

# **Saemix:** Open Source R package for mixed effects modeling

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# Welcome to mixed effects modeling in R

The `saemix` project is an R package ([Comets et al., 2017](#)) available in CRAN that implements the Stochastic Approximation of the EM (SAEM) algorithm introduced in ([Kuhn and Lavielle, 2004](#)). This algorithm is state-of-the-art method for fitting, possibly non linear, models in agronomy, animal breeding or Pharmacokinetics-Pharmacodynamics (PKPD) analysis.

Thus far, the main area using the package thus far is Pharmacology, especially to understand how drugs, under development, behave in the body or how the body reacts to a drug during clinical trials (see ([Karimi et al., 2020](#)) or ([Samson et al., 2006](#))) but we ought to aim at a more general audience of biostatisticians dealing with nonlinear mixed effects modeling.

`saemix` is licensed under [GPL-2](#) | [GPL-3](#) [expanded from: GPL ( $\geq 2$ )].



# Chapter 1

## Introduction

Longitudinal data arise in many fields, such as agronomy, spatial analysis, imagery, clinical trials, and have been particularly prominent in the field of pharmacokinetics (PK) and pharmacodynamics (PD), where increasingly complex models involving mechanistic and empirical processes have been developed to describe the time course of and responses to drugs. Nonlinear models pose unique challenges in terms of estimation methods, and have driven the research to provide better estimation of parameters as well as the associated uncertainty, diagnostics of model misspecification and more informative designs. The SAEM algorithm, based on two highly cited publications by one of our project members Marc Lavielle, see (Delyon et al., 1999) and (Kuhn and Lavielle, 2004), was implemented in R in 2011 in the `saemix` R package~(Comets et al., 2017). Several applications of SAEM in agronomy, animal breeding and PKPD analysis have been published using `saemix`, see (Karimi et al., 2020) or (Samson et al., 2006).

PK/PD analyses are now a fundamental element of the registration file submitted to health authority for the approval of new drugs, but NLMEM are also increasingly applied to other areas. In clinical trials, they complement the point analyses by offering a unique understanding of the evolution of disease or treatment action. In cohort studies, they allow to model trajectories such as growth or cognitive decline. Joint models are now routinely used to link the evolution of markers with the occurrence of an event. Making use of S4 classes and methods to provide user-friendly interaction, `saemix` provides a new maximum likelihood estimation tool with a powerful exact algorithm to the R community.





## Chapter 2

# Installation

`saemix` can be installed and used on several platforms. Installation can range from easy to challenging, depending on the platform. We are in the process of streamlining this process, and any help or suggestions are greatly appreciated!

### 2.1 CRAN

You can either install the package directly from your R console

```
install.packages("saemix")
```

### 2.2 Github: Build from source

You can also build it from Github using the R package “devtools”

```
library("devtools")
```

```
install_github("saemixdevelopment/saemix" )
```

```
library(saemix)
```



## Chapter 3

# Materials

### 3.1 User's Guide

- Saemix User's Guide: PDF

### 3.2 Posters and Presentations

- PAGE 2011, Athens, Greece: PosterPAGE

Various other publications can be found [here](#).



## Chapter 4

# Case Studies

Some basic Case Studies are demonstrated in this chapter; the vignettes will be discussing the application in more depth.

### 4.1 A two-compartment PK model

```
library(saemix)
?saemix
```

#### Read the Data

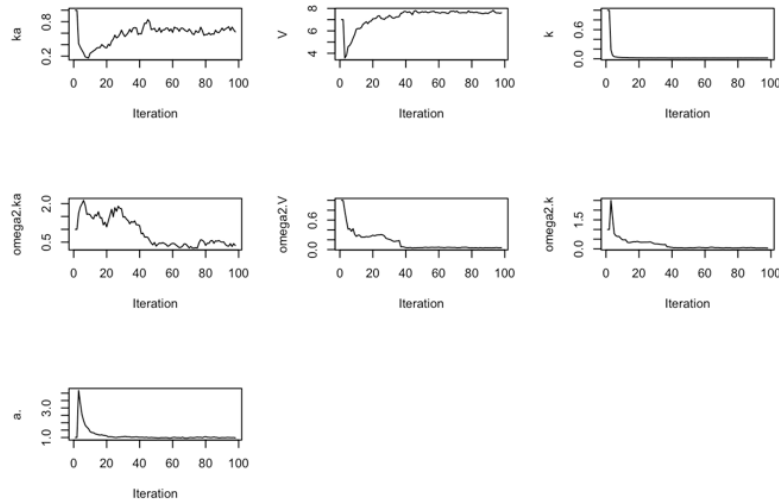
```
warfa_data <- read.table("data/warfarin_data.txt", header=T)
saemix.data<-saemixData(name.data=warfa_data,header=TRUE,sep=" ",
  na=NA, name.group=c("id"),name.predictors=c("amount","time"),
  name.response=c("y1"), name.X="time")
```

#### Create the Model

saemix models are contained in a R function with one blocks:

```
model1cpt<-function(psi,id,xidep) {
  dose<-xidep[,1]
  tim<-xidep[,2]
  ka<-psi[id,1]
  V<-psi[id,2]
  k<-psi[id,3]
  CL<-k*V
  ypred<-dose*ka/(V*(ka-k))*(exp(-k*tim)-exp(-ka*tim))
  return(ypred)
}
```

```
## Running main SAEM algorithm
## [1] "Fri Jan 31 10:52:18 2020"
## .
```



```
saemix.model<-saemixModel(model=model1cpt,description="warfarin",
  type="structural",psi0=matrix(c(1,7,1,0,0,0),ncol=3,byrow=TRUE,
  dimnames=list(NULL, c("ka","V","k"))),transform.par=c(1,1,1),
  omega.init=matrix(c(1,0,0,0,1,0,0,0,1),ncol=3,byrow=TRUE),
  covariance.model=matrix(c(1,0,0,0,1,0,0,0,1),ncol=3,
  byrow=TRUE))
```

### Run the SAEM algorithm

```
K1 = 200
K2 = 100
```

#### #Run SAEM

```
options<-list(seed=39546,map=F,fim=F,ll.is=F,
  nbiter.mcmc = c(2,2,2), nbiter.saemix = c(K1,K2),nbiter.sa=0,
  displayProgress=TRUE,save.graphs=FALSE,nbiter.burn =0)
fit<-saemix(saemix.model,saemix.data,options)
```

## 4.2 A categorical data model with regression variables

### 4.2.1 mlxR: simulate synthetic data

```
library("mlxR")
catModel <- inlineModel(
  "[LONGITUDINAL]
  input = {beta0,gamma0,delta0, dose}
  dose = {use=regressor}
  EQUATION:
  lm0 = beta0+gamma0*t + delta0*dose
  D = exp(lm0)+1
  p0 = exp(lm0)/D
  p1 = 1/D

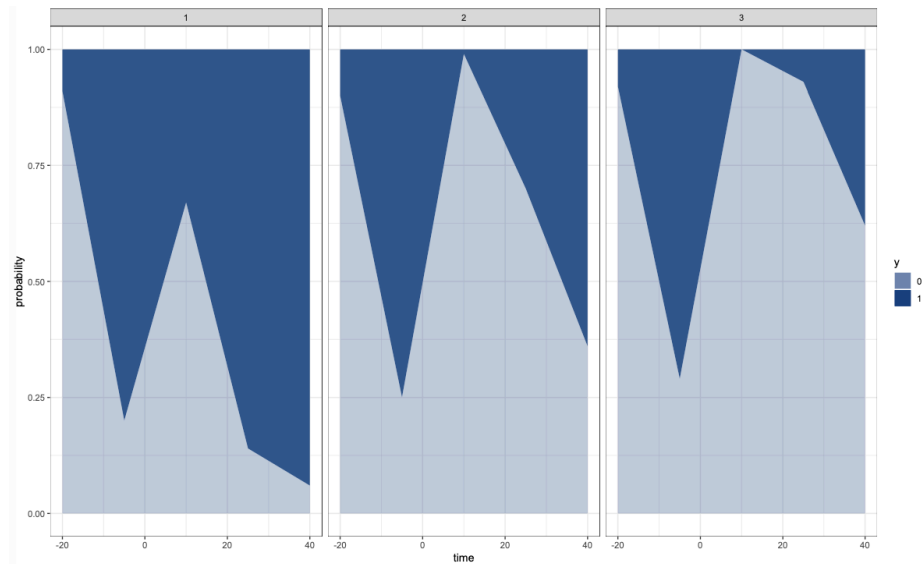
  DEFINITION:
  y = {type=categorical, categories={0, 1},
       P(y=0)=p0,
       P(y=1)=p1}
  [INDIVIDUAL]
  input={beta0_pop, o_beta0,
         gamma0_pop, o_gamma0,
         delta0_pop, o_delta0}
  DEFINITION:
  beta0  = {distribution=normal, prediction=beta0_pop, sd=o_beta0}
  gamma0 = {distribution=normal, prediction=gamma0_pop, sd=o_gamma0}
  delta0 = {distribution=normal, prediction=delta0_pop, sd=o_delta0} ")

nobs = 15
tobs<- seq(-20, 50, by=nobs)
reg1 <- list(name='dose',
             time=tobs,
             value=10*(tobs>0))

reg2 <- list(name='dose',
             time=tobs,
             value=20*(tobs>0))

reg3 <- list(name='dose',
             time=tobs,
             value=30*(tobs>0))

out  <- list(name='y', time=tobs)
```



```

N <- 100
p <- c(beta0_pop=-4, o_beta0=0.3,
      gamma0_pop= -0.5, o_gamma0=0.2,
      delta0_pop=1, o_delta0=0.2)

g1 <- list(size=N,regressor = reg1)
g2 <- list(size=N,regressor = reg2)
g3 <- list(size=N,regressor = reg3)
g <- list(g1,g2,g3)
res <- simulx(model=catModel,output=out, group=g,parameter=p)
plot1 <- catplotmlx(res$y)

```

### 4.2.2 saemix: fit the noncontinuous data model

Create the saemix.data object

```

saemix.data<-saemixData(name.data=res,header=TRUE,sep=" ",
  na=NA, name.group=c("id"),name.predictors=c("amount","time"),
  name.response=c("y1"), name.X="time")

```

Create the model

saemix models are contained in a R function with one blocks:

```

cat.model<-function(psi,id,xidep) {
  level<-xidep[,1]

```



```

dose<-xidep[,2]
time<-xidep[,3]
th1 <- psi[id,1]
th2 <- psi[id,2]
delta0 <- psi[id,3]
lm0 <- th1+th2*time + delta0*dose
D <- exp(lm0)+1
P0 <- exp(lm0)/D
P1 <- 1/D

P.obs = (level==0)*P0+(level==1)*P1
return(P.obs) }

saemix.model<-saemixModel(model=cat.model,description="cat model",
  type="likelihood", psi0=matrix(c(2,1,2),ncol=3,byrow=TRUE,
  dimnames=list(NULL,c("th1","th2","th3"))),transform.par=c(0,0,0),
  covariance.model=matrix(c(1,0,0,0,1,0,0,0,1),ncol=3,byrow=TRUE),
  omega.init=matrix(c(2,0,0,0,1,0,0,0,1),ncol=3,byrow=TRUE),
  error.model="constant")

```

#### Run the SAEM algorithm

```

K1 = 500
K2 = 100

options<-list(seed=39546,map=F,fim=F,ll.is=F,
  nbiter.mcmc = c(2,2,2), nbiter.saemix = c(K1,K2),nbiter.sa=0,
  displayProgress=TRUE,save.graphs=FALSE,nbiter.burn =0)
saemix.fit<-saemix(saemix.model,saemix.data,options)

```

## 4.3 A repeated time-to-event data model

### Read the Data

```

data(tte.saemix)
saemix.data<-saemixData(name.data=tte.saemix,header=TRUE,
  sep=" ",na=NA, name.group=c("id"),
  name.response=c("y"),name.predictors=c("time","y"),
  name.X=c("time"))

```

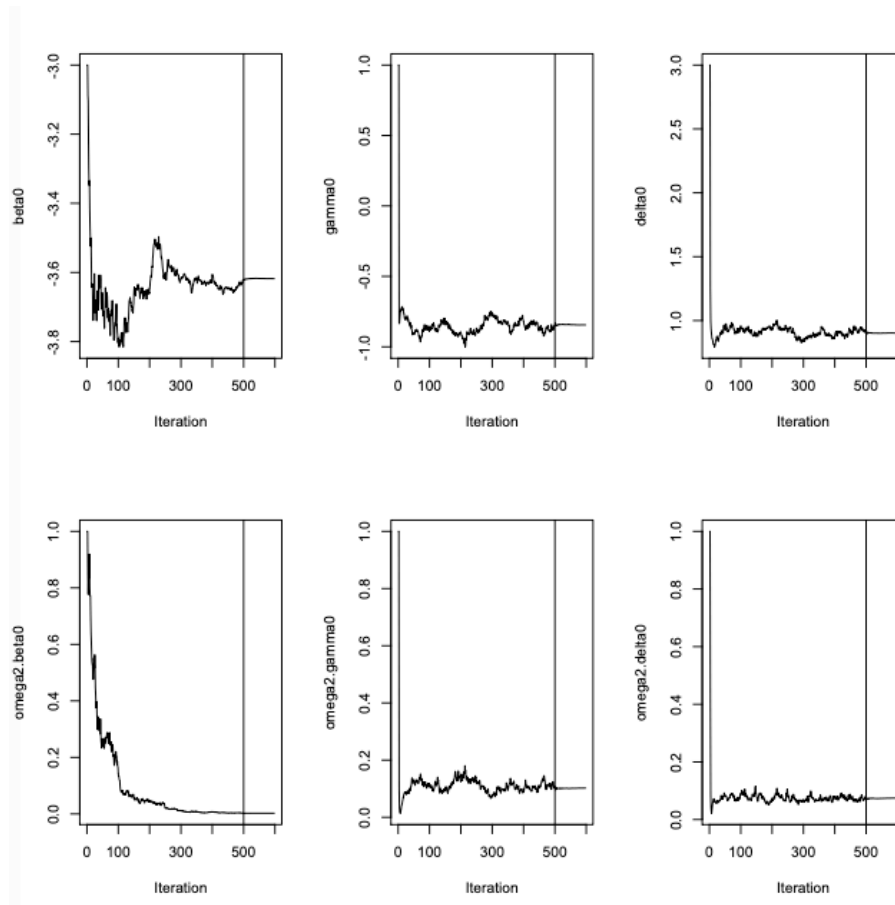
### Create the Model

saemix models are contained in a R function with one blocks:

```

timetoevent.model<-function(psi,id,xidep) {
  T<-xidep[,1]

```



```

N <- nrow(psi)
Nj <- length(T)
censoringtime = 20
lambda <- psi[id,1]
beta <- psi[id,2]
init <- which(T== 0)
cens <- which(T== censoringtime)
ind <- setdiff(1:Nj, append(init,cens))
hazard <- (beta/lambda)*(T/lambda)^(beta-1)
H <- (T/lambda)^beta
logpdf <- rep(0,Nj)
logpdf[cens] <- -H[cens] + H[cens-1]
logpdf[ind] <- -H[ind] + H[ind-1] + log(hazard[ind])
return(logpdf) }

saemix.model<-saemixModel(model=timetoevent.model,description="time model",
  type="likelihood", psi0=matrix(c(2,1),ncol=2,byrow=TRUE,
  dimnames=list(NULL, c("lambda","beta"))), transform.par=c(1,1),
  covariance.model=matrix(c(1,0,0,1),ncol=2, byrow=TRUE))

```

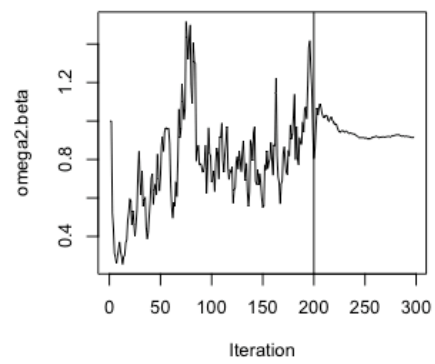
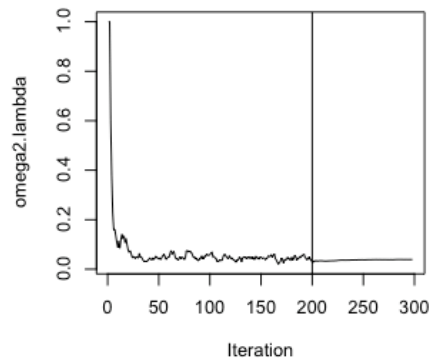
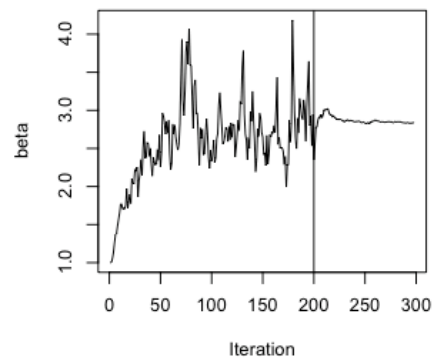
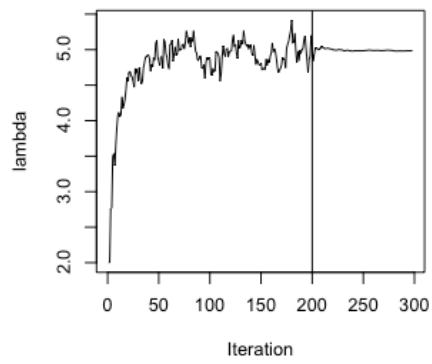
#### Run the SAEM algorithm

```

K1 = 200
K2 = 100

saemix.options<-list(map=F,fim=F,ll.is=F, nb.chains = 1,
  nbiter.saemix = c(K1,K2),displayProgress=TRUE,save.graphs=FALSE)
saemix.fit<-saemix(model,saemix.data,saemix.options)

```



# Contacts

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Please address any questions, bug notice or suggestions.



# Bibliography

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