

Estimation of fork length using cranial measurements of sablefish (*Anoplopoma fimbria*) in British Columbia

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(*ANOPILOPOMA FIMBRIA*) IN BRITISH COLUMBIA

by

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CONTENTS

ABSTRACT	v
RÉSUMÉ	vi
1 Introduction	1
2 Methods	2
2.1 Experimental Study 2016	2
2.1.1 Data Collection	2
2.1.2 Data Analysis	2
2.1.3 Gender test development	2
2.2 Fishery Pilot Study 2017	3
2.2.1 Data Collection	3
2.2.2 Data Analysis	3
3 Results	3
3.1 Experimental Study 2016	3
3.1.1 Genetic sex-determinations	4
3.2 Fishery Pilot study 2017	4
4 Discussion	4
5 Acknowledgments	5
6 Tables	6
7 Figures	9
APPENDICES	12
A IMAGES OF THE SIX CRANIAL DIMENSION MEASUREMENTS.	12

B SEX DETERMINATION BY OPERCULUM MARKING	13
8 References	14

ABSTRACT

Kathryn x. Temple, L.C. Lacko and Holt K.R. and MGL person 2022. Estimation of fork length using cranial measurements of sablefish (*Anoplopoma fimbria*) in British Columbia. Can. Tech. Rep. Fish. Aquat. Sci. nnn: vi + 14 p.

Routine biological sampling of whole round sablefish from commercial fishing operations in British Columbia began the early 1990's. Historically, specimens were obtained through the voluntary fishery catch sampling and tag recovery programs. We investigate the potential for obtaining sex and length information using heads, rather than the entire fish. In 2016, 438 sablefish (240-1080 mm) were sampled at sea and six different fish head measurements were collected. Genetic samples (137) were obtained to develop methods of DNA-based sex identification. A pilot study occurred in 2017 with 360 head-only samples collected and sexed by a commercial vessel, followed by scientific sampling on shore. Regression analysis results reveal that all six cranial dimensions can be used to accurately predict length. However, interorbital distance was not only a good predictor of length, but samplers ranked it the most efficient to measure and easily repeatable. Genomic DNA were successfully processed for 130 of the 137 samples. Fisher sex determination was accurate xx% of the time. Given the results, the 2018 sampling collection program was modified so that returns of whole round sablefish were replaced by head-only samples with knife cuts on the operculum to indicate sex.

RÉSUMÉ

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1 Introduction

Biological samples of British Columbia (BC) sablefish (*Anoplopoma fimbria*) from trap and hook & line fisheries have been collected from a fishery-based voluntary catch sampling program since 1995 (Haist and Wyeth 2001). Prior to 2018, samples of whole fish were collected dockside by Fisheries and Oceans Canada (DFO) port samplers and/or contracted service providers via the dockside monitoring program (DMP). In addition, tagged fish were collected whole from commercial fisheries (trap, trawl, hook & line) at the point of landing also by the dockside monitoring program. Biological data collected during catch sampling included fork length and sex. Otoliths were collected and archived for future ageing analyses. These data provide a fishery-dependent source of age and size composition data for the two-sex age structured operating model used as part of the BC Management Strategy Evaluation (MSE) (Cox et al. 2019, DFO 2020).

Between 2016 and 2017, DFO undertook research to evaluate the potential monitoring change to a 'head only' catch sampling program for sablefish trap and hook & line fisheries. Head-only sampling allows commercial crew to collect samples directly while at sea, thereby eliminating the need to rely on port samplers for dockside data collection as well as the need for fishing vessels to store whole fish. Instead, commercial crew J-cut the fish at-sea as per commercial practice, examine the gonads to determine sex, mark the sex with knife cuts on the operculum, and store the frozen head (and/or floy tag) for later sampling by science personnel. The key motivation for switching to a head-only sampling program was to increase the number and spatial distribution of fishery samples, while simultaneously maintaining the quality of biological data.

Evaluation of a new program necessitated the development and testing of methods. Previous research on other fish species has shown that lengths can be accurately estimated from head dimensions (Serafy et al. 1996; Park et al. 2007), head and mandible lengths (Isermann and Vandergoot 2005), and head height to eye diameter ratio (Richardson et al. 2015).

In this technical report, we describe the research investigation in 2016 that collected sablefish samples to: 1) identify cranial measurements that are reliable predictors of fork length; 2) identify cranial measurements that are practical to measure; and 3) develop DNA analysis techniques for gender detection. We also describe the fishery pilot study in 2017 to: 1) assess the ease of sample collection for commercial fishers; 2) determine the accuracy of fisher sex identification using DNA analysis; followed by 3) among-sampler measurements of each head morphometric.

2 Methods

2.1 Experimental Study 2016

2.1.1 Data Collection

Sablefish were randomly selected for sampling during the 2016 biennial DFO Groundfish Synoptic Bottom Trawl surveys, following a length stratified protocol. A total of 212 fish were sampled on the West Coast Vancouver Island survey (Williams et al. 2018) and 219 fish on the West Coast Haida Gwaii survey (Nottingham et al. 2018) (Figure 1). In addition, seven small sablefish were collected during the 2016 salmon survey (put reference here when obtain).

For each selected fish, fork length, round weight, sex and maturity were recorded at sea. The heads were removed, labelled with a unique ID and frozen. On shore, the cranial dimensions of upper jaw (L_{UJ}), eye diameter (L_{ED}), interorbital distance (L_{ID}), snout length (L_{SL}), post orbital to preoperculum distance (L_{PP}) and post orbital head length (L_{PO}) were measured using Mitutoyo Absolute® 500-762-20 coolant proof digimatic calipers (Table 1, Appendix A). Sagittal otoliths were collected for future ageing. The post orbital head length (L_{PO}) measurement was abandoned after testing 130 sablefish due to low sample quality and technical issues.

At the time of sampling, each cranial dimension was evaluated by experienced samplers (Kathryn, how many samplers) on a five point rating scale in terms of two distinct metrics: 1. ease of use and 2. repeatability. The ease of use metric focused on three key attributes of the measurement learn-ability (task understanding), efficiency (task-completion time) and degree of difficulty (task performance ease). The repeatability metric focused on ranking each measurement under repeated caliper placement, noting the presence of soft and hard head tissues.

Operculum clips (DNA) were collected from the first 137 fish measured and forwarded to the Molecular Genetics Laboratory (MGL) for analysis and gender test development.

2.1.2 Data Analysis

Using the `lm()` function in R (**R?**), fork lengths (L_{FL}) were estimated by a simple linear regression model, using the six cranial measurements as predictor variables to see how well each could predict fork length. Model fit was evaluated in terms of statistical significance (p-value less than 0.05) and coefficients of determination (r^2).

2.1.3 Gender test development

In the Molecular Genetics Laboratory (MGL), gender test methods were successfully developed. DNA multiplex polymerase chain reactions (PCRs) were performed using fluorescently labelled forward primers. X-insert and Y-insert specific primers developed by Rondeau et al. (2013) were used, but the X-insert forward and Y-nested reverse were redesigned to produce slightly smaller

PCR products (Table 2). Sex specific alleles were size fractionated in an ABI 3730 capillary DNA analyzer and were scored with ABI GeneMapper using an internal lane sizing standard.

2.2 Fishery Pilot Study 2017

2.2.1 Data Collection

In 2017, a pilot study was conducted with the commercial sector returning sablefish head-only samples. A total of 360 heads were collected from J-cut sablefish on a limited-entry fishery trip to the Cobb and Eickelberg seamounts (Figure 1). Each operculum was marked by commercial fishers with either one knife cut (male) or two knife cuts (female) (Appendix B).

Scientific sampling occurred on shore, with the first 99 heads of the pilot study measured by three experienced technicians for L_{ID} , L_{SL} , L_{UJ} and L_{PP} to determine the most repeatable measurement. The cranial dimensions of L_{ED} and L_{PO} were eliminated after the results from the 2016 experimental study.

2.2.2 Data Analysis

Given that three highly trained technicians were used to conduct the same measurements of the each head morphometric, the Index of Average Error (IAE) (Beamish and Fournier 2011) was calculated to find the most consistent cranial measurement. It is defined as:

$$IAE = \frac{1}{N} \sum_{j=1}^N \left[\frac{1}{R} \sum_{i=1}^R \frac{|X_{ij} - \bar{X}_j|}{\bar{X}_j} \right]$$

where N is the number of sablefish measured for each cranial measurement, R is the number of times each cranial measurement was taken, \bar{X}_j is the average cranial length for the jth sablefish, and X_{ij} is the ith cranial measurement of the jth sablefish.

Fin clips (95 of 99) were forwarded to the molecular genetics lab for an audit of fisher sex determination.

3 Results

3.1 Experimental Study 2016

A total of 438 specimens comprising 222 males and 216 females were evaluated for this study. The smallest fork length of the collected specimens was 240 mm, the largest was 1080 mm, and the average was 573.3 mm (Table 3).

Table 4 lists the statistics of the cranial dimensions (L_{UJ} , L_{ED} , L_{ID} , L_{SL} , L_{PP} , L_{PO}) as predictors of the response variable fork length (L_{FL}). All cranial dimensions were highly correlated with fork length with high R-squared values and significant p-values (Figure 2 and Figure 3).

The correlation coefficient (r) was highest for female measurements of L_{SL} (0.983) and L_{ID} (0.98) and male measurements of L_{UJ} (0.974) and L_{SL} (0.972). The R-squared values (r^2) were highest for female measurements of L_{SL} (0.966) and L_{ID} (0.96) and male measurements of L_{UJ} (0.949) and L_{SL} (0.945).

The aquatic science research technicians scored interorbital distance (L_{ID}) as the highest on the five point scale for ease of use and repeatable criteria, and eye diameter (L_{ED}) and postorbital head length (L_{PO}) were scored as the lowest (Table 5). L_{ID} (narrowest distance between the eye sockets) proved the easiest measurement as the tissue could be easily compressed with the caliper jaws to obtain bone measurements. L_{ED} (anterior-posterior diameter of eye socket) proved hard to repeat on soft tissue and L_{PO} (posterior inner edge of orbit to dorsal insertion of opercle) was difficult to measure since many opercula were damaged during head removal by J-cut.

3.1.1 Genetic sex-determinations

Genomic DNA (130 of 137 fin clips) were successfully PCR amplified by the MGL to determine sex. The accuracy of sex detection for the experimental study by the science technicians was 92% (119/130).

3.2 Fishery Pilot study 2017

The 360 heads received from the commercial vessel were in good condition and operculum cuts worked well to indicate sex. The first 99 heads (60 from Cobb Seamount, 39 from Eikelberg seamount) were measured once by three expert science technicians for each cranial dimension of L_{UJ} , L_{ID} , L_{SL} and L_{PP} (Table 6). The cranial measurements that produced the lowest mean error were upper jaw (L_{UJ}) and interorbital distance (L_{ID}) with IAE values of 1 % and 1.1 %, respectively (Table 6). The accuracy of the commercial fisher sex detection from the DNA gender analysis was xx% (x/95).

4 Discussion

All cranial dimensions collected in the research investigation were found to be good predictors of sablefish fork length (L_{FL}) with resulting r^2 values for male and female sablefish >0.88 . However, the catch sampling program implemented the interorbital distance (L_{ID}) length measurement as the new standard based on feedback from the experienced technicians. This measurement could be performed quickly, accurately and repeatedly. In general, they reported measurement errors were more probable when the calipers were placed on soft tissues, rather than bone. Other errors were a result of the manner in which the sablefish were handled before measurement, ie. the operculum was cut off. Samplers also reported that the smaller sablefish placed limitations on performing cranial measurements. Future studies could expand the size range to include small fish to determine those factors that influence predictions of L_{FL} . During this phase of the

research investigation, the Pacific Biological Station Molecular Genetics Laboratory successfully developed a genetic protocol to determine sex.

The pilot commercial fishing trip for collection of sablefish heads returned high quality samples and fishers were successful in fish sex determinations (I hope x %). These pilot samples were measured by three experienced technicians using a standardized protocol that determined L_{UJ} and L_{ID} had lower average errors over L_{SL} and L_{PP} . This matched the comments on the difficulty in caliper jaw positioning during the L_{SL} and L_{PP} measurements. Success with this pilot may be useful for other species such as Pacific halibut who experience predation problems on longline surveys, or difficult to measure deep water species.

As a result of our study, routine biological sampling procedures in 2018 were modified so that commercial fisheries are now only returning head samples, rather than the entire fish. These changes have improved the fishers ability to freeze, store and transport samples. We recommend that a follow-up review in a couple years to better examine how sampling rates, length composition and spatial distribution have changed to improve our overall data confidence.

5 Acknowledgments

We thank Schon Acheson and Kristina Castle for lending their skilled technical expertise for this report. We also thank the Molecular Genetics Laboratory at PBS for developing a sablefish gender test. A special thanks to the crew of the Pacific Viking for participating in the pilot project.

6 Tables

Table 1. List of head dimensions for upper jaw length (L_{UJ}), eye diameter (L_{ED}), interorbital distance (L_{ID}), snout length (L_{SL}), post orbital to preoperculum distance (L_{PP}) and post orbital head length (L_{PO}) measurement descriptions and specification of caliper jaw placement. Many follow the morphological measurements described in (Shaw and McFarlane 1997). The matching images are found in Appendix A.

Head dimension	Head description	Caliper jaw position
L_{UJ}	Tip of snout to the posterior edge of the maxilla.	Outside caliper jaw measurement from forward point and centre of snout to back of maxilla.
L_{ED}	Anterior-posterior diameter of eye socket.	Inside caliper jaw measurement firmly stretched against eye socket at vertical midpoint of eye.
L_{ID}	Narrowest distance between eye sockets, measured on dorsal surface.	Outside caliper jaw measurement of the horizontal midpoint of eyes on dorsal surface.
L_{SL}	Tip of snout to anterior inner edge of eye socket.	Outside caliper jaw measurement from forward point and centre of snout to horizontal midpoint of anterior edge of eye socket.
L_{PP}	Posterior inner edge of orbit to visual insertion point of preopercle.	Outside caliper jaw measurement from back of eye socket to preopercle bone insertion point. Preopercle must be lifted to expose preopercle bone underneath.
L_{PO}	Posterior inner edge of orbit to dorsal insertion of opercle.	Outside caliper jaw measurement from back of eye socket to bone underneath gill cover notch at dorsal insertion of the opercle. The operculum must be held taut.

Table 2. Primers used in the development of a genetic test for determining sablefish sex, developed by the Pacific Biological Station (PBS) Molecular Genetics Laboratory (MGL).

Locus	Sequence	Fragment Size
X-insert-DFO_F1	6FAM-CACCGCTCATGTACACTTTG	321
X-insert-2R	TGCTGCACTGTACCATCAAA	
Y-nested-1F	NED-GTCAGAAGGCAGTGGTGTAGT	234
Y-nested-MGL_2R	CGCTTGCAATACTACTGAATG	

Table 3. Summary of sablefish biological data collected during the 2016 experimental study. Tally of fork lengths (L_{FL}), round weights (RW), upper jaw lengths (L_{UJ}), eye diameters (L_{ED}), interorbital distances (L_{ID}), snout lengths (L_{SL}), post orbital to preoperculum distances (L_{PP}), post orbital head lengths (L_{PO}), females (F), males (M), sagittal otoliths and operculum clips (DNA) listed by survey.

Survey	L_{FL}	RW	L_{UJ}	L_{ED}	L_{ID}	L_{SL}	L_{PP}	L_{PO}	F	M	Otoliths	DNA	Total
2016 WCHG	219	219	219	219	218	219	207	52	111	108	219	59	219
2016 WCVI	212	212	211	212	212	211	212	78	105	107	212	78	212
Salmon	7	0	7	7	7	7	7	0	0	7	0	0	7
Total	438	431	437	438	437	437	426	130	216	222	431	137	438

Table 4. Statistics associated with simple linear regressions using measurements (mm) of cranial lengths: upper jaw length (L_{UJ}), eye diameter (L_{ED}), interorbital distance (L_{ID}), snout length (L_{SL}), post orbital to preoperculum length (L_{PP}) and post orbital head length (L_{PO}) as predictors of the fork length (L_{FL}) of 438 sablefish collected from research surveys in 2016. All models were significant at $P < 0.001$.

Cranial Measurement	Sex	N	Slope	SE	R^2	r	Predictor		Response L_{FL}	
							Mean	SD	Mean	SD
L_{UJ}	female	215	7.4	0.12	0.945	0.972	59.8	16.93	586.7	129.71
	male	222	8.1	0.13	0.949	0.974	56.2	13.16	560.3	109.45
L_{ED}	female	216	23.9	0.5	0.914	0.956	25.9	5.19	586.7	129.41
	male	222	20.1	0.51	0.877	0.936	25.9	5.09	560.3	109.45
L_{ID}	female	215	11.3	0.15	0.96	0.98	41.6	11.27	586.9	129.68
	male	222	12.2	0.25	0.916	0.957	39.2	8.59	560.3	109.45
L_{SL}	female	215	10.9	0.14	0.966	0.983	46.6	11.7	586.7	129.71
	male	222	11.9	0.19	0.945	0.972	43.1	8.93	560.3	109.45
L_{PP}	female	211	13.8	0.23	0.945	0.972	32.6	9.03	583	127.89
	male	215	15.7	0.31	0.921	0.96	30.4	6.73	559.9	109.84
L_{PO}	female	73	6.6	0.23	0.923	0.961	62.1	22.51	574.9	154.35
	male	57	7.8	0.34	0.904	0.951	59.2	12.81	555.6	104.97

Table 5. Ease of use and repeatable five point ranking scale recorded by experienced samplers for each cranial measurement for the 2016 experimental study, where 5 = Great; 4 = Good; 3 = Moderate; 2 = Questionable; 1 = Poor.

Head Dimension	5 Point Rank		Measurement			Considerations
	Ease of use	Repeatable	Caliper limitation	Bilateral	Bone	
L _{UJ}	3	4	x	x	x	End of the maxilla difficult to define. Caliper jaw position must be in center of snout.
L _{ED}	3	2		x		Caliper outside jaw position on soft tissue in eye socket may result in measurement differences.
L _{ID}	5	5			x	Tissue is compressed to obtain bone measurement. Easy to determine caliper jaw position.
L _{SL}	4	5		x	x	Caliper jaw position must be in center of snout.
L _{PP}	4	5		x	x	Caliper position on pre-operculum may result in measurement differences.
L _{PO}	3	2	x	x		Operculum damage from handling was observed on several fish.

Table 6. Summary of sablefish biological data collected from the 2017 pilot study to Cobb (n=60) and Eickelberg (n=39) seamounts, measured by three expert technicians using a standardized protocol for the 2017 pilot study. Index of Average Error (IAE) % values are reported for each of the cranial lengths.

Head Dimension	Sampler A	Sampler B	Sampler C	Sex		IAE
	Min - Max (mm)	Min - Max (mm)	Min - Max (mm)	Males	Females	
L _{UJ}	51.65 - 91.02	52.64 - 91.53	53.82 - 92.55	60	39	1
L _{ID}	35.43 - 61.18	35.91 - 60.81	34.94 - 60.72	60	39	1.1
L _{SL}	39.67 - 69.27	40.38 - 70.37	41.03 - 70.32	60	39	1.2
L _{PP}	27.69 - 49.88	26.83 - 52.51	27.25 - 52.15	60	39	2.3

7 Figures

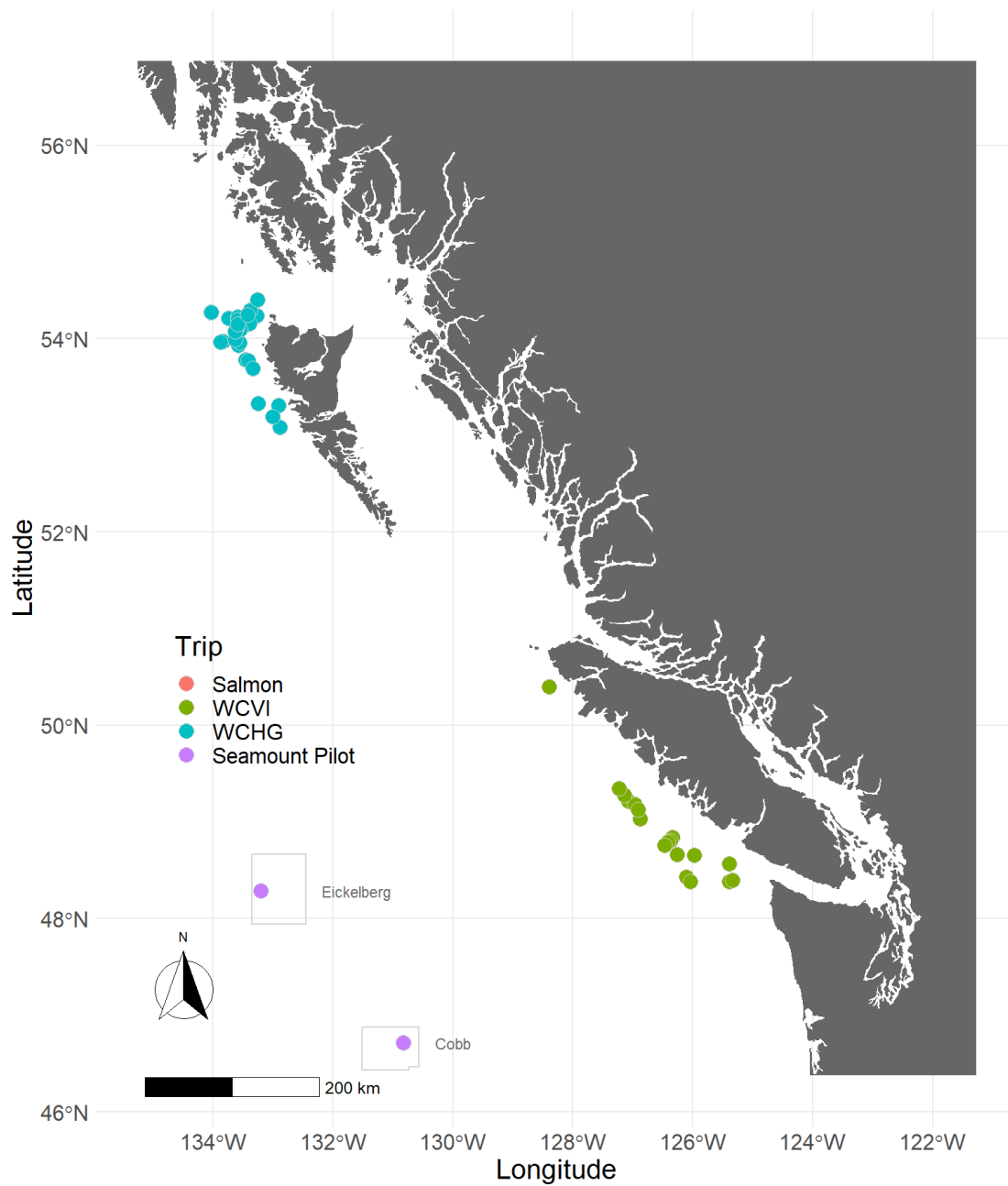


Figure 1. Sample locations from the 2016 WCVI, WCHG and salmon research surveys and 2017 pilot study.

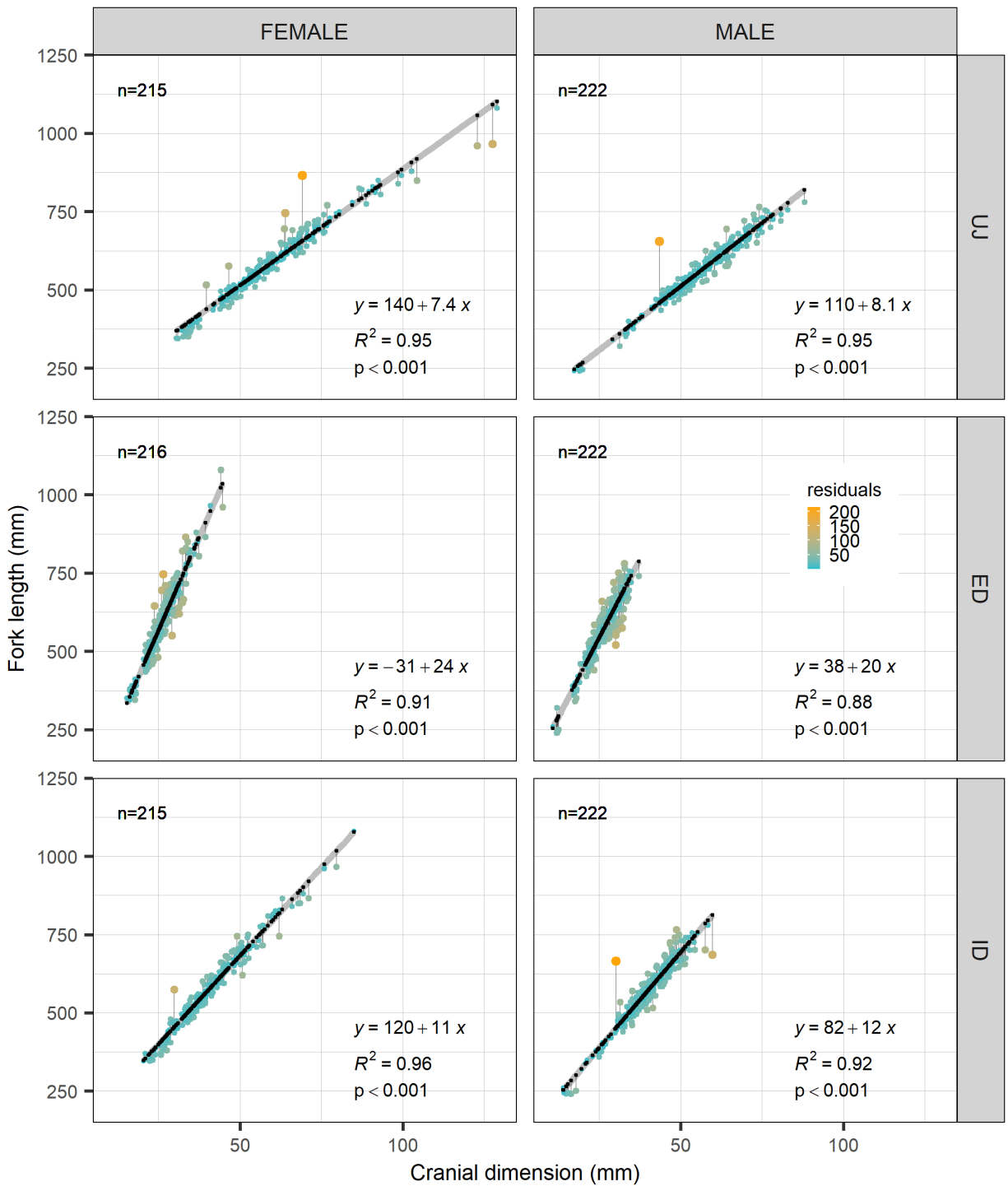


Figure 2. Relationship between cranial lengths (UJ, ED, ID) vs fork length in millimeters. Predicted points represented by black circles, measured values colored by residual scale.

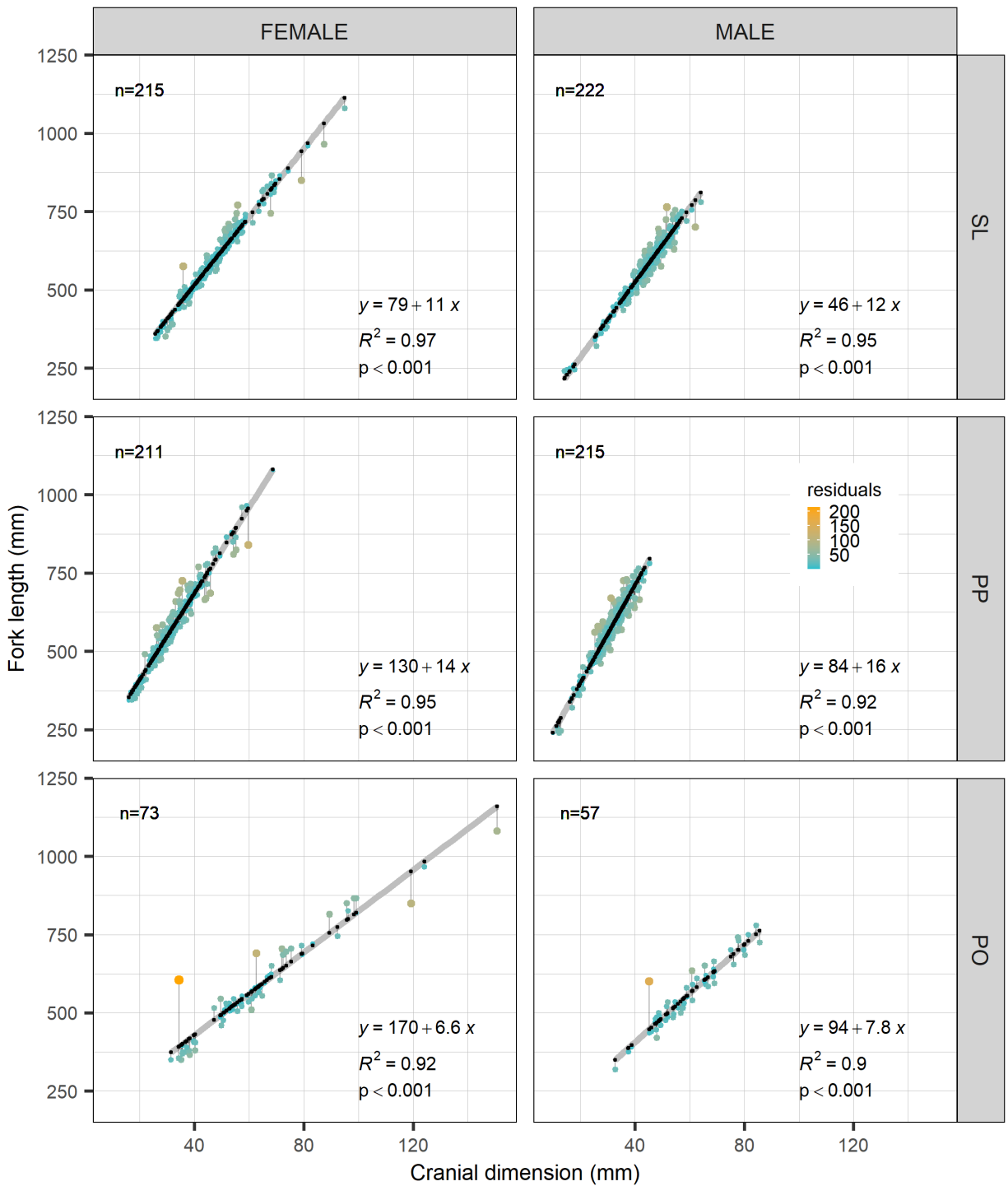
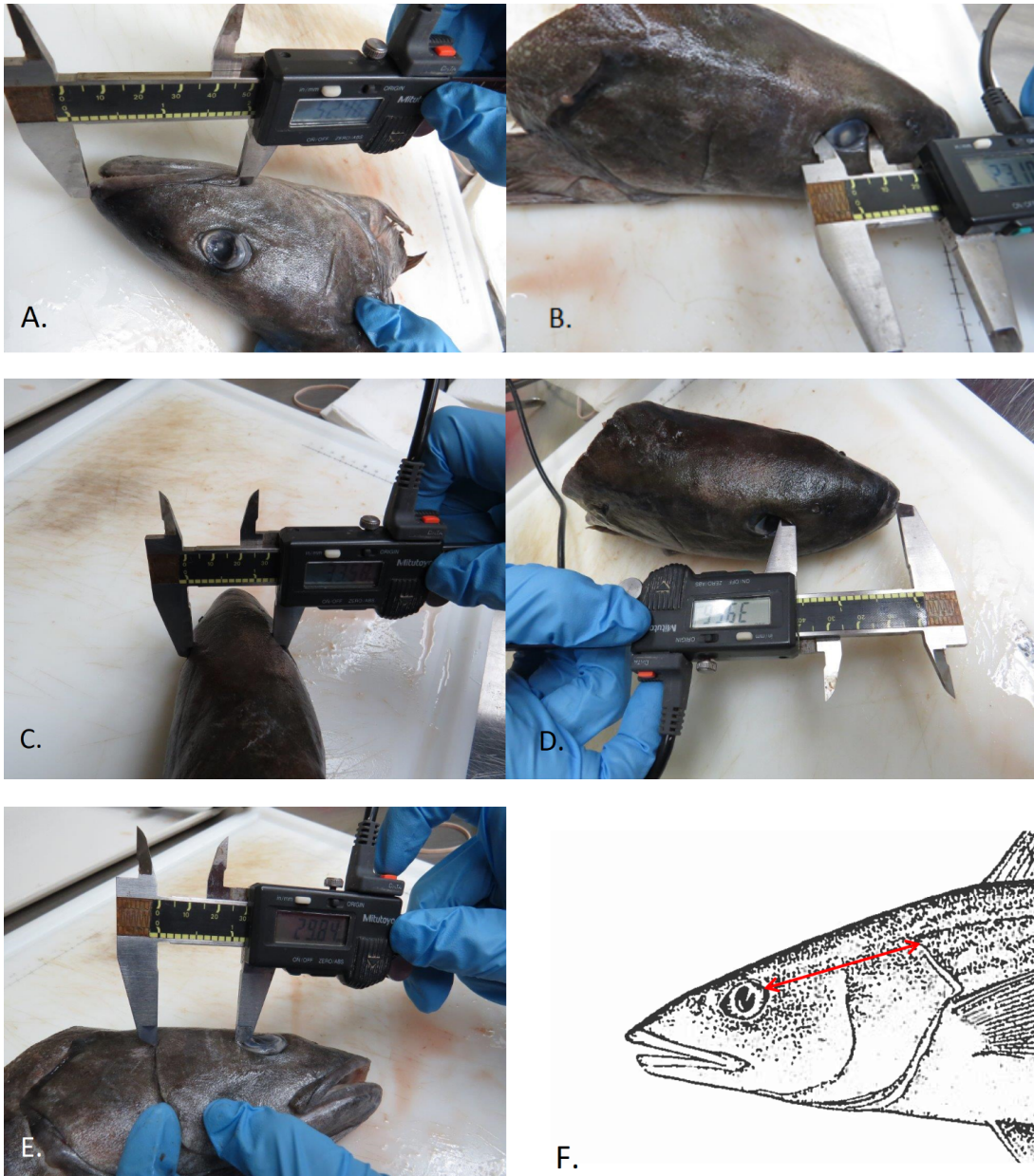


Figure 3. Relationship between cranial lengths (SL, PP, PO) vs fork length in millimeters. Predicted points represented by black circles, measured values colored by residual scale.

APPENDIX A IMAGES OF THE SIX CRANIAL DIMENSION MEASUREMENTS.

A. Upper jaw measurement (UJ); B. Eye diameter measurement (ED); C. Interorbital distance (ID); D. Snout length (SL); E. Post orbital to preoperculum length measurement (PP); F. Post orbital head length (PO).



APPENDIX B SEX DETERMINATION BY OPERCULUM MARKING

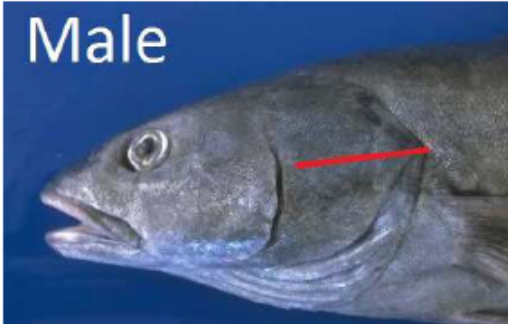
Instructions for sex determinations and operculum knife cuts for sablefish males and females.

MALES

Look for lobes that have an 'edge' to them. Due to the 'edge', they will not 'roll' easily in your fingers. The lobes are fused together at the end closest to the head.

Mark with one cut on the gill cover

Male



Sample photos for color and shape reference:

Juvenile: translucent-white



Developing: creamy-white



Post Spawning: brown or tan

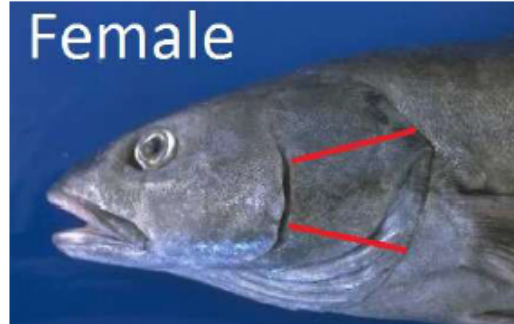


FEMALES

Look for tubes that are oval in cross section and will 'roll' in your fingers.

Mark with two cuts on the gill cover

Female



Sample photos for color and shape reference:

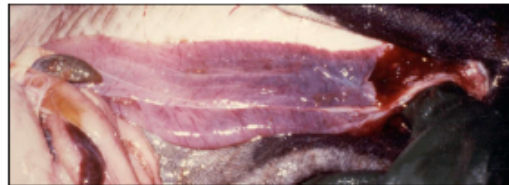
Juvenile: translucent pinky purple



Developing: creamy white, filled with eggs



Post Spawning: reddish- purple- brown, whitish sheen to the exterior surface



8 References

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