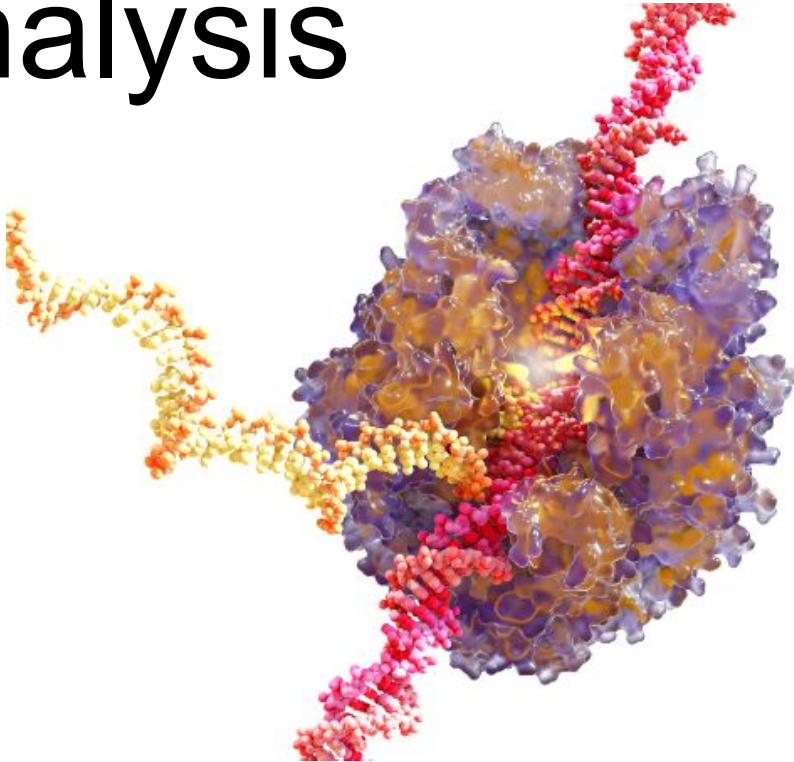


# Differential hnRNPC Binding Analysis



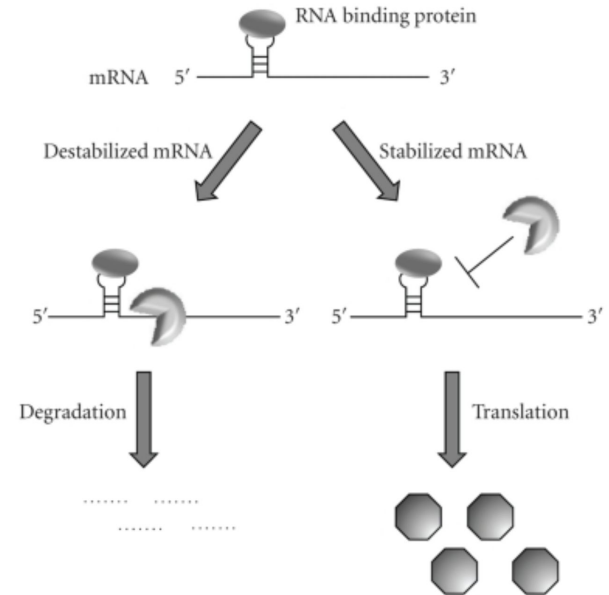
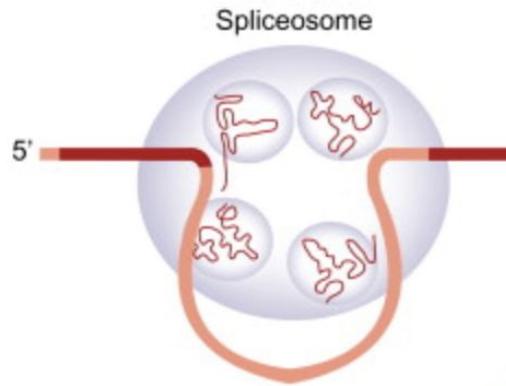
Davyd Sadovskyy  
PATH-GDS Rotation 3  
Dr. David Bentley Lab



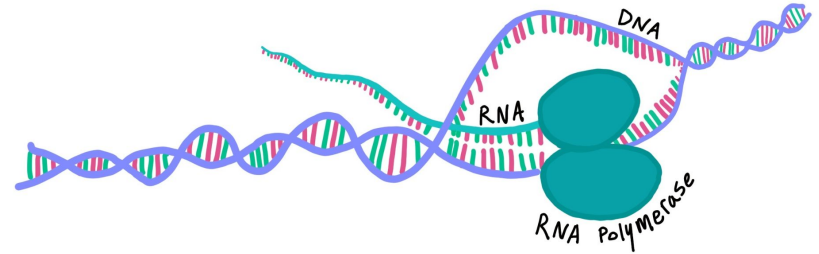
University of Colorado  
Anschutz Medical Campus

# What is hnRNP?

- hnRNP (heterogeneous nuclear ribonucleoprotein) is a an RNA-binding protein (RBP) that binds to newly transcribed RNA
- Functions in RNA stabilization, splicing, and sorting transcripts (mRNA Export)
- **hnRNPC** (my project) recognizes unstructured single-stranded RNA regions (~200-300 bp long)

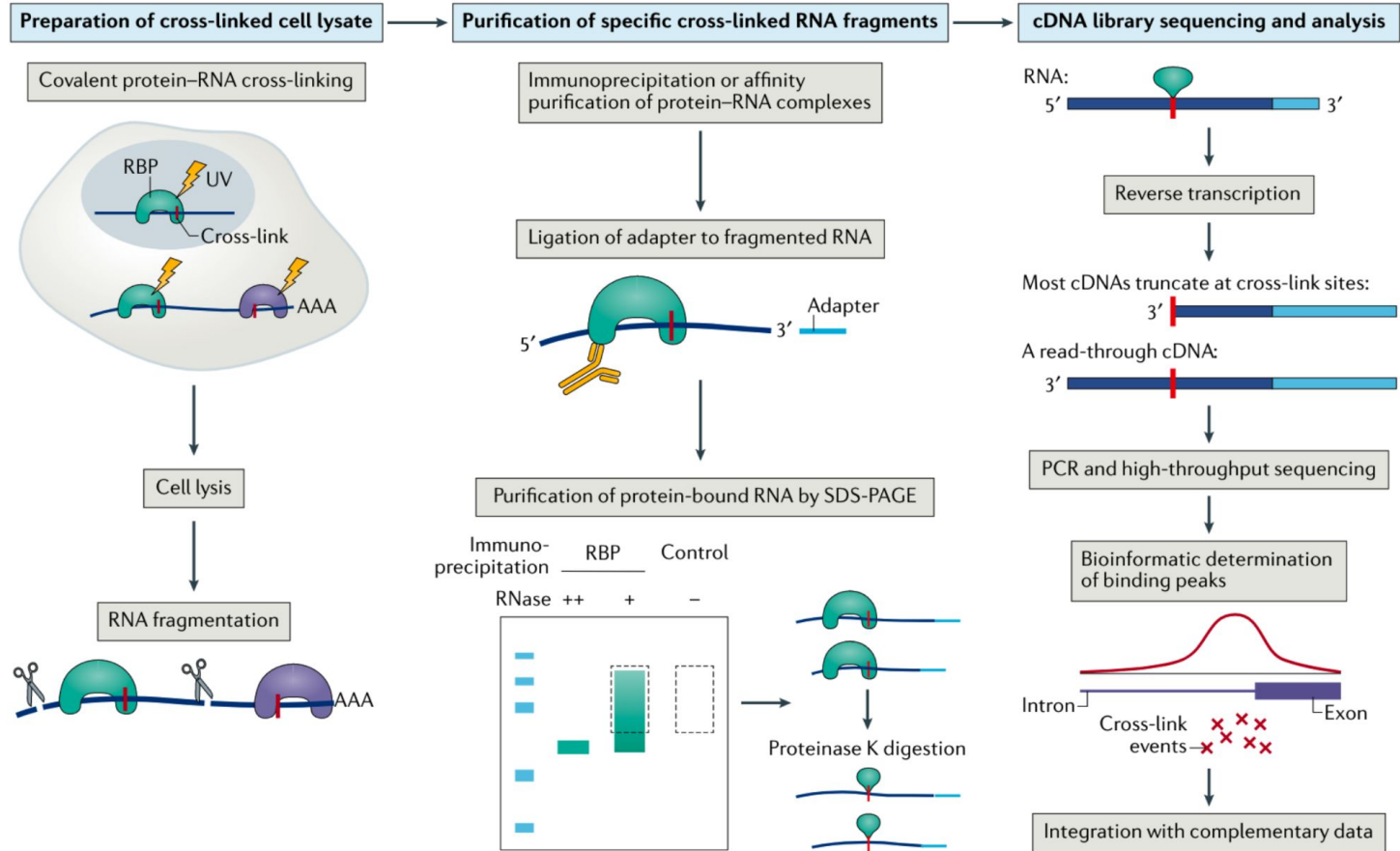


# Fast and Slow RNA polymerase



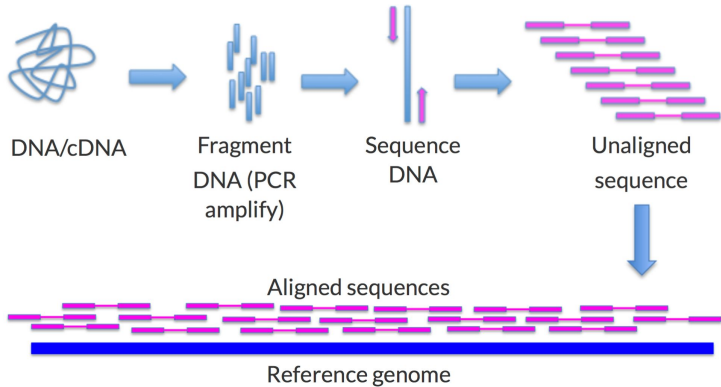
- Fast Pol II (Wild-Type)
  - ~3-4 kb per minute
  - Pre-mRNA folds before proteins can bind
- Slow Pol II (R749H Mutation / C4)
  - ~2 kb per minute
  - More unstructured RNA available for hnRNPC binding
- **Research Question**
  1. Is there differential binding of hnRNPC in fast vs slow pol?
  2. Where in genome are binding site occupancy differences?

# iCLIP Method



# iCLIP Raw Data

- The sequenced reads (cDNAs) are aligned to a reference genome using a tool like Bowtie or STAR.

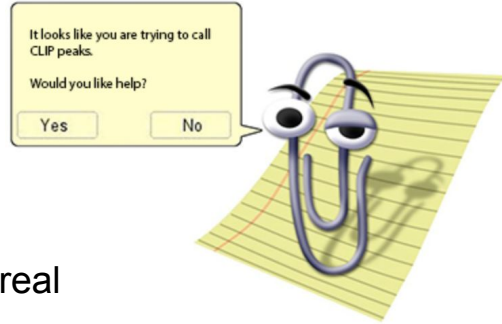


- **BAM File**

- Records every mapped cDNA read
- Chromosome
- BP position of alignment.
- Strand information (which direction it maps to).
- Sequence quality scores.

# Peak Calling - Clippy

.bam file



.bed file

- not all reads correspond to real cross-link events (random fragmentation, PCR bias, or sequencing errors)
- Different RNA regions have different background levels

- Peak calling identifies clusters of cross-linked reads, ranks them by intensity, and assigns binding confidence scores.

- start and end positions of peaks.
- Strand (+ or -).
- Peak intensity (more cross-links = higher score).

# Wilcoxon Rank-Sum Test

- **Does slow pol have different peak intensity compared to fast pol?**
- non parametric test - no assumption about distribution

## Steps for the test:

1. Rank all values together
2. Sum ranks for each group
3. Compute test statistic.  $R_1$  = sum of ranks for the smaller group.  
 $n_1$  = size of the smaller group

$$U = R_1 - \frac{n_1(n_1 + 1)}{2}$$

4. compare  $U$  to all possible rank sums that could occur if the groups were randomly shuffled. The p-value is the proportion of times the observed  $U$  value is as extreme (or more extreme) than expected under the null.

# Results

Total hnRNPC Binding  
Peaks in Each Condition

Condition	Total_Peaks
WT_Rep3	40855
C4_Rep3	115584
WT_Rep4	9116
C4_Rep4	29475

Shared hnRNPC Binding  
Peaks Between Conditions

Condition	Shared_Peaks
WT_Rep3	29363
C4_Rep3	29363
WT_Rep4	4556
C4_Rep4	4556

Unique hnRNPC Binding  
Peaks in Each Condition

Condition	Unique_Peaks
WT_Rep3	16651
C4_Rep3	91274
WT_Rep4	5435
C4_Rep4	25777

Average hnRNPC Binding Peak  
Score in Each Condition

Condition	Avg_Peak_Score
WT_Rep3	14.3153
C4_Rep3	16.1859
WT_Rep4	18.6286
C4_Rep4	15.0892

Wilcoxon Rank-Sum Test for Binding Strength Differences

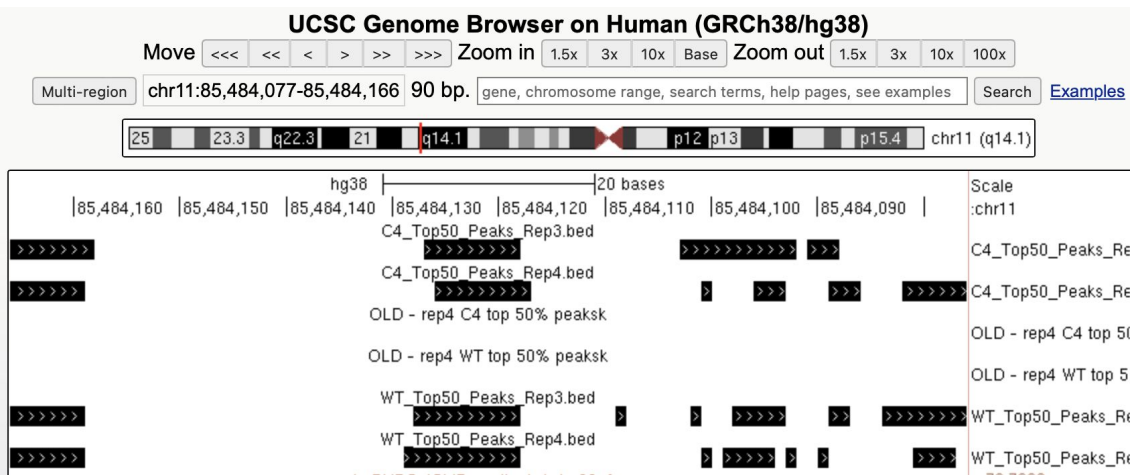
Comparison	Wilcoxon_Statistic	Wilcoxon_p_value
WT_Rep3 vs C4_Rep3	-19.91	0.0e+00
WT_Rep4 vs C4_Rep4	-4.74	2.1e-06

- Rep3: p-value =  $3.6e-88$  (VERY significant). C4 had higher peak scores (stronger hnRNPC binding with slow Pol II).
- Rep4: p-value =  $2.1e-6$  (still significant, but less extreme). C4 had lower peak scores than in Rep3 but still different from WT



# Next Steps

- Do further statistical analysis identify exact genes or genomic regions with different hnRNPC binding
- Validate differences in alternative splicing transcripts with more experiments.
  - RT-PCR or qPCR – Check if alternative exons are included or skipped differently in WT vs. C4 cells.
  - RNA-seq Analysis – Perform differential splicing analysis to identify transcripts with altered exon inclusion.



# Acknowledgments

Thank you to David Bentley and Ben Erickson for your help and guidance in this project