

CS481/CS583: Bioinformatics Algorithms

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KMER DATA STRUCTURES

Baseline problem

In-memory representation of a large set of short k-mers:

e.g.

ACTGAT

GTATGC

ATTAAA

GAATTG

...

Applications

- Genome assembly
- Error-correction of DNA sequencing data
- Detection of similarity between sequences
- Detection of distances between datasets
- Alignment
- Pseudoalignment / quasi-mapping
- Detection of taxonomy
- Indexing large collections of sequencing datasets
- Quality control
- Detection of events (e.g., SNPs, indels, CNVs, alt. transcription)
- ...

k-mers

Sequences of k consecutive letters, e.g. ACAG or TAGG for $k=4$

N.G. de Bruijn (1946),
de Bruijn sequences ¹



C. Shannon (1948),
information theory ²



Framing the problem

Large set of k-mers : 10^6 - 10^{11} elements

k in $[11; 10^3]$

Operations to support

- **Construction** (from a disk or stream)
- **Membership** (“is X in the set?”)
- **Iteration** (enumerate all elements in the set)
- ...

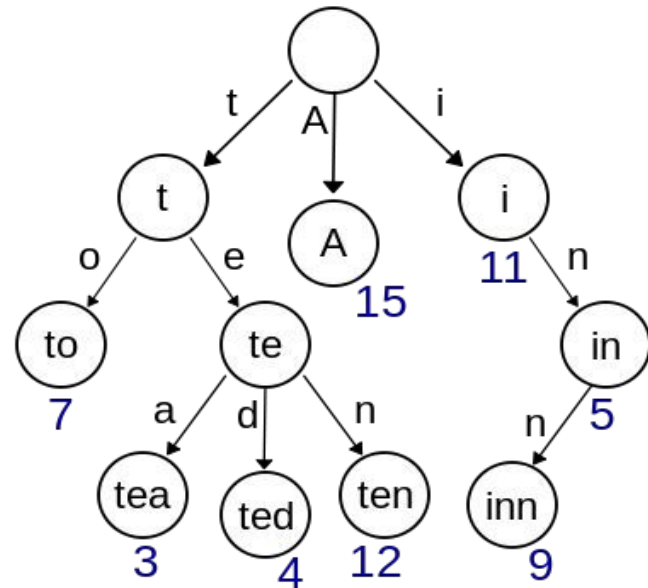
Extensions:

- Associate value(s) to k-mers (e.g. abundance)
- Navigate the de Bruijn graph

Membership test: Tries

Worst case

- $O(k)$ insertion
- $O(k)$ deletion
- $O(k)$ search



Also supports indexing

Membership test: Bloom filters

Init a bit array

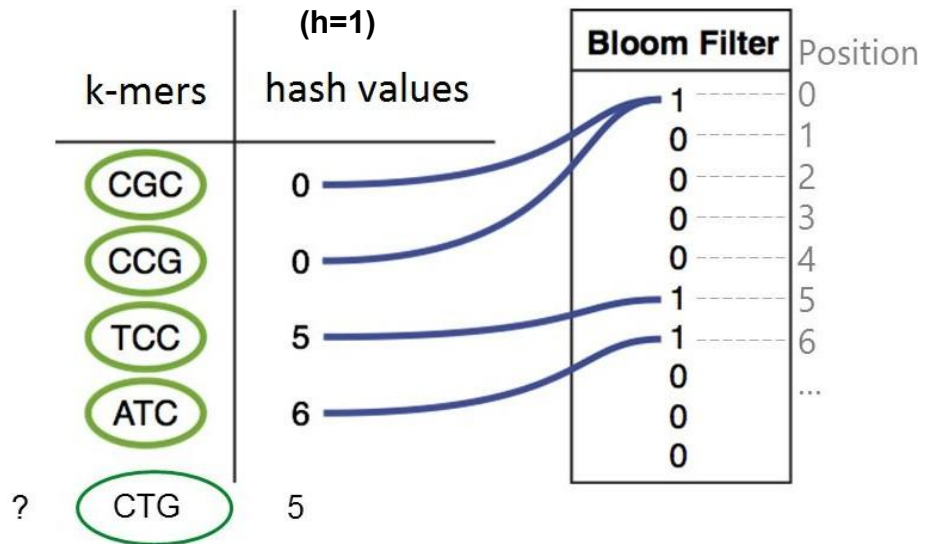
Take h hash

functions

Insertion: put 1's at positions given by hash functions

Query: are there 1's at all positions given by hash functions?

- $O(hk)$ insertion
- $O(hk)$ deletion
- $O(hk)$ query

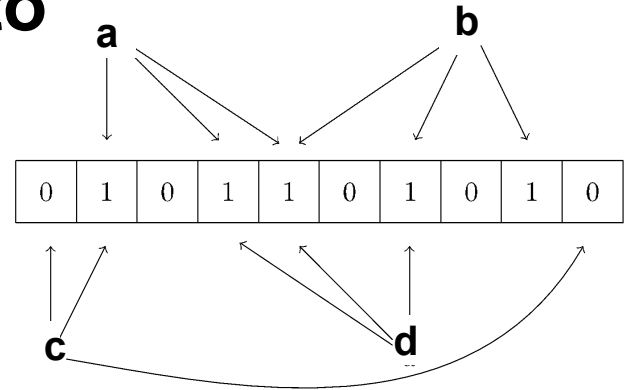


Bloom filter

- Bloom filter encodes a set of k-mers
 - Uses a bit array B of length m and d hash functions
 - to insert x , we set $B[h_i(x)] = 1$, for $i=1, \dots, d$
 - to query y , we check if $B[h_i(y)]$ all equal 1, for $i=1, \dots, d$
 - Need an estimate for n , the number of k-mers to insert
-

Bloom filter example

- a and b are inserted in to a Bloom filter with $m = 10$, $n=2$, $d=3$
- c is not in the set, since some bits are 0
- d has not been inserted, but is still reported in the set, a false positive
- Bloom filters have no false negatives



Bloom filter

- Storing n k -mers in m bit array with d hash functions has a false positive rate of

$$\approx (1 - e^{-d n/m})^d$$

- Given n and m , the optimal d is $\approx m/n \ln(2)$
- Example $m = 8n$, $d=5$ gives 2.16% fpr
 $m = 6n$, $d=4$ gives 5.6% fpr
 $m = 4n$, $d=3$ gives 14.6% fpr
- $m=8n$, corresponds to storing 1 byte per k -mer

Space: $m = 1.44n \log_2(\epsilon)$ where ϵ is the false positive rate

Counting k-mers

- Simple method

Store each k-mer in a
hash table with a counter

- Memory needed

- ❑ store canonical k-mers
- ❑ 2 bits for each of A,C,G,T
- ❑ $k/4$ bytes per k-mer ($k=31$, 8 bytes)
- ❑ 1-2 bytes per counter
- ❑ +10% hash table overhead

- For a genome of size G , expect to see up to G distinct k-mers (2.5-3 billion for Human)

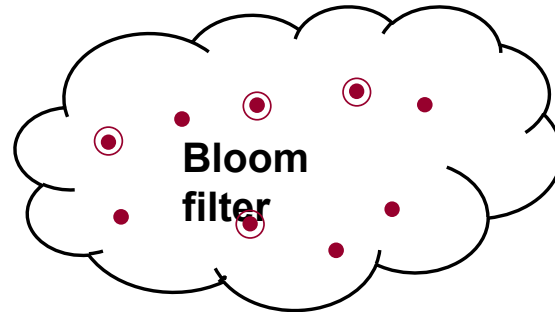
- ~ 36 Gb of memory

Memory-efficient k-mer counting

- Use a Bloom filter and a hash table

read 1 ATGAAGTGGG
k-mers ATGA ←
TGAA ←
GAAG ←
AAGT ←
AGTG →
GTGG →
TGGG →

read 2 AGTGGGTGAA
k-mers AGTG →
GTGG →
TGGG →
GGGT ←
GGTG ←
GTGA →
TGAA ←



First
Pass

Hash
table

TGGG	1
AGTG	1
GTGA	1
GTGG	1

Algorithm

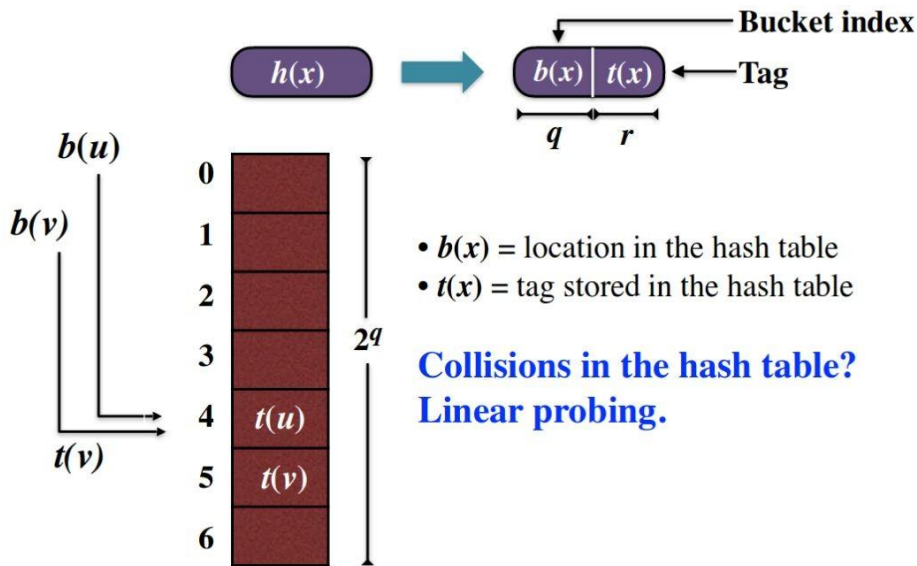
- This scheme guarantees
 - ❑ k-mers seen twice will be in the hash table
 - ❑ some unique k-mers will slip through
 - ❑ second pass gives accurate counts and allows to discard false positives
 - Memory usage
 - ❑ full for k-mers in hash table (~ 9 bytes)
 - ❑ minimal for k-mers in bloom filter ($\sim .5$ -1 bytes)
-

Results whole genome

- 25-mers in 36 bp reads
- 2.37 billion distinct 25-mers in hg18
- 12.18 billion 25-mers in the sequencing data
 - 9.35 billion unique
 - 2.83 billion with coverage 2 or greater

Program	Time (hrs)	Memory (G)
BFCOUNTER	23.82	42
Naïve	> 26.83	>128

Approximate membership test: Counting Quotient Filter



Hybrid between a
compact hash table and
a Bloom Filter.

**Approximate
membership**

- $O(k)$ insertion
- $O(k)$ deletion
- $O(k)$ query

Multi-set Membership Test: Sequence Bloom Trees

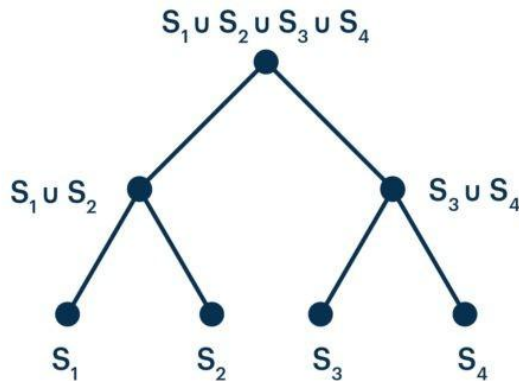


Fig: <https://www.sevenbridges.com/sequence-bloom-trees-principles/>

Leaf: Bloom Filter of a sequencing dataset

Internal nodes: Bitwise union of children BF's

- Representation of sets of k-mer sets
 - Approximate membership across all datasets in $O(\text{hits})$ instead of $O(\text{datasets})$
 - No k-mer iteration
 - Insertion/deletion of complete datasets
 - Whole structure resides on disk
- Solomon, Kingsford, *Nat Biotech* 2017
Sun, Harris, Chikhi, Medvedev, *RECOMB* 2017
Solomon, Kingsford, *RECOMB* 2017

Application: fast sequence search in 1000's of RNA-seq experiments

Multi-set Membership Test: Bloom Filter Tries

Add *gcgccaggaatc* -> BURST

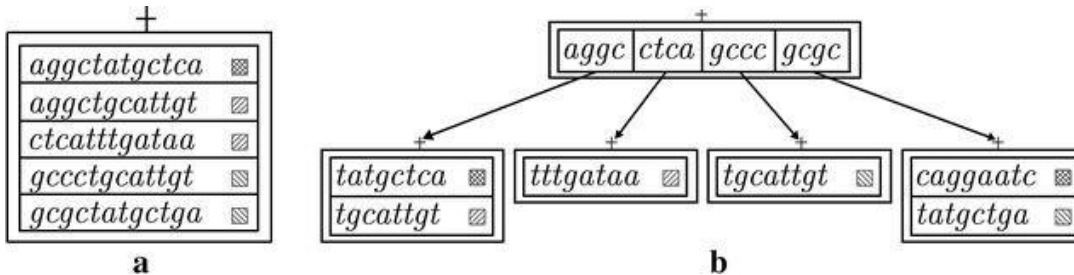


Fig: BFT
article

Principle: cut k-mers into chunks, insert in a burst trie, Bloom Filters added for speed

- Representation of sets of k-mer sets
- Tailored to pan-genomes: a single k-mer belongs to many sets
- Explicit DBG operations support

Application: indexing and compression of pan-genomes

Holley, Wittler, Stoye, *WABI* 2016

Alternative: colored de Bruijn graphs

Slide by Rayan Chikhi

De Bruijn Graphs

- n -dimensional directed graph of m symbols
 - m^n vertices: all possible length- n sequences of m symbols
 - Edges between vertices v and w if $sequence(w)$ can be generated by *shifting* $sequence(v)$ by one character and add one new character
 - $S = \{s_1, s_2, \dots, s_m\}$
 - $V = S^n = \{(s_1, \dots, s_1, s_1), (s_1, \dots, s_1, s_2), \dots, (s_m, \dots, s_m, s_m)\}$
 - $E = \{((v_1, v_2, \dots, v_n), (w_1, w_2, \dots, w_n)): v_2=w_1, v_3=w_2, \dots, v_n=w_{n-1}\}$

De Bruijn Graph for DNA Assembly

- $m = 4$ (A, C, G, T)
- $n = k$ (k-mer size)
- 4^k *potential* vertices
 - In reality if k is sufficiently large, upper bound is genome size
 - Twin vertices: vertices with sequences that are reverse-complement of each other
 - AAAA twin of TTTT

De Bruijn Assemblers

- Currently the most common for short read seq: Euler, ALLPATHS-LG, Velvet, ABySS, SOAPdenovo
 - Divide reads into k-mers
 - Build graph from k-mers
 - Put an edge if there is k-1 bp prefix-suffix match
 - Error correction
 - Eulerian path
 - The first parts (graph construction & correction) is essentially common to all these assemblers, with a few implementation differences (e.g., parallelization/distributed computing in ABySS)
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A quick example

TAGTCGAGGCTTTAGATCCGATGAGGCTTTAGAGACAG

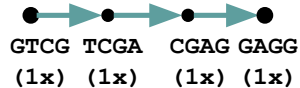
```
AGTCGAG CTTTAGA  CGATGAG CTTTAGA
  GTCGAGG  TTAGATC  ATGAGGC    GAGACAG
    GAGGCTC    ATCCGAT AGGCTTT  GAGACAG
AGTCGAG      TAGATCC ATGAGGC  TAGAGAA
TAGTCGA  CTTTAGA CCGATGA    TTAGAGA
      CGAGGCT  AGATCCG TGAGGCT  AGAGACA
TAGTCGA GCTTTAG TCCGATG  GCTCTAG
      TCGACGC    GATCCGA GAGGCTT AGAGACA
TAGTCGA      TTAGATC GATGAGG TTTAGAG
      GTCGAGG TCTAGAT  ATGAGGC  TAGAGAC
        AGGCTTT  ATCCGAT AGGCTTT  GAGACAG
AGTCGAG      TTAGATT  ATGAGGC    AGAGACA
        GGCTTTA  TCCGATG    TTTAGAG
      CGAGGCT TAGATCC  TGAGGCT    GAGACAG
AGTCGAG  TTTAGATC  ATGAGGC  TTAGAGA
      GAGGCTT  GATCCGA GAGGCTT  GAGACAG
```

A quick example

```
AGTCGAG CTTTAGA  CGATGAG CTTTAGA
  GTCGAGG  TTAGATC  ATGAGGC    GAGACAG
    GAGGCTC   ATCCGAT AGGCTTT GAGACAG
AGTCGAG    TAGATCC ATGAGGC  TAGAGAA
TAGTCGA  CTTTAGA CCGATGA    TTAGAGA
    CGAGGCT  AGATCCG TGAGGCT  AGAGACA
TAGTCGA GCTTTAG TCCGATG  GCTCTAG
    TCGACGC    GATCCGA GAGGCTT AGAGACA
TAGTCGA    TTAGATC GATGAGG TTTAGAG
  GTCGAGG TCTAGAT  ATGAGGC  TAGAGAC
    AGGCTTT  ATCCGAT AGGCTTT GAGACAG
AGTCGAG    TTAGATT  ATGAGGC    AGAGACA
    GGCTTTA  TCCGATG    TTTAGAG
    CGAGGCT TAGATCC  TGAGGCT    GAGACAG
AGTCGAG  TTTAGATC  ATGAGGC  TTAGAGA
    GAGGCTT  GATCCGA GAGGCTT  GAGACAG
```


A quick example

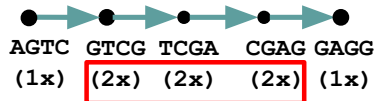
First read: GTCGAGG



A quick example

First read: GTCGAGG

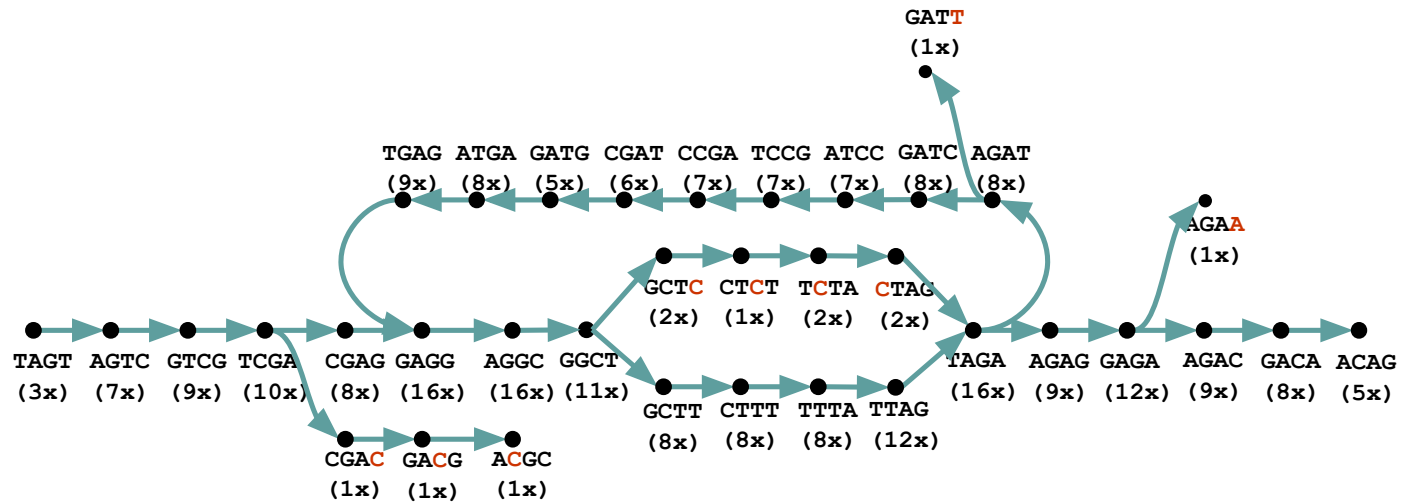
Second read: AGTCGAG



insert *increment counter*

A quick example

All the others...

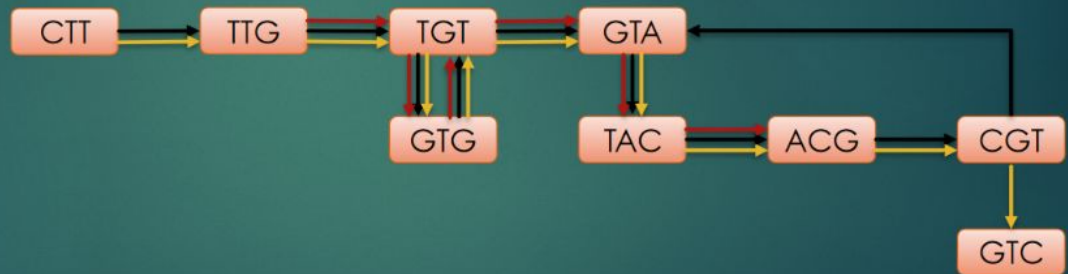


Colored de Bruijn Graphs

$K = 4$

Samples:

- CTTGTGTACGTA
- CTTGTGTACGTC
- TTGTGTACG



For more

- “Data structures based on k-mers for querying large collections of sequencing data sets”, Camille Marchet, Christina Boucher, Simon J. Puglisi, Paul Medvedev, Mikaël Salson and Rayan Chikhi
 - ▣ Genome Res, 2020:
<https://genome.cshlp.org/content/early/2020/12/16/gr.260604.119.abstract>
 - ▣ bioRxiv:
<https://www.biorxiv.org/content/10.1101/866756v3>
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