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College of Computing and Information Technology, AASTMT  
2017

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## DNA Sequencing Error Correction Algorithms

Student:

- *Salma Gomaa*

Supervisors:

- *Prof. Dr. Yasser El-sonbaty*
  - *Dr. Nahla Belal*
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# Table of Contents

- Introduction
- Problem Definition
- Objective
- Background
- Related Work
- K-mer Grouping Error Correction Proposal
- H-RACER Proposal
- Evaluation
- Conclusion
- Future Work

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- Problem Definition
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# Deoxyribonucleic Acid (DNA)



# DNA Sequencing

- DNA sequence - a single format onto which a broad range of biological phenomena can be projected for high-throughput data collection

```
>Gene1
TTTCGGCGGTGCGCTATCCGGCGGAACTTTTGCGCGTGATGGCGAGTTCGGTCGCGGAAAGACGACCCTCGTGAATCGCCTTCGCTTT
CGATCCGCCGAGGCGATCCAAGTATCCGCATCCGGGATCGGACTCGTCAATGCGCAACCTGTGGACCGCAAGGAGATCGAGCGCAGGTC
GCGCTATGTCCACGAGGATGACCTCTTTATCGCGTCCCTAACGCGCAGGGAACACCTGATTTTCCAACGCATGGTCGGGATCGCACGAC
ATCTGACCTATCGCGACGGAGTGCGCCCGGTGGATCAGGTGATCCAGGACGTTTTCCGTACGAAATGTCACGACACGATCATCGGTGTC
GCCGCGAGGGTGAAAGGTCTGTCCGCGGGAGAAAGGAACGGTCTGCGATTCCGCTCCGAGCGTCTAACC GATCCCGCCGTTCTGATCTG
GATGACGCCACCTCCGGA CTGGACTCCTTTACCGCCACACGGTCGTCCAGGTGCTGAAGAACGTGTCCGAGAAGGCGAAGACCGTCAT
CCTGACCATTATCACGCGTCTTCCGACGTGTTTGACGTCTTTGACAAGATCCTTCTGATGCGCGAGGCGAGGGTACGTTTCTTGGCGA
CTCCACGGAACGCGTCGACTTCTTTCTA
```

# DNA Sequencing

- Next-generation DNA sequencing has the potential to dramatically accelerate biological and biomedical research
- NGS generates too many reads in a suitable time, which negatively affected the output accuracy

# NGS Errors Corrections:

→ Nucleotide Error Types:

- Substitution
- Insertion
- Deletion

# Table of Contents

- Introduction
- **Problem Definition**
- Objective
- Background
- Related Work
- K-mer Grouping Error Correction Proposal
- H-RACER Proposal
- Evaluation
- Conclusion
- Future Work



# Problem Definition

- DNA Next Sequencing Generation - *generates reads with many errors with different types*
- DNA Reads Accuracy - *is a vital factor in all of the DNA reads processes*

# Table of Contents

- Introduction
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- Background
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- Evaluation
- Conclusion
- Future Work

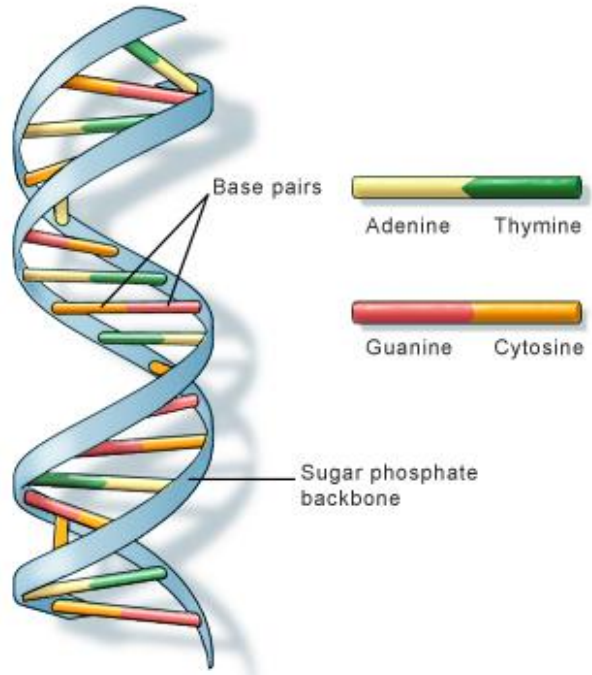
# Objective

- Raising the DNA reads accuracy
- Correcting all of the different types of errors
- Accomplishing the correction process within the shortest time

# Table of Contents

- Introduction
- Problem Definition
- Objective
- **Background**
- Related Work
- K-mer Grouping Error Correction Proposal
- H-RACER Proposal
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- Conclusion
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# Deoxyribonucleic Acid (DNA)



# Deoxyribonucleic Acid (DNA)

- Human DNA consists of about 3 billion bases
- The bases order determines the information available for building and maintaining an organism

# DNA Sequencing

→ DNA sequencing - outputs *fasta* or *fastq* file.

FASTA

>MISEQ-2:20:000000000-A61NM:1:1101:12299:1738 1:N:0:some\_name  
TGCGTCATCATCTTTGTCATCGTGTACTACGCCCTGATGGCTGGTGTGGTTTGGTTTGTGGTC

FASTQ

```
@MISEQ-2:20:000000000-A61NM:1:1101:12299:1738 1:N:0:some_name
TGCGTCATCATCTTTGTCATCGTGTACTACGCCCTGATGGCTGGTGTGGTTTGGTTTGTGGTC
+
AAAAADAFFFFFGGGFGGFGGFHFGFHHFGAEGIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII
```

# Next-Generation sequencing (NGS):

- NGS generates too many reads in a suitable time
- NGS introduced two painful issues:
  - Read Length Shortness
  - Reads Accuracy Decrement



# NGS Errors Corrections:

- Reads accuracy is a vital factor in all reads processes
- Detecting and Correcting errors is an essential step, and can be either:
  - Standalone Program
  - Process Preceding Step

# NGS Errors Corrections:

- Detecting and Correcting errors depends on:
  - Nucleotide Frequency
  - Nucleotide Quality Value
  
- Nucleotide Error Types:
  - Substitution
  - Insertion
  - Deletion

# NGS Errors - Substitution Error

T	C	T	C	G
---	---	---	---	---

# NGS Errors - Substitution Error

→ *Substitution Error*

T	C	T	C	G
T	C	<u>A</u>	C	G

# NGS Errors - Substitution Error

T	C	<u>A</u>	C	G
---	---	----------	---	---

# NGS Errors - Insertion Error

T	C	T	C	G
---	---	---	---	---

# NGS Errors - Insertion Error

T	C	T		C	G
T	C	<u>G</u>	T	C	G

# NGS Errors - Insertion Error

T	C	<u>G</u>	T	C	G
---	---	----------	---	---	---



# NGS Errors - Deletion Error

T	C	T	C	G
---	---	---	---	---

# NGS Errors - Deletion Error

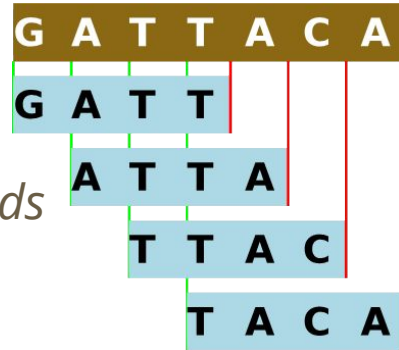
T	C	<b>T</b>	C	G
T	C	-	C	G

# NGS Errors - Deletion Error

T	C	C	G
---	---	---	---

# Correction Concepts and Definitions

→ K-mer - *All the possible sub-sequences (of length  $k$ ) from a read*



→ K-mer Frequency - *Number of a  $k$ -mer repetition in all the reads*

→ K-mer Frequency Threshold - *A preset threshold used in classifying  $k$ -mers*

# Correction Concepts and Definitions

- Coverage - *Number of reads that include a given nucleotide in the sequence*

$$C = LN/G$$

- Spectrum Alignment - *A filtration step that classifies the k-mers into strong and weak k-mers*
- Spectrum alignment depends on the k-mers frequencies and/or the nucleotides quality values

# Correction Concepts and Definitions

→ Correction can take place with:

- Spectrum Alignment - *by obtaining the nucleotides substitutions that leads to reduce the weak k-mers count*
- Tree Breadth-First Search - *by traversing multi out-going edges nodes, and removing fewer reads paths, then re-aligns them to the existing path*

# Correction Concepts and Definitions

→ Correction can take place with:

- Reads Alignments - *by aligning reads with a common k-mer, then fixing misaligned nucleotides based on their occurrences and quality values*
- Suffix Array - *built using a string of reads, and the correction takes place with the letter that appears most at each position*

# Correction Concepts and Definitions

→ Correction can take place with:

- Suffix Trie - *the edges are labelled with DNA letters, where the correction is based on the number of leaves in the sub-trie rooted at the node*
- K-mer Hashing Table - *by storing the total times each nucleotide appears before and after a k-mer, where the error is corrected via the counts*



# Correction Concepts and Definitions

→ Correction can take place with:

- K-mer Discontinuities - *the frequencies of adjacent k-mers, where the correction is based on the removal or minimizing the discontinuity*

# Table of Contents

- Introduction
- Problem Definition
- Objective
- Background
- **Related Work**
- K-mer Grouping Error Correction Proposal
- H-RACER Proposal
- Evaluation
- Conclusion
- Future Work

# Related Work

→ Correction Methodologies can be classified into two categories:

- Substitution only correction
- Substitution, insertion and deletions correction

# Related Work

→ Substitution only correction methodology, like:

	Euler	Velvet	AllPaths	SOAP	Quake	Reptile	CUDA	HiTEC	RACER	EC
Year	2004	2008	2008	2010	2010	2010	2010	2011	2013	2015
Used Concept	Spectrum Alignment	Tree BFS	Spectrum Alignment	Tree BFS	Spectrum Alignment	Spectrum Alignment	Spectrum Alignment	Suffix Array	Hash Table	Hash Table
Depends On	K-mer Freq.	Nuc. Freq.	K-mer Freq. with Nuc. QV	K-mer Freq.	K-mer Freq. with Nuc. QV	K-mer Freq. with Nuc. QV	K-mer Freq. with Votes	Nuc. Freq.	Nuc. Freq.	Nuc. Freq.

# Related Work

→ Substitution, insertion and deletions correction methodology , like:

	<b>HSHREC</b>	<b>Coral</b>	<b>Pollx</b>
<b>Year</b>	2010	2011	2015
<b>Used Concept</b>	Suffix Trie	Reads Alignment	K-mer Discontinuities
<b>Depends On</b>	Nuc. Freq.	Nuc. Freq. with QV	K-mer Freq.

# Related Work

→ RACER characteristics:

- Ability to correct data sets that have varying read lengths
- Hash table and k-mer nucleotides neighbours
- Fastest DNA error correction algorithm existent nowadays with a high accuracy
- Corrects substitutions only

# Table of Contents

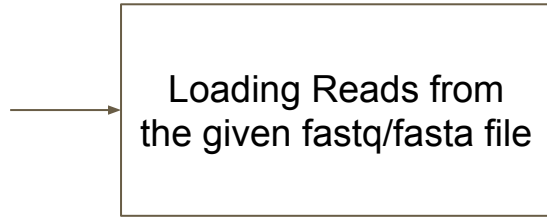
- Introduction
- Problem Definition
- Objective
- Background
- Related Work
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# K-mer Grouping Error Correction Proposal

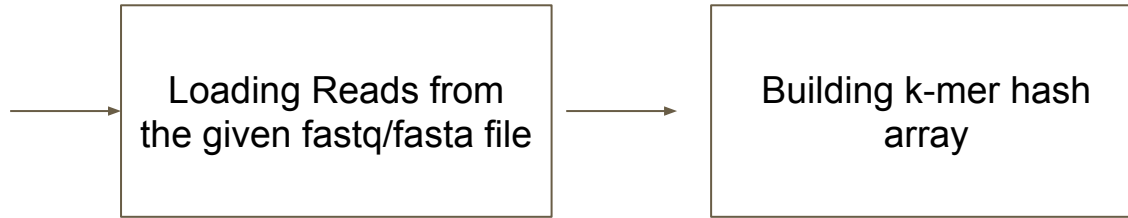
- Aiming to correct all types of errors
- Hashing the k-mers into integers
- Flexible to run more correction iterations



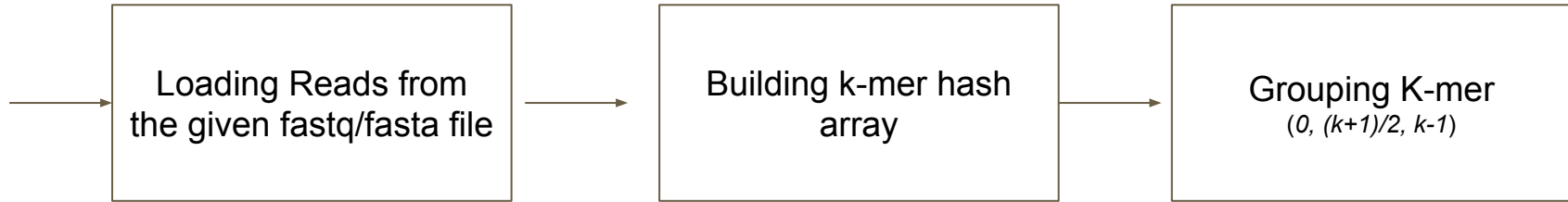
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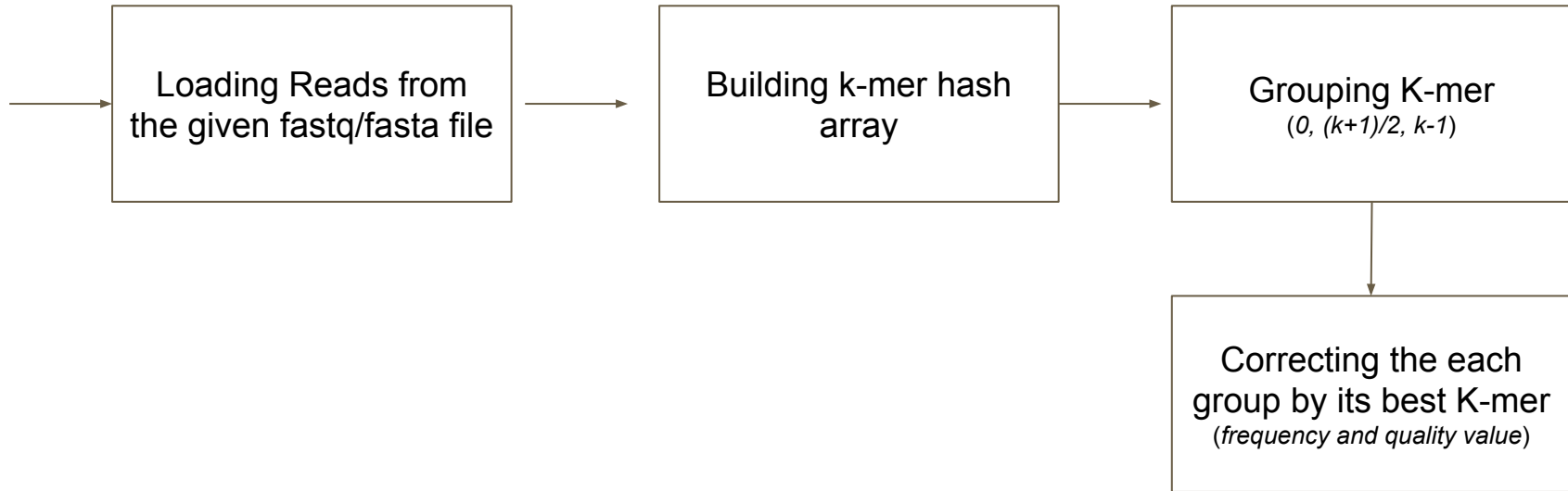
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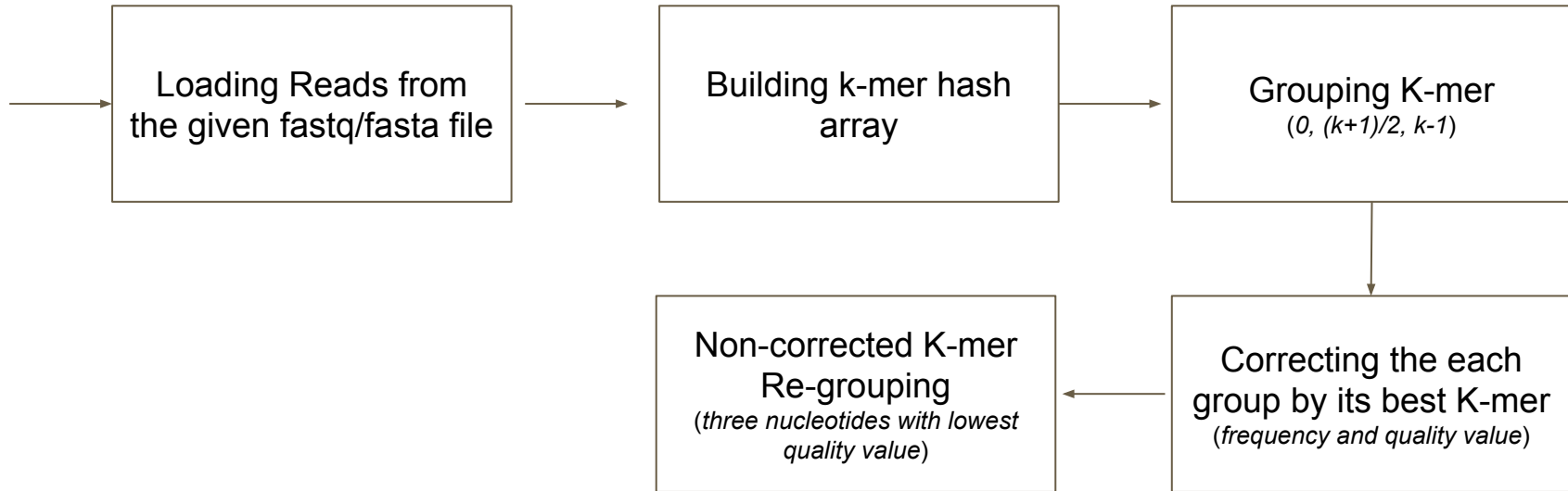
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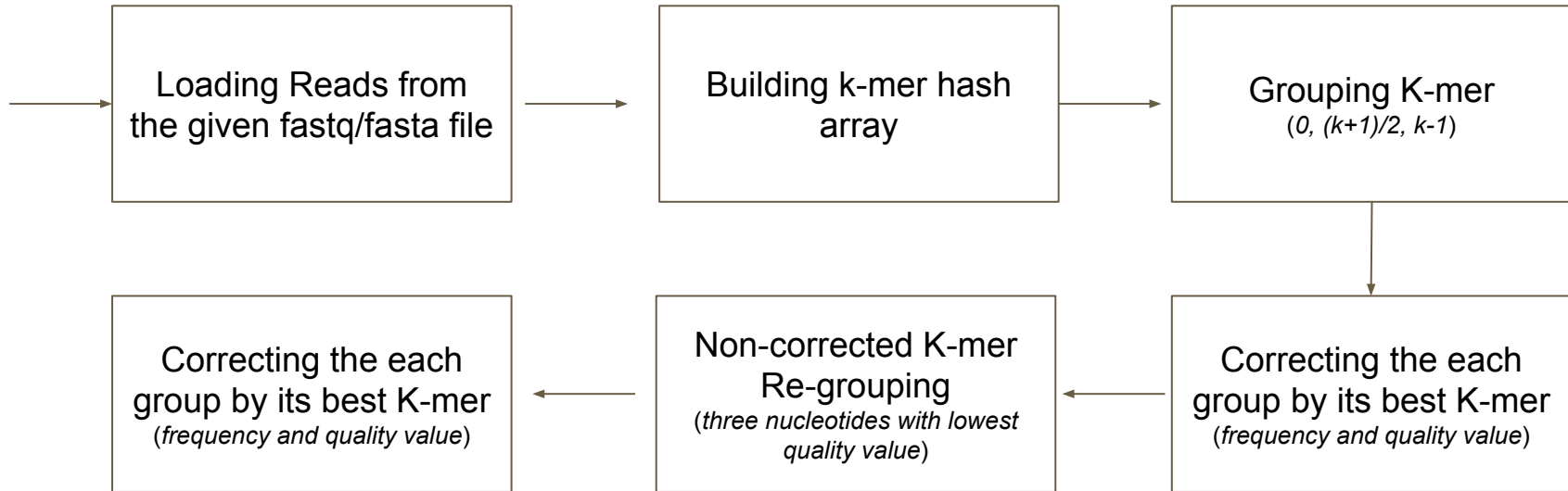
# K-mer Grouping Error Correction Proposal



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# K-mer Grouping Error Correction Proposal



# K-mer Grouping Error Correction Proposal

id	k-mer	Freq
0	CCGTAAT	9
1	CGCTACT	13
2	GTACGGT	8
3	AGCTACT	4
4	GTCCTGT	3
5	ACGTAAT	2
6	GGCCACT	2
7	GGCCTAA	1
8	TGCCACC	3

# K-mer Grouping Error Correction Proposal

id	k-mer	Freq	Gp
0	CCGTAAT	9	-1
1	CGCTACT	13	-1
2	GTACGGT	8	-1
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4	GTCCTGT	3	-1
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4	GTCCTGT	3	-1
5	<u>ACG</u> <u>TAAT</u>	2	-1
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4	GTCCTGT	3	-1
5	<u>ACG</u> <u>TAAT</u>	2	0
6	<u>GGCC</u> <u>ACT</u>	2	-1
7	GGCCTAA	1	-1
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4	GTCCTGT	3	-1
5	<u>ACG</u> <u>TAAT</u>	2	0
6	<u>GGCCACT</u>	2	1
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3	<u>AGCTACT</u>	4	1
4	GTCCTGT	3	-1
5	<u>ACG</u> <u>TAAT</u>	2	0
6	<u>GGCCACT</u>	2	1
7	GGCCTAA	1	-1
8	<u>IGCCACC</u>	3	-1

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1	<u>CGCTACT</u>	13	1
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3	<u>AGCTACT</u>	4	1
4	GTCCTGT	3	-1
5	<u>ACG</u> <u>TAAT</u>	2	0
6	<u>GGCCACT</u>	2	1
7	GGCCTAA	1	-1
8	<u>TGCCACC</u>	3	1

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id	k-mer	Freq	Gp
0	<u>CCG</u> <u>TAAT</u>	9	0
5	<u>ACG</u> <u>TAAT</u>	2	0

id	k-mer	Freq	Gp
1	<u>CGCT</u> <u>ACT</u>	13	1
3	<u>AGCT</u> <u>ACT</u>	4	1
6	<u>GGCC</u> <u>ACT</u>	2	1
8	<u>TGCC</u> <u>ACC</u>	3	1

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# K-mer Grouping Error Correction Proposal

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id	k-mer	Freq	Gp
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# K-mer Grouping Error Correction Proposal

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4	GT <u>CCT</u> GT	3	-1
7	GGCCTAA	1	-1

# K-mer Grouping Error Correction Proposal

id	k-mer	Freq	Gp
2	GT <u>AC</u> GGT	8	0
4	GTC <u>CT</u> TGT	3	0
7	GGCCTAA	1	-1

# K-mer Grouping Error Correction Proposal

id	k-mer	Freq	Gp
2	GT <u>AC</u> GGT	8	0
4	GT <u>CCT</u> GT	3	0

id	k-mer	Freq	Gp
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# K-mer Grouping Error Correction Proposal

id	k-mer	Freq	Gp
2	GT <u>AC</u> GGT	8	0
4	GT <u>CCT</u> GT	3	0

id	k-mer	Freq	Gp
7	GGCCTAA	1	-1

# K-mer Grouping Error Correction - Evaluation

→ Data sets

Name	Genome Length	Read Length	Number of Reads	Coverage
Lactococcus Lactis	2,598,144	36	4,370,050	60.55

# K-mer Grouping Error Correction - Evaluation

→ Lactococcus Lactis (G: 2,598,144 - L: 36, N: 4,370,050, C: 60.55)

	Coral	Pollux	SHSREC	KGEC
Accuracy in Percentage	91.45	94.15	95.34	95.39
Time in Minutes	5	3	15	61

# K-mer Grouping Error Correction Defects

- This algorithm is mainly dependent on the k-mers grouping
- kmers grouping takes place by generating all of the possible cases of the corrections of every kmer, and here goes the time defect (exponential)
- On removing the method with the exponential complexity, the accuracy of the algorithm has been greatly negatively affected.



# K-mer Grouping Error Correction Defects

- The main major step of the proposal implies to it's weakness point, which proves that this proposal won't get a better results
- So, it fails to run on big data
- Using real data sets to get a good indication of real life performance

# Table of Contents

- Introduction
- Problem Definition
- Objective
- Background
- Related Work
- K-mer Grouping Error Correction Proposal
- **H-RACER Proposal**
- Evaluation
- Conclusion
- Future Work

# H-RACER Proposal

- H-RACER is a newly correction approach for correcting all types of errors.
- H-RACER is inherited from RACER
- RACER is the fastest algorithm specialized in correction substitution errors only

# H-RACER Proposal

→ RACER characteristics:

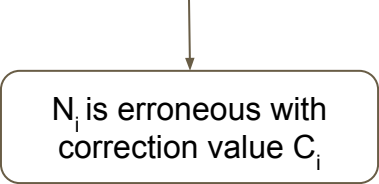
- Ability to correct data sets that have varying read lengths
- Hash table and k-mer nucleotides neighbours
- Fastest DNA error correction algorithm existent nowadays with a high accuracy
- Corrects substitutions only

# H-RACER Proposal

- H-RACER uses the same algorithm of RACER in detecting errors and deciding corrections values
- H-RACER detects the error type for an erroneous nucleotide by studying its correction value against its neighbours
- H-RACER decides the corrective action (substitute, insert, delete) according to the detected error type

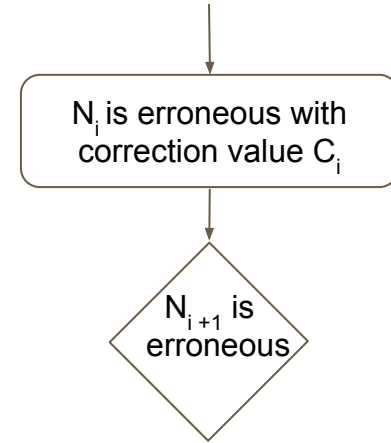
# H-RACER - Error Type Detection

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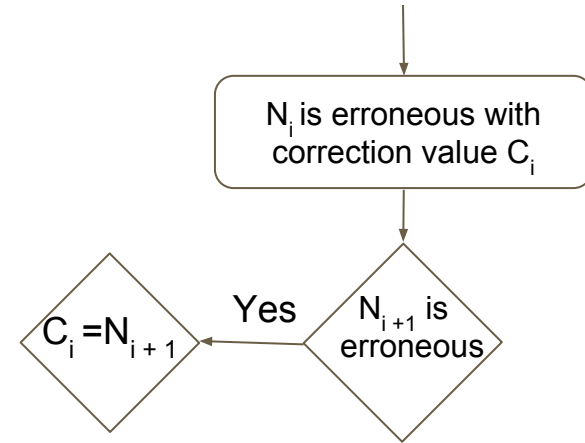
$N_i$  is erroneous with  
correction value  $C_i$

# H-RACER - Error Type Detection

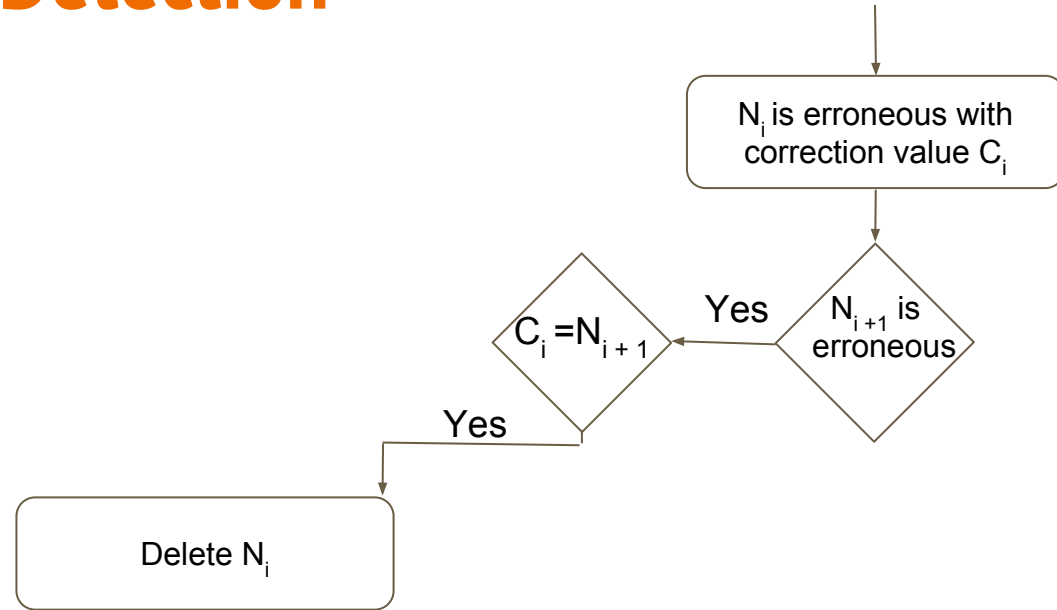




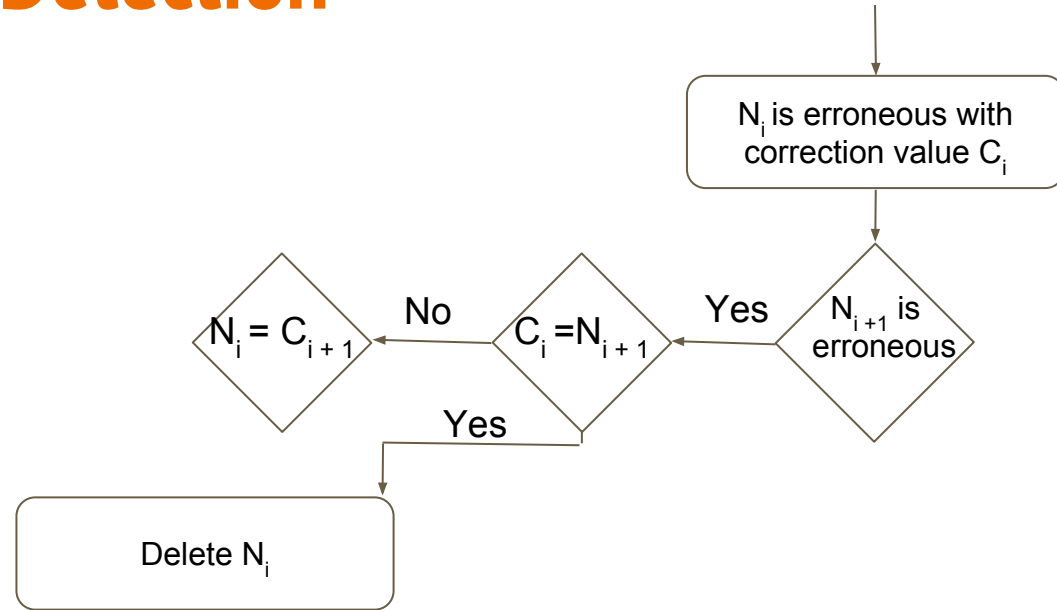
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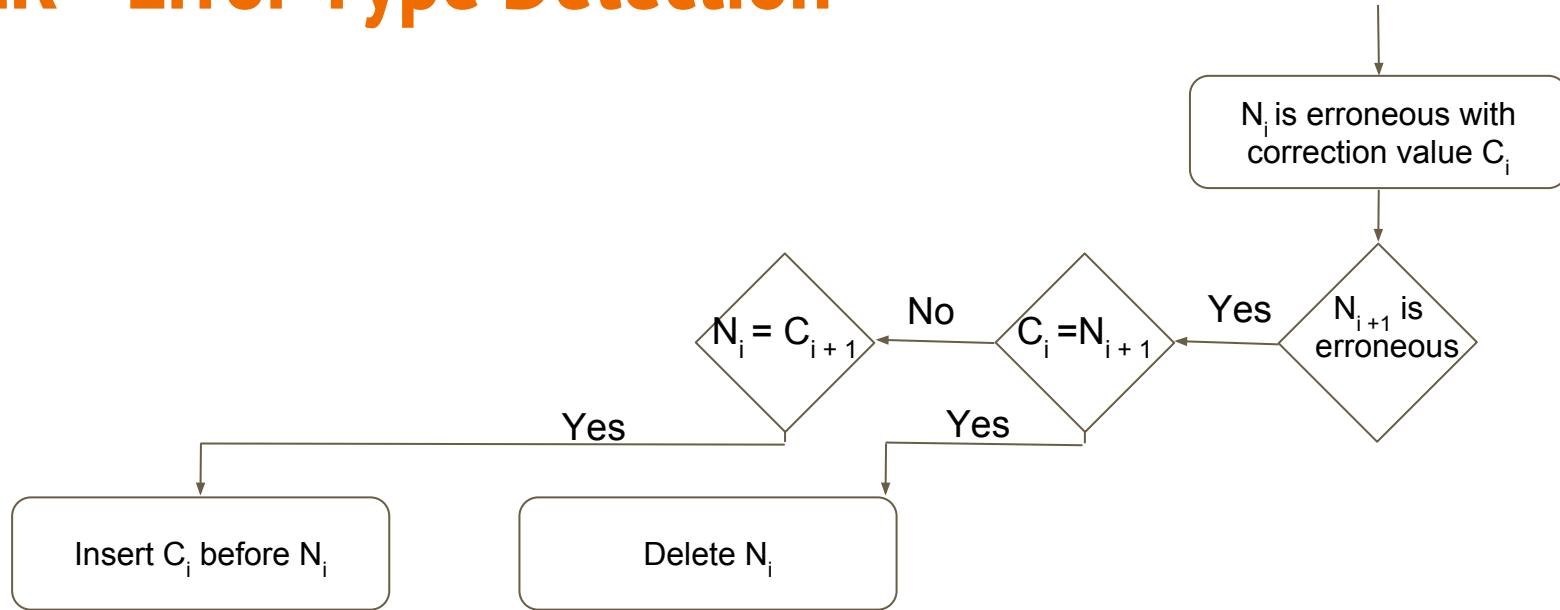
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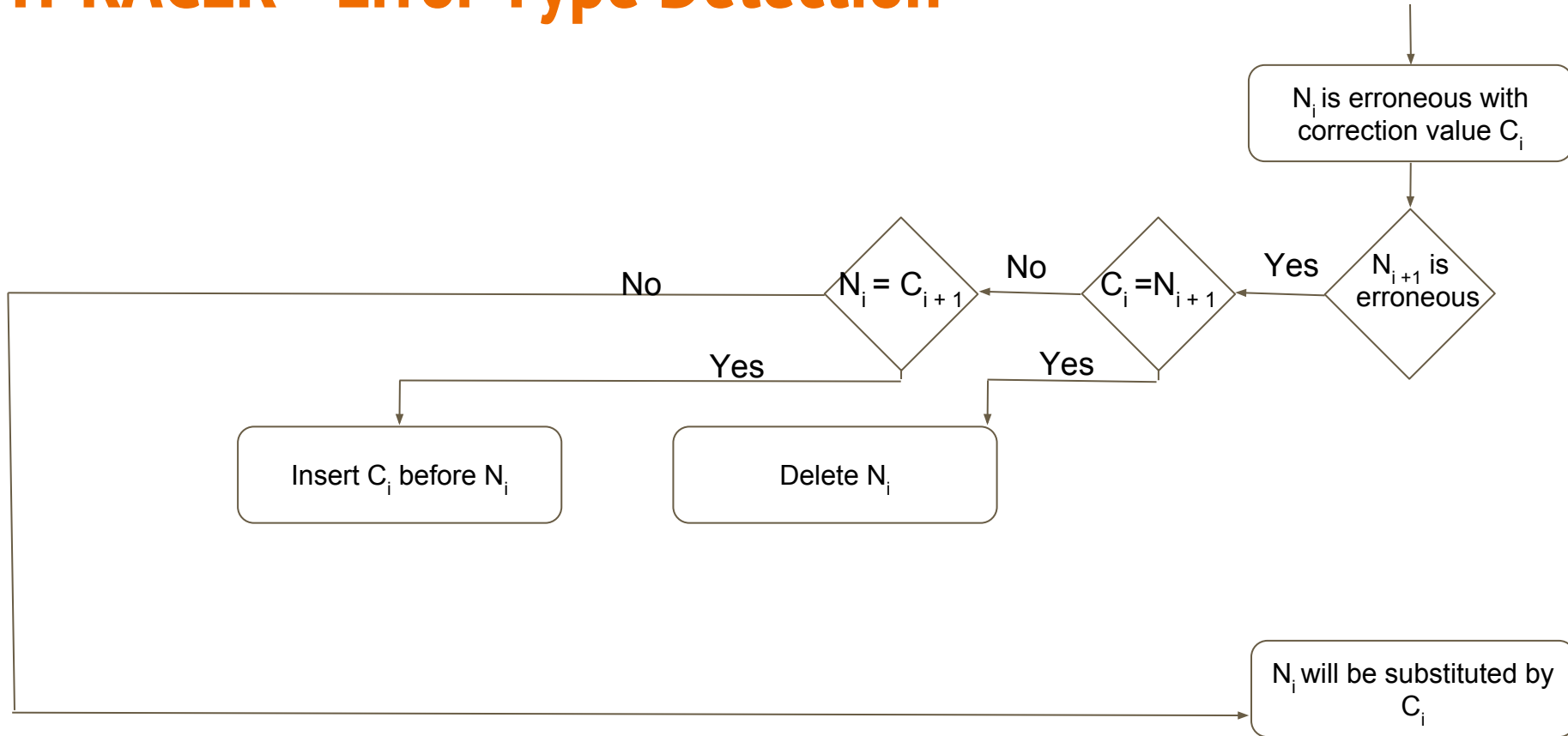
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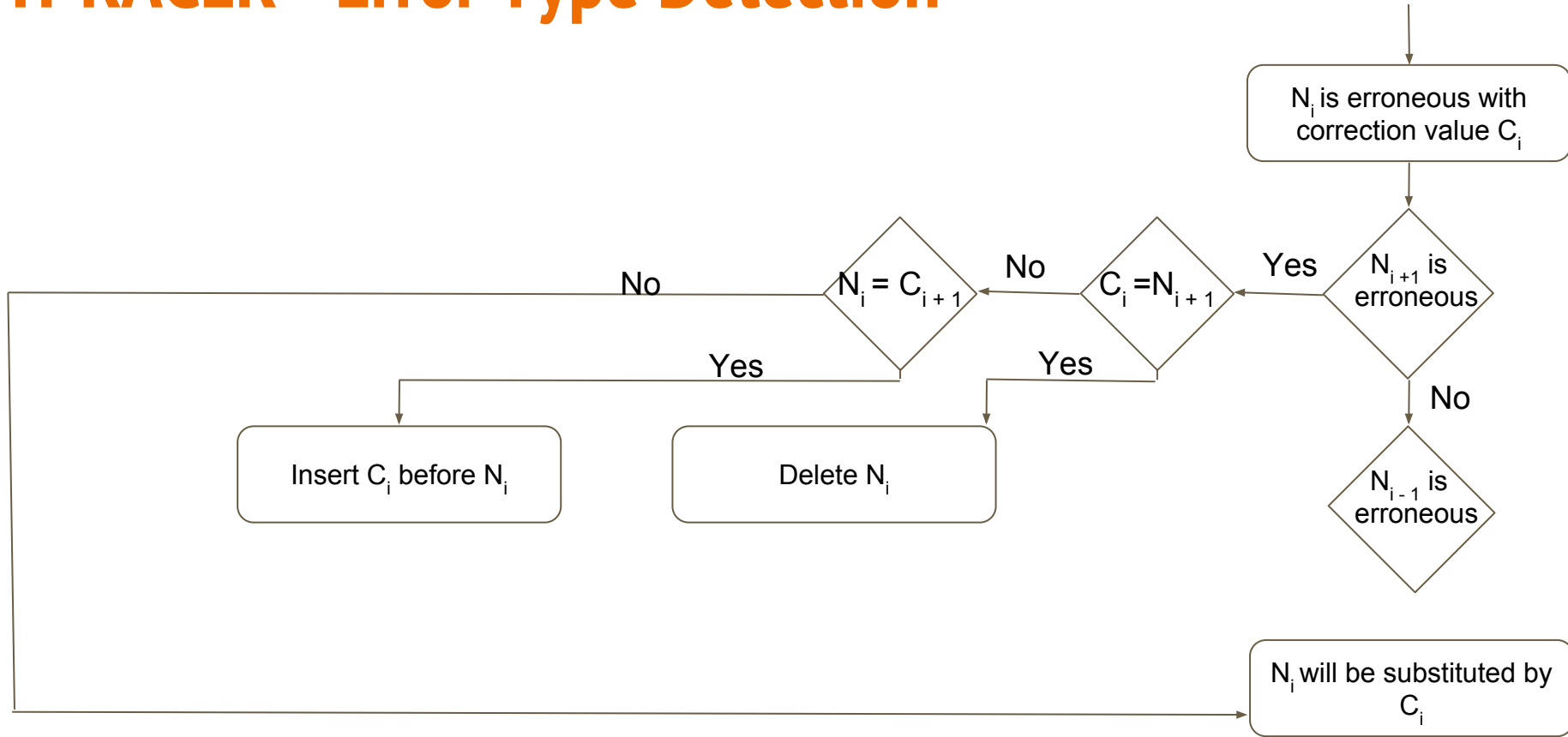
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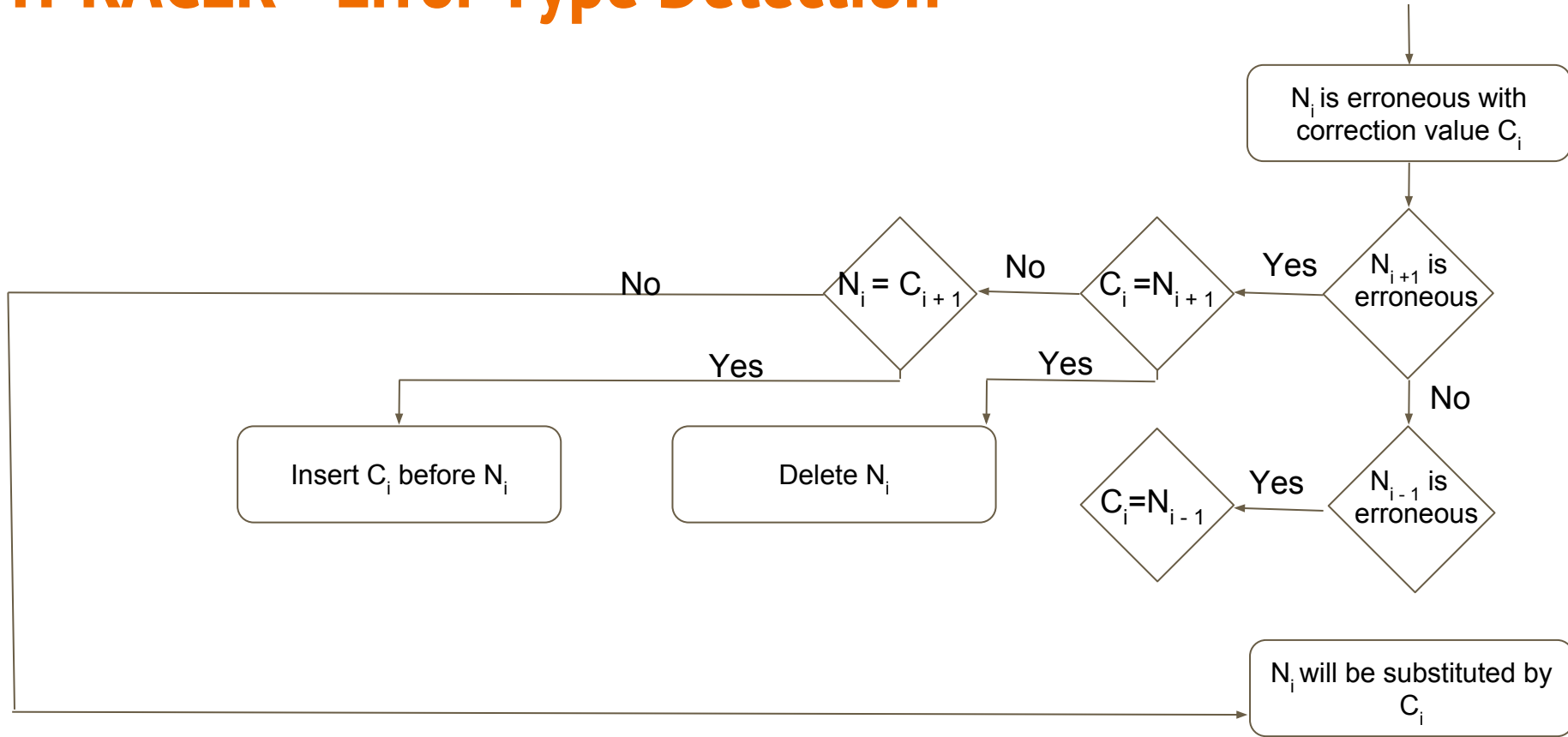
# H-RACER - Error Type Detection



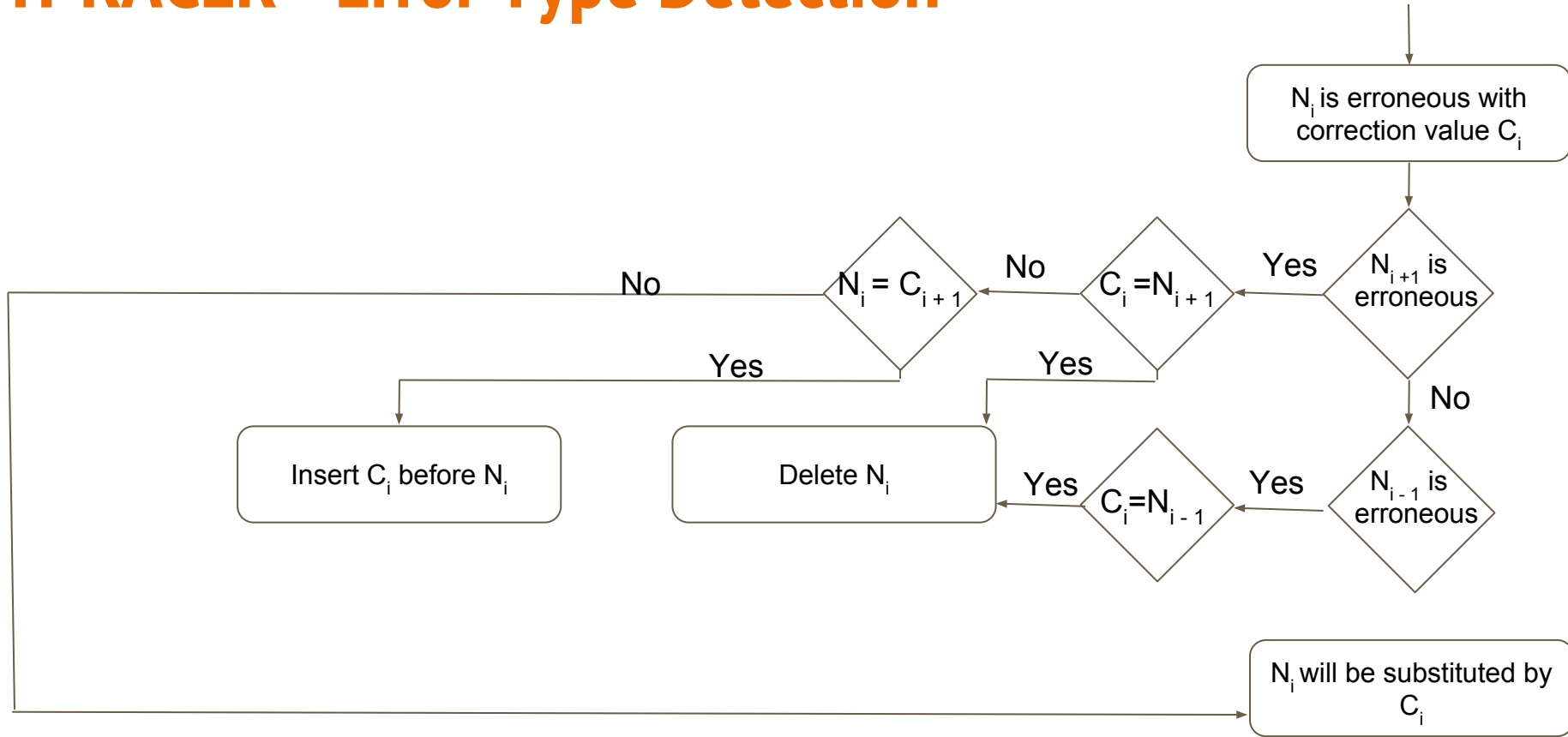
# H-RACER - Error Type Detection



# H-RACER - Error Type Detection

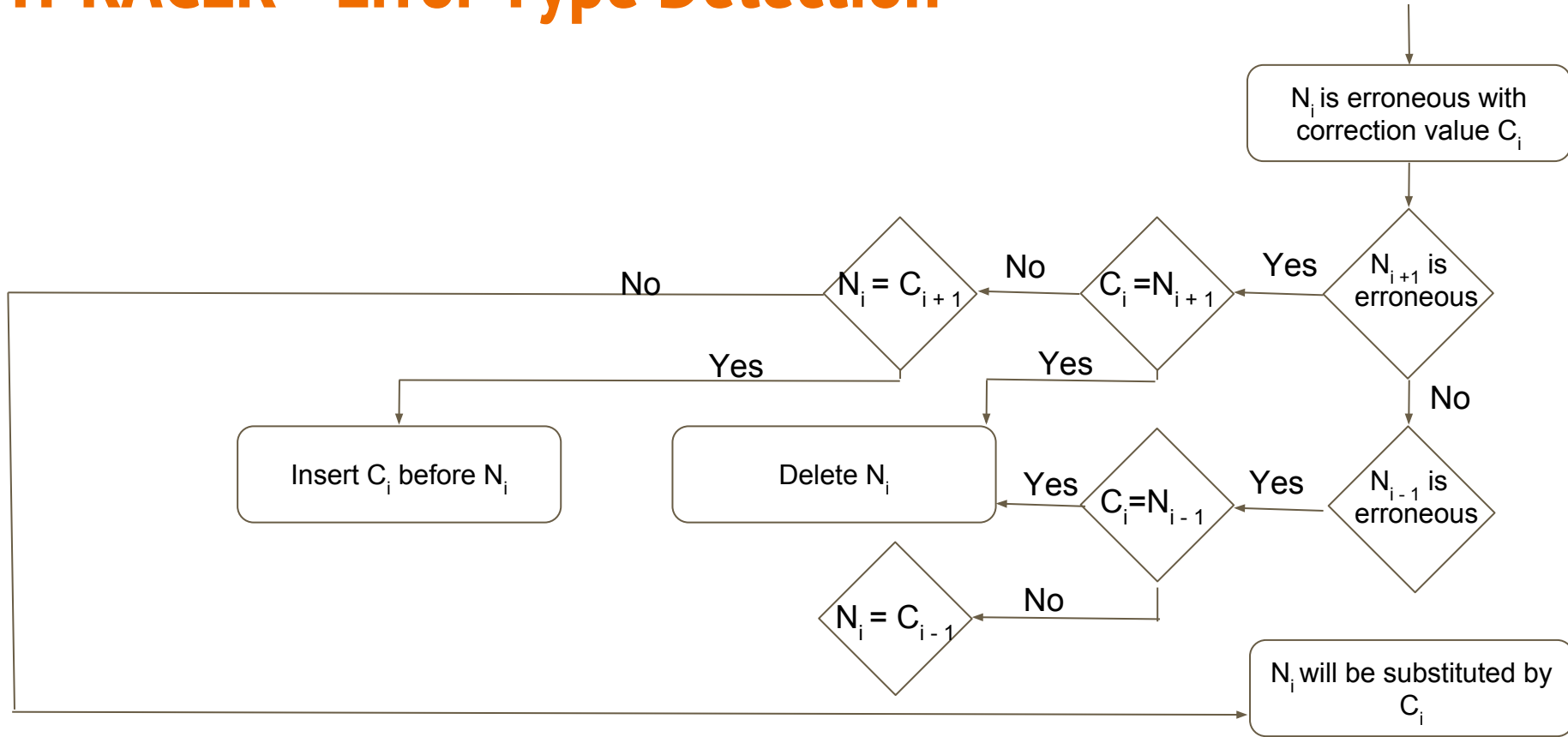


# H-RACER - Error Type Detection

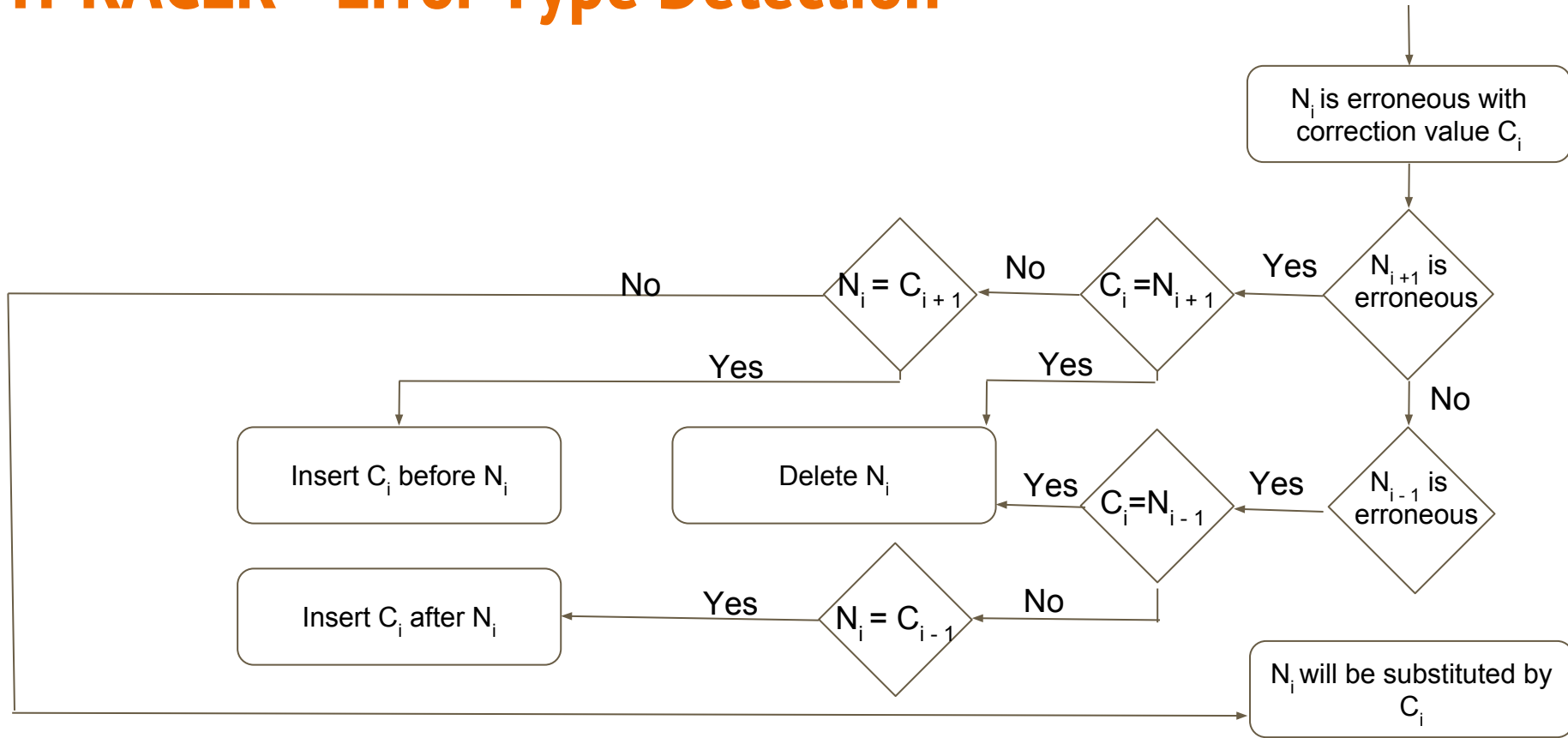




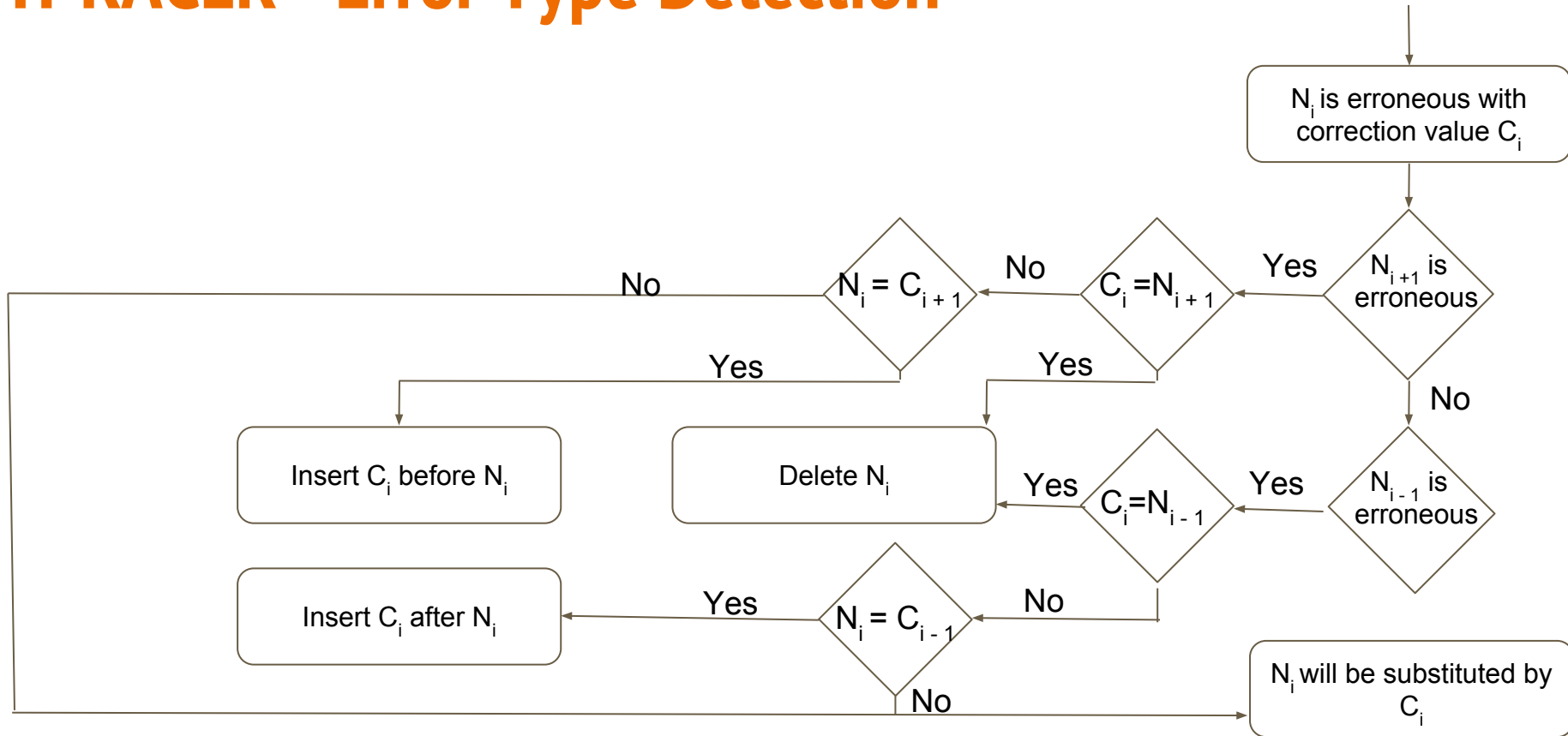
# H-RACER - Error Type Detection



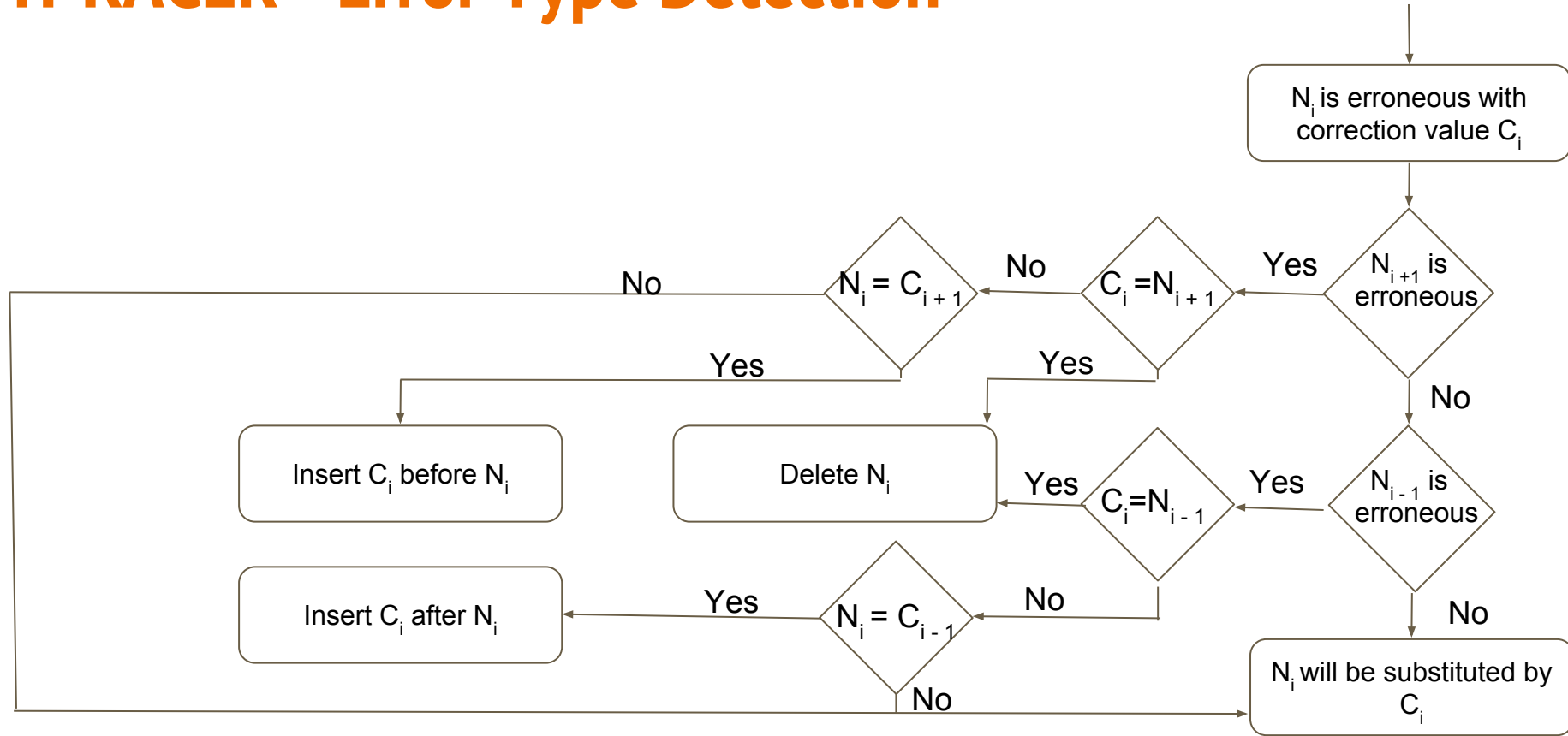
# H-RACER - Error Type Detection



# H-RACER - Error Type Detection



# H-RACER - Error Type Detection



# H-RACER - Error Type Detection

→ Erroneously Inserted Nucleotide

... A C C A T G ...

# H-RACER - Error Type Detection

→ Erroneously Inserted Nucleotide

... A C C A T G ...

# H-RACER - Error Type Detection

→ Erroneously Inserted Nucleotide

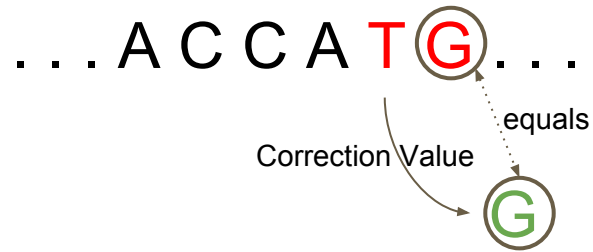
... A C C A T G ...

Correction Value

G

# H-RACER - Error Type Detection

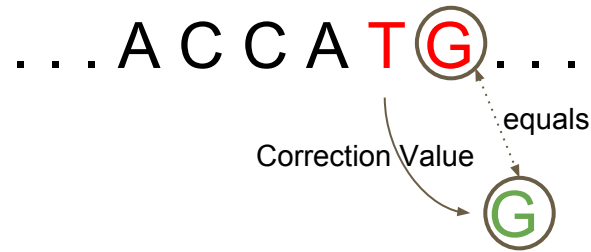
→ Erroneously Inserted Nucleotide





# H-RACER - Error Type Detection

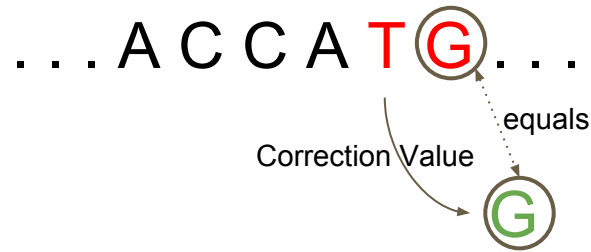
→ Erroneously Inserted Nucleotide



∴ T - *erroneously inserted nucleotide*

# H-RACER - Error Type Detection

→ Erroneously Inserted Nucleotide

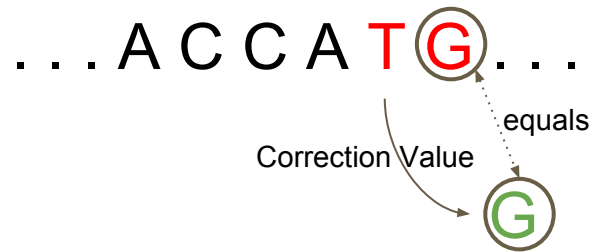


∴ **T** - *erroneously inserted nucleotide*

∴ Correction - *delete T*

# H-RACER - Error Type Detection

→ Erroneously Inserted Nucleotide



∴ **T** - *erroneously inserted nucleotide*

∴ Correction - *delete T*

... A C C A **G** ...

# H-RACER - Error Type Detection

→ Erroneously Deleted Nucleotide

. . . A A C C T G . . .

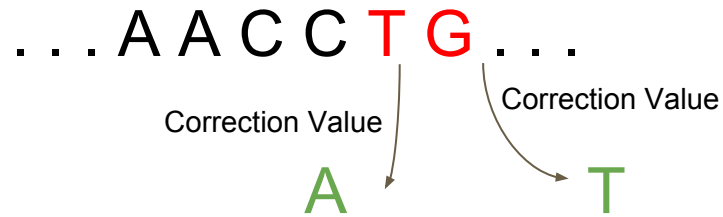
# H-RACER - Error Type Detection

→ Erroneously Deleted Nucleotide

. . . A A C C T G . . .

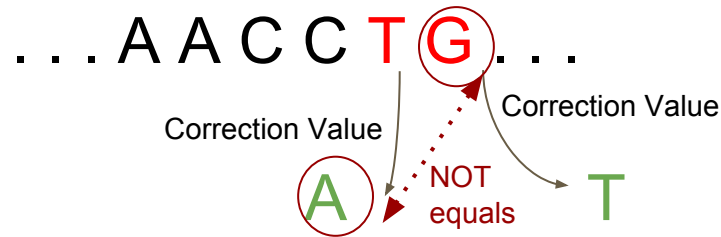
# H-RACER - Error Type Detection

→ Erroneously Deleted Nucleotide



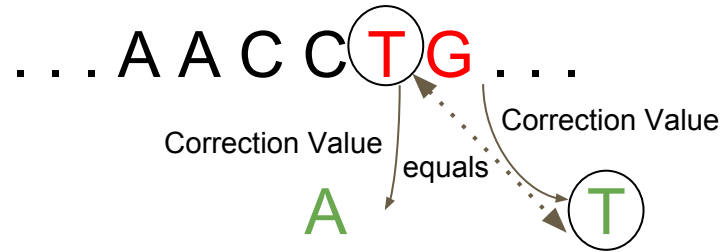
# H-RACER - Error Type Detection

→ Erroneously Deleted Nucleotide



# H-RACER - Error Type Detection

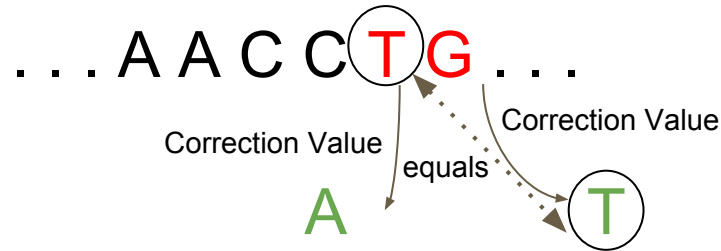
→ Erroneously Deleted Nucleotide





# H-RACER - Error Type Detection

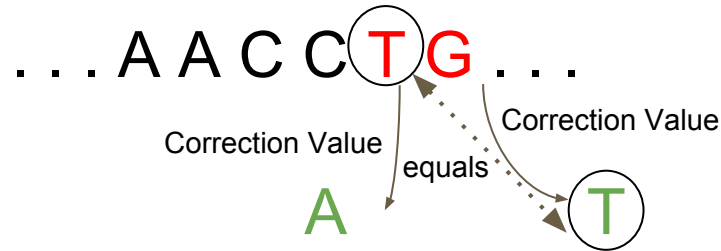
→ Erroneously Deleted Nucleotide



∴ *A - erroneously deleted nucleotide*

# H-RACER - Error Type Detection

→ Erroneously Deleted Nucleotide

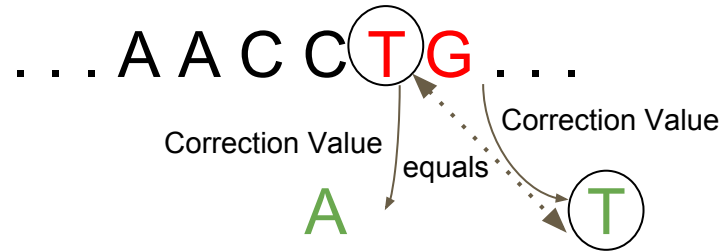


∴ *A* - *erroneously deleted nucleotide*

∴ *Correction - insert A*

# H-RACER - Error Type Detection

→ Erroneously Deleted Nucleotide



∴ *A* - erroneously deleted nucleotide

∴ Correction - insert *A*

... A A C C A T G ...

# H-RACER - Error Type Detection

→ Erroneously Substituted Nucleotide

... A A C C T G ...

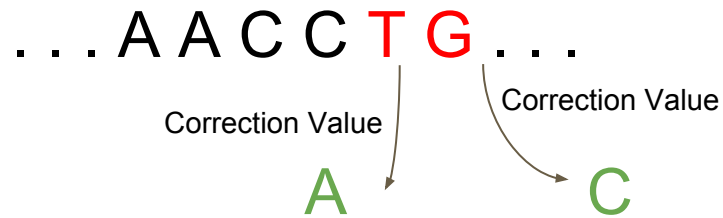
# H-RACER - Error Type Detection

→ Erroneously Substituted Nucleotide

. . . A A C C T G . . .

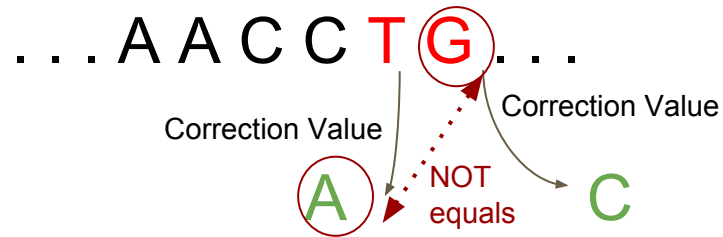
# H-RACER - Error Type Detection

→ Erroneously Substituted Nucleotide



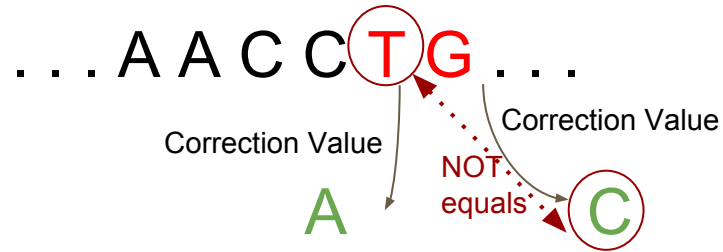
# H-RACER - Error Type Detection

→ Erroneously Substituted Nucleotide



# H-RACER - Error Type Detection

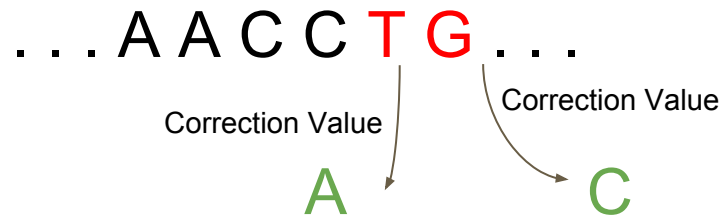
→ Erroneously Substituted Nucleotide





# H-RACER - Error Type Detection

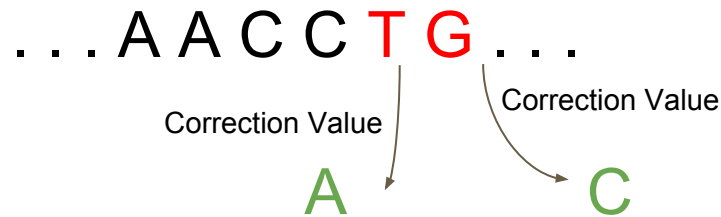
→ Erroneously Substituted Nucleotide



∴ A - erroneously substituted by T, and, C - erroneously substituted by G

# H-RACER - Error Type Detection

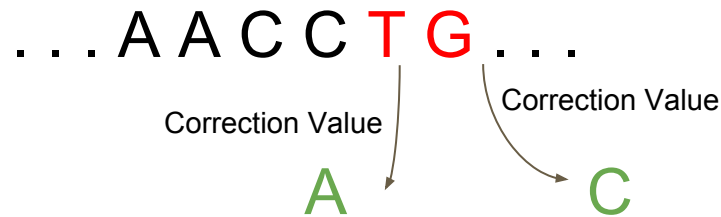
→ Erroneously Substituted Nucleotide



- ∴ `A` - erroneously substituted by `T`, and, `C` - erroneously substituted by `G`
- ∴ Correction - substitute `T` by `A` and `G` by `C`

# H-RACER - Error Type Detection

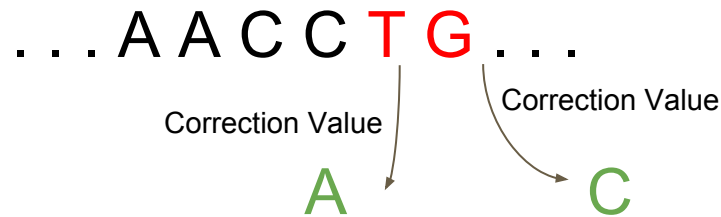
→ Erroneously Substituted Nucleotide



- ∴ `A` - erroneously substituted by `T`, and, `C` - erroneously substituted by `G`
- ∴ Correction - substitute `T` by `A` and `G` by `C`

# H-RACER - Error Type Detection

→ Erroneously Substituted Nucleotide



∴ `A` - erroneously substituted by `T`, and, `C` - erroneously substituted by `G`

∴ Correction - substitute `T` by `A` and `G` by `C`

`... A A C C A C ...`

# Table of Contents

- Introduction
- Problem Definition
- Objective
- Background
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- H-RACER Proposal
- **Evaluation**
- Conclusion
- Future Work

# Evaluation Definitions

- True Positive - *Properly detected as erroneous and properly corrected*
- False Positive - *Improperly counted as erroneous and improperly corrected*
- False Negative - *Improperly considering as not erroneous*

# Evaluation Definitions

→ True Negative - *Properly considering as not erroneous*

→ Sensitivity - *Ability to detect the erroneous nucleotides*

$$\text{Sensitivity} = TP / (TP + FN)$$

# Evaluation Definitions

→ Specificity - *Ability to properly corrects the erroneous nucleotides*

$$\text{Specificity} = \text{TN} / (\text{TN} + \text{FP})$$

→ Accuracy - *All over error rate*

$$\text{Accuracy} = (\text{TP} + \text{TN}) / (\text{TP} + \text{FP} + \text{FN} + \text{TN})$$



# H-RACER - Evaluation

- Data sets were brought from the National Center for Biotechnology Information (NCBI)
- Executing on amazon elastic cloud (AWS EC2) instance with 32 vCPU and 244GiB RAM, with Linux (Ubuntu) operating system
- Verified by a standalone C/C++ program implemented by RACER, that has the advantage of avoiding the interference of mapping/assembling programs

# H-RACER - Evaluation

→ Data sets

Name	Genome Length	Read Length	Number of Reads	Coverage
Lactococcus Lactis	2,598,144	36	4,370,050	60.55
Treponema Pallidum	1,139,417	35	7,133,663	219.13
E.coli 75a	4,639,675	75	3,454,048	55.83
E.coli 75b	4,639,675	75	4,341,061	70.17

# H-RACER - Evaluation

→ Lactococcus Lactis

	Coral	Pollux	HSRREC	H-RACER
True Positive in Millions	15.4	25.3	25.5	21.2
False Positive in Millions	2.0	7.7	6.1	0.02
False Negative in Millions	11.4	1.5	1.3	5.6
True Negative in Millions	128.5	122.8	124.5	130.5
Sensitivity in Percentage	57.43	94.46	95.25	79.22
Specificity in Percentage	98.44	94.08	95.36	99.98
Accuracy in Percentage	91.45	94.15	95.34	96.45
Time in Minutes	5	3	15	1

# H-RACER - Evaluation

→ Treponema Pallidum

	Coral	Pollux	HSRREC	H-RACER
True Positive in Millions	25.60	63.9	64.4	56.3
False Positive in Millions	3.5	8.8	8.1	0.2
False Negative in Millions	41.6	3.3	2.7	10.8
True Negative in Millions	179.1	173.7	174.4	182.4
Sensitivity in Percentage	38.08	95.15	95.95	83.87
Specificity in Percentage	98.10	95.16	95.55	99.88
Accuracy in Percentage	81.97	95.16	95.65	95.58
Time in Minutes	12	3	22	2

# H-RACER - Evaluation

→ E.coli 75a

	Coral	Pollux	HSRREC	H-RACER
True Positive in Millions	26.4	80.0	N/A	76.3
False Positive in Millions	5.6	31.7	N/A	0.03
False Negative in Millions	73.7	20.2	N/A	23.8
True Negative in Millions	153.4	127.2	N/A	158.9
Sensitivity in Percentage	26.40	79.87	N/A	76.21
Specificity in Percentage	96.51	80.07	N/A	99.98
Accuracy in Percentage	69.40	79.99	N/A	90.79
Time in Minutes	9	16	N/A	1

# H-RACER - Evaluation

→ E.coli 75b

	Coral	Pollux	HSRSEC	H-RACER
True Positive in Millions	13.3	99.4	N/A	81.06
False Positive in Millions	3.7	37.8	N/A	0.04
False Negative in Millions	108.5	22.4	N/A	40.8
True Negative in Millions	200.0	166.0	N/A	203.7
Sensitivity in Percentage	10.93	81.58	N/A	66.54
Specificity in Percentage	98.19	81.46	N/A	99.98
Accuracy in Percentage	65.55	81.50	N/A	87.47
Time in Minutes	13	21	N/A	2

# H-RACER - Evaluation

- H-RACER has the best results in accuracy and time, especially for long genomes
- H-RACER uses the bitwise orientation in implementation (inherited from RACER), so it shows the best time
- H-RACER error detection algorithm has a complexity  $O(r)$ , where  $r$  is the number of reads

# H-RACER - Evaluation

- H-RACER has the best accuracy, as it depends on:
  - Lowering false positive rate
  - Lowering sensitivity rate
- Lowering false positive rate negatively affects the true positive and false negative rates



# H-RACER - Evaluation

- Enhancing the reads overall accuracy is the main vital target. So, corrective algorithms should not introduce errors (represented in false positive rate).
- H-RACER has the best accuracy although it hasn't the best sensitivity rate

# H-RACER - Evaluation

- Using genomes with high coverage rate, negatively affects H-RACER by:
- Increasing error detection ambiguity
  - Raising false negative rate
  - Lowering accuracy

# H-RACER - Evaluation

- The comparisons were established between H-RACER and algorithms specialized in correcting all types of errors
- Using real data sets, to get a good indication of real life performance
- Different data sets, with different read length, genome size and coverage

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# Conclusion

- H-RACER acquires the major advantages of RACER in both aspects performance and time
- H-RACER added its elegant algorithm in detecting the errors types and properly applying their corrections

# Conclusion

- H-RACER is the fastest with the highest accuracy algorithm among the algorithms that corrects all types of errors
- H-RACER algorithm is an open source program implemented in C/C++
- H-RACER has been published in IWBBIO - April 2017

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# Future Work

- Enhancing the memory usage of H-RACER for long genomes, so as to be able to run long genomes within 244GiB RAM
- Implementing H-RACER with parallel threads, where both time and memory will be enhanced, especially for long genomes
- Inventing a new k-mer grouping algorithm for K-mer Grouping Error Correction algorithm with a reasonable complexity



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— **Thank you!** —

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