 Field, G. D. Additional sour Bialek, W. Hecht S, S Rieke, F., & Learning of write code	naterial provides background and additional context. It is linked in the Canvas page for this assignment. "A Sampath, A. P. (2017). Behavioural and physiological limits to vision in mammals. Philosophical Transactions of the Royal Society B, 372(1717). "Deserting for the curious, but not required reading: (2012). Biophysics. Princeton University Press. Chapter 2. Inlaer S, Pirenne MH. (1942). Energy, quanta, and vision. J. Gen. Physiol. 25, 819–840. Baylor, D. (1998). Single-photon detection by rod cells of the retina. Reviews of Modern Physics, 70(3), 1027–1036. bjectives to generate simulated streams of photons, edifferent probability distributions
 predict the fit a theore Introduct Experiments in probability of severying degree To model the process of the probability of the process of the pro	number of photos required for detection based on experimental results tically derived curve to experimental data ion photon detection require producing a dim flash of light that is right at the limit of what is perceivable. A counter intuitive aspect of detection is that it's probabilistic. For seemingly the same light intensity, sometimes the flash is seen and sometimes it's not. What changes is being the flash. The question is, where does this randomness come from? Is it the inherent variability of biology? Are there sources of noise? Is the observer, perhaps unconsciously, adjusting their criterion? Could it be in the light itself? Like most things in biology, the answay, all of the above. The production of the election, we first have to describe the stream of photons. This stream is inherently random. For a light source and for reflected light in the natural world, there isn't a highly precise clock emitting an exact number photons with femtosecond precision. The spontaneously, at controlled rates, and are independent of each other, which means that they arrive at random intervals. If they didn't, they wouldn't be independent.
EXERCISE import numpy from scipy i from matplot import ipywi import math	as np
We will simulate notebook) to see 1a. Rando Write a function def randtime return in def plotflas # unit h	a stream of photons with a Poisson process, which can be modeled in two different ways. One is to select the event times randomly, the other is to generate random intervals between the events. When you run your code, run it multiple times (using control-enter in the just that there is a wide variety of patterns. Sometimes there are wide gaps, other times the events are tightly clustered, occasionally they are more evenly spread out. In times In randtimes (N; t1, t2) to simulate a Poisson process by generating N random times in the interval $[t_1, t_2)$. Write a function plotflash to plot your results as a stem plot of the times with unit heights. In (N:int=100, t1:float=0, t2:float=1): p. random.uniform(t1, t2, N) In (T:np.array, title:str="Photon Release based on Poisson Distribution", xlim:list=[None, None]):
<pre># plot plt.stem if xlim[plt.xlab plt.ylim plt.ylab plt.titl plt.show</pre>	(T, ones) 0] is not None: xlim(xlim) el("Photon Release Distribution") ([-0.1, 2]) el("Relative Weight of photons") e(title) () s(t1=0, t2=50)
1.75 - 1.50 - 1.25 - 1.00 - 0.75 - 0.25 -	
A different way where λ is the	The photon Release Distribution $\frac{1}{20}$ $\frac{1}{30}$ $\frac{1}{40}$ $\frac{1}{50}$ $\frac{1}{50}$ $\frac{1}{9}$ Photon Release Distribution $\frac{1}{9}$
def randinte t = t1; for n in T.ap t = return n	<pre>range(N): pend(t + np.random.exponential(1/lam)) T[-1] p.array(T)</pre> rvals(lam=10)
1.75 - 1.50 - 1.25 - 1.00 - 0.75 - 0.25 -	
separate signa At any given tir	the flash Terences between the two methods above. At the visual limit, seeing a flash involves just a small number of photons. Each photoreceptor is capable of detecting single photos, but to "see" the flash requires detecting a minimum number of photons within a certain interval from noise. Explain why this is inherently probabilistic. The cones and rods of the eye experience levels of noise. In order to separate this noise from signal and respond to the signal, they require a constant stream of photons for a given time. This is inherently probabalistic because of the non-deterministic nature of the photon of the process of t
2a. The pro	ating the probability detection obability of K photons istribution specifies the probability of observing n events at rate λ within a unit time interval $p(n \lambda) = \frac{\lambda^n}{n!}e^{-\lambda}, n=0,1,2,\dots$ time period, we simply scale the rate: a period that is twice as long will see twice as many events. $p(n \lambda,T) = \frac{(\lambda T)^n}{n!}e^{-\lambda T}, n=0,1,2,\dots$ Integration period for visual detection is ~100 msecs. Use the Poisson distribution pdf to calculate the probability of receiving a specific number of photons within the period.
Make a bar plo Double the pho @widgets.int def probkpho # arrang K = np.a P = np.a # plot plt.bar(plt.ylab	ton rate (i.e. the light intensity) and then double it again plotting both results. Observe how the probability of seeing the flash increases. eract(lam=(0,40), T=(0.0,2.0)) tons(lam=10, T=0.1): e values range(0, 20) rray([(lam*T)**k / math.factorial(k) * np.exp(-lam*T) for k in K]) K, P)
interactive(Here we can se evidence to sug 2b. The pre The Poisson di	children=(IntSlider(value=10, description='lam', max=40), FloatSlider(value=0.1, description='T', It T to be 0.1 (100 msec) and change the value of lambda to approximate the probability of observing an event. At lambda=15, it is obvious that the probability of observing a rod responding to a photon is not 0, but is very close to it. Doubling lambda to 30, provides reason great that a cell might respond to a consistent stream of at least 6 photons. Obability of K or more photons Stribution specifies the probability of observing exactly n events. Of course, we would also see the flash for any number of events exceeding the threshold, so the actual probability of seeing would be the sum of all probabilities at or above some threshold K . $p(n \ge K \lambda,T) = \sum_{n=K}^{\infty} p(n \lambda,T)$ plement of the cumulative distribution function $cdf(x)$ where $x = K - 1$ which we can use to calculate the detection probability. (Note that we need to be careful at the threshold to use $K - 1$, because the cdf is defined by the sum <i>through x</i>).
<pre>def detection return 1 detectionpro 0.0839179420 K = [0, 1, 2] B = np.array plt.bar(K, B plt.xlabel(" plt.ylabel(" plt.title("P</pre>	, 3, 4, 5, 6, 7, 8, 9] ([detectionprob(k, 40, 0.1) for k in K])
1.0 - 0.8 - 0.6 - 0.4	Probability of Observing greater than K events
0.4 0.2 0.0	ng the threshold from experimental data details we need to address before we can apply model to experimental data.
detected. This coneeding to specific second is to the incident photostate actually are actually aa. Simula Define a function	duration of the flash. Experimental results show that if we have a shorter flash with the same number of photons, we have the same probability of detecting it. This makes sense because they could all arrive simultaneously (but at different rhodopsin molecules) and would doesn't hold if we make the flash too long (> ~200 ms) the photons will be too spread out, the resulting currents in the retinal circuitry wouldn't sum, and we wouldn't see it. Thus, it makes sense to talk about the intensity of the flash in terms of the total number of photons city the duration. that only a fraction of the photons arriving at the eye are actually detected by rods. Some are scattered, some are absorbed, some pass through the retina without being detected. Measurements in original paper by Hecht, Shlaer, and Pirenne (1942) (HSP) estimated that oncome are reflected by the cornea, 50% are absorbed by the lens and other ocular media, and at least 80% passes through the retina without being absorbed. Overall, Hecht et al estimated that, for a typical flash, the range of 54 to 148 photons that arrive at the cornea, only absorbed by retinal rods. **Ting the photon stream** In lightflash(\(\lambda\); t1=0.8, t2=2.2) that returns an array of random (photon) time points at rate \(\lambda\) starting and t1 and stopping at t2. Write a function that simulates and plots the stream of photons for the three following stages: In stream at a rate of 100 photons / msec from times f1 to f2.
3. The subse Use this to simple aligned and def lightflatettettettettettettettettettettettettet	ppend(t) np.random.exponential(1/lam) p.array(PT)
<pre># full s P = ligh plotflas # s1 to P = P[(P plotflas # detect P = np.a</pre>	tflash(lam, f1, f2) h(P, "Photon Stream from f1 to f2", [f1-0.1, f2+0.1]) s2 - through shutter > s1) & (P < s2)] h(P, "Photon Stream through shutter", [f1-0.1, f2+0.1]) sed by retina rray([p for p in P if np.random.uniform() < alpha]) h(P, "Photons detected by rods", [f1-0.1, f2+0.1])
1.75 - 1.50 - 1.00 - 1.00 - 0.75 -	
2.00 1.75 -	8 1.0 1.2 1.4 1.6 1.8 2.0 2.2 Photon Release Distribution Photon Stream through shutter
Relative Weight of photons - 25.1	8 1.0 1.2 1.4 1.6 1.8 2.0 2.2 Photon Release Distribution
2.00	Photons detected by rods
3b. Probal	1.8 1.0 1.2 1.4 1.6 1.8 2.0 2.2 Photon Release Distribution Dility of seeing se combined effects, let I be the total number of photons arriving at the cornea and α be the fraction absorbed, then the average number detected is αI . This gives a revised expression for the detection probability
def probseei return 1 probseeing(5 0.0839179420 3c. Plotting A key insight from	3130343 7 % detected vs light intensity for different parameters 9 mm HSP was that it was possible to estimate the detection threshold K from the data of human subjects (who sat for long hours, in a completely dark chamber, detecting the dimmest possible flashes of light). Here we will reproduce a figure from Bialek (2012) which shows
Write a function so it's easier to Observe how co def plotdete # plot a for i in P = plt. # plot h if HSP i	e curve depends on K but not on α . In plot detection curve (α =0.5, K=6) that plots the percentage of light flashes detected as a function of the intensity I . You will want to write this is a way that allows you to overlay multiple curves on the same plot for different values of α and K . Make the x-axis log compare the shapes of different curves. The x-axis should range from 0.01 to 100. The transposition of the intensity I is a way that allows you to overlay multiple curves on the same plot for different values of α and K . Make the x-axis log compare the shapes of different curves. The x-axis should range from 0.01 to 100. The transposition of the intensity I is a way that allows you to overlay multiple curves on the same plot for different values of α and K . Make the x-axis log compare the shapes of different curves. The x-axis should range from 0.01 to 100. The transposition of the intensity I is a way that allows you to overlay multiple curves on the same plot for different values of α and K . Make the x-axis log compare the shapes of different curves. The x-axis should range from 0.01 to 100. The transposition of the intensity I is a way that allows you to overlay multiple curves on the same plot for different values of α and K . Make the x-axis log compared to α and α is a way that allows you to overlay multiple curves on the same plot for different values of α and K . Make the x-axis log compared to α and α is a way that allows you to overlay multiple curves on the same plot for different curves. The x-axis should range from 0.01 to 100. The transposition of α is a way that allows you to overlay multiple curves on the same plot for different curves. The x-axis log compared to α and α is a way that allows you to overlay multiple curves on the same plot for α and α is a way that allows you to overlay multiple curves. The x-axis log compared to α and α is a way that allows you to overlay multiple (α and α is a way that allows you to overla
plt.ylab plt.xlab	
p(Detection Flash)	—— alpha=0.5, K=6
Here we will re Use your plot fo 24.1, 37 0.0, 4	parameters to experimental data produce the figure 2 from the review by Rieke and Baylor (1998), which also references the classic HSP paper. Inction above to plot curves (overlaid) for the following pairs: $(\alpha=0.02, K=2)$ and $(\alpha=0.13, K=12)$. Now also overlay the following data points from HSP subject SS ("S" in HSP): 1.6, 58.6, 91.0, 141.9, 221.3 # SS: average photons at cornea 1.0, 18.0, 54.0, 94.0, 100.0 # SS: percent seen 1.2 evalues of α and K that best fit the data. Wrap your code above in a function plotfit (α, K) to make this easier. Use the fact that α and K will tend to co-vary (e.g. try plotfit $(\alpha=3, K=3)$), since increasing α will increase the number of photons reaching the restriction threshold must also increase to maintain the same performance curve.
# set data alpha = np.a K = np.array HSP = [[24.1 [0.00] # plot data plotdetection	, 37.6, 58.6, 91.0, 141.9, 221.3], 0, 0.040, 0.180, 0.540, 0.940, 1.000]] ncurve(alpha, K, (0, 250), HSP) Probability of Detection of a Flash - alpha=0.02, K=2 - alpha=0.13, K=12
p(Detection Flash)	HSP scatter
<pre># optima opt_alph opt_K = min_mse # search for a in for</pre>	<pre>a = None None</pre>
<pre># return return o a, k = optim print(f"opti optimal alph def plotfit(</pre>	mse = (1/len(probs)) * np.sum((probs-HSP[1])**2) # re-store if (min_mse is None) or (mse < min_mse):
opt_a, o # plot plotdete plotfit(sear	24.1, 37.6, 58.6, 91.0, 141.9, 221.3], [0.000, 0.040, 0.180, 0.540, 0.940, 1.000]] pt_k = optimalfit(HSP, alpha, K) if search else (alpha, K) ctioncurve([opt_a], [opt_k], (24.1-50, 221.3+50), HSP) ch=True) Probability of Detection of a Flash
p(Detection Flash)	alpha=0.09090909090909091, K=8 HSP scatter 0 50 100 150 200 250
0.8 -	htensity ch=False, alpha=(3), K=(3)) Probability of Detection of a Flash
b(Detection Flash) 0.0 - 0.0 -	— alpha=3, K=3 HSP scatter 0 50 100 150 200 250 ch=False, alpha=0.15, K=8)
b(Detection Flash) 1.0 - 0.0 - 0.0 - 0.4 -	Probability of Detection of a Flash
0.2 -	alpha=0.15, K=8 HSP scatter 0 50 100 150 200 250 ch=False, alpha=0.091, K=9) Probability of Detection of a Flash - alpha=0.091, K=9
1.0 -	HSP scatter
1.0 -	