

# Saemix 3 - time-to-event data models

Emmanuelle

09/2022

## Version

Use saemix version  $\geq 3.2$

## Objective

Run TTE and RTTE models in **saemix**

This notebook uses additional code from the **saemix** development github, not yet integrated in the package. The *workDir* folder in the next chunk of code points to the folder where the user stored this code, and is needed to run the notebook (*workDir* defaults to the current working directory). Specifically, the notebook loads:

- code for different bootstraps in non-linear mixed effect models (Comets et al. 2021 and submitted)
  - the bootstrap runs have been performed previously and are stored in files to be read
    - \* bootstraps can be run instead by switching the *runBootstrap* variable to TRUE in the first chunk of code
    - \* in the code, the number of bootstraps is set to 10 for speed but we recommend to use at least 200 for a 90% CI.
  - this can be changed in the following change of code by uncommenting the line *nboot<-200* and setting the number of bootstrap samples (this may cause memory issues in **Rstudio** with older machines, if this is the case we recommend executing the code in a separate script)
- code for the MC/AGQ provided by Sebastian Ueckert (Ueckert et al. 2017)
  - again if memory issues arise the code can be run in a separate script.

The current notebook can be executed to create an HTML or PDF output with comments and explanations. A script version containing only the R code is also given as *saemix3\_tteModel.R* in the same folder.

## TTE data

**Data description - lung cancer** The example chosen to illustrate the analysis of time-to-event data in **saemix** is the NCCTG Lung Cancer Data, describing the survival in patients with advanced lung cancer from the North Central Cancer Treatment Group (Loprinzi et al. 1994). Covariates measured in the study include performance scores rating how well the patient can perform usual daily activities. We reformatted the *cancer* dataset provided in the **survival** package in R in SAEM format: patients with missing age, sex, institution or physician assessments were removed from the dataset. Status was recoded as 1 for death and 0 for a censored event, and a censoring column was added to denote whether the patient was dead or alive at the time of the last observation. A line at time=0 was added for all subjects. Finally, subjects were numbered consecutively from 0 to 1.

We can plot the distribution of times as a histogram.

```
data(lung.saemix)
saemix.data<-saemixData(name.data=lung.saemix,header=TRUE,name.group=c("id"),
                        name.predictors=c("time","status","cens"),name.response=c("status"),
```

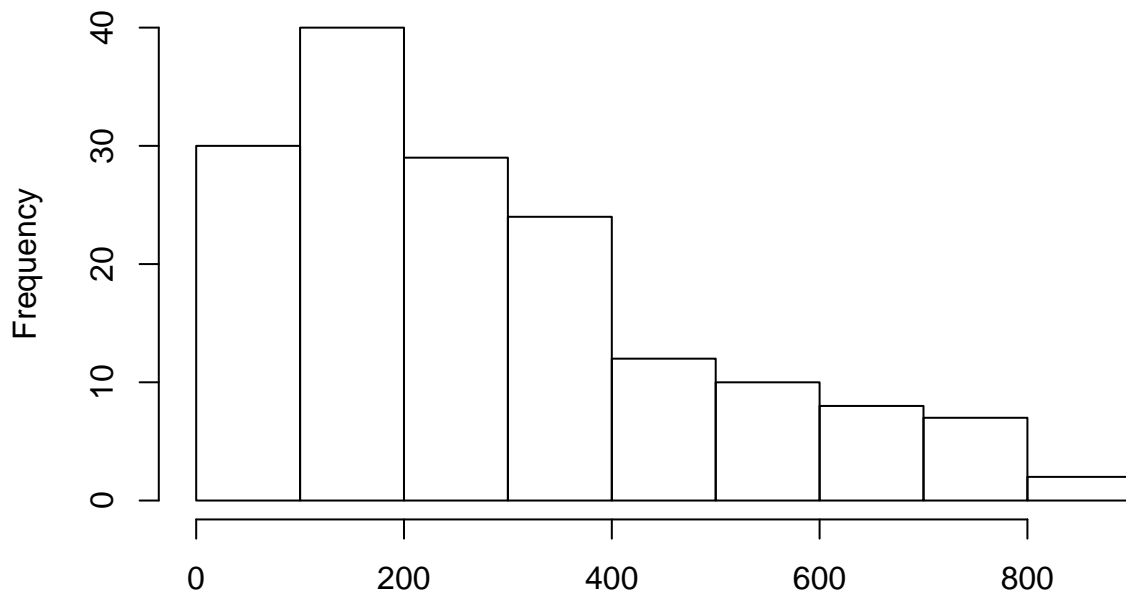
```
name.covariates=c("age", "sex", "ph.ecog", "ph.karno", "pat.karno", "wt.loss", "meal.cal"),
units=list(x="days", y="", covariates=c("yr", "", "-", "%", "%", "cal", "pounds")))
```

```
##
##
## The following SaemixData object was successfully created:
##
## Object of class SaemixData
##   longitudinal data for use with the SAEM algorithm
## Dataset lung.saemix
##   Structured data: status ~ time + status + cens | id
##   X variable for graphs: time (days)
##   covariates: age (yr), sex (), ph.ecog (-), ph.karno (%), pat.karno (%), wt.loss (cal), meal.cal
##   reference class for covariate sex : 0
```

```
# Histogram
```

```
hist(lung.saemix$time[lung.saemix$status==1])
```

### Histogram of lung.saemix\$time[lung.saemix\$status == 1]



lung.saemix\$time[lung.saemix\$status == 1]

```
# Note: missing data in pat.karno, wt.loss and meal.cal
```

```
if(FALSE)
  print(summary(lung.saemix))
```

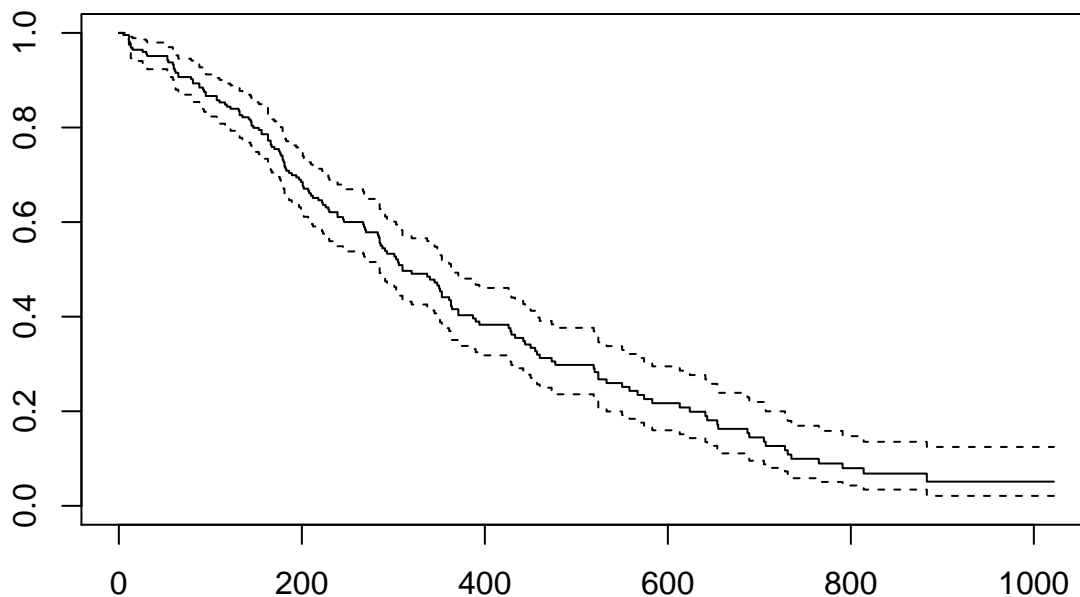
```
lung.surv<-lung.saemix[lung.saemix$time>0,]
lung.surv$status<-lung.surv$status+1
Surv(lung.surv$time, lung.surv$status) # 1=censored, 2=dead
```

Kaplan-Meier plot

```
## [1] 306 455 1010+ 210 883 1022+ 310 361 218 166 170 654
```

```
## [13] 728 567 144 613 707 61 88 301 81 624 371 394
## [25] 520 574 118 390 12 473 26 533 107 53 122 814
## [37] 965+ 93 731 460 153 433 145 583 95 303 519 643
## [49] 765 735 189 53 246 689 65 5 132 687 345 444
## [61] 223 175 60 163 65 208 821+ 428 230 840+ 305 11
## [73] 132 226 426 705 363 11 176 791 95 196+ 167 806+
## [85] 284 641 147 740+ 163 655 239 88 245 588+ 30 179
## [97] 310 477 166 559+ 450 364 107 177 156 529+ 11 429
## [109] 351 15 181 283 201 524 13 212 524 288 363 442
## [121] 199 550 54 558 207 92 60 551+ 543+ 293 202 353
## [133] 511+ 267 511+ 371 387 457 337 201 404+ 222 62 458+
## [145] 356+ 353 163 31 340 229 444+ 315+ 182 156 364+ 291
## [157] 179 376+ 384+ 268 292+ 142 413+ 266+ 194 320 181 285
## [169] 301+ 348 197 382+ 303+ 296+ 180 186 145 269+ 300+ 284+
## [181] 350 272+ 292+ 332+ 285 259+ 110 286 270 81 131 225+
## [193] 269 225+ 243+ 279+ 276+ 135 79 59 240+ 202+ 235+ 224+
## [205] 239 237+ 173+ 252+ 221+ 185+ 92+ 13 222+ 192+ 183 211+
## [217] 175+ 197+ 203+ 116 188+ 191+ 105+ 174+ 177+
```

```
nonpar.fit <- survfit(Surv(time, status) ~ 1, data = lung.surv)
plot(nonpar.fit)
```



**Model for TTE data** We can use a Weibull model for the hazard, parameterised as  $\lambda$  and  $\beta$ . For individual  $i$ , the hazard function of this model is:

$$h(t) = \frac{\beta}{\lambda} \left( \frac{t}{\lambda} \right)^{\beta-1}$$

And the parametric survival function is given by:

$$S(t) = e^{-\left(\frac{t}{\lambda}\right)^\beta}$$

```
weibulltte.model <- function(psi, id, xidep) {
  T <- xidep[, 1]
  y <- xidep[, 2] # events (1=event, 0=no event)
  cens <- which(xidep[, 3] == 1) # censoring times (subject specific)
```

```

init <- which(T==0)
lambda <- psi[id,1] # Parameters of the Weibull model
beta <- psi[id,2]
Nj <- length(T)

ind <- setdiff(1:Nj, append(init,cens)) # indices of events
hazard <- (beta/lambda)*(T/lambda)^(beta-1) # ln(H')
H <- (T/lambda)^beta # ln(H)
logpdf <- rep(0,Nj) # ln(l(T=0))=0
logpdf[cens] <- -H[cens] + H[cens-1] # ln(l(T=censoring time))
logpdf[ind] <- -H[ind] + H[ind-1] + log(hazard[ind]) # ln(l(T=event time))
return(logpdf)
}

saemix.model<-saemixModel(model=weibulltte.model,description="time model",modeltype="likelihood",
  psi0=matrix(c(1,2),ncol=2,byrow=TRUE,dimnames=list(NULL, c("lambda","beta"))),
  transform.par=c(1,1),covariance.model=matrix(c(1,0,0,0),ncol=2, byrow=TRUE))

##
##
## The following SaemixModel object was successfully created:
##
## Nonlinear mixed-effects model
## Model function: time model
## Model type: likelihood
## function(psi,id,xidep) {
## T<-xidep[,1]
## y<-xidep[,2] # events (1=event, 0=no event)
## cens<-which(xidep[,3]==1) # censoring times (subject specific)
## init <- which(T==0)
## lambda <- psi[id,1] # Parameters of the Weibull model
## beta <- psi[id,2]
## Nj <- length(T)
##
## ind <- setdiff(1:Nj, append(init,cens)) # indices of events
## hazard <- (beta/lambda)*(T/lambda)^(beta-1) # ln(H')
## H <- (T/lambda)^beta # ln(H)
## logpdf <- rep(0,Nj) # ln(l(T=0))=0
## logpdf[cens] <- -H[cens] + H[cens-1] # ln(l(T=censoring time))
## logpdf[ind] <- -H[ind] + H[ind-1] + log(hazard[ind]) # ln(l(T=event time))
## return(logpdf)
## }
## Nb of parameters: 2
## parameter names: lambda beta
## distribution:
## Parameter Distribution Estimated
## [1,] lambda log-normal Estimated
## [2,] beta log-normal Estimated
## Variance-covariance matrix:
## lambda beta
## lambda 1 0
## beta 0 0
## No covariate in the model.
## Initial values

```

```

##          lambda beta
## Pop.CondInit      1    2

saemix.options<-list(seed=632545,save=FALSE,save.graphs=FALSE, displayProgress=FALSE)
tte.fit<-saemix(saemix.model,saemix.data,saemix.options)

## Nonlinear mixed-effects model fit by the SAEM algorithm
## -----
## ----          Data          ----
## -----
## Object of class SaemixData
##   longitudinal data for use with the SAEM algorithm
## Dataset lung.saemix
##   Structured data: status ~ time + status + cens | id
##   X variable for graphs: time (days)
##   covariates: age (yr), sex (), ph.ecog (-), ph.karno (%), pat.karno (%), wt.loss (cal), meal.cal
##   reference class for covariate sex : 0
## Dataset characteristics:
##   number of subjects:      225
##   number of observations: 450
##   average/min/max nb obs: 2.00 / 2 / 2
## First 10 lines of data:
##   id time status cens status.1 age sex ph.ecog ph.karno pat.karno wt.loss
## 1  1    0      0  0          0 74  0      1      90      100      NA
## 2  1 306      1  0          1 74  0      1      90      100      NA
## 3  2    0      0  0          0 68  0      0      90       90      15
## 4  2 455      1  0          1 68  0      0      90       90      15
## 5  3    0      0  0          0 56  0      0      90       90      15
## 6  3 1010     0  1          0 56  0      0      90       90      15
## 7  4    0      0  0          0 57  0      1      90       60      11
## 8  4 210      1  0          1 57  0      1      90       60      11
## 9  5    0      0  0          0 60  0      0     100       90       0
## 10 5 883      1  0          1 60  0      0     100       90       0
##   meal.cal mdv cens.1 occ ytype
## 1      1175  0      0  1      1
## 2      1175  0      0  1      1
## 3      1225  0      0  1      1
## 4      1225  0      0  1      1
## 5         NA  0      0  1      1
## 6         NA  0      0  1      1
## 7      1150  0      0  1      1
## 8      1150  0      0  1      1
## 9         NA  0      0  1      1
## 10        NA  0      0  1      1
## -----
## ----          Model          ----
## -----
## Nonlinear mixed-effects model
##   Model function:  time model
##   Model type:     likelihood
## function(psi,id,xidep) {
##   T<-xidep[,1]
##   y<-xidep[,2] # events (1=event, 0=no event)
##   cens<-which(xidep[,3]==1) # censoring times (subject specific)
##   init <- which(T==0)

```

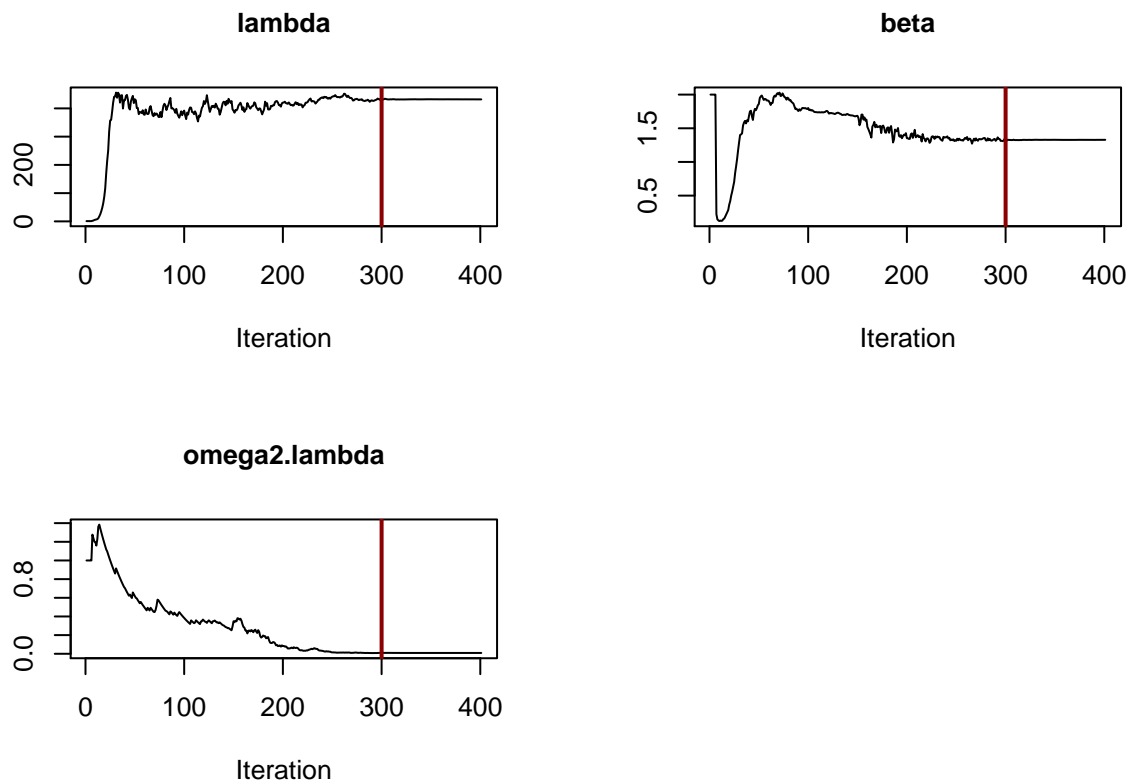
```

## lambda <- psi[id,1] # Parameters of the Weibull model
## beta <- psi[id,2]
## Nj <- length(T)
##
## ind <- setdiff(1:Nj, append(init,cens)) # indices of events
## hazard <- (beta/lambda)*(T/lambda)^(beta-1) # ln(H')
## H <- (T/lambda)^beta # ln(H)
## logpdf <- rep(0,Nj) # ln(l(T=0))=0
## logpdf[cens] <- -H[cens] + H[cens-1] # ln(l(T=censoring time))
## logpdf[ind] <- -H[ind] + H[ind-1] + log(hazard[ind]) # ln(l(T=event time))
## return(logpdf)
## }
## <bytecode: 0x557afccda190>
## Nb of parameters: 2
## parameter names: lambda beta
## distribution:
## Parameter Distribution Estimated
## [1,] lambda log-normal Estimated
## [2,] beta log-normal Estimated
## Variance-covariance matrix:
## lambda beta
## lambda 1 0
## beta 0 0
## No covariate in the model.
## Initial values
## lambda beta
## Pop.CondInit 1 2
## -----
## ---- Key algorithm options ----
## -----
## Estimation of individual parameters (MAP)
## Estimation of standard errors and linearised log-likelihood
## Estimation of log-likelihood by importance sampling
## Number of iterations: K1=300, K2=100
## Number of chains: 1
## Seed: 632545
## Number of MCMC iterations for IS: 5000
## Simulations:
## nb of simulated datasets used for npde: 1000
## nb of simulated datasets used for VPC: 100
## Input/output
## save the results to a file: FALSE
## save the graphs to files: FALSE
## -----
## ---- Results ----
## -----
## ----- Fixed effects -----
## -----
## Parameter Estimate SE CV(%)
## [1,] lambda 431.8 51.60 12
## [2,] beta 1.3 0.19 14
## -----
## ----- Variance of random effects -----
## -----

```

```
##      Parameter      Estimate SE   CV(%)
## lambda omega2.lambda 0.009   0.17 1858
## -----
## ----- Correlation matrix of random effects -----
## -----
##              omega2.lambda
## omega2.lambda 1
## -----
## ----- Statistical criteria -----
## -----
## Likelihood computed by linearisation
##      -2LL= 5189.352
##      AIC = 5197.352
##      BIC = 5211.017
##
## Likelihood computed by importance sampling
##      -2LL= 2269.357
##      AIC = 2277.357
##      BIC = 2291.021
## -----
```

```
plot(tte.fit, plot.type="convergence")
```



**Simulation function** Simulating from a TTE model is slightly more complicated than for the other non Gaussian models. When the hazard function has an inverse, we can use the inverse CDF technique (or inverse transformation algorithm) for generating a random sample. The method uses the fact that a continuous cumulative density function,  $F$ , is a one-to-one mapping of the domain of the cdf into the interval  $(0,1)$ . Therefore, if  $U$  is a uniform random variable on  $(0,1)$ , then  $X = F^{-1}(U)$  has the distribution  $F$ .

For the single event Weibull model:

$$F = 1 - e^{-\int_0^T h(u)du} = 1 - e^{-\left(\frac{T}{\lambda}\right)^\beta} \sim \mathcal{U}(0,1)$$

Assuming we simulate  $U = 1 - V$  from  $\mathcal{U}(0,1)$ , we can obtain a sample from the Weibull parametric model as:

$$T = \lambda \left( -\ln(V) + \left( \frac{T}{\lambda} \right)^\beta \right)^{1/\beta}$$

In the following we assume the first column of *xidep* contains the observed times, and that there is a common censoring time (the maximum observed time). We could also assume a common censoring (function *simulateWeibullTTE.maxcens()* below) but simulating from this function shows an excess of times simulated at the censoring limit compared to the original dataset.

```
# Simulate events based on the observed individual censoring time
simulateWeibullTTE <- function(psi,id,xidep) {
  T<-xidep[,1]
  y<-xidep[,2] # events (1=event, 0=no event)
  cens<-which(xidep[,3]==1) # censoring times (subject specific)
  init <- which(T==0)
  lambda <- psi[,1] # Parameters of the Weibull model
  beta <- psi[,2]
  Nj <- length(T)
  ind <- setdiff(1:Nj, append(init,cens)) # indices of events
  tevent<-T
  Vj<-runif(dim(psi)[1])
  tsim<-lambda*(-log(Vj))^(1/beta) # nsuj events
  tevent[T>0]<-tsim
  tevent[tevent[cens]>T[cens]] <- T[tevent[cens]>T[cens]]
  return(tevent)
}

# Checking the simulation function
xidep1<-saemix.data@data[,saemix.data@name.predictors]
nsuj<-saemix.data@N
psiM<-data.frame(lambda=rnorm(nsuj, mean=tte.fit@results@fixed.effects[1], sd=2), beta=tte.fit@results@
id1<-rep(1:nsuj, each=2)
simtime<-simulateWeibullTTE(psiM, id1, xidep1)

par(mfrow=c(1,2))
hist(saemix.data@data$time[saemix.data@data$time>0], breaks=30, xlab="Time", main="Original data")
hist(simtime[simtime>0], breaks=30, xlim=c(0,1000), xlab="Time", main="Simulated data")

# Ignoring the cens column and assuming a common censoring time instead
simulateWeibullTTE.maxcens <- function(psi,id,xidep) {
  etime<-xidep[,1]
  censoringtime <- max(etime)
  lambda <- psi[,1]
  beta <- psi[,2]
  N<-dim(psi)[1]
  Vj<-runif(N)
  T<-lambda*(-log(Vj))^(1/beta)
  T[T>censoringtime]<-censoringtime
  etime[etime>0]<-T
}
```

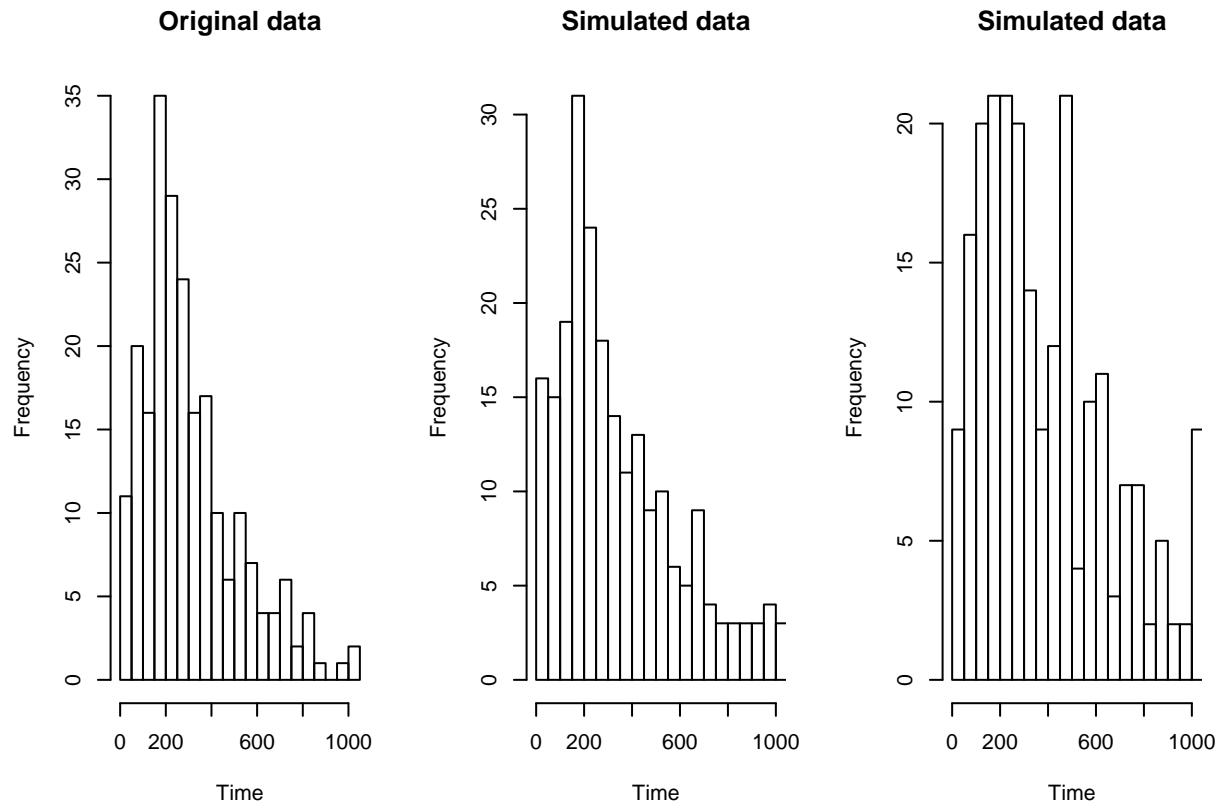


```

    return(etime)
}
simtime.maxcens<-simulateWeibullTTE.maxcens(psiM, id1, xidep1)

par(mfrow=c(1,3))
hist(saemix.data@data$time[saemix.data@data$time>0], breaks=30, xlab="Time", main="Original data")
hist(simtime[simtime>0], breaks=30, xlim=c(0,1000), xlab="Time", main="Simulated data")
hist(simtime.maxcens[simtime.maxcens>0], breaks=30, xlim=c(0,1000), xlab="Time", main="Simulated data")

```



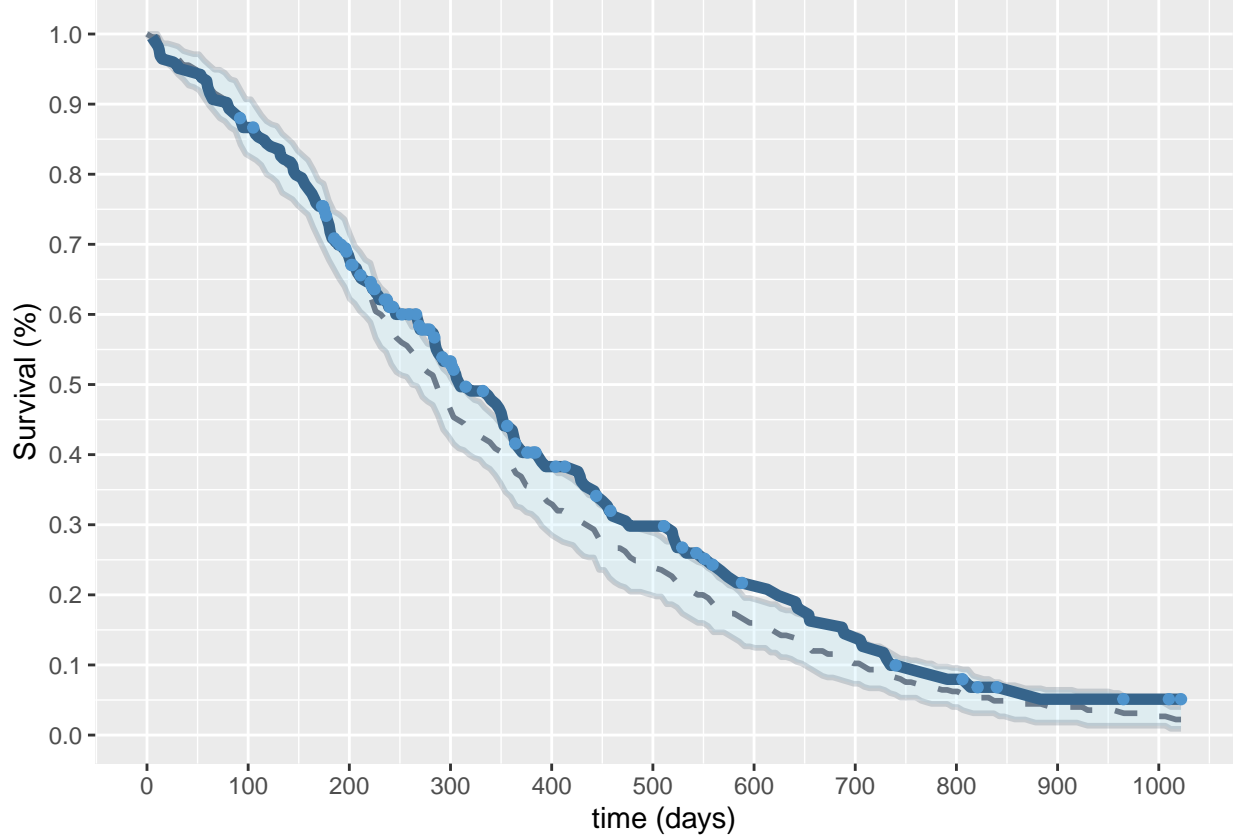
We then use the simulation function defined above to simulate from the fitted model and plot VPC.

```

tte.fit@model@simulate.function <- simulateWeibullTTE
simtte.fit <- simulateDiscreteSaemix(tte.fit, nsim=100)

gpl <- discreteVPC(simtte.fit, outcome="TTE")
plot(gpl)

```



Note that there are some specialised packages such as the **survsim** and the **simsurv** package that could be leveraged for this exercise.

Exact FIM by AGQ (code by Sebastian Ueckert)

For non-Gaussian models, the exact FIM should be computed, and two approaches have been proposed using either numerical integration by a combination of MC and adaptive Gaussian quadrature (MC/AGQ, Ueckert et al 2017) or stochastic integration by MCMC (Rivière et al. 2017).

Both these approaches are computationally intensive.

Here we use code provided by Sebastian Ueckert implementing the MC/AGQ approach, as the MCMC requires the installation of rStan. In this approach, the information matrix (FIM) over the population is first decomposed the sum of the individual FIM:

$$FIM(\Psi, \Xi) = \sum_{i=1}^N FIM(\Psi, \xi_i)$$

where  $\xi_i$  denotes the individual design in subject  $i$ . Assuming  $Q$  different elementary designs, the FIM can also be summed over the different designs weighted by the number of subjects  $N_q$  in design  $q$  as:

$$FIM(\Psi, \Xi) = \sum_{q=1}^Q N_q FIM(\Psi, \xi_q)$$

In the following, we first load the functions needed to compute the exact FIM. We then define a model object with the following components:

- *parameter\_function*: a function returning the list of parameters as the combination of fixed and random effects

- *log\_likelihood\_function*: using the parameters, computes the log-likelihood for all y in the dataset
- *simulation\_function*: using the parameters, computes the log-likelihood and produces a random sample from the corresponding distribution
- *inverse\_simulation\_function*: supposed to be the quantile function but not quite sure :-/ (here, returns the category in which is urand)
- *mu*: the fixed parameters
- *omega*: the variance-covariance matrix

For *mu* and *omega*, we use the results from the saemix fit.

TODO ?

```
# Code Sebastian
source(file.path(dirAGQ,"default_settings.R"))
source(file.path(dirAGQ,"helper_functions.R"))
source(file.path(dirAGQ,"integration.R"))
source(file.path(dirAGQ,"model.R"))

saemix.fit <- tte.fit
```

## Diagnostics

**Comparison to the KM fit** With TTE data the First-Order approximation for the FIM doesn't seem to perform too badly. We can use the delta-method to obtain standard errors around the value of the survival function, using the following vector of derivatives:

$$\begin{pmatrix} \frac{\delta S}{\delta \lambda} \\ \frac{\delta S}{\delta \beta} \end{pmatrix} = \begin{pmatrix} \frac{\beta}{\lambda} \left(\frac{t}{\lambda}\right)^{\beta} e^{-\left(\frac{t}{\lambda}\right)^{\beta}} \\ -\ln\left(\frac{t}{\lambda}\right) \left(\frac{t}{\lambda}\right)^{\beta} e^{-\left(\frac{t}{\lambda}\right)^{\beta}} \end{pmatrix}$$

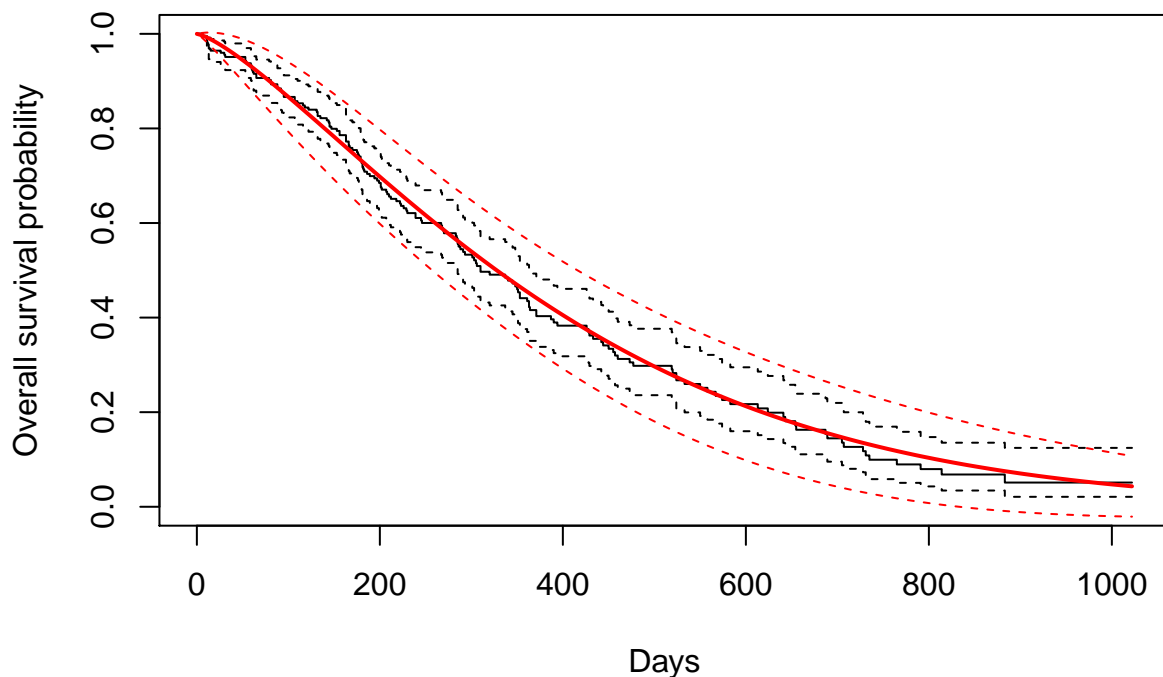
We overlay the parametric fit and its confidence interval in red over the previous non-parametric KM estimate, and find a good concordance between the two.

```
ypred<-predict(tte.fit)

# Use survival package to assess Survival curve
xtim<-seq(0,max(lung.saemix$time), length.out=200)
estpar<-tte.fit@results@fixed.effects
estse<-tte.fit@results@se.fixed
ypred<-exp(-(xtim/estpar[1])^(estpar[2]))

# Computing SE for the survival curve based on linearised FIM (probably not a good idea) through the de
invfim<-solve(tte.fit@results@fim[1:2,1:2])
xcal<- (xtim/estpar[1])^estpar[2]
dsdbeta<- -log(xtim/estpar[1]) * xcal *exp(-xcal)
dsdalpha<- estpar[2]/estpar[1] * xcal *exp(-xcal)
xmat<-rbind(dsdalpha, dsdbeta)
# x1<-t(xmat[,1:3]) %*% invfim %*% xmat[,1:3]
sesurv<-rep(0,length(xcal))
for(i in 1:length(xcal))
  sesurv[i]<-sqrt(t(xmat[,i]) %*% invfim %*% xmat[,i])

# Comparison between KM and parametric fit
plot(nonpar.fit, xlab = "Days", ylab = "Overall survival probability")
lines(xtim,ypred, col="red",lwd=2)
lines(xtim,ypred+1.96*sesurv, col="red",lwd=1, lty=2)
lines(xtim,ypred-1.96*sesurv, col="red",lwd=1, lty=2)
```



```
# Adding the simulation function to the model component of the fit
tte.fit@model@simulate.function <- simulateWeibullTTE
ysim.tte <- simulateDiscreteSaemix(tte.fit, nsim=100)
```

## VPC TODO

**VPC using Ron's package** A recent package was developed by Ron Keizer to implement VPC for different types of data. For survival data, we can use the `vpc_tte()` function from this package to produce the KM-VPC plot.

```
library(vpc)

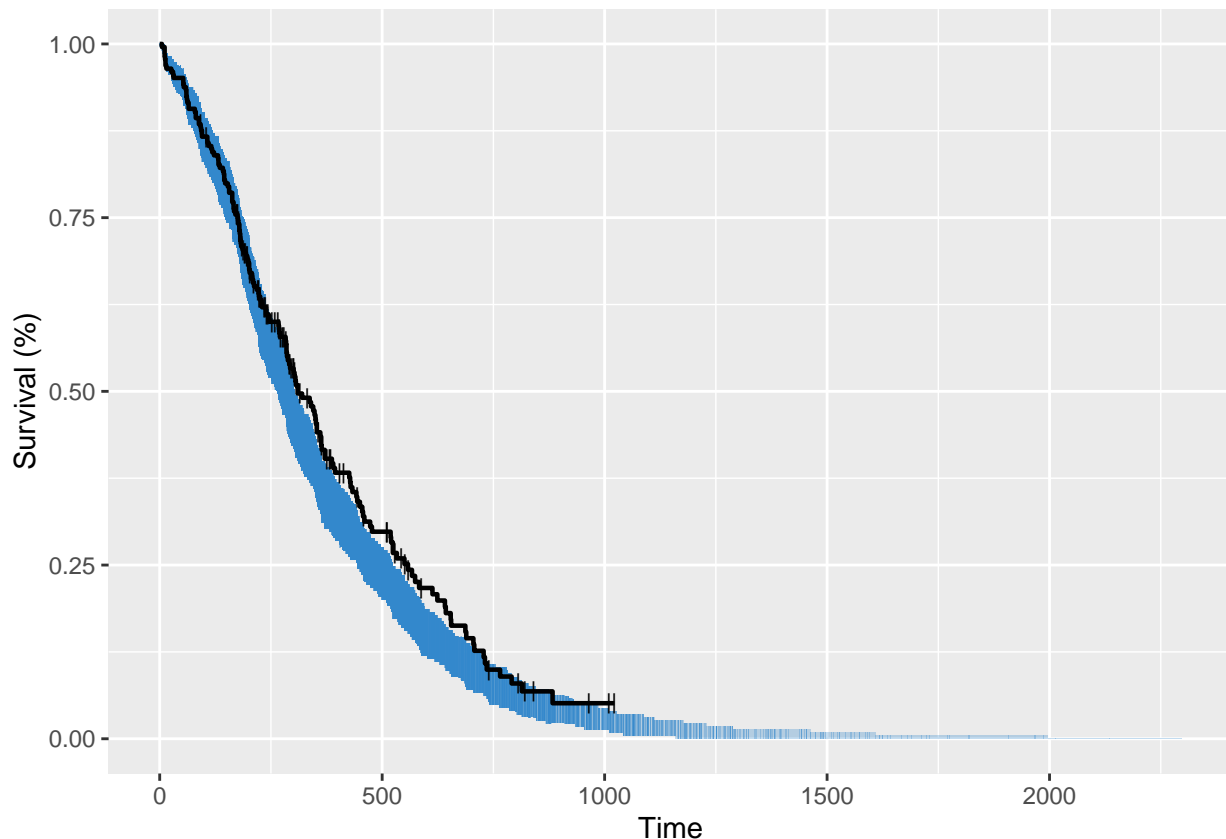
n<-dim(ysim.tte@sim.data@datasim)[1]
simron<-cbind(ysim.tte@sim.data@datasim,dv=rep(c(0,1), n/2))
colnames(simron)<-c("id","sim","time","dv")

vpc_tte(sim = simron,
        obs = lung.saemix,
        # rtte = FALSE,
        sim_cols=list(id="id", dv = "dv", idv = "time"), obs_cols=list(id="id", dv="status", idv = "time"))

## Initializing.
## Detected column 'cens' with censoring information in observation data, assuming 1=censored event, 0=
## Warning: `group_by()` was deprecated in dplyr 0.7.0.
## Please use `group_by()` instead.
## See vignette('programming') for more help
## This warning is displayed once every 8 hours.
## Call `lifecycle::last_lifecycle_warnings()` to see where this warning was generated.
## Calculating simulation stats.
```

```
## Warning: `arrange_()` was deprecated in dplyr 0.7.0.
## Please use `arrange()` instead.
## See vignette('programming') for more help
## This warning is displayed once every 8 hours.
## Call `lifecycle::last_lifecycle_warnings()` to see where this warning was generated.

## =====
##
## Plotting.
## Warning: Removed 1 rows containing missing values (geom_rect).
```



```
if(FALSE) { # Setting late simulated times to the maximum event time (a censored event)
  simron$time[simron$time>max(lung.saemix$time)]<-max(lung.saemix$time)
  vpc_tte(sim = simron,
    obs = lung.saemix,
    #       rtte = FALSE,
    sim_cols=list(id="id", dv = "dv", idv = "time"), obs_cols=list(id="id", dv="status", idv = "t
}
```

## RTTE model

In this section we simulate repeated time-to-event data from a Weibull model and fit it. To simulate from a RTTE model, we simulate repeated events starting from the previous one using the inverse CDF technique. Because we don't know in advance the number of events in each subject, we lose the efficient vectorisation from **R** and this function can be considerably slower than the single event TTE.

```
# Simulating RTTE data by simulating from U(0,1) and inverting the cdf
simul.rtte.unif<-function(psi) { # xidexp, id not important, we only use psi
```

```

censoringtime <- 3
maxevents <- 30
lambda <- psi[,1]
beta <- psi[,2]
simdat<-NULL
N<-nrow(psi)
for(i in 1:N) {
  eventTimes<-c(0)
  T<-0
  Vj<-runif(1)
  # T <- (-log(Vj)*lambda[i])^(beta[i])
  T<-lambda[i]*(-log(Vj))^(1/beta[i])
  nev<-0
  while (T < censoringtime & nev<maxevents){
    eventTimes <- c(eventTimes, T)
    nev<-nev+1
    Vj<-runif(1)
    # T <- T+(-log(Vj)*lambda[i])^(beta[i])
    # T<-(-log(Vj)*lambda[i] + T^(1/beta[i]))^(beta[i])
    T<-lambda[i]*(-log(Vj) + (T/lambda[i])^(beta[i]))^(1/beta[i])
  }
  if(nev==maxevents) {
    message("Reached maximum number of events\n")
  }
  eventTimes<-c(eventTimes, censoringtime)
  cens<-rep(1,length(eventTimes))
  cens[1]<-cens[length(cens)]<-0
  simdat<-rbind(simdat,
                data.frame(id=i, T=eventTimes, status=cens))
}
return(simdat)
}

# Subjects
set.seed(12345)
param<-c(2, 1.5, 0.5)
# param<-c(4, 1.2, 0.3)
omega<-c(0.25,0.25)
nsuj<-200
risk<-rep(0,nsuj)
risk[(nsuj/2+1):nsuj]<-1
psiM<-data.frame(lambda=param[1]*exp(rnorm(nsuj,sd=omega[1])), beta=param[2]*exp(param[3]*risk+rnorm(nsuj,sd=omega[2])),
simdat <- simul.rtte.unif(psiM)

## Reached maximum number of events
simdat$risk<-as.integer(simdat$id>(nsuj/2))

saemix.data<-saemixData(name.data=simdat, name.group=c("id"), name.predictors=c("T"), name.response="status")

##
##
## The following SaemixData object was successfully created:
##
## Object of class SaemixData

```

```

##      longitudinal data for use with the SAEM algorithm
## Dataset simdat
##      Structured data: status ~ T | id
##      Predictor: T ()
##      covariates: risk (-)
##      reference class for covariate risk : 0

rtte.model<-function(psi,id,xidep) {
  T<-xidep[,1]
  N <- nrow(psi) # nb of subjects
  Nj <- length(T) # nb of events (including 0 and censoring times)
  # censoringtime = 6
  censoringtime = max(T) # same censoring for everyone
  lambda <- psi[id,1]
  beta <- psi[id,2]
  tinit <- which(T==0) # indices of beginning of observation period
  tcens <- which(T==censoringtime) # indices of censored events
  tevent <- setdiff(1:Nj, append(tinit,tcens)) # indices of non-censored event times
  hazard <- (beta/lambda)*(T/lambda)^(beta-1)
  H <- (T/lambda)^beta
  logpdf <- rep(0,Nj)
  logpdf[tcens] <- -H[tcens] + H[tcens-1]
  logpdf[tevent] <- -H[tevent] + H[tevent-1] + log(hazard[tevent])
  return(logpdf)
}

saemix.model.base<-saemixModel(model=rtte.model,description="Repeated TTE model",modeltype="likelihood",
                                psi0=matrix(c(1,2),ncol=2,byrow=TRUE,dimnames=list(NULL, c("lambda","beta")),
                                transform.par=c(1,1),covariance.model=matrix(c(1,0,0,1),ncol=2, byrow=TRUE))

##
##
## The following SaemixModel object was successfully created:
##
## Nonlinear mixed-effects model
## Model function: Repeated TTE model
## Model type: likelihood
## function(psi,id,xidep) {
##   T<-xidep[,1]
##   N <- nrow(psi) # nb of subjects
##   Nj <- length(T) # nb of events (including 0 and censoring times)
##   # censoringtime = 6
##   censoringtime = max(T) # same censoring for everyone
##   lambda <- psi[id,1]
##   beta <- psi[id,2]
##   tinit <- which(T==0) # indices of beginning of observation period
##   tcens <- which(T==censoringtime) # indices of censored events
##   tevent <- setdiff(1:Nj, append(tinit,tcens)) # indices of non-censored event times
##   hazard <- (beta/lambda)*(T/lambda)^(beta-1)
##   H <- (T/lambda)^beta
##   logpdf <- rep(0,Nj)
##   logpdf[tcens] <- -H[tcens] + H[tcens-1]
##   logpdf[tevent] <- -H[tevent] + H[tevent-1] + log(hazard[tevent])
##   return(logpdf)
## }

```

```

## Nb of parameters: 2
##      parameter names:  lambda beta
##      distribution:
##      Parameter Distribution Estimated
## [1,] lambda    log-normal    Estimated
## [2,] beta      log-normal    Estimated
## Variance-covariance matrix:
##      lambda beta
## lambda      1    0
## beta        0    1
##      No covariate in the model.
##      Initial values
##      lambda beta
## Pop.CondInit      1    2

saemix.model<-saemixModel(model=rtte.model,description="Repeated TTE model",modeltype="likelihood",
                          psi0=matrix(c(1,2),ncol=2,byrow=TRUE,dimnames=list(NULL, c("lambda","beta"))),
                          transform.par=c(1,1),covariate.model=matrix(c(0,1),ncol=2),
                          covariance.model=matrix(c(1,0,0,1),ncol=2, byrow=TRUE))

##
##
## The following SaemixModel object was successfully created:
##
## Nonlinear mixed-effects model
## Model function:  Repeated TTE model
## Model type:  likelihood
## function(psi,id,xidep) {
##   T<-xidep[,1]
##   N <- nrow(psi) # nb of subjects
##   Nj <- length(T) # nb of events (including 0 and censoring times)
##   # censoringtime = 6
##   censoringtime = max(T) # same censoring for everyone
##   lambda <- psi[id,1]
##   beta <- psi[id,2]
##   tinit <- which(T==0) # indices of beginning of observation period
##   tcens <- which(T==censoringtime) # indices of censored events
##   tevent <- setdiff(1:Nj, append(tinit,tcens)) # indices of non-censored event times
##   hazard <- (beta/lambda)*(T/lambda)^(beta-1)
##   H <- (T/lambda)^beta
##   logpdf <- rep(0,Nj)
##   logpdf[tcens] <- -H[tcens] + H[tcens-1]
##   logpdf[tevent] <- -H[tevent] + H[tevent-1] + log(hazard[tevent])
##   return(logpdf)
## }
## Nb of parameters: 2
##      parameter names:  lambda beta
##      distribution:
##      Parameter Distribution Estimated
## [1,] lambda    log-normal    Estimated
## [2,] beta      log-normal    Estimated
## Variance-covariance matrix:
##      lambda beta
## lambda      1    0
## beta        0    1

```



```

## Covariate model:
##      lambda beta
## [1,]      0      1
##      Initial values
##      lambda beta
## Pop.CondInit      1      2
## Cov.CondInit      0      0

saemix.options<-list(seed=632545,save=FALSE,save.graphs=FALSE, fim=FALSE, displayProgress=FALSE)
rtte.fit<-saemix(saemix.model,saemix.data,saemix.options)

## Nonlinear mixed-effects model fit by the SAEM algorithm
## -----
## ----          Data          ----
## -----
## Object of class SaemixData
##      longitudinal data for use with the SAEM algorithm
## Dataset simdat
##      Structured data: status ~ T | id
##      Predictor: T ()
##      covariates: risk (-)
##      reference class for covariate risk : 0
## Dataset characteristics:
##      number of subjects:      200
##      number of observations: 967
##      average/min/max nb obs: 4.83 / 2 / 32
## First 10 lines of data:
##      id          T status risk mdv cens occ ytype
## 1  1 0.0000000      0    0  0    0  1      1
## 2  1 0.7520145      1    0  0    0  1      1
## 3  1 0.8775847      1    0  0    0  1      1
## 4  1 2.4331650      1    0  0    0  1      1
## 5  1 3.0000000      0    0  0    0  1      1
## 6  2 0.0000000      0    0  0    0  1      1
## 7  2 1.3712351      1    0  0    0  1      1
## 8  2 3.0000000      0    0  0    0  1      1
## 9  3 0.0000000      0    0  0    0  1      1
## 10 3 2.8564910      1    0  0    0  1      1
## -----
## ----          Model          ----
## -----
## Nonlinear mixed-effects model
##      Model function: Repeated TTE model
##      Model type: likelihood
## function(psi,id,xidep) {
##      T<-xidep[,1]
##      N <- nrow(psi) # nb of subjects
##      Nj <- length(T) # nb of events (including 0 and censoring times)
##      # censoringtime = 6
##      censoringtime = max(T) # same censoring for everyone
##      lambda <- psi[id,1]
##      beta <- psi[id,2]
##      tinit <- which(T==0) # indices of beginning of observation period
##      tcens <- which(T==censoringtime) # indices of censored events
##      tevent <- setdiff(1:Nj, append(tinit,tcens)) # indices of non-censored event times

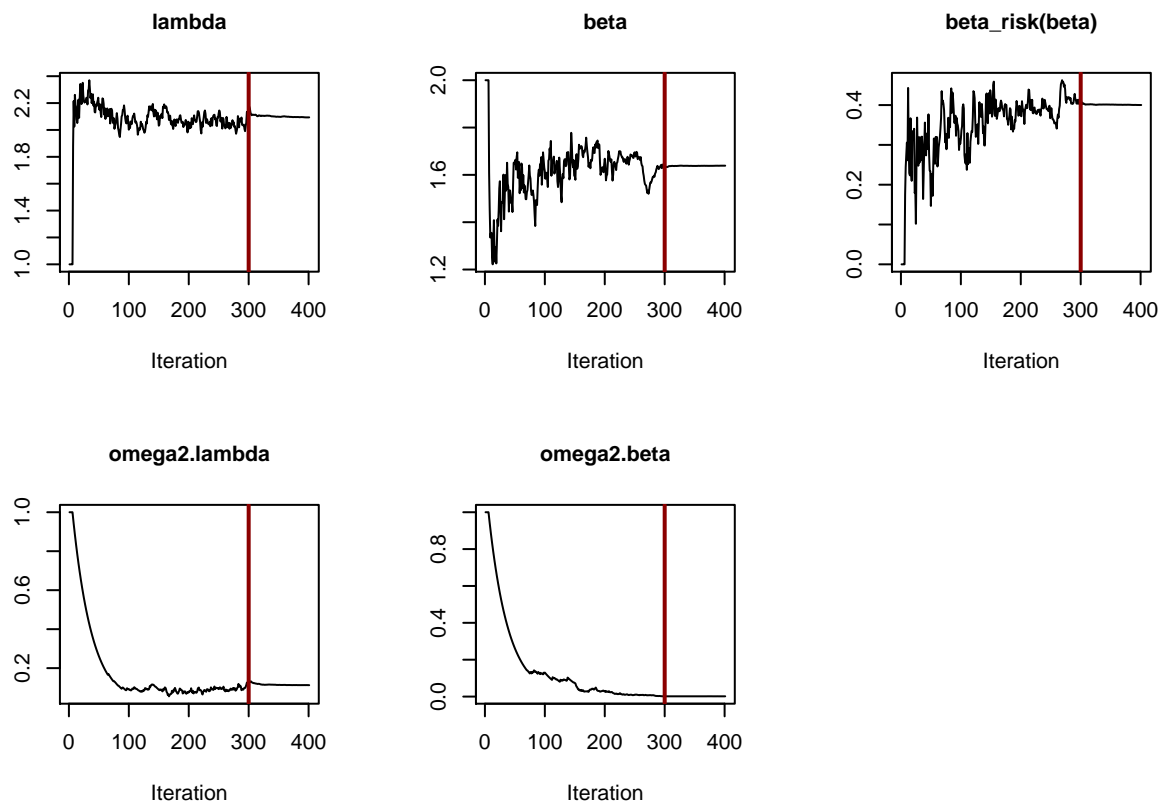
```

```

## hazard <- (beta/lambda)*(T/lambda)^(beta-1)
## H <- (T/lambda)^beta
## logpdf <- rep(0,Nj)
## logpdf[tcens] <- -H[tcens] + H[tcens-1]
## logpdf[tevent] <- -H[tevent] + H[tevent-1] + log(hazard[tevent])
## return(logpdf)
## }
## <bytecode: 0x557afcfb90f0>
## Nb of parameters: 2
## parameter names: lambda beta
## distribution:
## Parameter Distribution Estimated
## [1,] lambda log-normal Estimated
## [2,] beta log-normal Estimated
## Variance-covariance matrix:
## lambda beta
## lambda 1 0
## beta 0 1
## Covariate model:
## [,1] [,2]
## risk 0 1
## Initial values
## lambda beta
## Pop.CondInit 1 2
## Cov.CondInit 0 0
## -----
## ---- Key algorithm options ----
## -----
## Estimation of individual parameters (MAP)
## Estimation of log-likelihood by importance sampling
## Number of iterations: K1=300, K2=100
## Number of chains: 1
## Seed: 632545
## Number of MCMC iterations for IS: 5000
## Simulations:
## nb of simulated datasets used for npde: 1000
## nb of simulated datasets used for VPC: 100
## Input/output
## save the results to a file: FALSE
## save the graphs to files: FALSE
## -----
## ---- Results ----
## -----
## ----- Fixed effects -----
## -----
## Parameter Estimate
## [1,] lambda 2.1
## [2,] beta 1.6
## [3,] beta_risk(beta) 0.4
## -----
## ----- Variance of random effects -----
## -----
## Parameter Estimate
## lambda omega2.lambda 0.1125

```

```
## beta      omega2.beta      0.0015
## -----
## ----- Correlation matrix of random effects -----
## -----
##              omega2.lambda omega2.beta
## omega2.lambda 1              0
## omega2.beta   0              1
## -----
## ----- Statistical criteria -----
## -----
##
## Likelihood computed by importance sampling
##      -2LL= 690.2485
##      AIC = 702.2485
##      BIC = 722.0384
## -----
plot(rtte.fit, plot.type="convergence")
```



**Statistical model** A nice review of the more frequent hazard functions used in parametric models of TTE data has recently been van Wijk and Simonsson (*CPT:PSP* 2022), including a Shiny app to explore their shape and how to set initial parameters. These models are very sensitive to the initial parameter estimates and their variance, therefore using

## References

**Comets E**, Rodrigues C, Jullien V, Ursino M (2021). Conditional non-parametric bootstrap for non-linear mixed effect models. *Pharmaceutical Research*, 38: 1057-66.

**Keizer R** (2021). vpc: Create Visual Predictive Checks. *R package* version 1.2.2. <https://CRAN.R-project.org/package=vpc>

**Morina D**, Navarro A (2014). The R package survsim for the simulation of simple and complex survival Data. *Journal of Statistical Software*, 59(2), 1–20.

**Ueckert S**, Mentré F (2017). A new method for evaluation of the Fisher information matrix for discrete mixed effect models using Monte Carlo sampling and adaptive Gaussian quadrature. *Computational Statistics and Data Analysis*, 111: 203-19. 10.1016/j.csda.2016.10.011

**van Wijk R**, Simonsson U (2022). Finding the right hazard function for time-to-event modeling: A tutorial and Shiny application. *Clinical Pharmacokinetics and Therapeutics: Pharmacometrics and Systems Pharmacology*