

BarnebyLives: an R package to create herbarium specimen labels and clean spreadsheets

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Abstract

Premise: Depositing specimens to herbaria is a time consuming task. Many institutions have reduced the amount of funding for herbaria, and universities have reduced the amount of education dedicated to curatorial tasks and specimen deposition. Despite this, the continual generation of herbaria specimens are essential for current and future research in evolution and ecology. In order to facilitate the continued growth of herbaria BarnebyLives was developed as tool to supplement collection notes, perform geographic and, taxonomic informatic processes, enact spell checks, produce labels, and submit digital data for fast accessioning of specimens

Methods and Results: BarnebyLives uses geospatial data from the U.S. Census Bureau to provide political jurisdiction information, and data from other sources, including the United States Geological Survey, to supplement collection notes by providing information on abiotic site conditions. It uses inhouse spell checks to verify the spelling of a collection at all taxonomic ranks, the IPNI standard author database to check standard author abbreviations, and the Royal Botanic Garden Kews ‘Plants of the World Online’ to check for nomenclatural innovations. Optionally the package writes driving directions to sites using Google Maps. The package outputs data in a tabular format, as well as a spatial format, for review by the user to accept or confirm changes, before dynamically rendering labels using LaTeX.

Conclusions: BarnebyLives provides accurate political and physical information, reduces typos, provides users the most current taxonomic opinions, generates driving directions to sites, and produces aesthetically appealing labels and shipping manifests in a matter of minutes.

Nearly 400 million specimens are housed in herbaria around the globe (Thiers, 2021). However, The rate of accessioning new collections to herbaria diminished in the 20th century as priorities in biology shifted

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away from describing and documenting earths biodiversity and towards understanding cellular and molecular processes (Prather et al., 2004; Pyke and Ehrlich, 2010; Daru et al., 2018). This shift, among other factors, lead to a decline in the funding allocated to collections based research, the number of staff maintaining and accessioning new collections, and educating students in these practices (Funk, 2014). istorically specimens have been used to describe the taxonomic diversity of plants and document the worlds floristic diversity (Greve et al., 2016; James et al., 2018; Brewer et al., 2019; Rønsted et al., 2020). A renewed interest in herbarium data through ‘big data approaches’, such as museomics, has brought herbarium collections back to the forefront of the natural sciences (Rønsted et al., 2020; Marsico et al., 2020).

Innovations in computing, specimen digitization, data sharing, DNA sequencing, and statistics have likely brought about greater use of herbarium specimens than ever before (Greve et al., 2016; James et al., 2018; Brewer et al., 2019; Rønsted et al., 2020). The current uses of specimens and derived data extend far beyond their traditional roles in systematics and floristics, and studies utilizing collections are regularly carried out to better understand the ecological niches, phenological processes, and interactions of plants (Rønsted et al., 2020; Davis, 2023). We anticipate that collections have yet to gain their full, fostered by novel approaches become more accessible, such as remote and electronic sensing, meta-barcoding, and community science (Tosa et al., 2021). While image based or purely observational (rather than collections based) citizen science approaches (e.g. iNaturalist, BudBurst) have dovetailed to meet many of these research needs specimens contain rich data which are not accessible via images. Specimens have the additional potential to: provide samples of DNA, secondary metabolites, or proteins, notes on the status and composition of the biotic and abiotic settings at time of collection, material for measuring (micro-)morphological attributes (Borges et al., 2020), and seeds or pollen. These factors will ensure that specimens will remain the ultimate botanical data source into perpetuity.

However, despite this renewed recognition of the utility of collections, efforts to continually grow them appear slow (Prather et al., 2004). We conjecture this is in part because collecting and depositing specimens is a fundamentally slower process, especially for novice collectors, when compared to taking photographs via professionally developed apps which can be run on smartphones (Daru et al., 2018; Mishler et al., 2020; Manzano and Julier, 2021). While many young botanists are capable of using dichotomous keys to reliably identify - and able to collect satisfactory - material exist, we have observed that they face difficulties navigating several aspects of data collection and preparation of labels for submission to herbaria. Apparent problems include the lack of dedicated time at a field seasons end to process specimens, a general lack of education on cartography and orienteering, natural history (e.g. geology, geomorphology), nomenclature, familiarity with various computer programs (e.g. Microsoft Office suite), and increasingly - foundational knowledge of plant

systematics (Woodland, 2007; Barrows et al., 2016; Nanglu et al., 2023). In the absence of suitable mentors this results in not only the delay in the deposition of many specimens, but in a failure for many specimens to be accessioned at all, and increasingly ever collected.

The generation of an herbarium specimen includes many steps which are easy to take for granted (Forman and Bridson, 1989). For example while acquiring appropriate political information for a collection site appears simple, young collectors rarely have the adequate cartographic resources (printed topographic maps, or GIS software) at their disposal. In topographically complex areas, where borders are often associated with hydrologic basins and the ridges defining them, collectors are liable to misinterpret their true geographic position. Even finding appropriate sites names can rarely be solved without a printed map, as many software maps now consider many features which would serve as site names extraneous in the era of GPS. Similarly, the rate at which taxonomic innovations are occurring has made it difficult to find more recently applied names (Hitchcock and Cronquist, 2018). Furthermore formatting a label correctly (e.g. abbreviations) is a time consuming process and likely to introduce several errors in formatting. Anecdotally, many mail merge templates still require collectors to modify many variables by hand, e.g. applying italicization. Even if a collector navigates all of these hurdles, the time allocated to each step is quite large.

As a result of these concerns, we have developed an R package, *BarnebyLives*, that aims to increase both the data quality of labels, and to speed up the process of producing them. It rapidly provides political and administrative boundary information for a collection site using data from the U.S. Census Bureau (Walker (2024)), the Public Land Survey System, and ownership details of public lands via the Protected-Areas Database (PAD-US) from (Gap Analysis Project (GAP) (2024)). Site names are suggested via finding the closest unambiguously named place feature via the Geographic Name Information System (GNIS), and by precise calculation of the distance and azimuth from these localities to the collection site (Survey (2023)). Using the GMBA Mountain Inventory V. 2, a standardized named mountain data set with global coverage, which we have supplemented with over XXXX valleys allows for a relevant descriptor of the general region with less ambiguity (Snethlage et al. (2022)). Spell checks on all scientific names (including associated species) are performed using a copy of the World Checklist of Vascular Plants, and the collected species may be searched via Kew’s Plant of the World Online for relevant synonyms (Govaerts et al. (2021), POWO (2024)). Author abbreviations are verified using IPNI’s Standard Author Abbreviation Checklist and also returned by Kew’s Plants of the World Online to ensure proper abbreviation of authorities (The Royal Botanic Gardens and Herbarium (2024), POWO (2024)). Checks are performed to search for common issues associated with spreadsheets, or transcription, such as the auto-filling of coordinate and date columns. After final review of the data generated by the package, it allows for the option to export spreadsheets which are usable for mass

upload of data to multiple common herbarium databases, as well as the generation of herbarium labels.

Currently the label generation functionality is provided explicitly by two programs PLabel, and Symbiota, as well as commonly by the Microsoft Word tool Mail Merge (Perkins (2020), Gries et al. (2014)). The office suite costs money, and in our experience is finicky, further it's functionality ends with label creation.

PLabel is a standalone program which has greatly enhanced functionality relative to a Mail Merge, allowing users to specify the layout and formatting of label components using an intuitive and local graphical user interface (GUI) functionality, unfortunately it does not include data cleaning functionalities beyond verifying nations of collection; while some sources indicate it can only be used on Microsoft we expect it can be accessed on Linux and Mac using Windows emulators like Wine. The popular Symbiota biodiversity data management software not only provides label generation capabilities but also provides data cleaning functionality, in an attractive GUI web portal allowing for live management of collections, and bypassing the need for a local installation, allowing it to work on all operating systems. Symbiota offers functionality similar to the first four of our five stages of our 'Taxonomic' module and to our knowledge a check of the 'Political Boundaries' as well (see FIG). However, not all herbaria use Symbiota and many have original database systems which they maintain (e.g. Harvard University Herbarium, https://kiki.huh.harvard.edu/databases/specimen_index.html, Missouri Botanical Garden <https://tropicos.org/specimen/Search>, and The Consortium of Pacific Northwest Herbaria <https://www.pnwherbaria.org/>). However, many collectors like to generate their own labels, especially as they are likely to be sending different sets of collections to different institutions. Accordingly, Symbiota's functionality should exist in an ecosystem with alternative systems. In scenarios where users want to keep rendering labels in either of the three existing alternatives, they can easily export data in the appropriate formats after utilizing BLs data cleaning utilities.

BarnebyLives was named for plant taxonomist Rupert Charles Barneby (1911-2000), who published over 6,500 pages of text, described over 750 taxa, and is notable for balancing his studies at the William & Lynda Steere Herbarium at the New York Botanical Garden with annual collection trips in Western North America from 1937-1970, and sporadically until his passing in 2000 (Welsh (2001)). Select accolades of Rupert include the 1989 Asa Gray Award from the American Society of Plant Taxonomists (ASPT), the 1991 Engler Silver Medal from the International Association of Plant Taxonomists (IAPT), as well as being one of eight recipients of the International Botanical Congress's (IBC) Millennium Botany Award (1999) (Welsh (2001)). Most importantly, Rupert was remembered as being generous with his time to assist younger botanists with the more arcane aspects of field botany and taxonomy (Holmgren and Holmgren (1988)).

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and to create a shipping manifest and transfer notice.

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charge alternatives such as: LibreOffice, OpenOffice, or via the cloud on Googlesheets. The latter two options are documented here and in package vignettes, detailed descriptions of the required and suggested input columns are located on the Github page (<https://github.com/sagesteppe/BarnebyLives> ‘*Input Data Column Names*’) and over 300 real-world examples are on a Google Sheets accessible from the page. BarnebyLives is atypical of R packages in that it requires a considerable amount of data to operate (Table 1). Virtually all the on-disk memory associated with these data are used in storing spatial data, setting up a local instance of the program - at whichever scale a user desires (see Figure XX) is fully documented in the package documentation. Functions which require the on-disk data require a path to the data as an argument. Manually supplying the path argument allows for users to judiciously decide a storage location suitable for their needs.

We anticipate BarnebyLives usage will require less than a couple gigabytes of memory (ours covering all of the conterminous Western U.S. at 90m resolution is ~16 GiB), and the processing takes relatively little RAM, hence we believe installations can work on hardware as limited as Chromebooks, while having the data stored entirely on thumb-drives. The final steps of BarnebyLives, generating the labels require working installations of Rmarkdown, a LaTeX installation (e.g. pdfTeX, LuaTeX, XeLaTeX), and the open source command line tools pdfjam and pdftk. While these steps are run through a shell scripting language like bash, we have wrapped them in R functions which bypass the need to enter the commands directly into a terminal. Several commands in BarnebyLives require the output from previous functions, and a workflow which satisfies these requirements is presented in Figure 1.

Functionality

BarnebyLives can be thought of as consisting of five main modules (Figure 1): spatial, taxonomic, formatting, manual review, and data exporting.

The spatial module has five required functions, and two optional functions. *autofill_checker* searches for patterns in the input latitude and longitude data associated with autofilling from various spreadsheet programs, and will emit a warning if they are encountered. *coords2sf* will create a spatially explicit simple feature (sf) geometry data set of the input data. *political_grabber* will determine many levels of administrative ownership including land management, and public land survey system sections. *physical_grabber* provides various geographic data such as elevation, landform position, and aspect using 90m resolution spatial data. *site_writer* will write directions from an officially named and located placename to the collection site. *dms2dd* is an optional function used to convert from coordinates denoted in the degrees minutes and second format (e.g. 42°08’39.9”N 87°47’08.3”W) to decimal degree format (e.g. 42.14439, -87.78569). *directions_grabber* is an optional function which writes driving directions from a reasonably sized town to the closet drivable area

to the site using the Google Maps API; this will require a valid Google account, which is free per month for most personal and smaller academic usages.

The taxonomic module has four required functions, and one optional function. *spell_check* will perform a spell check on the entered scientific name based on a local copy of Kew Plants of the World Database filtered to the local continents, or a user specific backbone. *spell_check_family* performs a spell check on the family entered for each scientific name. *author_check* ensures authors are entered in a valid format, e.g. the correct standard abbreviations are used. *associates_check* will perform a spell check on all of the associated species using the local taxonomic database. *powo_searcher* can be used tandem to the functions *spell_check_family*, and *author-check*, but we use it in lieu of them; this function will search the current Plants of the World Online to determine relevant synonyms and alternative higher taxonomy for the focal species. No API key or any registration is required to use *powo_searcher*.

The formatting modules has three functions, the first two we will detail are technically optional, but they are run locally and so quickly there is no reason to skip them. *associate_dropper* if present this function will silently remove the collected species from the list of associated species, however it will search for the species to remove using scientific name entered initially by the user rather than returned via spell checks etc. *field_lengths* will emit messages for any fields which we suspect will create an ‘overflow’ on the physical label and should be truncated for clarity. *date_parser* is mandatory and will parse an input date into various formats for notating collection and determination dates on labels.

The manual review process technically only has one function which is optional, and may be ran during the spatial process (after *coords2sf*), but the importance of manual review is important enough that it warrants explicit mention. *geodata_writer* will write out a spatial copy of the data set to any geospatial format supported by the sf package, but defaults to writing out ‘kmls’ which are readily used with Google Earth, and can also be opened in several other free geographic information system (GIS) softwares such as QGIS. Notably many of the flags which BarnebyLives generates will be placed into columns (e.g. NAMES OF COLUMNS FROM FLAGS), and can be manually reviewed by the analyst, many of these issues can be resolved by simply addressing the relevant issues in the original data input spreadsheet.

The data exporting module contains three functions, which interact with LaTeX templates and require slightly more advanced R user interactivity such as setting up mapping functions using the tidyverses purrr package. *labels_skeleton* is an R script which will require a few modification steps to tailor to each instiution, these r scripts will put data into a TEMPLATE...

Herbarium Collections

The package was released into beta testing using the primary authors collections from 2023. The testing of the package within this manuscript was performed using a subset of their collections from 2018-2022, *all* of which are un-accessioned. Only collections which had identifications to the level of species or lower, and transcribed collection dates and coordinates were used. This results in a data set of 819 records for testing, from 204 sites located across Western North America (Figure 2). In total 615 species (with 557 sets of authors), with 66 infraspecies (22 authors) in 73 families were used for testing.

BarnebyLives took roughly three minutes (192.424sec) to run all local steps, and roughly twelve minutes (703.167sec) to search Plants of the World Online, and 73.73sec to search Google Maps and write directions to sites. Most of the local run time is attributable to the spatial (176.162sec), and taxonomic operations (14.733sec), while formatting data for labels took 1.529sec. The spell check operation of the scientific name accounted for nearly all of the time (14.506sec) spent performing local taxonomic operations. The generation of labels consumed around seven minutes (424.042sec) for the rendering, 50.54sec to combine individual labels four per single sheet of landscape orientated paper, and 2.97sec to combine the 205 sheets to a single Portable Document Format (PDF).

The total computer run time for processing these 819 specimens was 16 minutes.

RESULTS

Even on data which had been manually cleaned and error-checked by a human several times BarnebyLives was able to reduce transcription errors, identify typos, make nomenclature suggestions, and reformat text elements for downstream use. While none of the 73 families were misspelled, BarnebyLives made 24 suggestions on naming, identified 16 typos, identified 2 instances where an incorrect family was entered, and 0 instances where an outdated circumscription was applied. At the level of family BarnebyLives flagged 6 records where the author follows an alternative taxonomy, and flagged 0 records in error.

In the 292 genera analysed BarnebyLives identified 57 discrepancies at the level of genus between user submitted and processed data. In 36 of these instances the user supplied an outdated name (20 unique genera) flagged 5 records where the author follows an alternative taxonomy (3 genera total), and flagged 0

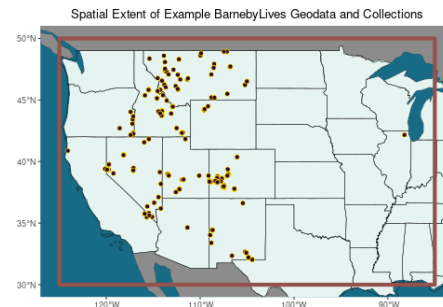


Figure 1: The spatial extent (orange), and herbarium collection sites (burgundy) tested in this manuscript.

records in error.

Of the 819 species analysed (615 distinct species) BarnebyLives flagged 55 records, and detected 28 instances of misspelled epithets (28 unique species). In 15 of these instances the user supplied an outdated name (15 unique species). It also flagged 2 records where the author follows an alternative taxonomy (2 unique species), and flagged 9 records in error. The final record was an egregious error where the order of the specific epithet and the genus name.

The number of author abbreviations which were not in the appropriate format were XX (% percent), in nearly all cases the presence or absence of a period were the issue. 5 records were appropriately flagged for issues with auto fill increment of the longitude value, and 3 records were also auto-flagged for increases in latitude values (% of records).

DISCUSSION

While numerous tools have been developed for cleaning of existing herbarium and museum records, few help with ensuring that the entered data are accurate (Patten et al. (2024)). We argue that the original collectors are the most qualified individuals to perform quality control checks, and BarnebyLives allows them to do so in a relatively fast and streamlined format. By utilizing both R and LaTeX, and having publicly available source code on Github, this program allows most users immediate familiarity with the system for troubleshooting issues, and implementing upgrades and modifications on project branches.

Accessioning often times relies on the use of the Office Suite of programs, and may utilize other costly software, such as ArcPro, or Adobe Acrobat. While BarnebyLives does not have it's own graphic user interface, the functionality of commonly used Interactive Development Environments (IDE's), such as Rstudio and VisualStudio (VS) Code, now offer functionality to readily view and filter data sets using familiar spreadsheet like formats.

LaTeX offers well documented and detailed functionality for customizing labels for individuals and institutions.

Data Sources for Package					
Variable	Usage	Source	Name	Data Model	Size (GiB)
County	Political	US Census Bureau	Counties	Vector	0.073
State			States		0.0*
Ownership		US Geological Survey	Protected Areas Database		0.435
TRS			Public Land Survey System		0.816
Place Names	Site Name		Geographic Names Information System		0.081
Mountains	Site Name	EarthEnv	GMBA Mountain Inventory v2		0.004
Elevation	Site Characteristics	Open Topography	Geomorpho90m - Elevation	Raster	4.2
Slope			Geomorpho90 - Slope		4.6
Aspect			Geomorpho90m - Aspect		4.1
Geomorphons			Geomorpho90m - Geomorphons		0.455
Surficial Geology		US Geological Survey	State Geologic Map Compilation	Vector	0.708
Taxonomic Spellings	Spell Checks	World Flora Online	World Flora Online	Text	0.002
Author Abbreviations		IPNI	International Plant Names Index		0.001

*Counties and States are merged into the same dataset while setting up the package. The value for "County" includes State.

Figure 2: Sources of Data required for operations

Anecdotaly, using its default settings it is able to produce more aesthetically pleasing results than the typical word processors. Very good documentation of LaTeX capabilities is offered in multiple areas for instance via the Overleaf) project.

CONCLUSIONS

BarnebyLives is an R package which can rapidly acquire relevant geographic, and taxonomic data. It is also capable of performing specialized spell checks, and assorted curatorial tasks to produce both digital and analog data. The package relies on no licensed Software, such as the Microsoft suite, and is suitable for install on all major operating systems (Windows, Mac, Linux), with a small amount of use of the command line, which may be called from the Rstudio rather than a ‘traditional’ terminal.

AUTHOR CONTRIBUTIONS

The project was conceptualized by R.C.B. The program was written by R.C.B. Data collection and analysis were performed by R.C.B. R.C.B. & J.B.F wrote the manuscript, and both authors approved the final version of the manuscript.

ACKNOWLEDGEMENTS

The Bureau of Land Management are graciously acknowledged as providers of funding to R.C.B for most of his specimen collection activities. Two anonymous peer reviewers who increased the quality of this manuscript are thanked. Sofia Garcia is acknowledged for creating the ‘Valleys’ data set which place naming in the package relies

on. Several prominent associated collectors of specimens used in this study are thanked: Dani Yashinovitz, Dakota Becerra, Hannah Lovell, Caitlin Miller & Hubert Szczygiel. Rosalind Rowe is thanked for providing useful feedback during the later development stages of the program.

DATA AVAILABILITY STATEMENT

The BarnebyLives R package is open source, the development version is available on GitHub (<https://github.com/sagesteppe/BarnebyLives>), and the stable version is available on CRAN. The package includes three real use-case vignettes (tutorials) on usage. One vignette “setting_up_files” explores setting up a instance for a certain geographic area. Another vignette “running_pipeline” showcases the usage of the package for processing data entered on a spreadsheet. A final vignette “creating_labels” shows the usage of an R, and Bash script launched from RStudio to produce print-ready labels. All data used in this manuscript are available at: https://github.com/sagesteppe/Barneby_Lives_dev/manu script.

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REFERENCES

- Barrows, C. W., M. L. Murphy-Mariscal, and R. R. Hernandez. 2016. At a crossroads: The nature of natural history in the twenty-first century. *BioScience* 66: 592–599.
- Borges, L. M., V. C. Reis, and R. Izbicki. 2020. Schrödinger’s phenotypes: Herbarium specimens show two-dimensional images are both good and (not so) bad sources

of morphological data. *Methods in Ecology and Evolution* 11: 1296–1308.

Brewer, G. E., J. J. Clarkson, O. Maurin, A. R. Zuntini, V. Barber, S. Bellot, N. Biggs, et al. 2019. Factors affecting targeted sequencing of 353 nuclear genes from herbarium specimens spanning the diversity of angiosperms. *Frontiers in plant science* 10: 1102.

Daru, B. H., D. S. Park, R. B. Primack, C. G. Willis, D. S. Barrington, T. J. Whitfeld, T. G. Seidler, et al. 2018. Widespread sampling biases in herbaria revealed from large-scale digitization. *New Phytologist* 217: 939–955.

Davis, C. C. 2023. The herbarium of the future. *Trends in Ecology & Evolution* 38: 412–423.

Forman, L., and D. Bridson. 1989. The herbarium handbook. Royal Botanic Gardens Kew.

Funk, V. A. 2014. The erosion of collections-based science: Alarming trend or coincidence. *The Plant Press* 17: 1–13.

Gap Analysis Project (GAP), U. S. G. S. (USGS). 2024. Protected areas database of the united states (PAD-US) 4.0.

Govaerts, R., E. Nic Lughadha, N. Black, R. Turner, and A. Paton. 2021. The world checklist of vascular plants, a continuously updated resource for exploring global plant diversity. *Scientific data* 8: 215.

Greve, M., A. M. Lykke, C. W. Fagg, R. E. Gereau, G. P. Lewis, R. Marchant, A. R. Marshall, et al. 2016. Realising the potential of herbarium records for conservation biology. *South African Journal of Botany* 105: 317–323.

Gries, C., M. E. E. Gilbert, and N. M. Franz. 2014. Symbiota—a virtual platform for creating voucher-based biodiversity information communities. *Biodiversity data journal*.

Hitchcock, C. L., and A. Cronquist. 2018. Flora of the pacific northwest: An illustrated manual. University of Washington Press.

Holmgren, N., and P. Holmgren. 1988. Intermountain flora v. 7. The New York Botanical Garden Press, New York.

James, S. A., P. S. Soltis, L. Belbin, A. D. Chapman, G. Nelson,
D. L. Paul, and M. Collins. 2018. Herbarium data: Global
biodiversity and societal botanical needs for novel research.
Applications in plant sciences 6: e1024.

Manzano, S., and A. C. Julier. 2021. How FAIR are plant
sciences in the twenty-first century? The pressing need for
reproducibility in plant ecology and evolution. *Proceedings of
the Royal Society B* 288: 20202597.

Marsico, T. D., E. R. Krimmel, J. R. Carter, E. L. Gillespie,
P. D. Lowe, R. McCauley, A. B. Morris, et al. 2020. Small
herbaria contribute unique biogeographic records to county,
locality, and temporal scales. *American journal of botany* 107:
1577–1587.

Mishler, B. D., R. Guralnick, P. S. Soltis, S. A. Smith, D.
E. Soltis, N. Barve, J. M. Allen, and S. W. Laffan. 2020.
Spatial phylogenetics of the north american flora. *Journal of
Systematics and Evolution* 58: 393–405.

Nanglu, K., D. de Carle, T. M. Cullen, E. B. Anderson, S. Arif,
R. A. Castañeda, L. M. Chang, et al. 2023. The nature of
science: The fundamental role of natural history in ecology,
evolution, conservation, and education. *Ecology and Evolution*
13: e10621.

Patten, N. N., M. L. Gaynor, D. E. Soltis, and P. S. Soltis.
2024. Geographic and taxonomic occurrence r-based scrubbing
(gatoRs): An r package and workflow for processing biodiversity
data. *Applications in Plant Sciences* 12: e11575.

Perkins, K. 2020. Plabel.

POWO. 2024. Geographic names information system (GNIS)
- USGS national map downloadable data collection: U.s. Ge-
ological survey.

Prather, L. A., O. Alvarez-Fuentes, M. H. Mayfield, and C. J.
Ferguson. 2004. The decline of plant collecting in the united
states: A threat to the infrastructure of biodiversity studies.
Systematic Botany 29: 15–28.

Pyke, G. H., and P. R. Ehrlich. 2010. Biological collections and

ecological/environmental research: A review, some observations
 and a look to the future. *Biological reviews* 85: 247–266.

Rønsted, N., O. M. Grace, and M. A. Carine. 2020. Integrative
 and translational uses of herbarium collections across time,
 space, and species. *Frontiers in Plant Science* 11: 1319.

Snethlage, M. A., J. Geschke, A. Ranipeta, W. Jetz, N. G.
 Yoccoz, C. Körner, E. M. Spehn, et al. 2022. A hierarchical
 inventory of the world’s mountains for global comparative
 mountain science. *Scientific data* 9: 149.

Survey, U. S. G. 2023. Geographic names information system
 (GNIS) - USGS national map downloadable data collection:
 U.s. Geological survey.

The Royal Botanic Gardens, H. U. H. & L., Kew, and A. N.
 Herbarium. 2024. International plant names index.

Thiers, B. M. 2021. The world’s herbaria 2021: A summary
 report based on data from index herbarium.

Tosa, M. I., E. H. Dziedzic, C. L. Appel, J. Urbina, A. Massey,
 J. Ruprecht, C. E. Eriksson, et al. 2021. The rapid rise of next-
 generation natural history. *Frontiers in Ecology and Evolution*
 9: 698131.

Walker, K. 2024. Tigris: Load census TIGER/line shapefiles.

Welsh, S. L. 2001. Rupert c. Barneby (1911-2000). *Taxon*.

Woodland, D. W. 2007. Are botanists becoming the dinosaurs
 of biology in the 21st century? *South African Journal of Botany*
 73: 343–346.

SUPPORTING INFORMATION

Additional supporting information can be found online in the
 Supporting Information section at the end of this article.

Appendix S1. A table of all time trials for each function.