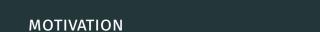
COMPUTATIONAL ANALYSIS OF CELL-TO-CELL HETEROGENEITY IN SINGLE-CELL RNA-SEQUENCING DATA REVEALS HIDDEN SUBPOPULATIONS OF CELLS

Buettner et al., (2015) Nature Biotechnology, 1–32. doi:10.1038/nbt.3102

Saket Choudhary January 29, 2015 BISC 542

OUTLINE

- Motivation
- · Methods[Overview]
- · Results
- · Challenges & Possibilities
- · Conclusion



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- · Profiling hundreds of cells: How to identify subpopulations?
- · Identify regulatory landscape of each cell population
- Account for hidden confounding factors that might explain cell heterogeneity

WHAT WAS IT ALL ABOUT?

- · Single cell transcriptomics heterogeneity: many single cells at the same time
- Accounting for technical noise was a solved problem; how do you account for other sources of variability: cell cycle, differentiation state etc.
- · Given expression levels, how do you infer the effect of *latent* variables

Key focus: How does cell cycle affect expression levels? Given the apriori nature of genes(association with cell-cycle), is it possible to remove the effect of cell cycles?

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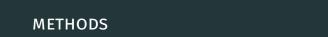
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- · Discovering previously unidentified cell types?!



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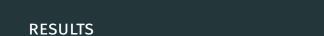
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- · Regress out effects of hidden factors
- · Corrected gene expression level



CELL CYCLE AFFECTS GLOBAL GENE EXPRESSION

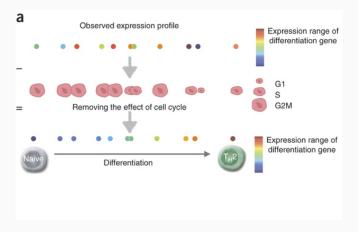


Figure: Observed Expression = Effect of differentiation + Effect of state of cell(G1, S, G2)

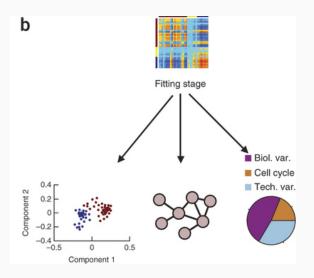


Figure: Infer the cell-cell covariance matrix using hidden factors such as the

RESULTS IN A GIST

- · Before applying scLVM the cells looked like a variable population
- scLVM corrected expression data showed there existed two sub-populations
- These two sub-populations were infact found to be associated with T-cell differentiation stages

The method is not about single cell transcriptomics. It is a general approach to isolate, model and understand sources of variability

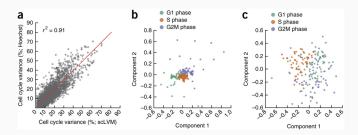


Figure: a. Gold standard v/s scLVM agreement b.Non Linear PCA on scLVM corrected data: no separation c.nonlinear PCA on unorrected data: separation by cell cycle!

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- The results were consistent even if the size of annotated genes was reduced to 10
- · The gold standard and scLVM results are in sync

EFFICACY: HOW ABOUT JUST THROWING OUT THOSE KNOWN GENES?

If the genes are known to be associated with cell cycle, why not simple throw them out to rule out the effect of cell cycle?

A non-linear PCA of datasets with cell-cylce associated genes thrown out gave a clear separation, and this separation was later validated to be two different cell cycles => scLVM accounts for the latent factors which cannot be simply accounted by throwing away those informative genes.

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- To further validate: non linear PCA with and without scLVM correction. Without correction: no obvious subgroups
- One of the two sub-populations post scLVM correction were found to have genes that marked completion of differentiation

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- It is possible to account for more than one factor, as long as the corresponding annotated gene sets are available
- When multiple confounding factors are considered, in order to ensure robust analysis it is important to ensure the statistical significance if the factors are weak or nonindependent
- · No formal tests exist for testing the presence of a particular factor

THE UNDERLYING MODEL

Let *N* = Number of cells

G = Number of variable genes(determined using a T-test on pre-processed count data) G_h = Set of marker genes(cell-cycle related) Y_h = $[y_1, y_2, ...y_h]$ = Vector of gene expressions where y_g represents gene g's expression acorss cells

$$Y_h = \mu + CU + XW + \psi \tag{1}$$

U, *W* = Weight of linear covariance model where *C* models *Q* known covariates and *W* models unknown covariates

 ψ models the rest of noise

C, *X* are determined using a bayesian approach assuming both U,W as gaussians prior.

$$P(Y_h|\sigma_u^2, \nu^2, X, C) = \Pi_{g=1}^G N(y_g|\mu_g, \sigma_u^2 CC^T + XX^T + \nu^2)$$
 (2)

The parameters are then estimated using maximum likelihood approach. Once XX^T is modeled, the genes are modeled as sume of mean and random effect:

$$y_g = \mu + \sum_{h=1}^{H} u_h + \psi_e + \psi_t \tag{3}$$

where $P(u_h) = N(\mu_h|0, \sigma_{gh}^2 \sigma_h)$, the last two terms accounting for residual and technical noise

and hence:

$$y_{corrected} = y_g - y_g^{(hidden)}$$
 where $y_g^{(hidden)}$ is the posterior estimation: $y_g^{hidden} = \sigma[\sigma + \nu_g]^{-1}(y_g - \mu_g)$