#### **BGGN-213: FOUNDATIONS OF BIOINFORMATICS**

The find-a-gene project assignment <a href="http://thegrantlab.org/bggn213/">http://thegrantlab.org/bggn213/</a>
Dr. Barry Grant

#### Overview:

The find-a-gene project is a required assignment for BGGN-213. You should prepare a written report in **PDF** format that has responses to each question labeled **[Q1] - [Q10]** below. You may wish to consult the scoring rubric at the end of this document and the example report provided online.

The objective with this assignment is for you to demonstrate your grasp of database searching, sequence analysis, structure analysis and the R environment that we have covered in class.

## **Due Date:**

Your responses to questions Q1-Q4 are due at the beginning of **Week 5**. Note that these answers can be obtained very quickly (at best within 10 or 15 minutes), so if you don't succeed at first, just keep trying.

The complete assignment, including responses to all questions, is due at the beginning of **Week 10**. Late responses will not be accepted under any circumstances.

## **Submission instructions:**

Submit your PDF document to GradeScope as directed on our class website. Please do make sure your document is in PDF format and named something like BGGN213\_F20\_[yourUCSDname].pdf for example, my document would be named BGGN213\_F20\_bjgrant.pdf

## Be sure to include your UCSD email and PID number on the first page of your report.

Submit your preliminary report with answers to Q1-Q4 at the beginning of **week 5** so we can determine if you have found a novel gene. Submit this preliminary report as one document with screen shots of the results inserted appropriately.

See the demonstration report linked to on the course website for an example of format. I will email you my decision; proceed with subsequent questions only after we are sure you have found a novel gene.

For the final report add your results for Q5-Q10 to the preliminary report and submit a final document containing the results for all questions. <u>Please do not submit only Q5-Q10 answers as the final report</u>.

## Samuel Rivera

## PID: A53272335

## **Questions**:

[Q1] Tell me the name of a protein you are interested in. Include the species and the accession number. This can be a human protein or a protein from any other species as long as it's function is known.

If you do not have a favorite protein, select human RBP4 or KIF11. Do not use beta globin as this is in the worked example report that I provide you with online.

Name: HSPE1

ACCESSION CAG28616 ORGANISM Homo sapiens

Function: This gene encodes a major heat shock protein which functions as a

chaperonin.

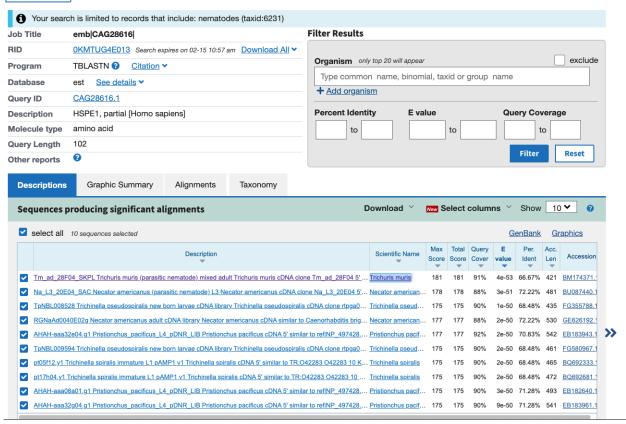
[Q2] Perform a BLAST search against a DNA database, such as a database consisting of genomic DNA or ESTs. The BLAST server can be at NCBI or elsewhere. Include details of the BLAST method used, database searched and any limits applied (e.g. Organism).

Method: TBLASTN

Database: Expressed Sequence Tags Organism: Nematodes (Taxid: 6231)

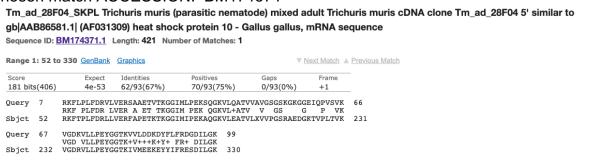
Also include the output of that BLAST search in your document. If appropriate, change the font to Courier size 10 so that the results are displayed neatly. You can also screen capture a BLAST output (e.g. alt print screen on a PC or on a MAC press \mathbb{H}-shift-4. The pointer becomes a bulls eye. Select the area you wish to capture and release. The image is saved as a file called Screen Shot [].png in your Desktop directory). It is not necessary to print out all of the

blast results if there are many pages.



On the BLAST results, clearly indicate a match that represents a protein sequence, encoded from some DNA sequence, that is homologous to your query protein. I need to be able to inspect the pairwise alignment you have selected, including the E value and score. It should be labeled a "genomic clone" or "mRNA sequence", etc. - but include no functional annotation.

#### Chosen match ACCESSION: BM174371



[Q3] Gather information about this "novel" <u>protein</u>. At a minimum, show me the protein sequence of the "novel" protein as displayed in your BLAST results from [Q2] as FASTA

format (you can copy and paste the aligned sequence subject lines from your BLAST result page if necessary) or translate your novel DNA sequence using a tool called EMBOSS Transeq at the EBI. Don't forget to translate all six reading frames; the ORF (open reading frame) is likely to be the longest sequence without a stop codon. It may not start with a methionine if you don't have the complete coding region. Make sure the sequence you provide includes a header/subject line and is in traditional FASTA format.

## Chosen sequence:

>Tm\_ad\_28F04\_SKPL Trichuris muris (parasitic nematode) mixed adult Trichuris muris cDNA clone Tm\_ad\_28F04 5' similar to gb|AAB86581.1| (AF031309) heat shock protein 10 - Gallus gallus, mRNA sequence-

RKFTPLFDRLLVERFAPETKTKGGIMIPEKAQGKVLEATVLXVVPGSRAEDGKTVPLTV KVGDRVLLPEYGGTKIVMEEKEYYIFRESDILGK

Name- Tm\_ad\_28F04\_SKPL Trichuris muris (parasitic nematode) mixed adult Trichuris muris cDNA clone Tm\_ad\_28F04 5' similar to gb|AAB86581.1| (AF031309) heat shock protein 10 - Gallus gallus, mRNA sequence

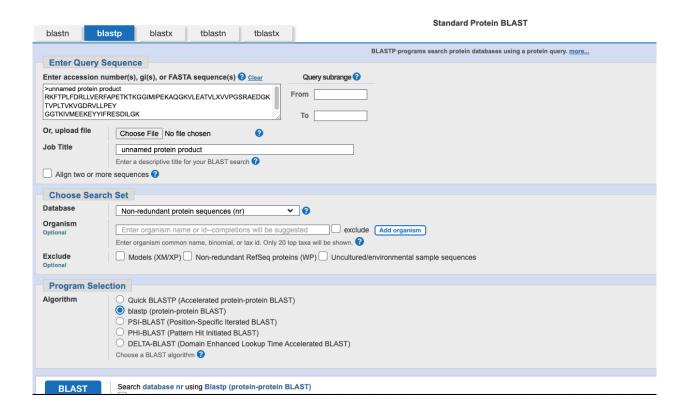
Species: Trichuris muris

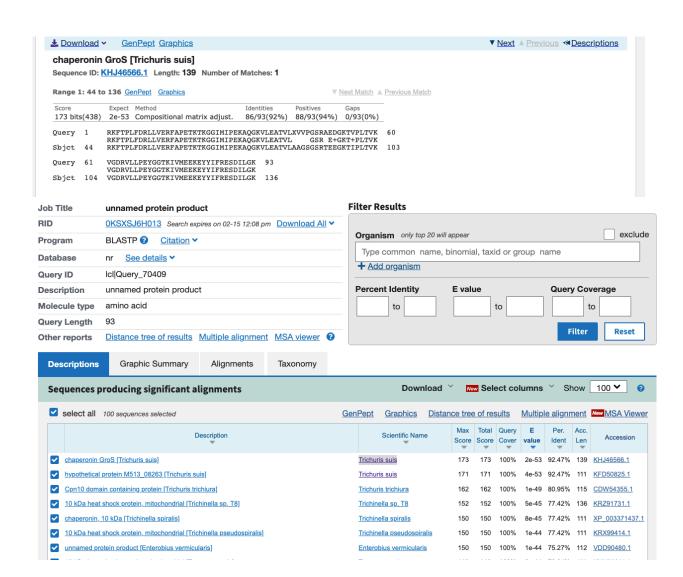
[Q4] Prove that this gene, and its corresponding protein, are novel. For the purposes of this project, "novel" is defined as follows. Take the protein sequence (your answer to [Q3]), and use it as a query in a blastp search of the nr database at NCBI.

- If there is a match with 100% amino acid identity to a protein in the database, from the same species, then your protein is NOT novel (even if the match is to a protein with a name such as "unknown"). Someone has already found and annotated this sequence, and assigned it an accession number.
- If the top match reported has less than 100% identity, then it is likely that your protein is novel, and you have succeeded.
- If there is a match with 100% identity, but to a different species than the one you started with, then you have likely succeeded in finding a novel gene.

• If there are no database matches to the original query from [Q1], this indicates that you have partially succeeded: yes, you may have found a new gene, but no, it is not actually homologous to the original query. You should probably start over.

BLASTP against NR database, no perfect match, top match from Trichuris suis, see alignment below.





[Q5] Generate a multiple sequence alignment with your novel protein, your original query protein, and a group of other members of this family from different species. A typical number of proteins to use in a multiple sequence alignment for this assignment purpose is a minimum of 5 and a maximum of 20 - although the exact number is up to you. Include the multiple sequence alignment in your report. Use Courier font with a size appropriate to fit page width.

Side-note: Indicate your sequence in the alignment by choosing an appropriate name for each sequence in the input unaligned sequence file (i.e. edit the sequence file so that the species, or short common, names (rather than accession numbers) display in the output alignment and in the subsequent answers below). The goal in this step is to create an interesting an alignment for building a phylogenetic tree that illustrates species divergence.

>Human chaperonin 10-related protein, partial [Homo sapiens]

AGQAFRKFLPLFDRVLVERSAAETVTKGGIMLPEKSQGKVLQATVVAVGSGSKGKGG EIQPVSVKVGDKVLLPEYGGTKVVLDDKDYFLFRDGDILG

>Trichuris muris (parasitic nematode) mixed adult Trichuris muris cDNA clone Tm\_ad\_28F04 5' similar to gb|AAB86581.1| (AF031309) heat shock protein 10 - Gallus gallus, mRNA sequence-

RKFTPLFDRLLVERFAPETKTKGGIMIPEKAQGKVLEATVLXVVPGSRAEDGKTVPLTV KVGDRVLLPEYGGTKIVMEEKEYYIFRESDILGK

>Mouse chaperonin 10 [Mus musculus]

MAGQAFRKFLPLFDRVLVERSAAETVTKGGIMLPEKSQGKVLQATVVAVGSGGKGKS GEIEPVSVKVGDKVLLPEYGGTKVVLDDKDYFLFRDSDILGKYVD >Rat heat shock 10 kDa protein 1 (chaperonin 10) [Rattus norvegicus]

## MAGQAFRKFLPLFDRVLVERSAAETVTKGGIMLPEKSQGKVLQATVVAVGSGGKGKG GEIQPVSVKVGDKVLLPEYGGTKVVLDDKDYFLFRDGDILGKYVD

>Zebra Fish chaperonin Cpn10, partial [Danio rerio]

## MQAFRKFLPMFDRVLVERLAAETVSRGGIMIPEKSQAKVLQATVVAVGPG

Alignment: EBI MUSCLE:

CLUSTAL multiple sequence alignment by MUSCLE (3.8)

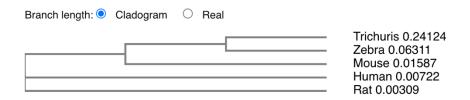
Trichuris -----RKFTPLFDRLLVERFAPETKTKGGIMIPEKAQGKVLEATVLXVVPGSRAEDGKT
Mouse MAGQAFRKFLPLFDRVLVERSAAETVTKGGIMLPEKSQGKVLQATVVAVGSGGKGKSGEI
Human -AGQAFRKFLPLFDRVLVERSAAETVTKGGIMLPEKSQGKVLQATVVAVGSGSKGKGGEI
Rat MAGQAFRKFLPLFDRVLVERSAAETVTKGGIMLPEKSQGKVLQATVVAVGSGGKGKGGEI
Zebra --MQAFRKFLPMFDRVLVERLAAETVSRGGIMIPEKSQAKVLQATVVAVGPG------

Trichuris VPLTVKVGDRVLLPEYGGTKIVMEEKEYYIFRESDILGK--Mouse EPVSVKVGDKVLLPEYGGTKVVLDDKDYFLFRDSDILGKYVD
Human QPVSVKVGDKVLLPEYGGTKVVLDDKDYFLFRDGDILG---Rat QPVSVKVGDKVLLPEYGGTKVVLDDKDYFLFRDGDILGKYVD
Zebra

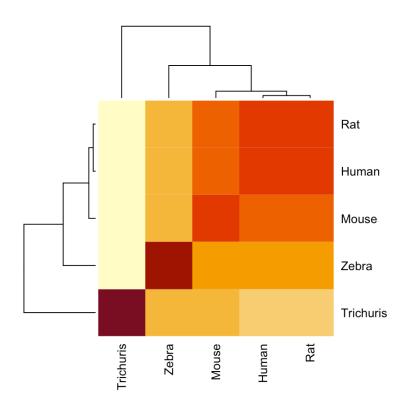
[Q6] Create a phylogenetic tree, using either a parsimony or distance-based approach. Bootstrapping and tree rooting are optional. Use "simple phylogeny" online from the EBI or any respected phylogeny program (such as MEGA, PAUP, or Phylip). Paste an image of your Cladogram or tree output in your report.

# Phylogenetic Tree

This is a Neighbour-joining tree without distance corrections.



[Q7] Generate a sequence identity based heatmap of your aligned sequences using R.



[Q8] Using R/Bio3D (or an online blast server if you prefer), search the main protein structure database for the most similar atomic resolution structures to your aligned sequences.

List the top 3 *unique* hits (i.e. not hits representing different chains from the same structure) along with their Evalue and sequence identity to your query. Please also add annotation details of these structures. For example include the annotation terms PDB identifier (structureId), Method used to solve the structure (experimentalTechnique), resolution (resolution), and source organism (source).

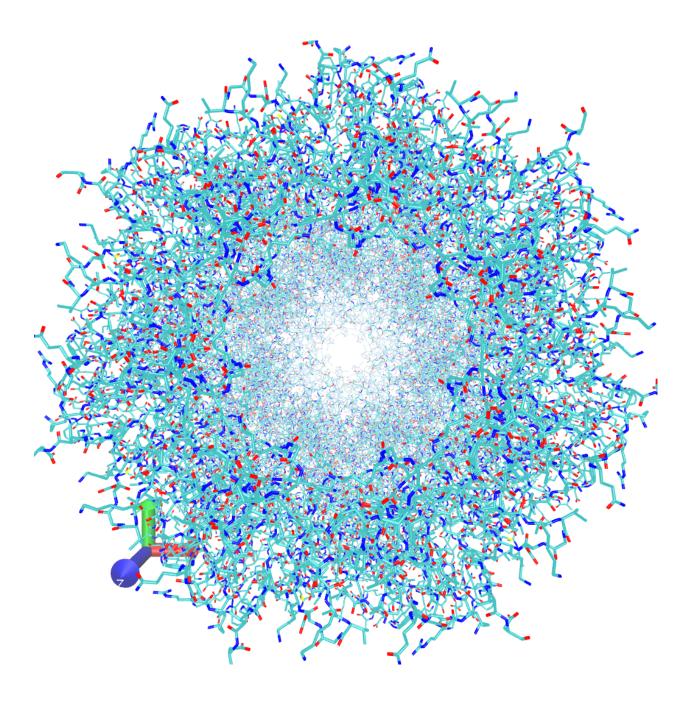
I used the Mus musculus sequence as it had the highest identity to the others.

ID	Technique	Resolution	Source	E value	Identity
Q64433	Electron Microscopy	2.9	Mus musculus	2E-56	100%
P26772	X-ray Diffraction	5.42	Mus caroli	2E-55	99.02%
Q9JI95	X-ray Diffraction	2.312	Zalophus californianus	2E-55	95.1%

[Q9] Generate a molecular figure of one of your identified PDB structures using **VMD**. You can optionally highlight conserved residues that are likely to be functional. Please use a white or transparent background for your figure (i.e. not the default black).

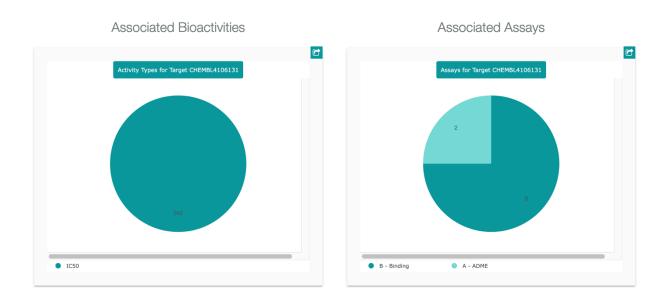
Based on sequence similarity. How likely is this structure to be similar to your "novel" protein?

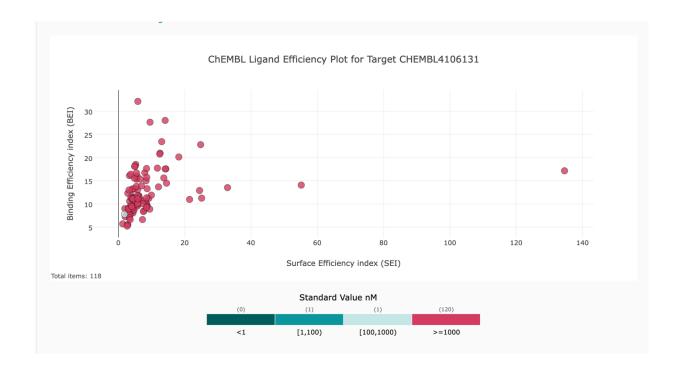
I used the human ADP-bound for this rendering. Based on my sequence alignment they certainly have significant regions of overlap but also several differences, so I would imagine that the overall 3D structure of these proteins is similar but likely these changes could make things like binding to certain molecules and activity difficult to predict.



[Q10] Perform a "Target" search of ChEMBEL ( <a href="https://www.ebi.ac.uk/chembl/">https://www.ebi.ac.uk/chembl/</a>) with your novel sequence. Are there any Target Associated Assays and ligand efficiency data reported that may be useful starting points for exploring potential inhibition of your novel protein?

There are 342 associated bioactivities, 2 ADME assays and 6 binding assays for the human HSPE1 version of my novel protein. Because, as we previously discussed there are significant differences between the human HSPE1 and my novel protein it is difficult to predict if these compounds would effectively interact with my novel protein, however they are a good place to start and could be promising at least as a starting point if interested in designing a molecule.





## **Scoring Rubric**:

[45 total points available]

<b>Q1</b> (4 points) Protein name	1	
Species	1	
Accession number	1	
Function known	1	
Q2 (6 points)		
Blast method	1	
Database searched	1	
Limits applied	1	
Search output list (top hits)	1	
Alignment of choice	1	
Evalue and other alignment stats	1	
Q3 (3 points)		
Protein sequence of choice matches Subject above	1	
Name in header	1	
Species	1	
<b>Q4</b> (3 point)		
Blastp output list with identities & Evalue	1	
Top alignment shown with alignment statistics	1	
Results indicates a "novel" gene found	1	
<b>Q5</b> (3 points)		
MSA labeled with useful names	1	
MSA trimmed appropriately (i.e. no gap overhangs)	1	
Pasted MSA fits report page width (i.e. font, format)		
<b>Q6</b> (1 point)		
Figure illustrates sequence clustering pattern	1	

## **Q7** (10 points)

Heatmap figure included in report	5
Heatmap is legible (i.e. no labels obscured)	5
<b>Q8</b> (10 points)	
PDB identifiers from multiple species reported	5
Annotation of PDB source, resolution and technique 4 Annotation of Evalue and Sequence Identity	1
<b>Q9</b> (4 points)	
Structure figure provided	2
Uses white background for molecular figure	1
Figure of high resolution (i.e. not just snapshot)	1
<b>Q10</b> (1 point)	
Evidence of ChEMBEL searches	1