipae cDNA mRNA sequence	Botrytis tulipae	99.0	99.0	91%	2e-23	48.91%	656	FQ083057.1
<input checked="" type="checkbox"/>	FQ105848 Botryotinia ficariaum CBS17663 Botryotinia ficariaum cDNA mRNA sequence	Botryotinia ficaria...	95.5	95.5	91%	2e-22	48.18%	574	FQ105848.1
<input checked="" type="checkbox"/>	FQ948614 Botryotinia fuckeliana 2 days old mycelia in rich medium cDNA library Botrytis cinerea cDNA mRNA se...	Botrytis cinerea	95.1	95.1	91%	2e-22	48.18%	563	FQ948614.1

Figure 2: BLAST results, original query

Alignment of choice: MgA0137f MgA Library Zymoseptoria tritici cDNA clone MgA0137 5', mRNA sequence

E Value: 3e-46

Percent Identity: 59.84%

Percent Coverage: 91%

Question 3

Here is some information about the homolog I am looking into:

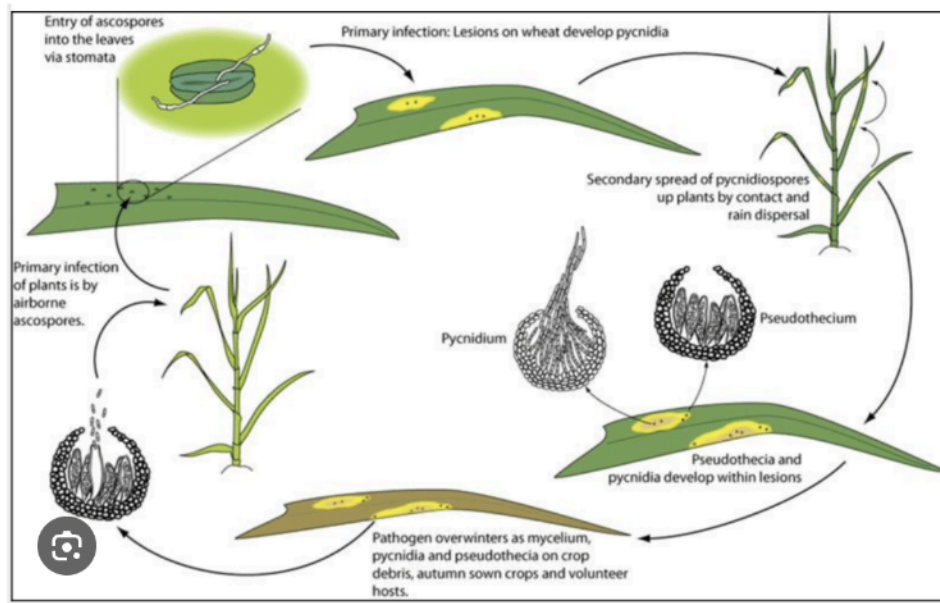


Figure 3: *Zymoseptoria tritici*

Name: MgA0137f MgA Library *Zymoseptoria tritici* cDNA clone MgA0137 5', mRNA sequence

Species Derived from: *Zymoseptoria tritici* : this is a pathogenic fungus that attacks wheat plants. It is resistant to multiple fungicides, and causes septoria leaf blotch.

FASTA format sequence, translated using EMBOSS Transeq:

AW180074.1_1 MgA0137f MgA Library *Zymoseptoria tritici* cDNA clone MgA0137 5', mRNA sequence

```
RQLSVNSQGNHYAEIHRQEAERALVGASALKSPTGSQRESFFSHLRKRARRRLSGRNSGVI
TPSMDAMETSAGCVPWAANKQTTFDTHSIASAAADPSSDPNFAELDRALQSVRYSLDAAA
NATQQARKPTNRVVEQPSLKRHHSLPHGVRHKTNPTTVYHDEH*STPRAADTRPPTKKKN
SRRSHELASARRTAFSX
```

```
DNYQSTHRAITTPKFTGRKLSVLWLAQALSSHRLAAKEKASSLICARGREDFPAATQVSS
HLQWMLWKPALGAFLGLLTNKPPSTPTRSLPQPIRHQTPISLSWIVHCKVYDTAWMPPR
TRLNKLGLSLRTALSNHHSVTTRFLTALDTRPTQPPYTTTSTEARHEQPIQDPRRRRI
LDEVMNSAHLAARRSR
```

TIISQLTGQSLRRNSPAGSACSGWRKRSQVTDWQPKRKLSSAQEGEKTFRPQLRCHH
 TFNGCYGNQRWVRSLGCTNHLRHPLDRVCRSRSVIRPQFRAGSCTAKCTIQGCRRE
 RDSTSEAYEPRSATIEASPLASSRRTQDQPNHRIPRRALKHATSSRYKTPDEEEEF
 STKSTQRISPHGVLX

RERRAARCAEFMTSSRILLRRGSCIGCSWRASVLVVVYGGWVGLVSNVAVRKRVVTLQ*W
 LLNYAVRRLPSLLSRVRGGIQAVSYYTLQCTIQLSEIGV**RIGCGRDRVGVGGFLVSS
 PRNAPSAGFHSIHRCDDTVAAGKSSRPLAQMREEAFSLAASRLESACANQSTLSFLP
 VNFGVVIALVDLS

SRTPCGEMRVHDFVENSSSSGVLYRLVACFSARRGIRWLGWSCVRREEASGDASMM
 VAQLRGSASLVESRRRHPGCIVHFAVHDPQRNWGLMTDRLRQTRSSGCRRWFVCQ
 PKERTQRWFPHPKLVHLSCGRKVFSPSCADERRSFLFGCQSVTERLRQPEHAQLPA
 GEFRRSDCPVSLIIVX

ENAVRRDALSSLRREFFFFVGGGLVSAARGVLQCSSWYTVVGLVLCLTPGSEWRFNDG
 CSTTRFVGFLACVAFAAASRLYRTLCSARSSSAKLGSDDGSAAADAIEWVSKVVCLLAA
 QGTHPALVSIASIEGVMTELPESLLALLRR*EKKLWLVPVGDRLALAPTRARSASCR
 ISALPCELTDCR

Question 4

To determine if this protein is novel:

I used NCBI blastp, in the nr database.

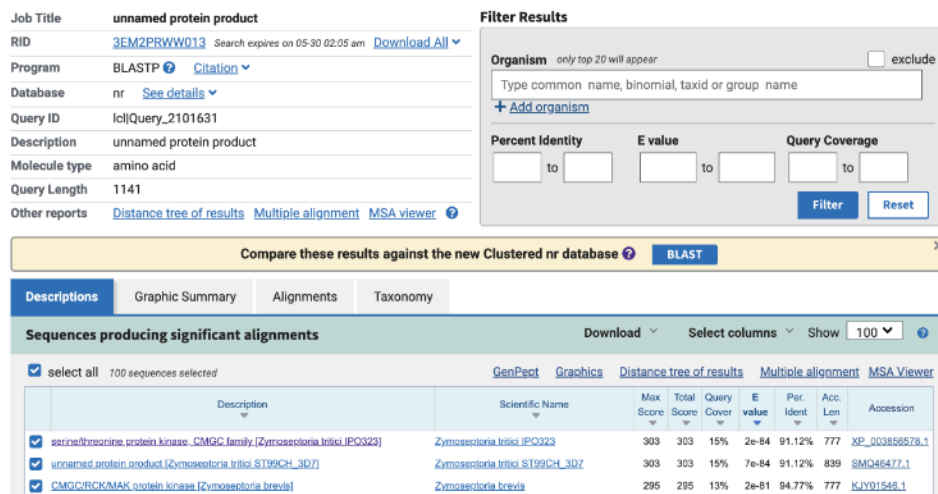


Figure 4: BLAST results, novel query

There is no match with 100% identity!

Question 5

I will use MUSCLE at EBI to produce a multiple sequence alignment of the following proteins:

My novel protein

>Zymoseptoria unknown protein (novel protein from BLAST results)

My original sequence for ophiocordyceps:

>ADI72911.1 serine/threonine-protein kinase MAK, partial [Ophiocordyceps unilateralis]

Other proteins of interest, based on BLAST results of original and novel proteins:

1. >KAK4508075.1 hypothetical protein PRZ48_001812 [Zasmidium cellare]
2. >XP_047755397.1 Sporulation protein kinase pit1 [Fulvia fulva]
3. >KJY01546.1 CMGC/RCK/MAK protein kinase [Zymoseptoria brevis]
4. >KAK4981489.1 hypothetical protein LTR28_003093, partial [Elasticomyces elasticus]
5. >KAI5369935.1 putative serine/threonine-protein kinase, active [Septoria linicola]

Alignment

Here is the alignment I obtained after running the above 7 proteins through MUSCLE at EBI (labeled by species), displayed in a code chunk because PDF formatting was giving me issues

```
"
Zymoseptoria      LLRRGSCIGCSWRASVLVVVYGGWGLVSNVVRKRVVTLQWLLNYAVRRLPSLLSRVRGG
Elasticomyces     -----VASHGNHYADHRHEAEQALN-----
Zymoseptoria_br   -----VNSQGNHYAEIHRQEAERALV-----
Ophiocordyceps    -----QEAERALS-----
Septoria          -----VNSQGNHYAELHRQEAERALN-----
Zasmidium         -----VNSQGNHYADIHRQEAERALT-----
Fulvia            -----VNSQGNHYADMHRQEAERALT-----

Zymoseptoria      IQAVSYTLQCTIQLSEIGVRIGCGRRDRVGVEGGLFVSSPRNAPSAGFHSIHRCDTVA
Elasticomyces     -----GRNG---LASPTSSQRGSFFAHLRKRARRLS
Zymoseptoria_br   -----GASA---LKSPTGSQRESFFSHLRKRARRLS
Ophiocordyceps    -----GANG---RKSPTGTLLSFHSHLRKRARRLS
Septoria.         -----GASG---LKSPTGSQRESFFSHLRKRARRFS
Zasmidium         -----GANG---LKSPTGSQRESFFSHLRKRARRLS
```


Fulvia	-----GATG---LQSPTGSQRESFFSHLRKRARRLS
Zymoseptoria	GKSSRPLAQMREEAFSLAASRLSACANQSTLSFLPVNFGVVIALVDLSSRTPCGEMRVH
Elasticomyces	GRNQAPVSPSVDD-----IEASAG-----CAP----
Zymoseptoria_br	GRNSGVITPSMDA-----METNAG-----CVP----
Ophiocordyceps	GRNQGPMSPGAED-----LEANAG-----CAP----
Septoria	GKPSGLASPTAED-----MEANVG-----CAP----
Zasmidium	GRNQGPMSPGAED-----IEASVG-----CAP----
Fulvia	GRNSGPMSPSAED-----AEANVG-----CAP----
Zymoseptoria	DFVENSSSSSGVLYRLLVACFSARRGIRLWGWSCVRREEASGDASMMVAQLRGSASLVES
Elasticomyces	-----WAS-NRQSMAIE-----
Zymoseptoria_br	-----WAA-NKQPVF-D-----
Ophiocordyceps	-----WSS-NRGSIQ-E-----
Septoria	-----WTT-NRQSIP-D-----
Zasmidium	-----WSTNNRGSIQ-E-----
Fulvia	-----WASNRRQSVQ-E-----
Zymoseptoria	RSRRHPGCIVHFAVHDPAQNRWGLMTDRLRQTRSSGCRRWFVCQPKERTQRWFPHPPLKVH
Elasticomyces	-----SLAITTHATDPSSDPNFAELDRALQNVRYSLDAGSYSNNNVQK-----PVQKV-
Zymoseptoria_br	-----THSVASAAADPSSDPNFAELDRALQSVRYSLDAAANATQQARK-----PTNRV-
Ophiocordyceps	-----PQPIEAVASDPSSDPNFAELDRALQNVRYSLDATANTSNNQPK-----HPTKM-
Septoria	-----AQAIAPTAADPSVDPNFAELDRALQSVRYSLDATAGAMPTQPK-----PPVKM-
Zasmidium	-----PQPIAPAAADPSTDPNFAELDRALQNVRYSLDAAANPANMQPK-----HPTKM-
Fulvia	-----PQSIASVAVDPSSDPNFAELDRALQNVRYSLDAAAGAANPQPK-----QPTKM-
Zymoseptoria	LSCGRKVFSPSCADERRSFLFGCQSVTERLRQPEHAQLPAGEFRRSDCPVSLIIVXENAV
Elasticomyces	-----PSNPMLKR-----HHSLPFGQDERISPVPAVNGPISSRT
Zymoseptoria_br	-----VSNPSLKR-----HHSLPHGVDDKTQ-----ANHRIP
Ophiocordyceps	-----ASNPSLKR-----HQSSHSG-----
Septoria	-----ASNPA LKR-----HHSLPYGKEEL-----SVVNRT
Zasmidium	-----PSNQSLKR-----HHSLPYGKEEIMSQT---GGSTSNRT
Fulvia	-----ASNPTLKR-----HHSVPCSKEEVTSNP-----SMANRT
Zymoseptoria	RRDALSSLRREFFFFVGGLVSAARGVLQCSSWYTVVGLVLCLTPGSEWRFNDCSTTRFV
Elasticomyces	RRS-----VRQAPHGHRYPDEEEEELL
Zymoseptoria_br	RRA-----LKHATSS--RYETPDEEEEELL
Ophiocordyceps	-----
Septoria	RRS-----VKQAPSN-IRYETPCEEDELL
Zasmidium	RRS-----LRHAPSS--RYETPCEEDELL
Fulvia	RRS-----LRHAPSS--RYETPCEEDELL

Zymoseptoria	GFLACVAFAAASRLYRTLCSARSSSAKLGSDDGSAADAIEWVSKV--VCLLAAQGTHPA
Elasticomyces	DEVLASAHRAARRLDRIYIQQDNSPLPSVTSQGERARPPVQVTS DP--GCFVPYLTPSPS
Zymoseptoria_br	DEVMTSAHLAARRL-----DNEQLSRPPLPHVISEPVTVYTAPYLTPSHS
Ophiocordyceps	-----PAPS
Septoria	DEAIASAHQAVTRL-----DNGITQPARPHLPHVTSEP--TYNVPYLTPSPS
Zasmidium	DEALASVHAAATRL-----DKGTS-NVAGTYQPSRPPIGHVTSEP--VYPAPYLTPSHS
Fulvia	DEALSNAHQAAQNL-----DNAPSMNTTATYQPARPLLQAISEP--AYAAPYLTPSPS
Zymoseptoria	LVSIA SI
Elasticomyces	KDRNG--
Zymoseptoria_br	KDQMSLD
Ophiocordyceps	RKP----
Septoria	KDHMAVD
Zasmidium	KDQMNV T
Fulvia	KDQMNI S
"	

```
[1] "\nZymoseptoria      LLRRGSCIGCSWRASVLVVVYGGWVGLVSNAVRKRVTTLQWLLNYAVRRLPSLLSRVRGG\nEla
```

Question 6

Given the alignment above, I can now create a phylogenetic tree using the EBI's Simple Phylogeny tool.

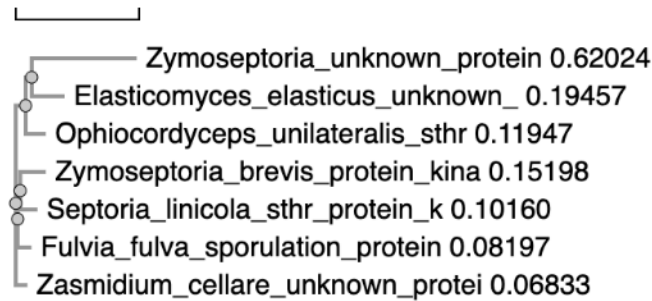


Figure 5: Phylogenetic tree for various fungal proteins

Question 7

To generate a heatmap, the msa must be in fasta format. One of the MUSCLE outputs is 'the alignment in FASTA format converted by Seqret', so I used that.

```
library(bio3d)

# Read multiple sequence alignment into R

msa <- read.fasta("finalseq.fasta")

# Calculate sequence identity matrix

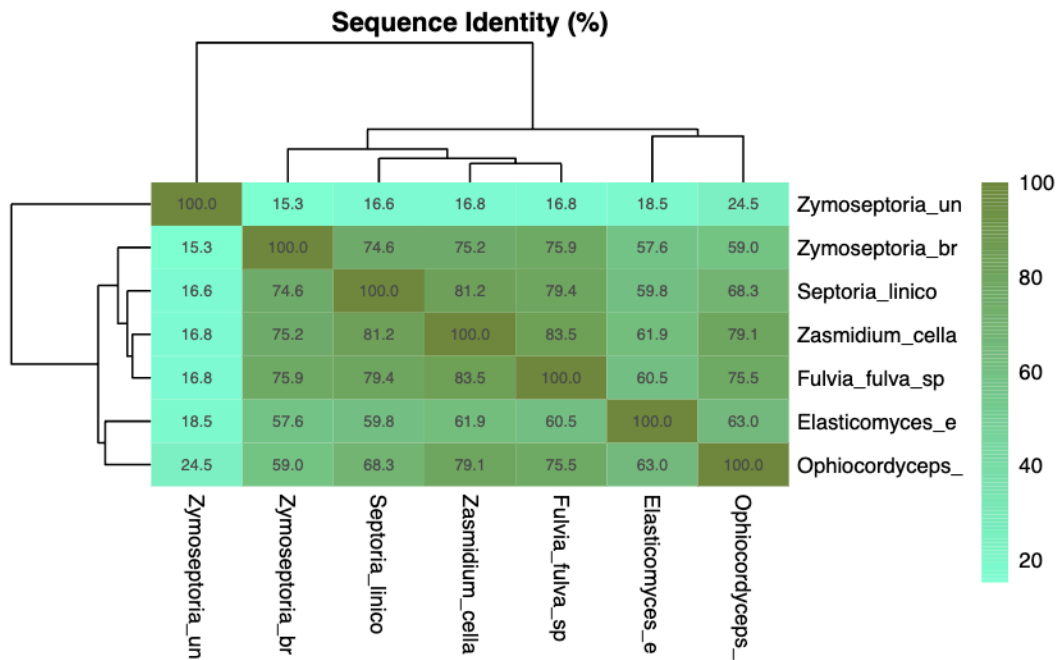
seq_id_m <- seqidentity(msa)

library(pheatmap)

#Generate a heatmap of the msa (nicer enhancements courtesy of ChatGPT)

idm_percent <- seq_id_m * 100
rownames(idm_percent) <- substr(msa$id, 1, 15)
colnames(idm_percent) <- substr(msa$id, 1, 15) #shortening the labels

pheatmap(idm_percent,
  display_numbers = TRUE,          # show exact % in each cell
  number_format = "%.1f",         # format: 1 decimal place
  color = colorRampPalette(c("aquamarine", "darkolivegreen4"))(100), # color gradient
  clustering_distance_rows = "euclidean",
  clustering_distance_cols = "euclidean",
  main = "Sequence Identity (%)",
  fontsize = 8,
  border_color = NA)              # cleaner look without gridlines
```

Quick note: because the sequence of my novel protein is quite long (over 2000 characters), some shortening is needed before it can be processed by many of the tools that I'll be using going forward.

I was unsure of the best way to shorten my sequence, so I asked ChatGPT to do it for me. It identified a central 1000-amino acid segment that avoids the N-terminal signal peptide and C-terminal tail, which are less structured and may not be essential for the core function.

The resulting novel sequence, printed below, will be saved as “zymo_novel”.

```
zymo_novel <- "VITPSMDAMETSAGCVPWAANKQTTFDTHSIAAAADPSSDPNFAELDRALQSVRYSLDAAANATQQARKPTNRVVE"
```

Question 8

I am going to try to create a consensus for all my aligned sequences (msa), using the Bio3d package `consensus()`.

```
cons <- consensus(msa)
```

Because this sequence appears to have a lot of gaps in it, I will use the protein sequences that I aligned instead. I have saved them as text in some of the code chunks above.

I will use the `blast.pdb()` function to search the seven fungal protein sequences against the pdb database.

```
# blast.pdb(zymo_novel) yielded no results  
# blast.pdb(ophiocordyceps) yielded no results  
# blast.pdb(zasmidium)
```

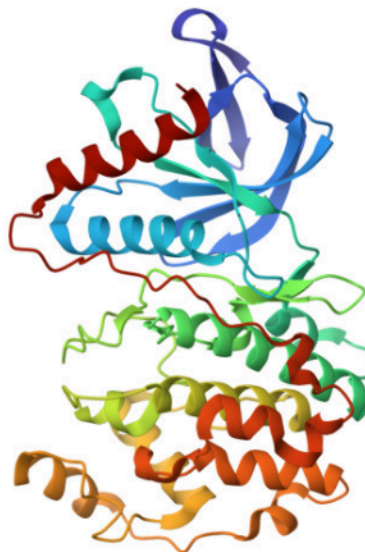


Figure 6: pdb match 1, 7W5C

Sequence details

PDB ID: 7W5C

E Value: 6.985e-33

Experimental technique: X-RAY DIFFRACTION

Resolution: 2.20 Å

Source organism: *Arabidopsis thaliana*

% identity: 34%

```
# blast.pdb(fulvia)
```

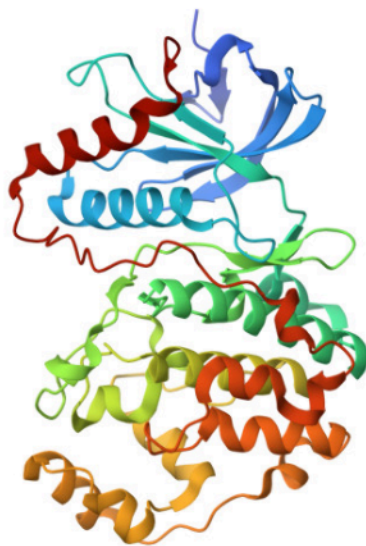


Figure 7: pdb match 2, 7E75

Sequence details

PDB ID: 7E75

E Value: 2.141e-28

Experimental technique: X-RAY DIFFRACTION

Resolution: 2.48 Å

Source organism: Homo Sapiens

% identity: 34%

```
# blast.pdb(z_brevis)
```

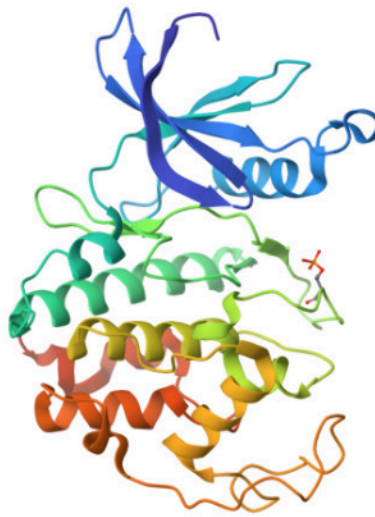


Figure 8: pdb match 3, 7NJ0

Sequence details

PDB ID: 7NJ0

E Value: 1.682e-40

Experimental technique: ELECTRON MICROSCOPY

Resolution: 3.60 Å

Source organism: Homo Sapiens

% identity: 34%

```
# blast.pdb(elasticomyces) yielded no results
```

```
# blast.pdb(septoria) yielded no results with a higher identity score than those above
```

Question 9

I will run “zymo_novel” through AlphaFold.

It took a couple hours, but my output included several files that were saved to my project directory.

I then ran “/test_62382/test_62382_unrelaxed_rank_001_alphafold2_ptm_model_2_seed_000.pdb” through the online Mol* viewer (the conserved regions are rendered in spacefill):

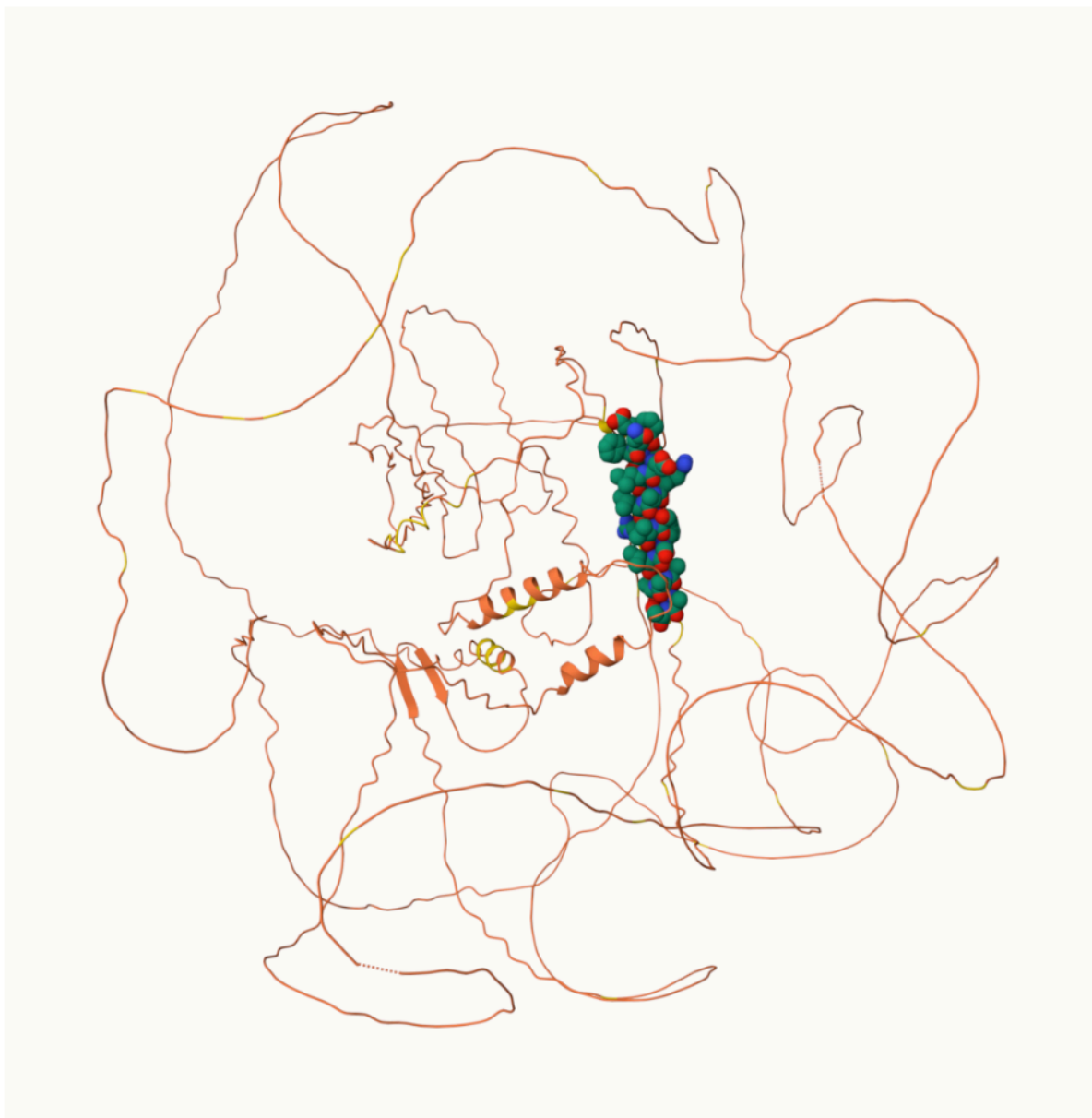


Figure 9: Final Unknown Protein Structure- colored by PLDDT confidence

Here it is without the spacefill, so the PLDDT coloring can be seen more clearly:

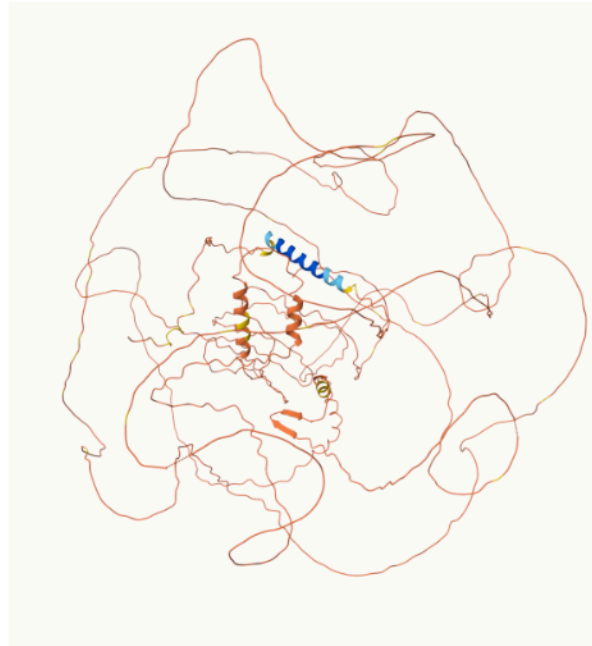


Figure 10: Final Protein Structure (no spacefill)

Question 10

- (i) I attempted to use the CASTPfold server to predict binding sites in my protein, but got the following result:

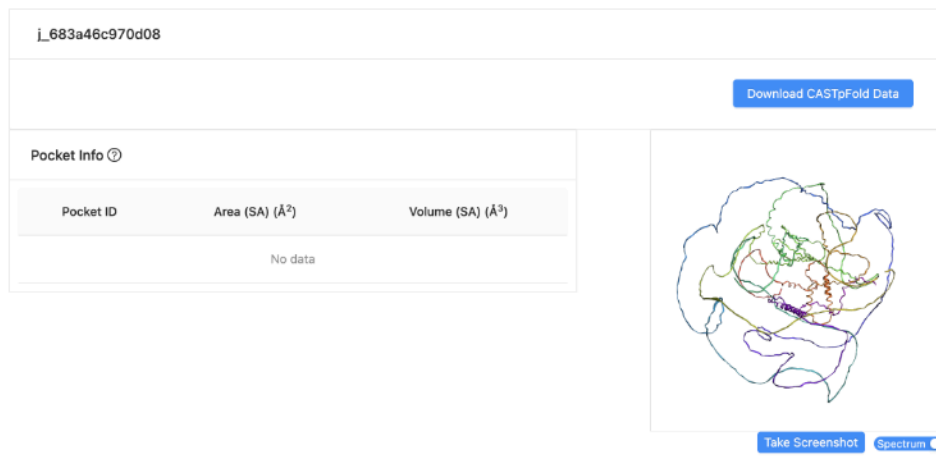


Figure 11: CASTPfold for my novel protein

So, unfortunately my protein appears to have no small molecule binding sites.

- (ii) I then performed a ChEMBL “target” search of my novel sequence. The resulting list consisted of 7 targets. I decided to focus on targets 4 and 5 (shown below) because they have the highest percent identity.

#	E-Value	Positives %	Identities %	Score (bits)	Score	Length	ChEMBL ID	Name	UniProt Accessions	Type	Organism	Compounds	Activities
1.	0.03	36.8	25.7	36.1	87	1321	CHEMBL1926492	Tau-tubulin kinase 1	Q5TCY1	SINGLE PROTEIN	Homo sapiens	142	185
2.	0.21	39.7	24.8	34.7	78	329	CHEMBL1255150	G-protein coupled bile acid receptor 1	Q80SS6	SINGLE PROTEIN	Mus musculus	482	786
3.	2.6	41.9	27.4	31.6	70	1288	CHEMBL1163123	Mitogen-activated protein kinase kinase 6	Q95382	SINGLE PROTEIN	Homo sapiens	447	537
4.	4.2	52.9	47.1	30.8	68	759	CHEMBL3317339	Polymerase basic protein 2	P03428	SINGLE PROTEIN	Influenza A virus (strain A/Puerto Rico/8/1934 H1N1)	100	131
5.	4.2	52.9	47.1	30.8	68	759	CHEMBL4523676	RNA-directed RNA polymerase	P03433 , P03431 , P03428	PROTEIN COMPLEX	Influenza A virus (strain A/Puerto Rico/8/1934 H1N1)	3	4
6.	5.3	47.5	27.9	30.4	67	571	CHEMBL2069163	Dual specificity testis-specific protein kinase 2	Q96553	SINGLE PROTEIN	Homo sapiens	173	173
7.	9.8	47.3	29.1	29.6	65	910	CHEMBL1250401	Potassium/sodium hyperpolarization-activated cyclic nucleotide-gated channel 1	Q88704	SINGLE PROTEIN	Mus musculus	14	22

Figure 12: My ChEMBL search

Target 4 appears to have the most useful data, as it has a defined compound that is effective against it, called Ribavirin.

RIBAVIRIN

Compound

Name and Classification

STRUCTURE SEARCH

ID: CHEMBL1643

Name: RIBAVIRIN

Max Phase: Approved [Learn more](#)

First Approval: 1985

Molecular Formula: C8H12N4O5

Molecular Weight: 244.21

Molecule Type: Small molecule

Synonyms and Trade Names:

ChEMBL Synonyms (11):

COTRONAK

NSC-163039

RIBAVIRIN

RIBAVIRINA

RIBAVIRIN BIOPARTNERS

- ALL (6 MORE) +

Figure 13: Details about Ribavirin

This compound functions as an RNA polymerase inhibitor.

- (iii) Overall, I do not believe my protein is druggable. My sequence is very long, and it was cut down so alphafold could process it. The resulting predicted structure did not have many conserved regions that I could use as starting points for further research. My CASTPfold search also did not yield any pockets. My ChEMBL searches yielded interesting results, but the highest percent identity listed was 47.1%, which does not place much confidence in the results. Perhaps if the whole sequence was run through a server with the capability to process it, a more accurate structure could be predicted.