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Program leave-one-out posterior predictive checking in R

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1 Introduction

2 Theory

2.1 Leave-one-out cross-validation

Leave-one-out cross-validation, or short *loo-cv* is a method for checking the predictive out-of-sample accuracy of a model. The idea is to omit the i -th observation and to fit the model for the remaining observations and then to predict the y_i as an out of sample point. This prediction is denoted as $\hat{y}_i^{(i)}$. Thus, we can overcome – at least to some extent – the problem of overfitting. The loo-cv method can be used in several ways for model evaluation and selection. We will use it in two ways. First, we will use it to calculate the *log pointwise prediction density* using the loo-cv, which is defined as

$$\text{lppd}_{\text{loo-cv}} = \sum_{i=1}^n \log p_{\text{dens}(-i)}(y_i). \quad (2.1)$$

That is, we calculate the densities of all y_i using the parameters we obtain from the n bayesian models, applying the loo-cv method. Then taking the logarithm and summing up. In the case at hand we deal with the normal density, since $y_i \sim \mathcal{N}(X_{(-i)}\beta_{(-i)}, \sigma_{(-i)}^2)$. Furthermore, multiplying with -2 , i.e. $-2 \text{lppd}_{\text{loo-cv}}$, gives us a measure that is comparable with other measures of predictive accuracy (e.g. AIC or WAIC) (Gelman, 2013). Besides the MSE we will use $-2 \text{lppd}_{\text{loo-cv}}$ for model selection within the Bayesian framework.

Second, we will need it in the context of comparing the predictive out-of-sample accuracy of the Bayesian and the frequentist model using Cook's distance.

2.2 Cook's Distance

Cook's Distance, or short *Cook's D*, is a measure to detect influential points in a linear regression. It was invented by Cook (1977) and is defined as

$$D_i = \frac{\sum_{j=1}^n (\hat{y}_j - \hat{y}_j^{(i)})^2}{p \text{ MSE}} \stackrel{(\text{OLS})}{=} \frac{e_i^2}{p \text{ MSE}} \left(\frac{h_{ii}}{(1 - h_{ii})^2} \right), \quad (2.2)$$

where

$$\text{MSE} = \frac{1}{n - p} \sum_{i=1}^n (y_i - \hat{y})^2 \quad (2.3)$$

is the *mean squared error*, $e_i^2 = (y_i - \hat{y})^2$ is the error for each observation i and $h_{ii} = x_i(X^T X)^{-1}x_i^T$ is the i -th diagonal element of the projection matrix of an OLS regression model, also called the *leverage* of an observation, which indicates the influence of an single observation on the model fit.

As above, the $\hat{y}^{(i)}$ is the predicted value for y_i if we exclude the i -th observation and fit the model for the remaining observations. Note that the right hand side term of the model is an analytical expression. In this case there is no need to fit additional n models to get the $\hat{y}^{(i)}$ predictions. This yields to a great advantage in performance, especially for large numbers of observations. But the analytical expression is constraint to OLS models – indicated by the *OLS* over the equation sign – since the projection matrix is used. For this reason we will use the left hand expression to calculate Cook's D in the Bayesian case using the loo-cv method described above and the right hand expression for the frequentist OLS regression model.

3 Code

For both applications of the cross validation, the model evaluation and the calculation of Cook's distance, we need the Gibbs sampler and the leave-one-out cross validation (loo-cv) algorithm. Those two functions are explained in the first part. The second part is about the model evaluation and the third part compares the loo-cv Cook's distance with the analytic solution, which are defined in equation (2.2).

Note that we only displayed the relevant code chunks in the pdf file. Some chunks regarding the plotting are hidden and can be found in the rmd file.

3.1 Basics

The first function we define is the *Gibbs sampler*. The Gibbs sampler draws the parameters $\beta_0, \dots, \beta_p, \sigma^2$ from their posterior distributions. Those distributions (see equation (??) and (??)) need the design matrix X and labels Y , which are given as arguments. Furthermore the Gibbs sampler is not just drawing one time, it draws B times, where $B = R + b$. b is the number of burn in samples and R the number of used samples. If $b = 0$, we use all generated samples.

In figure 2 the traces of the Gibbs sampler is plotted for all parameters. It can be seen that the values, drawn by the sampler differ in some range. Moreover we can see that the initial value of $\sigma^2 = 1$ is corrected very fast to a specific range. The initial value of β is directly drawn in the range where most values are generated.

Furthermore in figure 3 the histogram and the estimated density (using kernel method) is plotted for every parameter. The red line marks the posterior mean point estimator for any parameters.

Both figures base on data for the whole dataset of the optimal model (see below).

```
gibbsSampler <- function(X, Y, R, b = 0, initSigma = 1) {  
  # Sample parameters from posterior distribution  
  # Prepared for baysean linear regression with ordinary least square approach (OLS)  
  #  
  # Args:  
  #   X: Design matrix  
  #   Y: Labels  
  #   R: Number of draws (without burn in), R is a scalar in this function  
  #   b: Number of burn in draws  
  #   initSigma: Initial value of sigma (default 1)  
  #  
  # Returns:  
  #   List containing sampled betas (number depends on design matrix) and sigmas  
  
  # Get number of overall draws (including burn in)  
  B <- R + b  
  
  # Size of design matrix  
  n <- nrow(X) # Number of data points  
  p <- ncol(X) # Number of parameters  
  
  # Variables to store the samples in (initialize sigma with initSigma)  
  betas <- matrix(nrow = B, ncol = p)  
  sigma <- c(initSigma, rep(NA, B-1))  
  
  # Sampling  
  for(i in 1:B){  
    # OLS of beta  
    V <- solve(t(X)%*%X) # (X^T X)^-1  
    beta_hat <- V%*%t(X)%*%Y # (X^T X)^-1 X^T Y
```

```

# OLS of sigma
sigma_hat <- t(Y-X%*%beta_hat)%*(Y-X%*%beta_hat)/(n-p)

# Sample beta from the full conditional
betas[i,] <- rmvnorm(1, beta_hat, sigma[i]*V)

# Sample sigma from the full conditional
if(i < B) {
  sigma[i+1] <- 1/rgamma(1, (n-p)/2, (n-p)*sigma_hat/2)
}
}

# Remove burn in, if there is some
if(b != 0) {
  betas <- betas[-(1:b),]
  sigma <- sigma[-(1:b)]
}

return(list(betas = betas, sigma = sigma))
}

```

As next, we define a function doing the leave-one-out cross validation. It deletes one data point and samples the parameters from the posterior distribution (using the `gibbsSampler` function). It is done n times, where n is the size (number of rows) of the data matrix, so there is a sampled posterior distribution for n combinations of the data. Additionally the function estimates the posterior mean (point estimation) of the parameters and predict the labels, using the data matrix X and the point estimators of the parameters.

Note that the argument R can be a scalar or a vector. If R is a scalar, the cross validation just runs usual. If R is a ordered vector the Gibbs sampler draws as often as the last value of the R vector. The size of the vector is indicated by variable `steps`. Afterwards the calculation of the posterior mean and the prediction is done for every step in the vector R . In the first step, just the first R_1 number of samples are used to estimate the parameters. In the second step, $R_2 > R_1$ number of samples are used, and so on.

```

crossValidation <- function(X, Y, R, b) {
  # Implementation of leave-one-out cross validation (LOO-CV)
  # Cross validate bayesian linear regression model with OLS
  #
  # Args:
  #   X: Design matrix
  #   Y: Labels
  #   R: Number of draws (without burn in), R can be a vector (ordered) in this function
  #   b: Number of burn in draws
  #
  # Returns:
  #   List containing estimated parameters and label predictions for every LOO-CV step

  # Size of design matrix
  n <- nrow(X) # Number of data points
  p <- ncol(X) # Number of parameters

  # Get size of R vector (steps = 1, if R is scalar)
  steps <- length(R)

  # Run gibbs sampler to get sampled parameters
  samples <- lapply(1:n, function(i) gibbsSampler(X[-i,], Y[-i], R[steps], b))
}

```

```

# Inititalize lists to store results of estimation and prediction
Sigma <- list()
Betas <- list()
Yhati <- list()

# Calculate sigma, betas and Yhati for every R step (do it once if R is scalar)
for(k in 1:steps) {
  Sigma[[k]] <- sapply(samples, function(sample) mean(sample$sigma[1:R[k]]))
  Betas[[k]] <- sapply(samples, function(sample) colMeans(sample$betas[1:R[k],]))
  Yhati[[k]] <- sapply(1:n, function(i) X[i,]%*%Betas[[k]][,i])
}

return(list(Sigma = Sigma, Betas = Betas, Yhati = Yhati))
}

```

3.2 Model evaluation

The cross validation algorithm can be used to evaluate a *best* mode, where best means, that the mean squared error (mse), defined in equation (2.3) or the *lppd*, defined in equation (2.1), is the lowest, compared to the other models. Both measures are obtained by loo-cv.

The `bayesModelEvaluation` function calls the cross validation function for every model. Then for every model the *mse* and *lppd* is calculated and returned.

```

bayesModelEvaluation <- function(models, Y, R, b) {
  # Model evaluation based on mean squared error (MSE) and log posterior
  # predictive density (lppd) using leave-one-out cross validation (LOO-CV)
  #
  # Args:
  #   models: A list of design matrices with different size and types of parameters
  #   Y: Labels
  #   R: Number of draws (without burn in), R can be a vector in this function
  #   b: Number of burn in draws
  #
  # Returns:
  #   List containing MSEs and LPPDs for every model

  # Evaluate multiple models and return results from all models
  n <- length(Y)
  k <- length(models)

  # Cross validate every model
  results <- lapply(1:k, function(i) crossValidation(models[[i]], Y, R, b))

  # Calculate Mean Squared Errors
  MSEs <- sapply(results, function(el) {
    return((1/(n-nrow(el$Betas[[1]])))*sum((Y-el$Yhati[[1]])^2))
  })

  # Calculate log posterior predictive density (log likelihood)
  #  $y \sim N(XB, s^2(X^T X)^{-1}) = N(\text{Yhat}, s^2(X^T X)^{-1})$ 
  LPPDs <- sapply(results, function(eva) sum(log(dnorm(Y, eva$Yhati[[1]], eva$Sigma[[1]]))))

  return(list(MSEs = MSEs, LPPDs = LPPDs))
}

```

To run the mode, we first chose `swiss` as a data set. We defined a list of 11 design matrices, containing different models. Afterwards the `bayesModelEvaluation` is called to run the cross

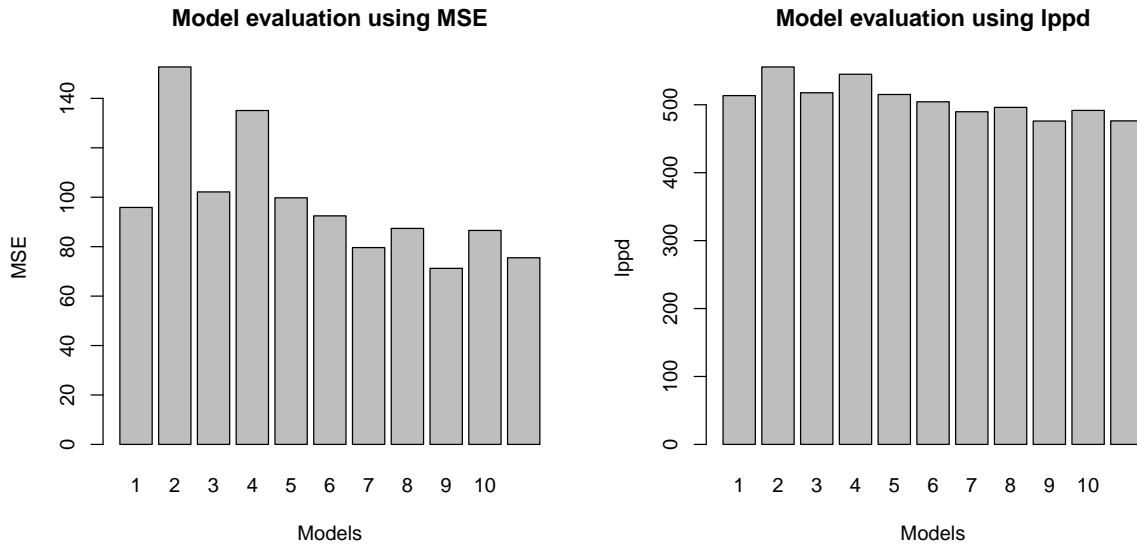


Figure 1: Model evaluation with leave-one-out cross validation (LOO-CV) using mean squared errors (MSE) and log posterior predictive density (lppd) to obtain best model.

validation and the calculation of *mse* and *lppd* for every model.

```
# Swiss data
dat <- swiss

# Response variable
Y <- dat$Fertility
n <- nrow(dat)

# Design matrices
models <- list(
  matrix(c(rep(1,n), dat$Education), nrow=n),
  matrix(c(rep(1,n), dat$Agriculture), nrow=n),
  matrix(c(rep(1,n), dat$Examination), nrow=n),
  matrix(c(rep(1,n), dat$Catholic), nrow=n),
  matrix(c(rep(1,n), dat$Education, dat$Agriculture), nrow=n),
  matrix(c(rep(1,n), dat$Education, dat$Examination), nrow=n),
  matrix(c(rep(1,n), dat$Education, dat$Catholic), nrow=n),
  matrix(c(rep(1,n), dat$Education, dat$Agriculture, dat$Examination), nrow=n),
  matrix(c(rep(1,n), dat$Education, dat$Agriculture, dat$Catholic), nrow=n),
  matrix(c(rep(1,n), dat$Education, dat$Examination, dat$Catholic), nrow=n),
  matrix(c(rep(1,n), dat$Education, dat$Agriculture, dat$Examination, dat$Catholic), nrow=n)
)

# Run model evaluation
criteria <- bayesModelEvaluation(models, Y, R = 50, b = 10) # R = 500, b = 100
```

The results in figure 1 show that model 9 seems to be the best model with lowest *mse* and lowest *lppd*, compared to the other 10 models. To check this quantitatively we get the minimum of the *mse* values and the minimum of the *lppd* values.

```
print(which.min(criteria$MSEs))

## [1] 9

print(which.min(-2*criteria$LPPDs))

## [1] 9
```


Indeed, both indicators favor model 9. We need to know, that, according to Gelman (2013), *mse* is just appropriate if the model is more or less normally distributed. Furthermore if the models are normal and have constant variances, *mse* and *lppd* should have the same proportion.

The proportions are calculated in table 1. It can be seen, that the proportion is in the same scale in all cases, but there are deviations. For example in model 2 and 9 the proportions are very different. This is probably due to the fact, that all the models are not well normal distributed.

	1	2	3	4	5	6	7	8	9	10	11
proportion	0.19	0.27	0.20	0.25	0.19	0.18	0.16	0.18	0.15	0.18	0.16

Table 1: Proportion of mse and lppd.

Finally we store the index of the optimal model in a variable. In the next step we want to calculate Cook's distance just for the best mode.

```
# Choose optimal modal
optIdx <- which.min(-2*criteria$LPPDs)
```

3.3 Cook's Distance

The second application of the cross validation is the approximate calculation of Cook's distance, which is mathematically defined in equation (2.2). The function `cooksDistance` calculates the approximate solution, using loo-cv, and the analytic solution, based on frequentist linear regression with ordinary least squares.

To calculate the approximation of Cook's distance with cross validation we make use of the implemented `crossValidation` function above. We choose `R` as a vector, such that the cross validation function calculates multiple estimates for many different sample sizes. Finally we just apply the formula for Cook's distance.

For the analytic solution, we calculate a frequentist linear regression, where $\hat{\beta}$ was solved by ordinary least square approach. After we obtained the point estimation, we can calculate Cook's distance.

In the end a list is returned, containing the analytic solution as a vector and the loo-cv solutions as a matrix, where the number of columns depend on the size of vector `R`.

```
cooksDistance <- function(X, Y, R, b) {
  # Calculate cooks distances for optimal model from model evaluation
  # Two methods are used:
  # (1) Use LOO-CV approach for bayesian linear regression model with OLS
  # (2) Use analytic solution for frequentist linear regression model with OLS
  #
  # Args:
  # X: Design matrix
  # Y: Labels
  # R: Number of draws (without burn in), R can be a vector in this function
  # b: Number of burn in draws
  #
  # Returns:
  # cooksBayesCV: Cook's distances calculated by LOO-CV (bayes model),
  #               where cook's distances are calculated for different sample sizes
  # cooksAnalytic: Cook's distances calculated by analytic solution
  # sample:       The gibbs sample from the bayes linear model (for plotting purpose)

  # R is usually a vector (but don't has to)
  B <- R + b
  steps <- length(R)
```

```

# Prepare projection matrix and number of parameters
H <- X%*%solve(t(X)%*%X)%*%t(X)
p <- ncol(X)

# Run cross validation with vector R
cv <- crossValidation(X, Y, R, b)

# Sample whole model (add B = R + b samples, remove b later)
sample <- gibbsSampler(X, Y, R[steps] + b)

# Cook's distance: Frequentist approach with analytic solution
betaHat <- solve(t(X)%*%X)%*%t(X)%*%Y
YhatLinReg <- X%*%betaHat
E <- Y-YhatLinReg
cooksAnalytic <- (E^2/((1/(n-p))*sum(E^2)*p))*(diag(H)/(1-diag(H))^2)

# Cook's distance: Bayesian approach with LOO-VC solution
cooksBayesCV <- matrix(nrow = n, ncol = steps)
for(k in 1:steps) {
  # Estimate posterior mean from betas
  betas <- colMeans(sample$betas[b:B[k],])

  # Predict values, using posterior mean
  Yhat <- X%*%betas

  # Calculate cook's distance
  dists <- apply(cv$Betas[[k]], 2, function(betas) {
    return((Yhat - X%*%betas)^2)
  })
  mse <- (1/(n-p))*sum((Y-Yhat)^2)
  cooksBayesCV[,k] <- colSums(dists)/(p*mse)
}

return(list(cooksBayesCV = cooksBayesCV, cooksAnalytic = cooksAnalytic, sample = sample))
}

```

We apply the function above to the optimal model, obtained by the model evaluation. We choose a specific sequence of sample sizes R and a burn in size.

```

# Number of samples
b <- 100
R <- seq(100,500,50) # seq(100,5000,50)

# Get optimal model and calculate projection matrix
cooks <- cooksDistance(models[[optIdx]], Y, R, b)

```

In the first step, we want to have a look at Cook's distances. In figure 4 the blue bars in all 3 plots are always the same analytic solution. They are compared to the the loo-cv solution using Bayes approach, which is shown in red. In the first plot 100 simulations where done, in the second plot 150 and 500 in the last plot. We can see, that the loo-cv solutions is coming closer to the analytic solution, the more simulations are done.

To see the convergence behavior of the loo-cv solution to the analytic solution, we developed a measure d to calculate the distance between both solutions. The Cook's distance value for both solutions where subtracted, squared and summed (squared distance) as follows

$$d = \sum_{i=1}^n (D_i^{\text{loo-cv}} - D_i^{\text{analytic}})^2, \quad (3.1)$$

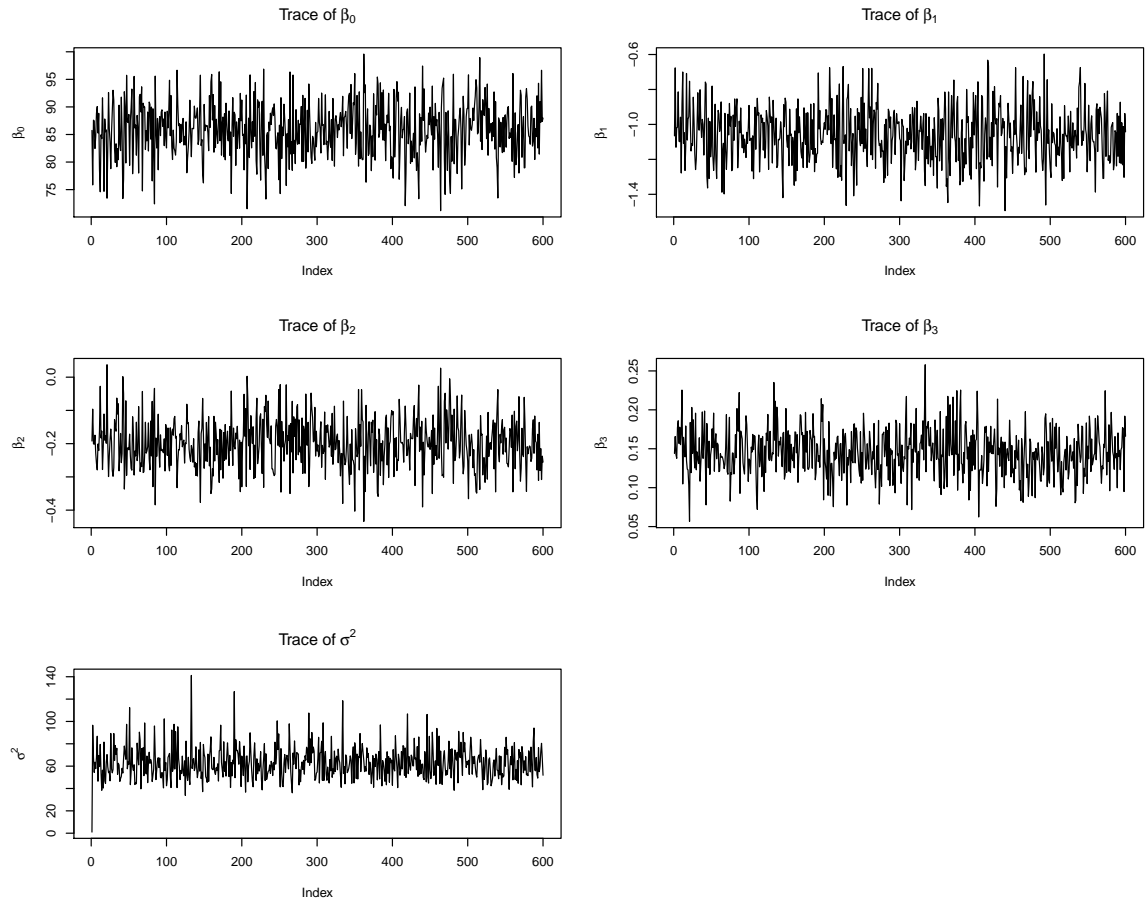


Figure 2: Traces of sampled parameters from posterior distributions regarding bayesian linear regression with ordinary least squares.

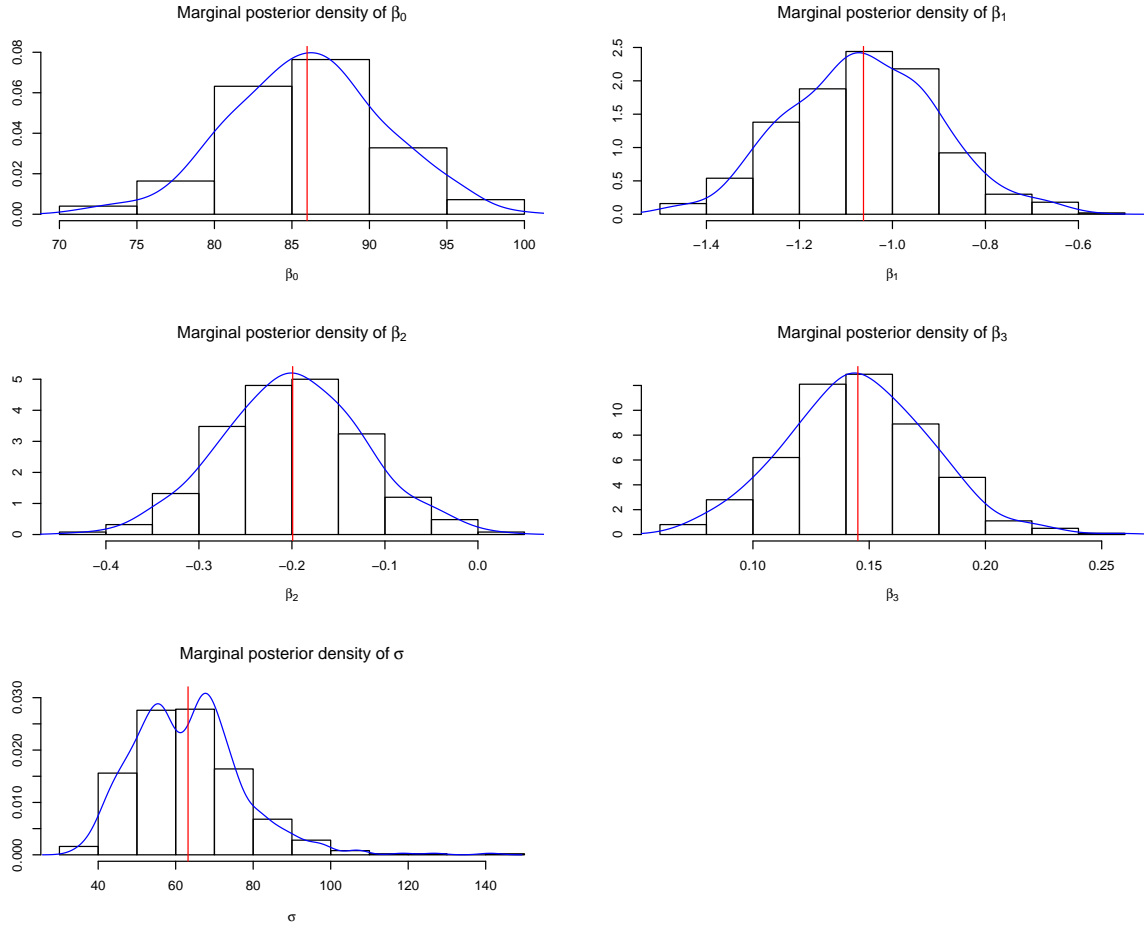


Figure 3: Posterior densities, histograms and posterior means (red vertical line) of sampled parameters regarding bayesian linear regression with ordinary least squares

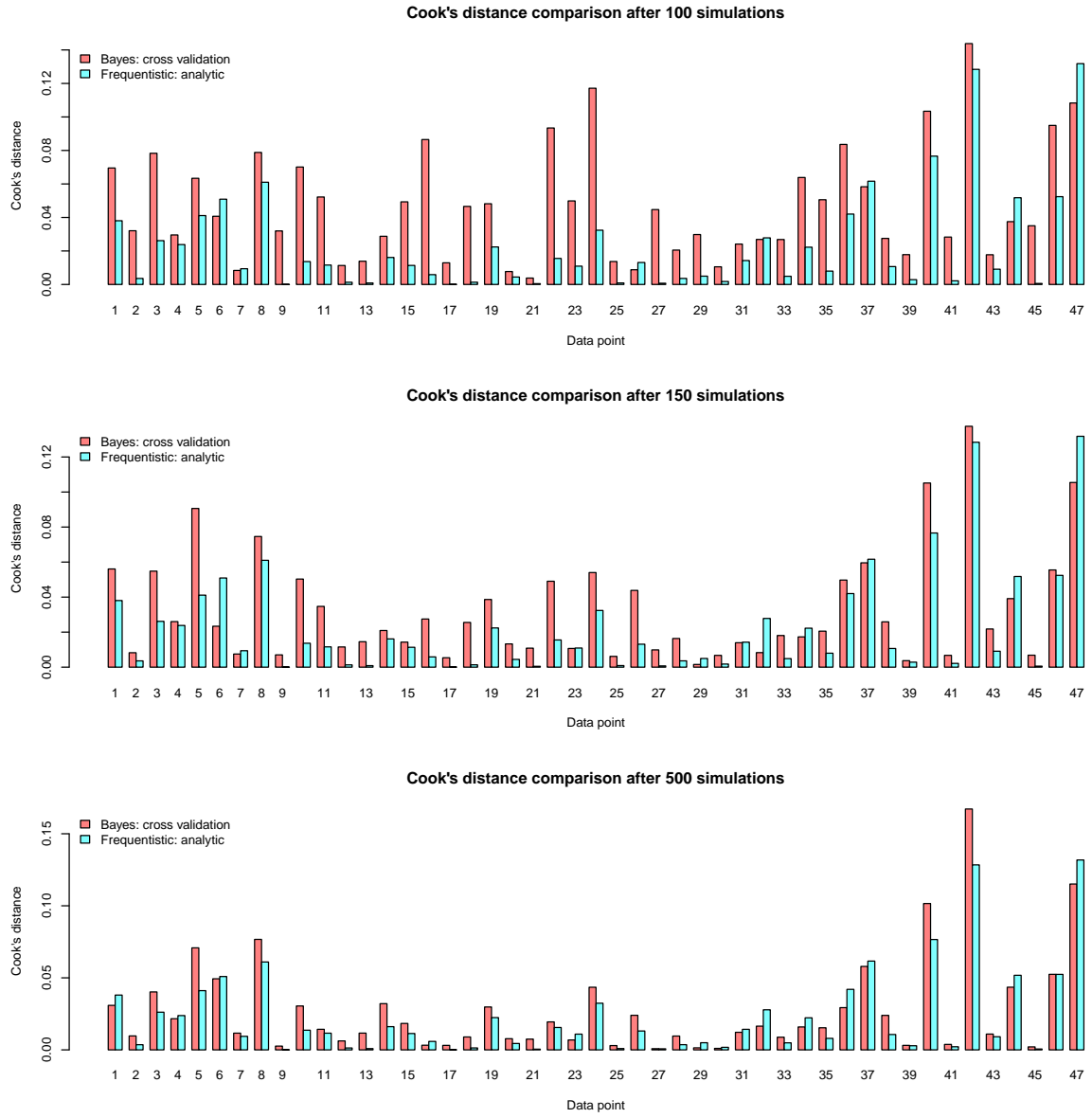


Figure 4: Pointwise cook's distance, calculated with three different sample sizes, compared to analytic cook's distance from frequentist linear regression model using least squares.

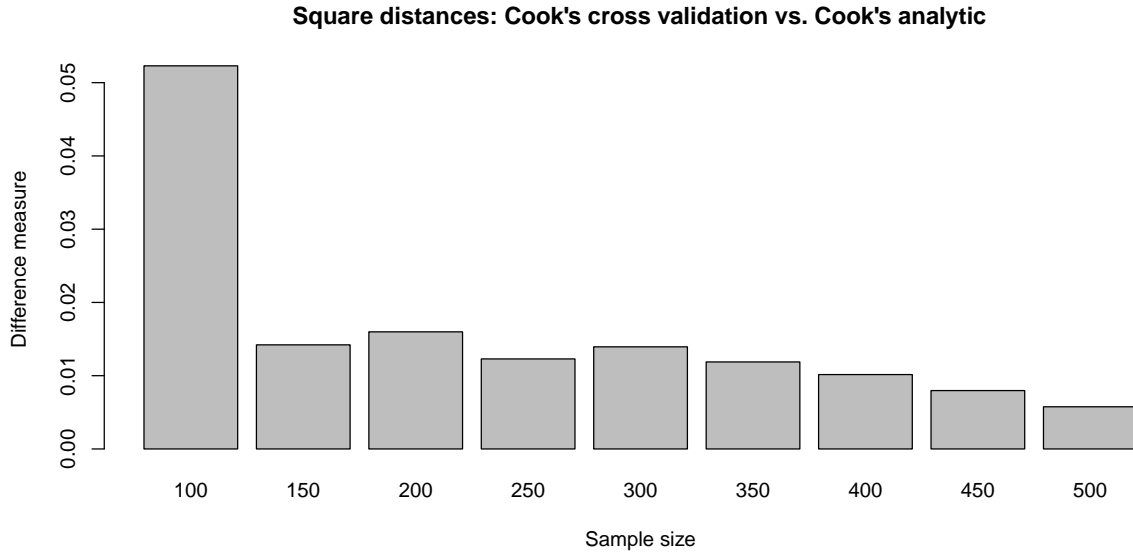


Figure 5: Cook's distance measure to quantify approximation from cross validated cook's distances of analytic cook's distances. The approximate solution with LOO-CV comes closer to the analytic solution, the more samples are drawn.

where n is the number of data points, $D_i^{\text{loo-cv}}$ Cook's distance for the loo-cv solution and D_i^{analytic} Cook's distance for the analytic solution for point X_i .

```
# Initialize matrices
steps <- length(R)
matBayesD <- cooks$cooksBayesCV
matFreqD <- matrix(rep(cooks$cooksAnalytic, steps), ncol=steps)

# Calculate differences between Cook's distance methods
cooksMeasure <- colSums((matBayesD-matFreqD)^2)
```

In figure 5 we can see that the defined distance d becomes lower. In other words: both solutions are coming closer together the more simulations are done. Therefore a convergence behavior can be seen.

4 References