Bee visitation, pollination services, and plant yield in commodity and hybrid seed canola

Samuel V. J. Robinson[1]*, Ralph V. Cartar[1],
Stephen F. Pernal[2], Riley Waytes[3], Shelley E. Hoover[4]

[1] Department of Biological Sciences, University of Calgary, Calgary, AB
[2] Agriculture and Agri-Food Canada, Beaverlodge Research Farm, Beaverlodge, AB
[3] Ministry of Land, Water, and Resource Stewardship, Smithers, BC
[4] Department of Biological Sciences, University of Lethbridge, Lethbridge, AB

Abstract

Insect-mediated pollination of crops is an important input to agricultural production, but pollination management suffers from key knowledge gaps that hinder its greater utility. While a solid theoretical and mechanistic framework for plant pollination exists, the pollination of agricultural crops is often treated as a "black box", without reference to the specific mechanisms underlying the processes of pollination and fruit production. We present a causal model that links insect visitation to pollination to three separate components of yield, using field data from two types of canola (*Brassica napus*) production systems. Our results demonstrate that yield in commodity canola fields is primarily determined by plant size, and we found no relationship between honey bee visitation and pollen deposition, or pollen deposition and seed yield. In contrast, yield in seed production canola fields was similarly controlled by plant size, but there was also a strong relationship between bee visitation and pollen deposition, as well as deposition and seed yield. Leafcutting bee visitation in particular strongly

*Corresponding author: samuel.robinson@ucalgary.ca

increased pollen deposition in seed canola fields, whereas honey bee visitation did not. This model serves as a step towards a dynamic model of pollination services, and points to the contextual importance of pollination services in seed canola production.

Introduction

Animal pollination of agricultural flowering plants is an important ecosystem service that contributes to about 10% of total crop production value worldwide (Gallai et al., 2009), and animal-pollinated plants produce a large number of important micronutrients in the human diet (Eilers et al., 2011). Pollination of these crops, however, is largely treated as a single step, while in reality there are many links in the chain of events that result in fruit production. Visitation of flowers by animals (typically bees or other insects) can increase the deposition of pollen onto the stigma of the flower, but in order for fruit set to occur, this must be followed by fertilization of the ovules, and maturation of fruit (Erbar, 2003; Goldberg et al., 1994). Each step has many other underlying constituent components (*e.g.* stigma receptivity, pollen tube growth, ovule abortion), but even at this coarse level of approximation, most studies of agricultural pollination fail to account for important underlying processes. Moving beyond this "black-box" model of agricultural pollination requires incorporation of these steps, and is needed for contextualizing the value of pollination services, as well as optimizing crop yield in novel circumstances.

Pollinator visitation is not uniform within fields: visitation rates typically decline with distance away from the pollinators' nest sites. Smaller bees fly shorter distances than larger bees (Greenleaf et al., 2007; Zurbuchen et al., 2010), so their presence at the centre of large fields can be limited (Isaacs and Kirk, 2010). Hence, managing the location and density of pollinators is important for optimal yield of pollinator-dependent crops (Fries and Stark, 1983; Cresswell and Osborne, 2004). Similarly, foragers will often specialize on certain species or morphs of flowers in order to forage more efficiently (floral fidelity, Heinrich, 1976; Goulson et al., 1997). Pollination of hybrid seed crops presents a challenge for specialized

foragers, as it requires movement of pollen between separate lines of plants, meaning that fidelity can reduce pollen transfer (~~Waytes, 2017;~~ Gaffney et al., 2019). Competition for floral resources can occur between different species of foragers, but they can spatially or temporally separate their foraging to reduce competition (Schaffer et al., 1979; Thomson et al., 1987, but see Steffan-Dewenter and Tscharntke, 2000), such as switching between floral morphs more often, or visiting less frequently (Heinrich, 1979; Greenleaf and Kremen, 2006). Thus, variation in visitation can be driven by distance, competition with other foragers, or floral specialization, as well as plant traits such as nectar production and flower size; however, this spatial variation is seldom considered, as commercial pollinators are often treated as a single agricultural "input" (akin to fertilizer).

Seed production can be limited by plant resources as well as pollen, meaning that the benefits of insect pollination depend on the resources available to the plant (Stephenson, 1981; Marini et al., 2015; Tamburini et al., 2017, 2019). Pollination can enhance fruit production in flowering plants by increasing either the quantity or quality of pollen deposited on the stigma (Stephenson, 1981; Burd, 1994; Aizen and Harder, 2007). Many flowering plant species are capable of self-pollination, but can produce more fruit or seeds from outcrossed pollen (Knight et al., 2005). However, the returns from this extra pollen deposition are diminishing (Plowright and Hartling, 1981), with very high numbers of pollen grains causing decreasing improvements in fruit production (Ashman et al., 2004; Harder et al., 2016). Pollen limitation can also occur at multiple levels within a plant, reducing the number of seeds per fruit or the number of mature fruit (Burd, 1994). Low pollen deposition can also cause flower abortion, where poorly-pollinated flowers are abcised from the plant (Stephenson, 1981); an indeterminate growth strategy may offset this by enabling the plant to continue producing more flowers (Lovett-Doust and Eaton, 1982; Lawrence, 1993; Sabbahi et al., 2006; Bos et al., 2007). Additional pollen may result in a greater number of seeds per fruit (Knight et al., 2006), which in turn may result in a reduction in the size per seed due to competition between ovules or fruit (Free and Nuttall, 1968; Mazer, 1987). Seed size can

also vary dramatically among plants, and can be affected by resources available to the plant (Mazer, 1987; Venable, 1992). Therefore, understanding how plants allocate reproduction under pollen or resource deficits are important for managing agricultural production (Bos et al., 2007; Tamburini et al., 2019).

Insect pollination is especially important in the production of canola (*Brassica napus* L.). Hybrid *commodity canola* (used for oil and meal production) is the offspring of two parental *seed canola* breeding lines, a male-sterile "female" and a hermaphroditic "male" line (Westcott and Nelson, 2001; Steffan-Dewenter, 2003; Clay, 2009). Seed canola production therefore requires a large number of pollinators to ensure pollen transfer from the male to the ~~femal~~ line (seeds from the male line are not harvested), but there are few studies that examine pollination in these seed production systems (but see Mesquida and Renard, 1981; Mesquida et al., 1991). Bee pollination may also increase the yield of commodity canola (Morandin and Winston, 2005; Rader, 2010; Bommarco et al., 2012; Bartomeus et al., 2015; Perrot et al., 2018), but this is unclear, as many key studies suffer from either a lack of realistic context, have a number of potential confounding variables, or infer plant-level outcomes from flower-level treatments (Ouvrard and Jacquemart, 2019). Greenhouse experiments typically involve unrealistically high levels of pollination, nutrient availability, and water, all of which can interact with yield (Bartomeus et al., 2015; Marini et al., 2015). Field studies often relate yield to indirect measures of pollination services, such as species richness or distance from sources of potential pollinators, or do not examine production at the plant level (Morandin and Winston, 2005; Ricketts et al., 2008). These proxies provide limited information about how plants dynamically respond to pollen exclusion or addition (but see Sabbahi et al., 2005). Net- or cage-treatments exclude insect visitation from certain plants or flowers, but can alter wind pollination, humidity, light, or pest pressure (Olsson, 1960; Neal and Anderson, 2004; Jauker and Wolters, 2008). All of these methods give an incomplete picture of how pollination relates to yield in canola crops (Ouvrard and Jacquemart, 2019), and obscure estimates of pollinator value in a globally valuable crop species (Melathopoulos

et al., 2015).

Seed production in canola involves a sequence of processes (visitation → pollen deposition → fruit production ← plant resources), that determine the magnitude of the link between pollination and the components of crop yield (seed size/number). However, other studies of canola pollination focus on individual processes, such as visitation and pollination (Cresswell, 1999; Thomson and Goodell, 2001), or visitation and yield (Steffan-Dewenter, 2003; Manning and Wallis, 2005; Hudewenz et al., 2013), but have not incorporated the links in a single framework (but see Sáez et al., 2018), and few have used realistic field data (Morandin and Winston, 2005; Isaacs and Kirk, 2010). In this study, we examine how distance influences pollinator visitation, which in turn influences pollen deposition and seed yield, using commodity and seed canola crops in Alberta, Canada. Using both commodity and seed canola provides an opportunity to compare two plant varieties that differ strongly in their pollination requirements, using a similar type of structural model, while accounting for agricultural differences between varieties. We expected that bee visitation would decrease with distance from each species' hive or shelter, and that lower visitation would result in lower pollen deposition. We also expected that higher pollination would increase seed production in commodity canola crops, but that the magnitude of the increase would be higher in seed canola, and that plant size (a proxy for plant resources) would similarly increase seed production. This study assesses the strength of the connections between visitation, pollination, and yield, in a globally important crop species, and identifies the relative importance of bee pollination for seed production, using in-field data from two distinct cropping systems.

Methods

Data collection

From June through August of 2014 and 2015, we surveyed 29 commodity canola fields (14 in 2014, 15 in 2015) near Beaverlodge, Alberta and 31 fields (17 in 2014, 14 in 2015) near

Lethbridge, Alberta. Commodity canola fields were selected based on the proximity of honey bee apiaries and site access. 28 of the 60 fields were stocked with Western honey bees (*Apis mellifera* L., hereafter HB) at the corner or side of the field (mean: 0.6 hives/hectare, SD: 0.58) while 32 fields were unstocked. 14 of 31 of the fields near Lethbridge were watered using central-pivot irrigation systems. Growers were also asked for canola variety information, but there was not enough replication to test for difference between varieties.

During 2015 and 2016, we also surveyed 35 hybrid seed canola fields (15 in 2015, 20 in 2016) near Lethbridge, Alberta, from June through August of each year, all of which had central-pivot irrigation, and were stocked with honey bee hives at a rate of 3.6 hives/ha (apiaries stationed in the corners of fields). In seed fields, bays of hermaphroditic (herafter "male") and female plants are typically planted in 1- and 6-m wide bays, respectively. Seed fields were also stocked with shelters (1.3 - 2.6 shelters/ha, Figure S2) containing alfalfa leafcutter bee cocoons (*Megachile rotundata* F., hereafter LCB) at a rate of 50-100,000 coocoons/ha.

In both field types, we established ~~set~~ of plots at varying ~~distance~~ from the ~~sources~~ of pollinators. ~~In commodity fields, plots were located at 1 m²~~ plots at 5, 20, 100, and 500 m (271 total plots), starting at the field edge closest to the set of honey bee hives or potential sources of natural pollinating insects in unstocked fields (forests, shrublands, or grasslands). We observed very few wild pollinators (Table S2), so these were excluded from the analysis. In seed fields, we established plots at 5, 20, 100 (250 m in 2016), and 400 m into the field along a transect from the nearest set of honey bee hives, using pairs of plots at the edge of adjacent male and female bays (Figure S3). To examine within-bay variation in visitation, we established a plot at the centre of the female bay at the 5m and 400m plot in each field. Distances to nearest shelters were measured using a Nikon™Laser 800S Rangefinder. Finally, we incorporated plot-level visitation data from Waytes (2017; same years and locations) to more accurately gauge the effect of distance from shelter on visitation rates (647 total plots). All surveys occurred on fair-weather days (median temperature: 24.5°C., range: 17–33) with

no rain and minimal wind (>30 km/hr).

Insect visitation and pollination data were collected ~~once during the main canola bloom at each field~~ (late June - late July), and plants were collected just prior to harvest (mid - late August). We recorded the number of insect visits that contacted the stigmas and anthers of flowers during 10 minutes of observation (5 min for seed fields during 2015), recorded the identity of the visitor, and counted the number of open, visitable flowers in each plot (petals had not yet started to dehisce, style had not elongated more than 3-4 millimeters beyond the anthers). To assess pollen deposition, we collected stigmas from five random flowers at each female plot, mounted them in fuchsin gel (Beattie, 1971) on depression slides, and counted the pollen on each stigma using a Leica™DME 13595 light microscope under 100x magnification (1294 commodity and 1050 seed canola stigmas). At the end of the growing season, we collected three plants from the same female plot and recorded the plant density per m^2 (789 commodity canola plants, 582 seed canola plants). After drying the plants, we weighed each plant, counted mature pods to estimate pod set, and counted the number of flower pedicels to estimate total flower production. We estimated seed size and seeds per pod by averaging the seed count and weight from five pods on each plant. Finally, we threshed all the pods for each plant by hand, winnowed them using an air separator, and weighed the total mass of cleaned seeds.

Analysis

To examine how yield is influenced by visitation, we used piecewise structural equation models (pSEM, Shipley, 2009). Structural equation models (SEMs) are a set of linear models arranged in a causal network, which provides a framework for testing hypotheses about complex systems (Grace et al., 2012; Lefcheck, 2015). SEMs provide a framework for empirical analysis of complex systems, as they allow causal relationships to be formally tested, can provide insight into alternative mechanisms, and are flexible in their assumptions (Shipley, 2009; Clough, 2012; Grace et al., 2012). SEMs represent an intermediate class of statistical

models, occupying a space somewhere between linear regression (or machine learning) models and dynamic linear models, making them ideal for generating and testing relationships between large sets of variables (Grace, 2006; Kline, 2013). Starting with a simple model (visitation $\rightarrow$ pollen deposition $\rightarrow$ fruit production $\leftarrow$ plant resources), we built a detailed model relating visitation to pollination to seed yield within a causal framework (see Figure 1 in Results). We also ran a linear model outside of each SEM to predict total yield (g per plant) as a function of the seed size, seeds per pod, and pods per plant (see Table S6). After the models were fit and validated, we simulated seed size and yield at varying distances from pollinators, using the coefficients from each model to integrate the effect of pollination on yield across all sub-models (while holding plant density constant).

The underlying models of each pSEM were specified as generalized linear mixed-effects models (GLMMs). GLMMs can model processes that occur at different levels; for example, we modeled seed size at the plant level, and pollen deposition at the flower level. Random intercepts were used to model variance at the field and plot level, but plot-level variance terms had poor traces and low effective sample sizes, indicating low plot-level variance; therefore, we omitted plot-level random effects for all models (except for pollen count and pod set in seed fields, see Table S5). Visitation models used log(time) as an "offset" variable to account for differences in observation times. We observed LCBs directly harassing HBs during the plot-level observations, attacking HBs both on the flowers and in the air (also seen by Batra, 1978 and Waytes, 2017), but HBs were never the aggressor in these interactions, so we treated LCB vistation as a potential cause of HB visitation (Figure 1). Pod set can suppress future vegetative growth and flower production (Stephenson, 1981), but SEMs do not allow for causal loops (Grace et al., 2012), so we included a path from pod set to flower production as the closest approximation of this process. The direction of the path could be reversed, implying that large flower production reduces the proportion of pod set, but we considered only the first scenario, as Sabbahi et al. (2006) showed that low pod set increases flower production. Plant and pollen samples were not collected from male bays in seed fields,

and there were no large differences in visitation rates for honey bees or LCBs (p = 0.32, 0.72, respectively) once other terms were included as covariates, so bay was excluded from further analysis. See Equation sets S1 and S2 for model specifications) Table S1 for a brief summary of the variables used in each SEM.

Structural equation models do not have paths between all variables, which may bias the model results if this independence assumption is incorrect; this requires testing a "basis set" of independence claims using Shipley's d-separation criteria (Shipley, 2000, 2009). The initial commodity canola pSEM was misspecfied, as several missing paths were identified (Fisher's C = 167.5, df = 64, p<<0.001), but the updated pSEM was deemed adequate (C = 52.0, df = 48, p = 0.32). Similarly, the initial seed field pSEM also had several missing paths (C = 105.6, df = 76, p = 0.01), which were added, making the updated pSEM adequate (C = 72.2, k = 72, p = 0.47). Basis sets were generated using the `dagitty` library in R 4.2.1.

All component GLMMs of the pSEM were written in Stan 2.18.1 and run using `rstan` 2.26.13 (Gelman et al., 2015; Stan Development Team). We used weakly informative normal priors ($\mu = 0, \sigma = 5$) for fixed effects and gamma priors ($\alpha = 1, \beta = 1$) for the variance components. Four separate chains were run with an adaptive phase of 1000 iterations, and a sampling phase of 1000 iterations, then checked for convergence of the chains ($\hat{R} \approx 1$) and low autocorrelation within chains (high N_{eff}). We assessed the underlying probability distribution functions of each model using posterior predictive checks (Gelman et al., 2013), and found that the probability distributions were properly specified (i.e. simulated data was close to actual data). p-values were calculated using the Z-scores (mean/SD) of the sampled posterior distributions; unless otherwise specified, listed model coefficents have a p-value less than 0.05, and are considered "strong" effects where p-values are < 0.01 (see Table S4 and S5 for more detailed model summaries). Predictions from models are taken from median values of the posterior, while uncertainty is derived from the 95% credible intervals (CIs). Figures were made using `ggplot2` and `ggpubr` (Wickham, 2016; Kassambara, 2020).

Results

Path analyses

The path analyses revealed that plant size and pollen deposition were the main drivers of seed production in seed canola, while plant size alone was the main driver in commodity canola (see Figure 1 for paths and effect sizes). LCB visitation strongly increased pollen deposition in seed canola, but there was no strong effect of HB visitation in either seed or commodity canola. Plant size increased with distance from the edge of the field in both crops, as well as plant density in seed fields. Plant size strongly increased the number of flowers per plant in both crops, and there were strong negative paths from pods per plant to flowers per plant, indicating that high flower survival suppressed future flower production in both crops. Plant size increased pod set and seeds per pod in both crop types, and also increased seed size in seed canola plants. Pollen deposition increased pod set and seeds per pod in seed canola, but did not affect seed or pod production in commodity canola, showing that plant resources matter strongly for both crops, but that pollen limitation matters only in seed canola. Finally, there was a direct effect (i.e. not mediated by visitation) of bay centre, distance from edge, and distance from LCB shelter on pollen count and pod set in seed fields, pointing to possible differences in pollen transfer even at the same rate of visitation. We consider each component of each path analysis in greater detail below.

Flower Visitation

Honey bee flower visitation (per plot) decreased with distance away from the edge of the field in both commodity and seed canola fields. In commodity fields, HB visitation decreased from 12 visits/hr per plot at the edge to 1.8 visits/hr at 400 m into the field (Figure 2a, apiary of 40 hives). Lower stocking rates decreased HB visitation at the edge of the field to 7.4 visits/hr at 20 hives, while unstocked field edges had an average of 0.8 visits/hr. Unsurprisingly (given the higher stocking rate of 160 hives per field), HB visitation was
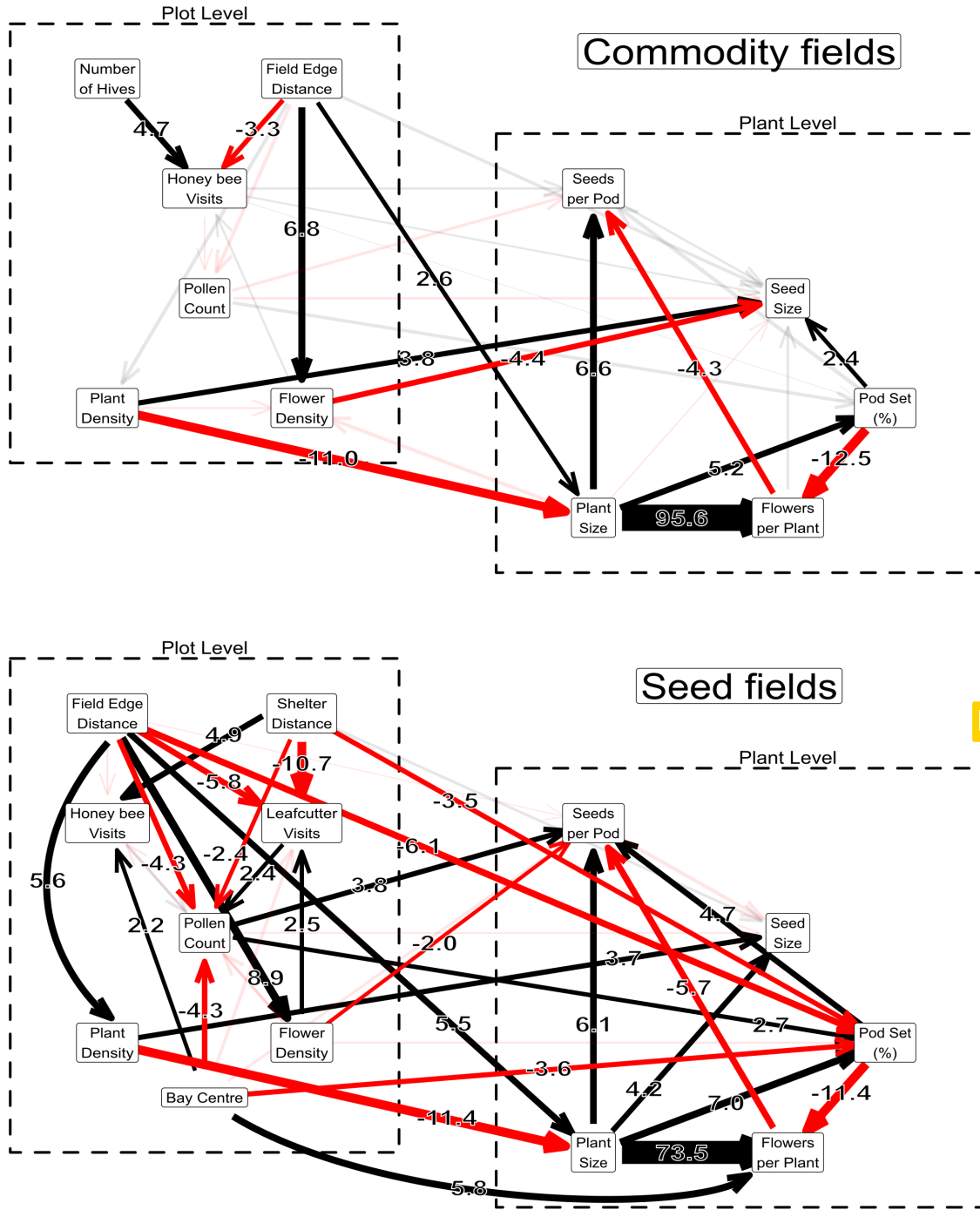
Figure 1: Results of path analyses in commodity and seed fields. *Field Edge Distance* refers to the distance to the edge of the field (and location of HB apiary) and *Shelter Distance* refers to the distance to the nearest LCB shelter. The width of each arrow is proportional to the effect size of each component path (number also displayed), with black and red lines representing positive and negative effects, respectively. Transparent arrows show path coefficients whose 95% posterior intervals overlapped zero.

Figure 2: Effect of distance from a) honey bee apiaries, b) leafcutter shelters, and c) female bay position on visitation rates. Commodity fields are shown at a stocking rate of 40 hives (black line), while stocking rates are at 160 hives in seed fields (yellow line).

much higher in seed canola fields, and decreased from 96.4 visits/hr at edge to 85.7 visits/hr at the centre of the field (Figure 2a). HB visitation was also much lower near LCB shelters, dropping from 81 visits/hr at 20 m to 31 at 2 m (Figure 2b), likely due to competition or aggression from LCBs.

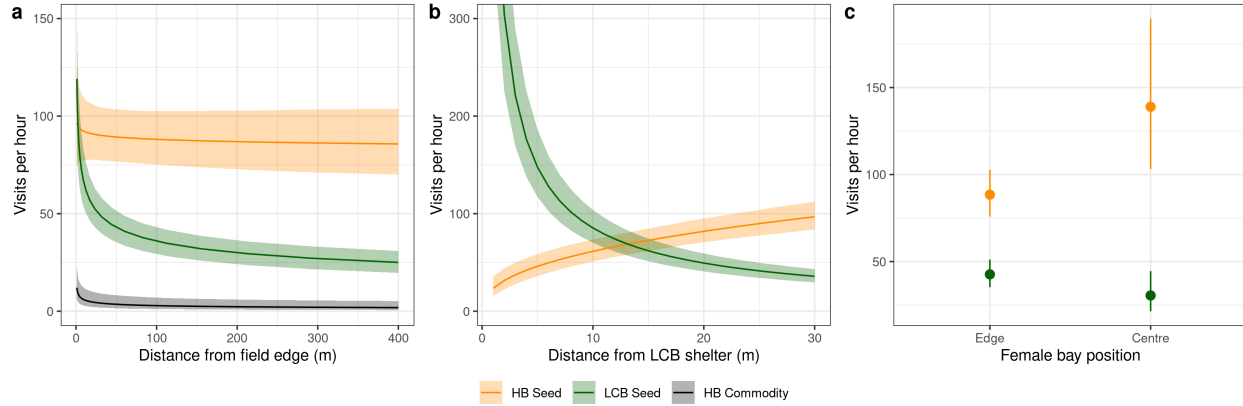LCB flower visitation was much higher close to their shelters, and surprisingly, was also higher at the edge than the centre of the field. Flower density was higher at further distances into the field in both field types, and had a positive effect on LCB visitation (but not HB visitation) in seed fields. LCB visitation dropped from 350 visits/hr at 2 m from the shelter to 50 visits/hr at 20 m (Figure 2b), and also decreased from 119 visits/hr at the edge of the field to 25 at the centre (Figure 2a, b) There was no large difference in LCB visitation between the edge and centre of the female bays, but HB visitation was higher at the centre of the bay (88 vs. 139 visits/hr, Figure 2c). Together, our results indicate that pollination is likely accomplished by different species in different parts of the field.

Pollen Deposition

LCB visitation increased pollen deposition in seed canola fields, but honey bees did not increase pollen deposition in commodity or seed canola fields (Figure 3a). Pollen deposition
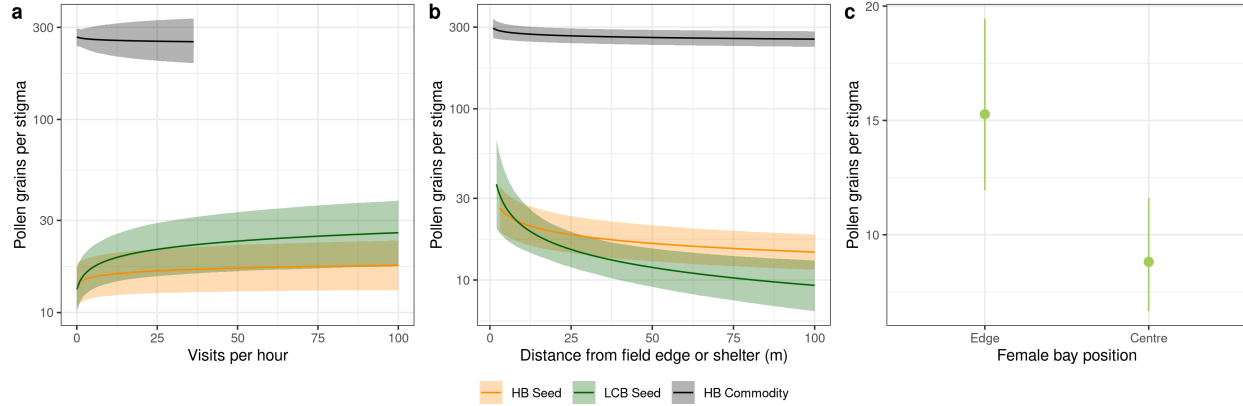
Figure 3: Effect of a) visitation rate, b) distance from apiary (HB) or shelter (LCB), and c) female bay position on pollen deposition.

on stigmas was high in commodity canola (mean: 293 grains/stigma, SD: 385, range: 0–3981), but honey bee visitation had no effect. Pollen deposition declined weakly with distance from the edge of the field (p=0.09), but this only amounted to a 6% average decrease (290 grains/stigma at the field edge vs. 240 at field centre, Figure 3b). In seed canola, pollen deposition was much lower overall (mean: 22 grains/stigma, SD: 43, range: 0–578), and decreased from 26.3 grains/stigma at the edge to 11.5 at the centre of the field (Figure 3b), independent of bee visitation. LCB visitation increased pollen deposition from 13 grains/stigma at 0 visits/hr to 25 pollen grains at 100 visits/hr (Figure 3a), but there was no effect of honey bee visitation (p=0.20), implying that most of the pollen deposition in seed fields is the result of LCB visitation. Pollen deposition also decreased from 15 grains/stigma at the edge of the female bay to 9 at the centre of the bay independent of visitation (Figure 3c), suggesting that bees crossing over into the male bays may deposit less pollen as they move towards the centre.

Seed Production

Commodity canola produced fewer flowers per plant than seed canola (mean: 196 vs. 461), and also produced fewer pods per plant (143 vs. 299, Table S1). In contrast, commodity canola also produced more seeds per pod than seed canola plants (mean: 23 vs. 16), but
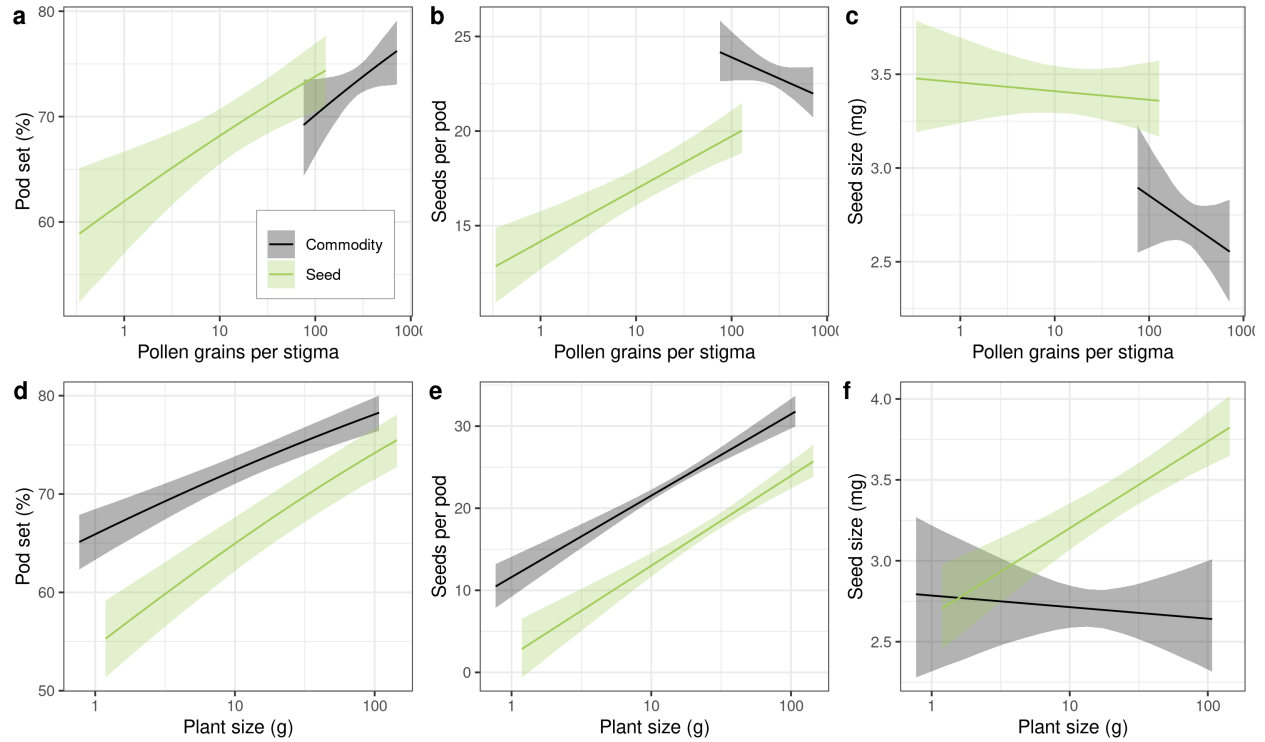
Figure 4: Marginal effects of pollen deposition (top row) and plant size (bottom row) on flower survival, seeds per pod, and seed size.

seed size was lower in commodity canola than seed canola (2.74 vs. 3.43 mg/seed). Finally, commodity plants were also lower-yielding than seed canola plants (mean: 6.8 vs. 9.5 g seed per plant), but produced more seeds per gram of vegetative biomass than seed canola plants (mean: 0.38 vs. 0.32), likely due to lower pollen limitation.

Plant size had by far the strongest positive effect on the number of pods produced (see large plant paths in Figure 1), and there was evidence of pod set suppressing flower production, but pollen only directly affected seed production in seed canola. Pollen deposition did not alter seed production in commodity canola (Figure 4a,b,c), nor was there a direct effect of honey bee visitation or distance from field edge, at least at the levels we observed. In both commodity and seed canola, plant size had a strong positive effect on % pod set and seeds per pod, but did not increase seed size in commodity canola (Figure 4d,e,f). Pod set in seed canola decreased with distance away from the field edge and LCB shelters, and was also lower at the centre of the female bay (independent of visitation and pollen deposition).

14

This distance effect was not seen in commodity canola, suggesting a more long-term pollen limitation effect in seed canola that was not captured by our snapshot of pollen and visitation. Plant density also had a positive direct effect on seed size in both crops; this likely indicates field areas with better microclimate or soil resources rather than density effects *per se*, as plants at edges of the field or in "field holes" are typically both smaller and less dense. Finally, pod set also had a strong negative influence on flowers per plant in both crop types, indicating that high pod set suppresses future flower production.

Yield simulations

Our path analysis simulations further confirmed the results of the analysis: HB visitation had little effect on yield in either crop type, but LCB visitation improved yield in seed crops. The simulations showed no large effect of HB stocking on seed size or total yield in commodity fields (Figure 5a and b, respectively), aside from the effect associated with distance from the edge of the field. Similarly, the simulations for seed fields showed a small increase in seed size and total yield with distance from the HB hives at the field edges, but the effect of distance from the leafcutter shelters was much more dramatic (compare the spacing of isolines along the x-axis and y-axis in Figures 5c and d). Simulations run separately for the edge and centre of the female bay revealed that not only are total seed size and total yield lower at the bay centre, but that the effect of HB distance is effectively *nil* at the bay centre (yield isolines are essentially horizontal in bay centre, and slightly tilted at the bay edge, Figure 5d and f). Interestingly, the simulations also showed evidence of nonlinear behaviour occurring in the centre of seed field bays, where a local minima in yield appears at approximately 25 m away from HB hives and 45 m away from LCB shelters (Figure 5)f), likely because of an interaction between the components of yield (seeds per pod, pods per plant, and weight per seed).

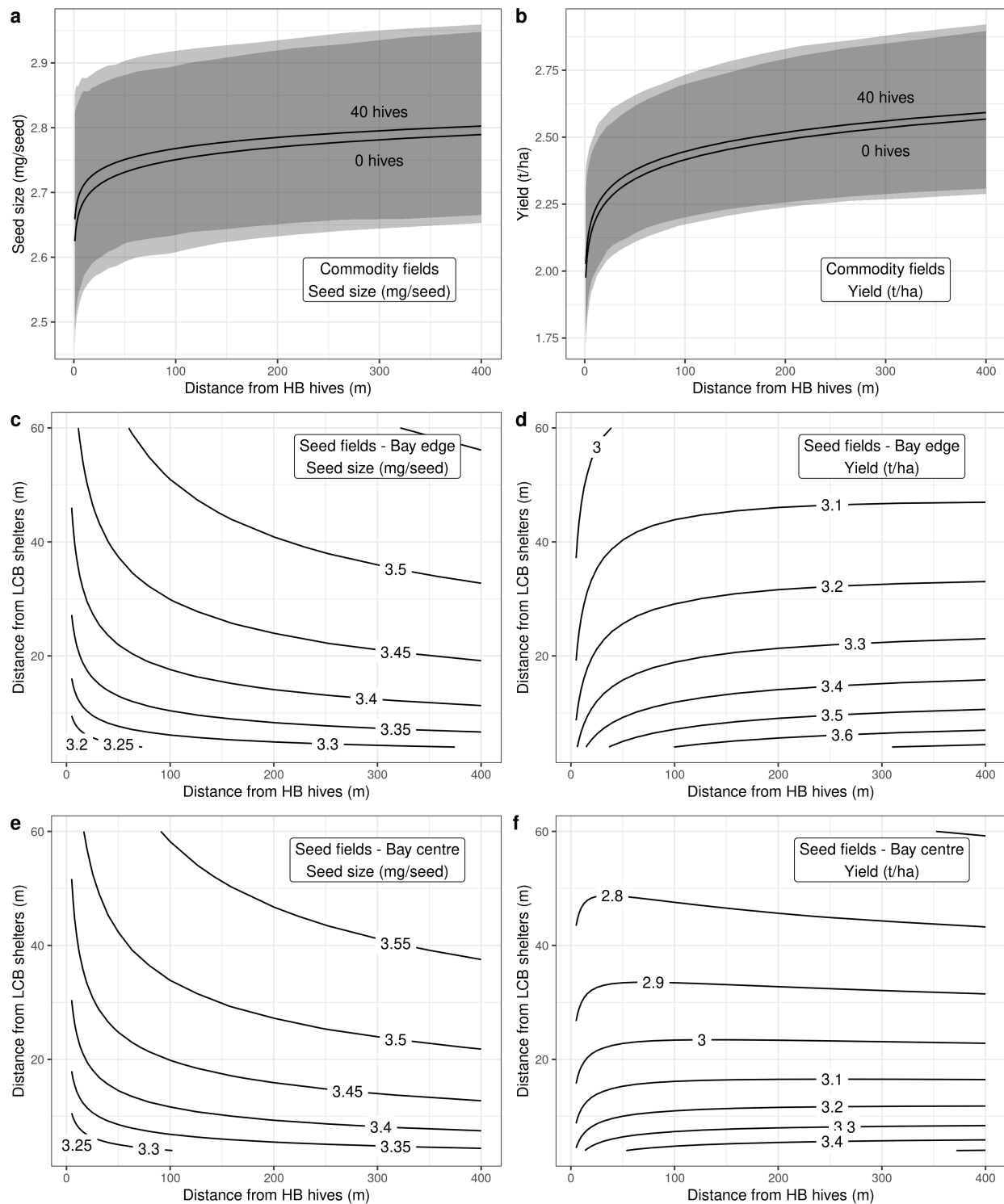Figure 5: Effect of honey bee distance and leafcutter distance on seed size (mg/seed, first column) and total yield (tonnes/hectare, second column), using coefficients from paths for simulation, while holding plant density constant at its mean value. Shaded areas shown in a & b are 95% quantiles for the simulations. Lines on panels c - f are isolines of constant seed size (c, e) or yield (d, f).

Discussion

We examined how bee visitation contributes to pollen deposition and seed yield in two different canola crops, and showed a strong link between visitation and yield in seed canola, but not in commodity canola. HB and LCB visitation decreased with distance from their respective hive or shelter, but unexpectedly, LCBs visited more frequently at the edge of the crop. HB visitation had little direct influence on pollen deposition in either crop type, while LCB visitation had a positive effect on pollen deposition in seed canola. We found that commodity canola production is largely limited by plant size, while seed canola production is limited by both pollen and plant size. Finally, our simulation results from the path analyses confirmed this, and showed the strong influence of LCB visitation on seed size and yield. These results provide more mechanistic information on the value of pollination services in commodity and seed canola crops, and show how a statistical-mechanistic model of seed production provides richer insight into the process of seed production than linear models alone.

Bee visitation

HB visitation in both field types declined with distance into the field, but overall visitation was much higher in seed fields than commodity fields, due to the higher HB stocking rate used in seed fields (3.6 vs. 0.6 hives/ha). Since HBs travel to the crop from their hive outside the edge of the field, this decline was not surprising (Robinson et al., 2022), but we did not expect a similar decrease in LCB visitation with distance into the field, as their shelters are located *within* the field. This decrease with distance into the field may have been caused by LCBs migrating from shelters at the centre of the field to the edge (Goerzen et al., 1995), as female LCBs are central-place foragers and tend to not forage far from their nests (Peterson and Roitberg, 2005; Pitts-Singer and Cane, 2011; Brunet et al., 2019). However, LCBs are not as constrained to a single nest as HBs are, and can drift between shelters within a given

field (Goerzen et al., 1995; Pitts-Singer, 2013). LCBs must also forage for leaf materials to create cocoons, but seem to prefer thinner leaves than canola, mainly plants in the families Fabaceae and Rosaceae (Sinu and Bronstein, 2018). Since seed canola fields have very few weeds within the crop, LCBs at the centre of the field may have few choices of leaf material, or alternative pollen and nectar sources. Therefore, LCBs likely move from the shelters at which they were released to the shelters at the edge of the field, so as to gain access to better leaf material or more diverse pollen and nectar sources (Horne, 1995a,b).

In seed fields, HB visitation was correspondingly lower in areas close to LCB shelters, suggesting competition between species. Lower HB visitation may have been caused by lower nectar and pollen resources, as areas close to leafcutter shelters can become depleted in nectar and pollen (Currie, 1997). HBs also suffer from direct interference by LCBs near to their shelters; during the study we observed LCBs directly harassing HBs during the plot-level observations, attacking HBs both on the flowers and in the air (also seen by Batra, 1978 and Waytes, 2017). HB visitation and foraging behaviour varied strongly between the edge and centre of the female bays in seed fields, while there was little difference between LCB visitation rates. HB visitation rates were almost twice as high at the centre of the female bay, but there was no corresponding difference in LCB visitation between male and female bays; this may reflect a greater need for pollen among LCBs than HBs (Cane et al., 2011), as female canola plants produce only nectar.

Very few HBs in the female bay were pollen foragers (1.4%, see Table S3), meaning that foragers who have recently come into contact with pollen are rare. Both Waytes (2017) and Gaffney et al. (2019) showed that HBs exhibit floral fidelity during foraging trips, with minimal crossing between male and female bays (∼5% of observations, Waytes, 2017), limiting pollen transfer between the male and female flowers. Side-working was a very common behaviour among HBs in commodity fields (65% of the total visits from HBs during 2015 were side-working), as well as the male bays of seed fields (36%), but not in the female bays (3%, Table S3). This behaviour is relatively common on male-fertile flowers of *Brassica*

(Free and Williams, 1973; Free and Ferguson, 1983; Delbrassine and Rasmont, 1988; Mohr and Jay, 1988) as well as other flowering crop flowers (Thomson and Goodell, 2001), and may be due to HBs avoiding contact with the stigmas to increase ease of access to nectar, or to reduce the amount of grooming needed during a nectar foraging bout. The foragers who were side-working tended to not switch to top-working (personal observation), so this is likely a consistent individual behaviour (at least in experienced foragers). Taken together, these pieces of evidence suggests that HBs have limited opportunities for pollen transfer in both commodity and seed fields, as they commonly engage in side-working behaviour in both field types, and pollen foragers typically avoid female bays in seed canola fields.

Pollen deposition

HB visitation did not increase pollen deposition in commodity canola fields compared to un-stocked fields, meaning that self-pollination are the likely agents of pollen transfer (*Brassica* flowers are not aligned for dispersal and deposition from the wind, so cross-pollination likely occurs mainly via insects (Mesquida and Renard, 1982; Cresswell et al., 2004)). However, wind-induced self-pollination (plant shaking) can increase yield in *Brassica* (Williams et al., 1986; Mesquida et al., 1988), and fields without bee pollination have outcrossing rates of about 20% within the field (Rakow and Woods, 1987; Becker et al., 1992). HBs can assist in deposition of self-pollen, as Ali et al. (2011) found that *Apis dorsata* and *Apis florea* can both deposit 100-200 grains of pollen per visit on a canola flower (*B. napus* var. *Bulbul*). However, Waytes (2017) used male-sterile flowers from seed canola fields and found that HB pollen foragers deposited far less outcrossed pollen than previously reported ($\sim$ 2 grains per visit), suggesting that much of the pollen deposited on commodity canola stigmas may be self-pollen. Because the overall pollen deposition rates found in this study were high, HB visitation seems to have made little difference in the amount of self-pollination, as large amounts of pollen were present on commodity canola stigmas even at the centre of unstocked fields. This suggests that commodity canola stigmas are largely saturated with self-pollen,

effectively "swamping" any extra pollen deposition by HBs.

Our models also revealed that pollen-transferring behaviour of HBs is limited in seed fields, as deposition strongly increased with LCB visitation but not HB visitation. HBs in seed fields tended to visit more in the centre of the female bay, even close to the edge of the field, yet pollen deposition was still about 40% lower. LCBs switch between male and female flowers more frequently than HBs, travel further between flowers, transport more viable pollen, and tend not to side-work canola flowers (Soroka et al., 2001; Parker et al., 2015; Waytes, 2017; Brunet et al., 2019), all of which may explain their higher pollination efficacy. Pollen deposition in seed fields also decreased with distance from the edge of the field, and was lower in the centre of the female bays independent of visitation, suggesting lower pollen transfer per visit. These may have been caused by two separate processes: a) low pollen carryover with distance into the female bays, and b) shorter trips between flowers at the centre of the field. Pollen carryover from the edge to the centre of the bay female is likely reduced as LCBs typically visit the edge of the female bay before venturing into the centre (Thomson, 1986; Pinnisch and McVetty, 1990). *Bombus* take shorter trips between flowers under nectar-rich conditions (Pyke, 1978; Heinrich, 1979), suggesting that HBs and LCBs may operate similarly at the centre of seed fields, resulting in lower pollen transfer. While manipulating nectar levels within a field to produce greater travel between flowers could be extremely difficult, this suggests that the optimal width of female bays within seed fields may be lower than is currently practiced.

While we found that HBs have no direct effect on pollen deposition in either crop type, they may have an indirect positive effect caused by: a) increasing airborne pollen from male-fertile flowers ~~Pierre et al. (2010),~~ b) lowering nectar and pollen standing crop, causing LCBs to travel further (Pyke, 1978; Heinrich, 1979), or c) lowering floral fidelity on higher-rewarding flowers in the male bays (Mesquida and Renard, 1978; Waytes, 2017; Gaffney et al., 2019). A simple test of this might be to remove HB hives from a seed field (mid-season) and observe changes in LCB visitation before and after the removal.

Seed production

Plant size increased the proportion of pod set in commodity canola, but both plant size and pollen deposition increased pod set in seed canola, suggesting that both pollen deposition and plant resources constrain pod production. This is similar to the findings of Mesquida and Renard (1981) and Steffan-Dewenter (2003), who found that pod set in male-sterile plants responded positively to visitation, while male-fertile plants had no response (but see Adegas and Nogueira Couto, 1992). Both pod set and seeds per pod can increase with extra pollination (Jauker and Wolters, 2008; Sabbahi et al., 2005, 2006; Durán et al., 2010), so the low influence of pollination in commodity fields may be due to a high overall level of pollen deposition (Figure 3). There was no additional effect of HB visitation on pod set, meaning that that HB visitation did not cause more pollen deposition or improve its quality through increased outcrossing (as in Rosa et al. 2011), but this may depend on the variety (Marini et al., 2015; Adamidis et al., 2019). Flowers per plant was reduced by pod set, implying that high pod set suppresses further flower production or increases competition among pods for plant resources. The first scenario is more likely, as Sabbahi et al. (2006) and Mesquida and Renard (1981) both found that canola plants compensated for experimental removal of flowers by increasing branch and flower production, until the plant reached about 170 pods per plant (our plants had a mean of 143 and 299 pods per plant in commodity and seed canola, respectively). There were also effects of distance on pod set independent of pollen: pod set decreased with distance from the edge of the field, distance to LCB shelters, and was lower at the centre of the female bays. Mesquida and Renard (1978) also found that pod set in male-sterile canola declined quickly with distance from the male-fertile plants (due to wind pollination), but the effect we found is likely related to the earlier effect of lower pollen carryover at the centre of the female bays, and possibly the centre of the field. Finally, the effect of plant size was roughly 6x larger than that of pollen deposition, indicating that the factors controlling plant size (e.g. fertilizer application, soil quality) likely constrain pod production more strongly than pollination alone (Marini et al., 2015; Tamburini et al., 2017;

Gagic et al., 2017; Tamburini et al., 2019).

We found that seeds per pod in commodity canola was mainly influenced by plant size, while both seed size and seeds per pod was influenced by both plant size and pollen deposition in seed canola. Extra pollination generally results in canola plants producing more, smaller-sized seeds (see review in Ouvrard and Jacquemart 2019, but see also Kołtowski 2005), as the plant re-allocates resources across a greater number of fertilized ovules. Similarly, canola plants can also increase the number of matured ovules depending on the available plant resources (Bouttier and Morgan, 1992; Kirkegaard et al., 2018). Our results in seed canola fields showed that pollen deposition had a smaller effect than plant size for seeds per pod ($Z = 3.8$ vs. 6.1), suggesting that plant resources limit seed production more strongly than pollen deposition, while plant size was the dominant term for commodity canola fields ($Z = 1.2$ vs. 6.6). Seed size was affected only by plant size in seed canola, and was not affected by pollen or plant size in commodity canola. Plant size is a measure of the source of photosynthates available to the seeds, while fertilized flowers act as sinks of nutrients, but both of these measures are crude, as both sources and sinks of seed nutrients vary across the season (Clarke, 1979; Zhang and Flottmann, 2018). Canola growth is indeterminate, and poorly-pollinated plants respond by making more branches (Mesquida and Renard, 1981; Sabbahi et al., 2006), so it may be that large, poorly-pollinated plants have more resources and low numbers of fertilized ovules per flower at the end of the season. While we did not quantify pod position on branches, the pods at the end of the branches did appear to have fewer seeds, meaning that variation within plants likely occurred because of late-season seed. Unexpectedly, plant density had a *positive* effect on seed size in both crop types, which may be caused by two different processes: a) plant density could be positively related to resource availability if plant survival is higher in plots with better growing conditions (i.e. Berkson's paradox, (Snoep et al., 2014)) or b) plant density may improve the microclimate conditions of the canola stand, reducing heat or desiccation stress on individual plants. We consider the first process more likely, as planting density under uniform conditions typically

has little effect on seed size (Angadi et al. 2003). In summary, our results add to a growing body of literature on the context-dependent value of pollination services (Marini et al., 2015; Tamburini et al., 2017, 2019).

Conclusion

This study has revealed some of the detailed aspects of pollination in commodity and seed canola crops. First, it shows how two important species of commercial pollinators decline with distance from their nesting sites, and how LCBs unexpectedly decline with distance from the edge of the field. Second, it sheds light on the relative importance of HBs and LCBs as pollinators, showing that levels of HB stocking used for honey production do not strongly influence yield in commodity canola crops, and the importance of LCBs as primary pollinators of seed canola. Finally, it contextualizes the value of pollination alongside plant resources, showing that plant resources and pollen limit production in seed canola, while only plant resources limit production in commodity fields. Our unique SEM approach allowed us to formulate and test relationships within a causal framework, which we consider to be extremely valuable, and highly underused in agricultural studies. Our model serves as an approximation of a dynamic process, but could be built upon by using dynamic linear models (Iwasa, 2000; Nord et al., 2011; Sáez et al., 2018) to examine how plants dynamically respond to changes in pollination. These would lend greater understanding to the process of hybrid seed production, and could be used to more accurately predict crop yields in novel scenarios.

References

Adamidis, G. C., Cartar, R. V., Melathopoulos, A. P., Pernal, S. F., and Hoover, S. E. 2019. Pollinators enhance crop yield and shorten the growing season by modulating plant functional characteristics: A comparison of 23 canola varieties. *Scientific Reports*, 9(1).

Adegas, J. and Nogueira Couto, R. 1992. Entomophilous pollination in rape (*Brassica napus* L. var *oleifera*) in Brazil. *Apidologie*, 23(3):203–209.

Aizen, M. A. and Harder, L. D. 2007. Expanding the limits of the pollen-limitation concept: effects of pollen quantity and quality. *Ecology*, 88(2):271–281.

Ali, M., Saeed, S., Sajjad, A., and Whittington, A. 2011. In search of the best pollinators for canola (*Brassica napus* L.) production in Pakistan. *Applied Entomology and Zoology*, 46(3):353–361.

Angadi, S., Cutforth, H., McConkey, B., and Gan, Y. 2003. Yield adjustment by canola grown at different plant populations under semiarid conditions. *Crop Science*, 43(4):1358–1366.

Ashman, T.-L., Knight, T. M., Steets, J. A., Amarasekare, P., Burd, M., Campbell, D. R., Dudash, M. R., Johnston, M. O., Mazer, S. J., Mitchell, R. J., Morgan, M. T., and Wilson, W. G. 2004. Pollen limitation of plant reproduction: ecological and evolutionary causes and consequences. *Ecology*, 85(9):2408–2421.

Bartomeus, I., Gagic, V., and Bommarco, R. 2015. Pollinators, pests and soil properties interactively shape oilseed rape yield. *Basic and Applied Ecology*, 16(8):737–745.

Batra, S. W. 1978. Aggression, territoriality, mating and nest aggregation of some solitary bees (Hymenoptera: Halictidae, Megachilidae, Colletidae, Anthophoridae). *Journal of the Kansas Entomological Society*, 51(4):547–559.

Beattie, A. 1971. A technique for the study of insect-borne pollen. *Pan-Pacific Entomologist*, 47:82.

Becker, H. C., Damgaard, C., and Karlsson, B. 1992. Environmental variation for outcrossing rate in rapeseed (*Brassica napus*). *Theoretical and Applied Genetics*, 84(3):303–306.

Bommarco, R., Marini, L., and Vaissière, B. E. 2012. Insect pollination enhances seed yield, quality, and market value in oilseed rape. *Oecologia*, 169(4):1025–1032.

Bos, M. M., Veddeler, D., Bogdanski, A. K., Klein, A.-M., Tscharntke, T., Steffan-Dewenter, I., and Tylianakis, J. M. 2007. Caveats to quantifying ecosystem services: Fruit abortion blurs benefits from crop pollination. *Ecological Applications*, 17(6):1841–1849.

Bouttier, C. and Morgan, D. G. 1992. Ovule development and determination of seed number per pod in oilseed rape (*Brassica napus* l.). *Journal of Experimental Botany*, 43(5):709–714.

Brunet, J., Zhao, Y., and Clayton, M. K. 2019. Linking the foraging behavior of three bee species to pollen dispersal and gene flow. *PLOS ONE*, 14(2):e0212561.

Burd, M. 1994. Bateman's principle and plant reproduction: the role of pollen limitation in fruit and seed set. *Botanical Review*, 60(1):83–139.

Cane, J. H., Gardner, D. R., and Harrison, P. A. 2011. Nectar and pollen sugars constituting larval provisions of the alfalfa leaf-cutting bee (*Megachile rotundata*) (Hymenoptera: Apiformes: Megachilidae). *Apidologie*, 42(3):401–408.

Clarke, J. M. 1979. Intra-plant variation in number of seeds per pod and seed weight in *Brassica napus* 'Tower'. *Canadian Journal of Plant Science*, 59(4):959–962.

Clay, H. 2009. Pollinating hybrid canola: the southern Alberta experience. *Hive Lights*, 22(3):14–16.

Clough, Y. 2012. A generalized approach to modeling and estimating indirect effects in ecology. *Ecology*, 93(8):1809–1815.

Cresswell, J. E. 1999. The influence of nectar and pollen availability on pollen transfer by individual flowers of oil-seed rape (*Brassica napus*) when pollinated by bumblebees (*Bombus lapidarius*). *Journal of Ecology*, 87(4):670–677.

Cresswell, J. E., Davies, T. W., Patrick, M. A., Russell, F., Pennel, C., Vicot, M., and Lahoubi, M. 2004. Aerodynamics of wind pollination in a zoophilous flower, *Brassica napus*. *Functional Ecology*, 18(6):861–866.

Cresswell, J. E. and Osborne, J. L. 2004. The effect of patch size and separation on bumblebee foraging in oilseed rape: implications for gene flow. *Journal of Applied Ecology*, 41(3):539–546.

Currie, R. W. 1997. *Pollen Biotechnology for Crop Production and Improvement*, chapter 6. Pollination constraints and management of pollinating insects for crop production, pages 121–151. Cambridge University Press.

Delbrassine, S. and Rasmont, P. 1988. Contribution à l'étude de la pollinisation du colza, *Brassica napus* L. var. *oleifera* (Moench) Delile, en Belgique. *Bulletin des recherches agronomiques de Gembloux*, 23:123–152.

Durán, X. A., Ulloa, R. B., Carrillo, J. A., Contreras, J. L., and Bastidas, M. T. 2010. Evaluation of yield component traits of honeybee pollinated (*Apis mellifera* L.) rapeseed canola (*Brassica napus* L.). *Chilean Journal of Agricultural Research*, 70(2):309–314.

Eilers, E. J., Kremen, C., Greenleaf, S. S., Garber, A. K., and Klein, A.-M. 2011. Contribution of pollinator-mediated crops to nutrients in the human food supply. *PLoS one*, 6(6):e21363.

Erbar, C. 2003. Pollen tube transmitting tissue: place of competition of male gametophytes. *International Journal of Plant Sciences*, 164(S5):S265–S277.

Free, J. and Nuttall, P. 1968. The pollination of oilseed rape (*Brassica napus*) and the behaviour of bees on the crop. *The Journal of Agricultural Science*, 71(01):91–94.

Free, J. B. and Ferguson, A. W. 1983. Foraging behaviour of honeybees on oilseed rape. *Bee World*, 64(1):22–24.

Free, J. B. and Williams, I. H. 1973. The foraging behaviour of honeybees (*Apis mellifera* L.) on brussels sprout (*Brassica oleracea* L.). *Journal of Applied Ecology*, 10(2):489–499.

Fries, I. and Stark, J. 1983. Measuring the importance of honeybees in rape seed production. *Journal of Apicultural Research*, 22(4):272–276.

Gaffney, A., Bohman, B., Quarrell, S., Brown, P., and Allen, G. 2019. Limited cross plant movement and non-crop preferences reduce the efficiency of honey bees as pollinators of hybrid carrot seed crops. *Insects*, 10(2):34.

Gagic, V., Kleijn, D., Báldi, A., Boros, G., Jørgensen, H. B., Elek, Z., Garratt, M. P., Groot, G. A., Hedlund, K., Kovács-Hostyánszki, A., Marini, L., Martin, E., Pevere, I., Potts, S. G., Redlich, S., Senapathi, D., Steffan-Dewenter, I., Świtek, S., Smith, H. G., Takács, V., Tryjanowski, P., van der Putten, W. H., van Gils, S., and Bommarco, R. 2017. Combined effects of agrochemicals and ecosystem services on crop yield across Europe. *Ecology Letters*, 20(11):1427–1436.

Gallai, N., Salles, J.-M., Settele, J., and Vaissière, B. E. 2009. Economic valuation of the vulnerability of world agriculture confronted with pollinator decline. *Ecological Economics*, 68(3):810–821.

Gelman, A., Carlin, J., Stern, H., Dunson, D., Vehtari, A., and Rubin, D. 2013. *Bayesian Data Analysis*. Chapman and Hall, New York, 3rd edition.

Gelman, A., Lee, D., and Guo, J. 2015. Stan. *Journal of Educational and Behavioral Statistics*, 40(5):530–543.

Goerzen, D. W., Richards, K. W., and Rank, G. H. 1995. Drifting in populations of the alfalfa leafcutting bee, *Megachile rotundata* (Fab.) (Hym., Megachilidae). *Journal of Applied Entomology*, 119(1-5):575–579.

Goldberg, R. B., de Paiva, G., and Yadegari, R. 1994. Plant embryogenesis: Zygote to seed. *Science*, 266:605–614.

Goulson, D., Ollerton, J., and Sluman, C. 1997. Foraging strategies in the small skipper butterfly, *Thymelicus flavus*: when to switch? *Animal Behaviour*, 53(5):1009 – 1016.

Grace, J. B. 2006. *Structural equation modeling and natural systems*. Cambridge University Press.

Grace, J. B., Schoolmaster, D. R., Guntenspergen, G. R., Little, A. M., Mitchell, B. R., Miller, K. M., and Schweiger, E. W. 2012. Guidelines for a graph-theoretic implementation of structural equation modeling. *Ecosphere*, 3(8):1–44. art73.

Greenleaf, S. S. and Kremen, C. 2006. Wild bees enhance honey bees' pollination of hybrid sunflower. *Proceedings of the National Academy of Sciences*, 103(37):13890–13895.

Greenleaf, S. S., Williams, N. M., Winfree, R., and Kremen, C. 2007. Bee foraging ranges and their relationship to body size. *Oecologia*, 153(3):589–596.

Harder, L. D., Aizen, M. A., and Richards, S. A. 2016. The population ecology of male gametophytes: the link between pollination and seed production. *Ecology Letters*, 19(5):497–509.

Heinrich, B. 1976. The foraging specializations of individual bumblebees. *Ecological Monographs*, 46(2):105–128.

Heinrich, B. 1979. Resource heterogeneity and patterns of movement in foraging bumblebees. *Oecologia*, 40(3):235–245.

Horne, M. 1995a. Leaf area and toughness: Effects on nesting material preferences of *Megachile rotundata* (Hymenoptera: Megachilidae). *Annals of the Entomological Society of America*, 88(6):868–875.

Horne, M. 1995b. Pollen preference and its relationship to nesting success of *Megachile rotundata* (Hymenoptera: Megachilidae). *Annals of the Entomological Society of America*, 88(6):862–867.

Hudewenz, A., Pufal, G., Bögeholz, A., and Klein, A. 2013. Cross-pollination benefits differ among oilseed rape varieties. *The Journal of Agricultural Science*, 152(5):770–778.

Isaacs, R. and Kirk, A. K. 2010. Pollination services provided to small and large highbush blueberry fields by wild and managed bees. *Journal of Applied Ecology*, 47(4):841–849.

Iwasa, Y. 2000. Dynamic optimization of plant growth. *Evolutionary Ecology Research*, 2:437–455.

Jauker, F. and Wolters, V. 2008. Hover flies are efficient pollinators of oilseed rape. *Oecologia*, 156(4):819–823.

Kassambara, A. 2020. *ggpubr: 'ggplot2' Based Publication Ready Plots*. R package version 0.4.0.

Kirkegaard, J. A., Lilley, J. M., Brill, R. D., Ware, A. H., and Walela, C. K. 2018. The critical period for yield and quality determination in canola (*Brassica napus* L.). *Field Crops Research*, 222:180–188.

Kline, R. B. 2013. *Structural equation modeling: A second course*, chapter Reverse arrow dynamics: Feedback loops and formative measurement, pages 41–79. Information Age Publishing, Charlotte, NC, second edition.

Knight, T. M., Steets, J. A., and Ashman, T.-L. 2006. A quantitative synthesis of pollen supplementation experiments highlights the contribution of resource reallocation to estimates of pollen limitation. *American Journal of Botany*, 93(2):271–277.

Knight, T. M., Steets, J. A., Vamosi, J. C., Mazer, S. J., Burd, M., Campbell, D. R., Dudash, M. R., Johnston, M. O., Mitchell, R. J., and Ashman, T.-L. 2005. Pollen limitation of plant reproduction: Pattern and process. *Annual Review of Ecology, Evolution, and Systematics*, 36:467–497.

Kołtowski, Z. 2005. The effect of pollinating insects on the yield of winter rapeseed (*Brassica napus* L. var. *napus* f. biennis) cultivars. *Journal of Apicultural Science*, 49(2):29–41.

Lawrence, W. S. 1993. Resource and pollen limitation: Plant size-dependent reproductive patterns in *Physalis longifolia*. *The American Naturalist*, 141(2):296–313.

Lefcheck, J. S. 2015. piecewiseSEM: Piecewise structural equation modelling in `R` for ecology, evolution, and systematics. *Methods in Ecology and Evolution*, 7(5):573–579.

Lovett-Doust, J. and Eaton, G. W. 1982. Demographic aspects of flower and fruit production in bean plants, *Phaseolus vulgaris* L. *American Journal of Botany*, 69(7):1156–1164.

Manning, R. and Wallis, I. R. 2005. Seed yields in canola (*Brassica napus* cv. karoo) depend on the distance of plants from honeybee apiaries. *Animal Production Science*, 45(10):1307–1313.

Marini, L., Tamburini, G., Petrucco-Toffolo, E., Lindström, S. A., Zanetti, F., Mosca, G., and Bommarco, R. 2015. Crop management modifies the benefits of insect pollination in oilseed rape. *Agriculture, Ecosystems & Environment*, 207:61–66.

Mazer, S. J. 1987. The quantitative genetics of life history and fitness components in *Raphanus raphanistrum* L. (Brassicaceae): Ecological and evolutionary consequences of seed-weight variation. *The American Naturalist*, 130(6):891–914.

Melathopoulos, A. P., Cutler, G. C., and Tyedmers, P. 2015. Where is the value in valuing pollination ecosystem services to agriculture? *Ecological Economics*, 109:59 – 70.

Mesquida, J., Pham-Délègue, M. H., Marilleau, R., Le Metayer, M., and Renard, M. 1991. La sécrétion nectarifère des fleurs de cybrides mâles-stériles de colza d'hiver (*Brassica napus* L.). *Agronomie*, 11(3):217–227.

Mesquida, J. and Renard, M. 1978. Entomophilous pollination of male-sterile strains of winter rapeseed (*Brassica napus* L. Metzger var. *oleifera*) and a preliminary study of alternating devices. *Proceedings of the IVth International Symposium on Pollination*, 4:49–57. University of Maryland.

Mesquida, J. and Renard, M. 1981. Pollinisation du colza d'hiver mâle-fertile et mâle-sterile (*Brassica napus* L. var *oleifera* metzger) par l'abeille domestique (*Apis m. mellifica* L.): effets sur la phénologie et le rendement. *Apidologie*, 12:345–362.

Mesquida, J. and Renard, M. 1982. Étude de la dispersion du pollen par le vent et de l'importance de la pollinisation anémophile chez le colza (*Brassica napus* L. var. *oleifera* Metzger). *Apidologie*, 13(4):353–366.

Mesquida, J., Renard, M., and Pierre, J.-S. 1988. Rapeseed (*Brassica napus* L.) productivity : the effect of honeybees (*Apis mellifera* L.) and different pollination conditions in cage and field tests. *Apidologie*, 19(1):51–72.

Mohr, N. A. and Jay, S. C. 1988. Nectar- and pollen-collecting behaviour of honeybees on canola (*Brassica campestris* L. and *Brassica napus* L.). *Journal of Apicultural Research*, 27(2):131–136.

Morandin, L. A. and Winston, M. L. 2005. Wild bee abundance and seed production in conventional, organic, and genetically modified canola. *Ecological Applications*, 15(3):871–881.

Neal, P. R. and Anderson, G. J. 2004. Does the 'old bag' make a good 'wind bag'?: Comparison of four fabrics commonly used as exclusion bags in studies of pollination and reproductive biology. *Annals of Botany*, 93(5):603–607.

Nord, E. A., Shea, K., and Lynch, J. P. 2011. Optimizing reproductive phenology in a two-resource world: a dynamic allocation model of plant growth predicts later reproduction in phosphorus-limited plants. *Annals of Botany*, 108(2):391–404.

Olsson, G. 1960. Self-incompatibility and outcrossing in rape and white mustard. *Hereditas*, 46:241–252.

Ouvrard, P. and Jacquemart, A. 2019. Review of methods to investigate pollinator dependency in oilseed rape (*Brassica napus*). *Field Crops Research*, 231:18–29.

Parker, A. J., Tran, J. L., Ison, J. L., Bai, J. D. K., Weis, A. E., and Thomson, J. D. 2015. Pollen packing affects the function of pollen on corbiculate bees but not non-corbiculate bees. *Arthropod-Plant Interactions*, 9(2):197–203.

Perrot, T., Gaba, S., Roncoroni, M., Gautier, J.-L., and Bretagnolle, V. 2018. Bees increase oilseed rape yield under real field conditions. *Agriculture, Ecosystems & Environment*, 266:39–48.

Peterson, J. H. and Roitberg, B. D. 2005. Impacts of flight distance on sex ratio and resource allocation to offspring in the leafcutter bee, *Megachile rotundata*. *Behavioral Ecology and Sociobiology*, 59(5):589–596.

Pierre, J., Vaissière, B., Vallée, P., and Renard, M. 2010. Efficiency of airborne pollen released by honeybee foraging on pollination in oilseed rape: a wind insect-assisted pollination. *Apidologie*, 41(1):109–115.

Pinnisch, R. and McVetty, P. B. E. 1990. Seed production of hybrid summer rape in the field using the pol cytoplasmic male sterility system: a first attempt. *Canadian Journal of Plant Science*, 70(3):611–618.

Pitts-Singer, T. L. 2013. Intended release and actual retention of alfalfa leafcutting bees (Hymenoptera: Megachilidae) for pollination in commercial alfalfa seed fields. *Journal of Economic Entomology*, 106(2):576–586.

Pitts-Singer, T. L. and Cane, J. H. 2011. The alfalfa leafcutting bee, *Megachile rotundata*: the world's most intensively managed solitary bee. *Annual Review of Entomology*, 56(1):221–237.

Plowright, R. C. and Hartling, L. K. 1981. Red clover pollination by bumble bees: A study of the dynamics of a plant- pollinator relationship. *The Journal of Applied Ecology*, 18(2):639–647.

Pyke, G. H. 1978. Optimal foraging: Movement patterns of bumblebees between inflorescences. *Theoretical Population Biology*, 13(1):72–98.

Rader, R. 2010. *The provision of pollination services to agroecosystems by a diverse assemblage of wild, unmanaged insect taxa*. PhD thesis, James Cook University.

Rakow, G. and Woods, D. L. 1987. Outcrossing in rape and mustard under Saskatchewan prairie conditions. *Canadian Journal of Plant Science*, 67(1):147–151.

Ricketts, T. H., Regetz, J., Steffan-Dewenter, I., Cunningham, S. A., Kremen, C., Bogdanski, A., Gemmill-Herren, B., Greenleaf, S. S., Klein, A. M., Mayfield, M. M., Morandin, L. A., Ochieng', A., and Viana, B. F. 2008. Landscape effects on crop pollination services: are there general patterns? *Ecology Letters*, 11(5):499–515.

Robinson, S. V. J., Hoover, S. E., Pernal, S. F., and Cartar, R. V. 2022. Optimal distributions of central-place foragers: honey bee foraging in a mass flowering crop. *Behavioral Ecology*, 33(2):386–397.

Rosa, A. d. S., Blochtein, B., and Lima, D. K. 2011. Honey bee contribution to canola pollination in southern Brazil. *Scientia Agricola*, 68:255 – 259.

Sabbahi, R., de Oliveira, D., and Marceau, J. 2005. Influence of honey bee (Hymenoptera: Apidae) density on the production of canola (Crucifera: Brassicacae). *Journal of Economic Entomology*, 98(2):367–372.

Sabbahi, R., De Oliveira, D., and Marceau, J. 2006. Does the honeybee (Hymenoptera: Apidae) reduce the blooming period of canola? *Journal of Agronomy and Crop Science*, 192(3):233–237.

Sáez, A., Morales, J. M., Morales, C. L., Harder, L. D., and Aizen, M. A. 2018. The costs and benefits of pollinator dependence: empirically based simulations predict raspberry fruit quality. *Ecological Applications*, 28(5):1215–1222.

Schaffer, W. M., Jensen, D. B., Hobbs, D. E., Gurevitch, J., Todd, J. R., and Schaffer, M. V. 1979. Competition, foraging energetics, and the cost of sociality in three species of bees. *Ecology*, 60(5):976–987.

Shipley, B. 2000. A new inferential test for path models based on directed acyclic graphs. *Structural Equation Modeling: A Multidisciplinary Journal*, 7(2):206–218.

Shipley, B. 2009. Confirmatory path analysis in a generalized multilevel context. *Ecology*, 90(2):363–368.

Sinu, P. A. and Bronstein, J. L. 2018. Foraging preferences of leafcutter bees in three contrasting geographical zones. *Diversity and Distributions*, 24(5):621–628.

Snoep, J. D., Morabia, A., Hernández-Díaz, S., Hernán, M. A., and Vandenbroucke, J. P. 2014. Commentary: A structural approach to berkson's fallacy and a guide to a history of opinions about it. *International Journal of Epidemiology*, 43(2):515–521.

Soroka, J., Goerzen, D., Falk, K., and Bett, K. 2001. Alfalfa leafcutting bee (Hymenoptera: Megachilidae) pollination of oilseed rape (*Brassica napus* L.) under isolation tents for hybrid seed production. *Canadian Journal of Plant Science*, 81(1):199–204.

Stan Development Team. RStan: the R interface to Stan. R package version 2.26.13.

Steffan-Dewenter, I. 2003. Seed set of male-sterile and male-fertile oilseed rape (*Brassica napus*) in relation to pollinator density. *Apidologie*, 34:227–235.

Steffan-Dewenter, I. and Tscharntke, T. 2000. Resource overlap and possible competition between honey bees and wild bees in central europe. *Oecologia*, 122(2):288–296.

Stephenson, A. G. 1981. Flower and fruit abortion: Proximate causes and ultimate functions. *Annual Review of Ecology and Systematics*, 12:253–279.

Tamburini, G., Bommarco, R., Kleijn, D., van der Putten, W. H., and Marini, L. 2019. Pollination contribution to crop yield is often context-dependent: A review of experimental evidence. *Agriculture, Ecosystems & Environment*, 280:16–23.

Tamburini, G., Lami, F., and Marini, L. 2017. Pollination benefits are maximized at intermediate nutrient levels. *Proceedings of the Royal Society of London B: Biological Sciences*, 284:20170729.

Thomson, J., Peterson, S., and Harder, L. 1987. Response of traplining bumble bees to competition experiments: shifts in feeding location and efficiency. *Oecologia*, 71(2):295–300.

Thomson, J. D. 1986. Pollen transport and deposition by bumble bees in *Erythronium*: influences of floral nectar and bee grooming. *The Journal of Ecology*, 74:329–341.

Thomson, J. D. and Goodell, K. 2001. Pollen removal and deposition by honeybee and bumblebee visitors to apple and almond flowers. *Journal of Applied Ecology*, 38(5):1032–1044.

Venable, D. L. 1992. Size-number trade-offs and the variation of seed size with plant resource status. *The American Naturalist*, 140(2):287–304.

Waytes, R. J. 2017. Pollinator movement and pollen transfer in hybrid seed canola. Master's thesis, University of Calgary.

Westcott, L. and Nelson, D. 2001. Canola pollination: an update. *Bee World*, 82(3):115–129.

Wickham, H. 2016. *ggplot2: Elegant Graphics for Data Analysis*. Springer-Verlag New York.

Williams, I. H., Martin, A., and White, R. 1986. The pollination requirements of oil-seed rape (*Brassica napus* L.). *The Journal of Agricultural Science*, 106(01):27–30.

Zhang, H. and Flottmann, S. 2018. Source-sink manipulations indicate seed yield in canola is limited by source availability. *European Journal of Agronomy*, 96:70–76.

Zurbuchen, A., Landert, L., Klaiber, J., Müller, A., Hein, S., and Dorn, S. 2010. Maximum foraging ranges in solitary bees: only few individuals have the capability to cover long foraging distances. *Biological Conservation*, 143(3):669–676.

Appendix A: Additional figures and tables

Table S1: Summary of variables used in structural equation models.

Field Type	Variable	Mean	Median	SD	Min	Max
Commodity	Number of hives	14.80	0.00	17.03	0.00	40.00
	Distance to edge (m)	137.48	20.00	195.02	1.00	500.00
	HB visitation (hr^{-1})	10.31	0.00	28.80	0.00	218.18
	Flower density (m^2)	470.33	448.00	231.32	52.00	1684.00
	Pollen per stigma	293.33	155.00	385.04	0.00	3891.00
	Plant density (m^2)	3.77	3.77	0.48	1.79	5.02
	Plant vegetative mass (g)	18.15	14.32	14.07	0.77	107.66
	Plant seed mass (g)	6.87	5.46	5.97	0.01	47.90
	Harvest index (g/g)	0.38	0.37	0.16	0.00	1.89
	Flowers per plant	196.09	156.50	150.96	13.00	1094.00
	Pods per plant	143.15	112.00	114.64	5.00	892.00
	Seeds per pod	22.96	23.60	4.97	4.60	35.40
	Seed size (mg)	2.74	2.73	0.80	0.39	5.35
Seed	Distance to edge (m)	162.70	100.00	146.18	3.00	400.00
	Distance to LCB shelter (m)	33.51	31.00	24.63	2.00	190.00
	HB visitation (hr^{-1})	112.79	24.00	187.30	0.00	1290.00
	LCB visitation (hr^{-1})	76.15	12.00	144.88	0.00	1272.00
	Bay Edge/Centre	0.13	0.00	0.34	0.00	1.00
	Flower density (m^2)	495.02	432.00	303.35	24.00	2686.40
	Pollen per stigma	21.76	7.00	42.53	0.00	578.00
	Plant density (m^2)	3.58	3.64	0.46	2.40	4.49
	Plant vegetative mass (g)	30.32	25.19	21.16	1.18	144.32
	Plant seed mass (g)	9.50	7.54	7.90	0.02	60.77
	Harvest index (g/g)	0.32	0.33	0.15	0.00	1.21
	Flowers per plant	461.32	362.50	326.59	26.00	2712.00
	Pods per plant	299.41	244.50	207.68	10.00	1410.00
	Seeds per pod	16.40	16.60	5.52	1.80	30.60
	Seed size (mg)	3.43	3.42	0.88	1.04	5.59
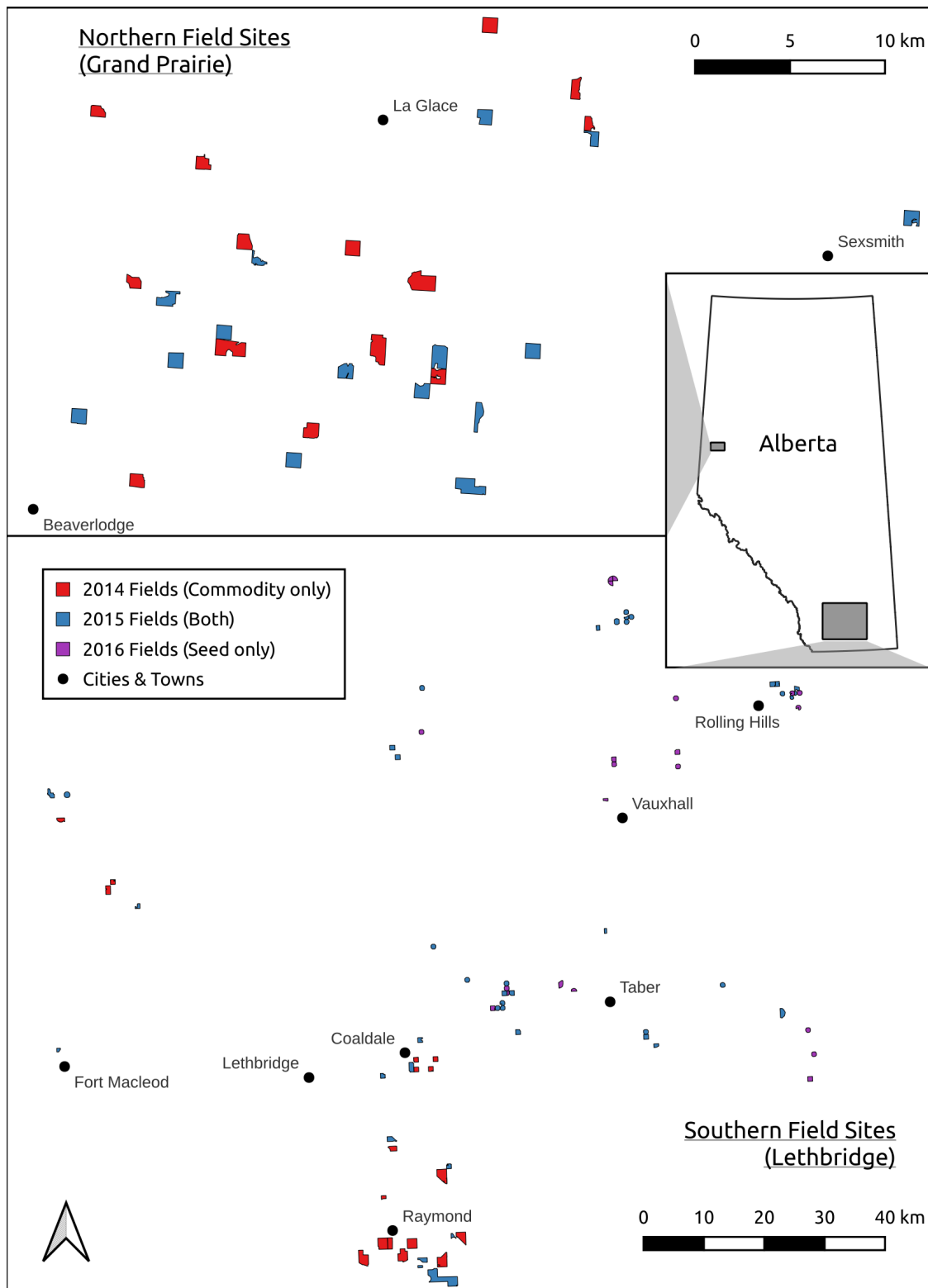
Figure S1: Map of sampled fields, showing locations of 29 commodity canola fields (14 during 2014, 15 during 2015), and 35 seed canola fields (15 during 2015, 20 during 2016). Seed canola is grown only in southern Alberta, while commodity canola is grown across both the northern and southern regions.

Figure S2: Hybrid seed field near Rainer, AB, showing the outlines of male and female bays in the foreground, with orange leafcutter bee shelters stationed throughout the field. The linear structure on the horizon is the central-pivot irrigation sprinkler.
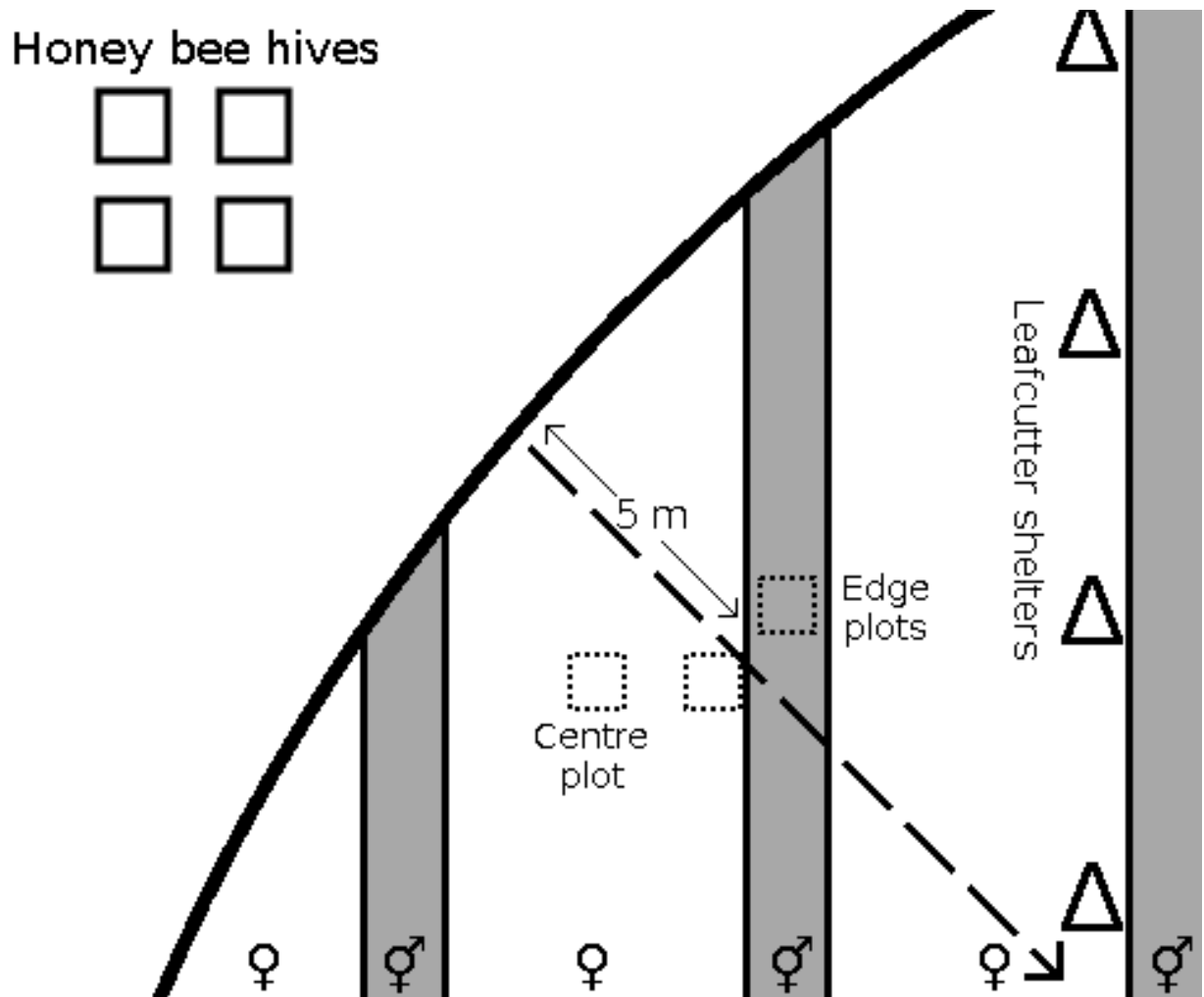
Figure S3: Plot arrangement for surveys in hybrid seed fields, showing hypothetical arrangement of leafcutter shelters (Δ), and male-fertile (♂) and female bays (♀) at 5m from the edge of the field. Plots were placed along a transect (dashed line) from the field edge nearest to the set of honey bee hives. Plots were placed side-by-side in the male bay and edge of the female bay ("edge" plots), and at the 5m and 400m distances, an additional plot was placed in the centre of the female bay ("centre" plots). Note: this figure shows a small section of a much larger circular field.

Appendix B: Additional information on insect visitors

In 2015, we recorded whether honey bees were top-working or side-working flowers (see also Free and Williams, 1973; Free and Ferguson, 1983; Mohr and Jay, 1988). Top-working bees landed on the top of the flower and inserted their proboscis down between the petals to access the nectaries of the flower, while side-working bees landed on the side of the flower and stole nectar by inserted their proboscis between the petals, avoiding contact with the stigma or anthers. Additionally, we recorded whether honey bees were pollen or nectar foragers (pollen foragers had a visible pollen load on their corbicula, while nectar foragers had none).

Pollen- and nectar-foraging honey bees had different patterns of side-working, both on commodity canola, and the male and female lines of seed canola. Side-working was common in nectar foragers, but was more common in commodity canola (64%) than in the male (36%) or female bays (2.8%) of seed canola, indicating that a large proportion of honey bees foraging on canola flowers may never come in contact with the stigmas. Pollen foragers were almost uniformly top-foragers in both commodity and seed fields (Table S3), and pollen foragers were much less common in the female bays (1.4%) than in the male bays (15%), or in commodity fields (18%). Therefore, foraging honey bees in seed canola fields tend to treat male-fertile flowers similar to commodity canola flowers, but seem to top-work flowers more in commodity canola than seed fields. Leafcutter bee foraging behaviours were not recorded, but seemed to almost exclusively top-work flowers in seed canola fields.

Table S2: Number of flower visitors recorded over a total of 44.8 hours of observation in commodity fields (2014 and 2015), and 46.9 hours of observation in the seed fields (2015 and 2016). "Fly" refers to larger calyptrate muscoid flies (families Muscidae, Anthomyiidae, Caliphoridae), while "Hover fly" refers to Syrphid flies. "Other bee" included Halictid and Andrenid bees, while "Bumble bee" was *Bombus* spp. "Butterfly" refers to all visiting Lepidopterans, mostly Pierids.

Taxon	Commodity fields		Seed fields	
	Visits	%	Visits	%
Honey bee	470	53.5	4850	77.1
Fly	222	25.3	74	0.878
Hover fly	94	10.7	151	1.79
Other bee	47	5.35	30	0.356
Bumble bee	25	2.85	0	0
Butterfly	16	1.82	0	0
Leafcutter bee	4	0.456	1675	19.9

Table S3: Foraging behaviours of honey bees on commodity and seed canola flowers, recorded during 2015. "Top" (top-working) indicates that the bee inserted their proboscis down between the petals from the top of the flower, while "side" (side-working) indicates that the bee fed from the side of the flower and did not contact the anthers or stigma. Pollen foragers had pollen visible on their corbicula, while nectar foragers had none.

	Commodity fields		Seed fields (female bay)		Seed fields (male bay)	
	Top	Side	Top	Side	Top	Side
Pollen forager	44	2	12	0	115	0
Nectar forager	75	138	832	24	428	242

Appendix C: Additional information on models

Commodity canola models

Formulas for commodity canola model using `lmer`-style R formulas. Terms on right side of $\sim$ indicate fixed effects, while terms in brackets indicate random effects (heirarchical intercepts). *distribution* indicates the type of probability distribution function used to model each variable.

$$\text{Plant Density} \sim \text{HB Distance} + (1|\text{Field}), \text{distribution} = \text{log-normal}$$
$$\text{Plant Size} \sim \text{Plant Density} + \text{HB Distance} + (1|\text{Field}), \text{distribution} = \text{log-normal}$$
$$\text{Flower Density} \sim \text{Plant Size} + \text{HB Distance} + \text{Plant Density} + (1|\text{Field}),$$
$$\text{distribution} = \text{square root-normal}$$
$$\text{HB Visits} \sim \text{offset}(log(\text{Time})) + \text{HB Distance} + \text{Hive Stocking} +$$
$$\text{Flower Density} + (1|\text{Field}), \text{family} = \text{negative binomial}$$
$$\text{Pollen per Stigma} \sim \text{HB Visits} + \text{HB Distance} + (1|\text{Field}), \text{distribution} = \text{negative binomial}$$
$$\text{Flowers per Plant} \sim \text{Plant Size} + \% \text{ Pod Set} + (1|\text{Field}), \phi \sim \text{Plant Size},$$
$$\text{distribution} = \text{negative binomial}$$
$$\% \text{ Pod Set} \sim \text{HB Visits} + \text{Pollen} + \text{Plant Size} + (1|\text{Field}), \text{distribution} = \text{beta-binomial}$$
$$\text{Seeds per Pod} \sim \text{HB Visits} + \text{Pollen} + \text{Plant Size} + \% \text{ Pod Set} + \text{Flowers per Plant} + (1|\text{Field}),$$
$$\text{distribution} = \text{exponential-normal}$$
$$\text{Weight per Seed} \sim \text{HB Visits} + \text{Pollen} + \text{Seeds per Pod} + \text{Plant Size} + \text{Plant Density} +$$
$$\text{HB Distance} + \% \text{ Pod Set} + \text{Flowers per Plant} + \text{Flower Density} + (1|\text{Field}),$$
$$\text{distribution} = \text{exponential-normal}$$

Table S4: Summary of parameters for commodity canola models

Model	Parameter	Mean	SD	Median	Min	Max	P-value	N_{eff}	$\hat{R}$
Plant density	Intercept	3.76	0.05	3.77	3.66	3.87	<0.0001	855	1.005
	HB distance	0.02	0.01	0.02	0.00	0.04	0.0868	10154	1.000
	Sigma	0.31	0.02	0.31	0.28	0.34	-	5039	0.999
	Sigma (field)	0.37	0.04	0.37	0.30	0.46	-	3774	1.000
Plant size	Intercept	5.24	0.24	5.24	4.77	5.72	<0.0001	2108	1.001
	Plant density	-0.70	0.06	-0.70	-0.82	-0.57	<0.0001	2097	1.001
	HB distance	0.03	0.01	0.03	0.01	0.06	0.0095	9212	1.000
	Sigma (field)	0.22	0.04	0.22	0.15	0.29	-	1456	1.000
	Sigma	0.64	0.02	0.64	0.61	0.68	-	7845	0.999
Flower density	Intercept	5.74	4.70	5.70	-3.62	15.01	0.222	1822	1.001
	Plant size	-1.66	0.98	-1.66	-3.60	0.25	0.0926	1632	1.001
	HB distance	0.75	0.11	0.75	0.53	0.96	<0.0001	6878	1.000
	Plant density	-0.41	0.74	-0.42	-1.86	1.06	0.5792	2243	1.001
	Sigma	3.52	0.17	3.51	3.19	3.87	-	4976	0.999
	Sigma (field)	3.55	0.39	3.53	2.88	4.40	-	4133	1.000
	Intercept	-1.98	0.45	-1.96	-2.89	-1.17	<0.0001	936	1.006

Model	Parameter	Mean	SD	Median	Min	Max	P-value	N_{eff}	$\hat{R}$
HB visits	HB distance	-0.31	0.10	-0.31	-0.50	-0.13	0.0011	3526	1.001
	Number of hives	0.72	0.15	0.72	0.43	1.04	<0.0001	1534	1.003
	Flower density	0.03	0.04	0.03	-0.06	0.12	0.541	2397	1.000
	Sigma (field)	1.44	0.34	1.43	0.80	2.14	-	494	1.014
	Phi	0.35	0.08	0.34	0.22	0.53	-	1630	1.003
Flowers per plant	Intercept	2.56	0.04	2.56	2.50	2.63	<0.0001	594	1.006
	Plant size	0.94	0.01	0.94	0.92	0.96	<0.0001	1709	1.002
	Pods per plant	-0.16	0.01	-0.16	-0.19	-0.14	<0.0001	2108	1.003
	Phi (field)	0.16	0.02	0.16	0.13	0.19	-	3461	1.000
	Intercept (Phi)	2.17	0.32	2.17	1.56	2.81	-	1517	1.001
	Plant size (Phi)	0.66	0.11	0.66	0.44	0.87	-	1587	1.002
	SigmaPhi (field)	0.66	0.12	0.65	0.45	0.90	-	844	1.005
Pods per plant	Intercept	0.66	0.08	0.66	0.50	0.82	<0.0001	1946	0.999
	HB visits	0.00	0.03	0.00	-0.05	0.05	0.96	3292	1.000
	Plant size	0.13	0.03	0.13	0.08	0.18	<0.0001	3345	1.000
	Pollen count	0.16	0.11	0.15	-0.05	0.38	0.1515	638	1.002
	Sigma (field)	0.29	0.03	0.29	0.23	0.37	-	2675	1.000
	Phi	3.35	0.07	3.35	3.21	3.48	-	3828	1.003
Seeds per pod	Intercept	25.07	1.77	25.07	21.64	28.54	<0.0001	2277	1.000
	HB visits	0.17	0.21	0.17	-0.24	0.60	0.4284	4417	1.000
	Pollen count	-0.97	0.84	-0.98	-2.60	0.68	0.2447	707	1.003
	Plant size	4.31	0.65	4.31	3.03	5.58	<0.0001	2229	1.000
	Pods per plant	0.56	0.29	0.56	0.00	1.12	0.0529	4888	0.999
	Flowers per plant	-2.80	0.65	-2.79	-4.08	-1.55	<0.0001	2105	1.000
	Sigma	4.06	0.12	4.06	3.83	4.29	-	6604	1.000
	Sigma (field)	1.98	0.26	1.96	1.51	2.53	-	2526	0.999
	Lambda	2.00	1.03	1.74	0.84	4.64	-	3265	1.000
Seed size	Intercept	0.64	0.54	0.64	-0.40	1.72	0.2367	1402	1.000
	HB visits	0.02	0.04	0.02	-0.05	0.09	0.6113	2748	1.000
	Pollen count	-0.15	0.17	-0.15	-0.48	0.20	0.3744	358	1.008
	Seeds per pod	0.00	0.01	0.00	-0.01	0.02	0.4508	3348	1.000
	Plant size	-0.03	0.12	-0.03	-0.26	0.20	0.7955	1633	1.000
	Plant density	0.30	0.08	0.30	0.14	0.45	0.0001	1704	1.001
	HB distance	0.02	0.01	0.02	-0.01	0.05	0.1686	1404	1.003
	Pods per plant	0.11	0.04	0.11	0.02	0.19	0.0159	3037	1.000
	Flowers per plant	0.11	0.12	0.11	-0.13	0.34	0.363	1526	1.000
	Flower density	-0.03	0.01	-0.03	-0.04	-0.02	<0.0001	2213	1.003
	Sigma	0.52	0.03	0.52	0.46	0.59	-	2100	1.000
	Sigma (field)	0.47	0.05	0.47	0.37	0.58	-	2828	0.999
	Lambda	2.71	0.48	2.63	2.08	3.86	-	1792	1.001

Seed canola models

Formulas for seed canola model using `lmer`-style R formulas. Terms on right side of $\sim$ indicate fixed effects, while terms in brackets indicate random effects (heirarchical intercepts). *distribution* indicates the type of probability distribution function used to model each variable.

$$\text{Plant Density} \sim \text{HB Distance} + (1|\text{Field}), \text{distribution} = \text{log-normal}$$

$$\text{Plant Size} \sim \text{HB Distance} + \text{Plant Density} + (1|\text{Field}), \text{distribution} = \text{log-t}$$

$$\text{Flower Density} \sim \text{HB Distance} + (1|\text{Field}), \text{distribution} = \text{square root-t}$$

$$\text{LCB Visits} \sim \text{offset}(log(\text{Time})) + \text{HB Distance} + \text{LCB Distance} + \text{Bay Centre}+$$
$$\text{Flower Density} + (1|\text{Field}), \text{family} = \text{ZI negative binomial}$$

$$\text{HB Visits} \sim \text{offset}(log(\text{Time})) + \text{HB Distance} + \text{LCB Distance} + \text{Bay Centre}+$$
$$\text{Flower Density} + (1|\text{Field}), \text{family} = \text{ZI negative binomial}$$

$$\text{Pollen per Stigma} \sim \text{HB Visits} + \text{LCB Visits} + \text{Bay Centre} + \text{HB Distance} + \text{LCB Distance}+$$
$$\text{Flower Density} + (1|\text{Field}) + (1|\text{Plot}), \text{family} = \text{negative binomial}$$

$$\text{Flowers per Plant} \sim \text{Plant Size} + \text{Bay Centre} + \% \text{ Pod Set} + (1|\text{Field}),$$
$$\phi \sim \text{Plant Size}, \text{family} = \text{negative binomial}$$

$$\% \text{ Pod Set} \sim \text{Pollen} + \text{Plant Size} + \text{Bay Centre} + \text{HB Distance} + \text{LCB Distance}+$$
$$\text{Flower Density} + (1|\text{Field}) + (1|\text{Plot}), \text{family} = \text{beta-binomial}$$

$$\text{Seeds per Pod} \sim \text{Pollen} + \text{Plant Size} + \text{Bay Centre} + \text{HB Distance} + \text{Flower Density}+$$
$$\% \text{ Pod Set} + \text{Flowers per plant} + (1|\text{Field}), \text{family} = \text{negative binomial}$$

$$\text{Weight per Seed} \sim \text{Pollen} + \text{Seeds per Pod} + \text{Plant Size} + \text{LCB Distance}+$$
$$\text{Plant Density} + (1|\text{Field}), \text{family} = \text{exponential-normal}$$

Table S5: Summary of parameters for seed canola models

Model	Parameter	Mean	SD	Median	Min	Max	P-value	N_{eff}	$\hat{R}$
Plant density	Intercept	3.60	0.06	3.60	3.47	3.73	<0.0001	524	1.004
	HB distance	0.06	0.01	0.06	0.04	0.07	<0.0001	5750	0.999
	Sigma	0.27	0.01	0.27	0.24	0.30	-	5199	0.999
	Sigma (field)	0.36	0.05	0.36	0.28	0.48	-	4182	1.000
Plant size	Intercept	6.08	0.25	6.08	5.57	6.56	<0.0001	2480	1.000
	Plant density	-0.79	0.07	-0.80	-0.92	-0.66	<0.0001	2470	1.000
	HB distance	0.07	0.01	0.07	0.05	0.10	<0.0001	6446	0.999
	Sigma	0.50	0.03	0.50	0.45	0.56	-	2332	1.000
	Sigma (field)	0.15	0.04	0.15	0.08	0.23	-	1053	1.001
	Nu (DF)	1.91	0.32	1.87	1.37	2.65	<0.0001	2700	1.000
Flower density	Intercept	0.48	0.56	0.46	-0.58	1.64	0.3827	661	1.005
	HB distance	1.11	0.12	1.11	0.87	1.36	<0.0001	5389	1.000
	Sigma	4.06	0.24	4.05	3.59	4.56	-	2404	1.001
	Sigma (field)	3.48	0.47	3.44	2.70	4.49	-	2640	1.001
	Nu (DF)	1.88	0.38	1.84	1.30	2.68	<0.0001	1500	1.004
HB visits	Intercept	3.08	0.10	3.08	2.87	3.28	<0.0001	2722	1.002
	Flower density	-0.01	0.01	-0.01	-0.03	0.01	0.5267	4150	1.000
	HB distance	-0.02	0.05	-0.03	-0.12	0.07	0.6229	4410	1.000
	LCB distance	0.42	0.09	0.42	0.24	0.58	<0.0001	5053	1.000
	Bay centre	0.45	0.21	0.45	0.07	0.88	0.0284	6486	0.999
	Phi	0.72	0.09	0.72	0.55	0.92	-	4354	1.001
	Theta (ZI)	0.32	0.03	0.32	0.26	0.38	-	3955	1.001
	Sigma (field)	0.41	0.11	0.41	0.22	0.64	-	600	1.001

Model	Parameter	Mean	SD	Median	Min	Max	P-value	N_{eff}	$\hat{R}$
LCB visits	Intercept	2.25	0.14	2.25	1.96	2.51	<0.0001	1734	1.002
	LCB distance	-0.79	0.07	-0.79	-0.94	-0.65	<0.0001	4016	1.000
	HB distance	-0.32	0.06	-0.32	-0.43	-0.21	<0.0001	5128	1.000
	Bay centre	-0.34	0.23	-0.34	-0.77	0.16	0.1458	6667	1.000
	Flower density	0.03	0.01	0.03	0.01	0.05	0.0115	4364	1.000
	Sigma (field)	0.63	0.13	0.62	0.40	0.91	-	1072	1.004
	Phi	0.70	0.11	0.69	0.50	0.93	-	3097	0.999
	Theta (ZI)	0.25	0.04	0.25	0.15	0.33	-	2383	1.000
Pollen count	Intercept	2.49	0.19	2.50	2.12	2.85	<0.0001	1141	1.002
	HB visits	0.05	0.04	0.05	-0.03	0.12	0.195	2317	1.000
	LCB visits	0.15	0.06	0.14	0.02	0.26	0.0155	2239	1.000
	Bay centre	-0.55	0.13	-0.55	-0.80	-0.30	<0.0001	2540	1.000
	HB distance	-0.17	0.04	-0.17	-0.25	-0.09	<0.0001	2265	1.003
	LCB distance	-0.35	0.14	-0.35	-0.63	-0.07	0.0164	2513	1.000
	Flower density	-0.03	0.02	-0.03	-0.06	0.01	0.1284	2058	1.001
	Sigma (field)	0.86	0.13	0.85	0.64	1.15	-	2978	1.000
	Sigma (plot)	0.65	0.06	0.65	0.53	0.78	-	1142	1.003
	Phi	0.82	0.04	0.82	0.74	0.90	-	5125	0.999
Flowers per plant	Intercept	3.05	0.04	3.05	2.96	3.14	<0.0001	3283	0.999
	Plant size	0.93	0.01	0.93	0.90	0.95	<0.0001	3712	0.999
	Bay centre	0.09	0.02	0.09	0.06	0.12	<0.0001	6790	0.999
	Pods per plant	-0.14	0.01	-0.14	-0.17	-0.12	<0.0001	5503	0.999
	Sigma (field)	0.08	0.01	0.08	0.06	0.11	-	2649	1.000
	Intercept (Phi)	2.24	0.32	2.24	1.62	2.87	-	2875	1.001
	Plant size (Phi)	0.38	0.10	0.39	0.19	0.57	-	2871	1.001
Pods per plant	Intercept	0.18	0.12	0.18	-0.05	0.40	0.1211	2261	1.000
	Pollen count	0.12	0.04	0.12	0.03	0.20	0.0074	915	1.004
	Plant size	0.19	0.03	0.19	0.14	0.24	<0.0001	4570	1.000
	Bay centre	-0.21	0.06	-0.21	-0.33	-0.10	0.0003	1886	1.000
	HB distance	-0.12	0.02	-0.12	-0.15	-0.08	<0.0001	1929	1.002
	LCB distance	-0.21	0.06	-0.21	-0.34	-0.10	0.0005	2637	1.000
	Flower density	0.00	0.01	0.00	-0.02	0.01	0.5456	2998	1.000
	Sigma (plot)	0.30	0.03	0.30	0.25	0.35	-	1367	1.004
	Sigma (field)	0.37	0.06	0.36	0.27	0.49	-	2905	1.000
	Intercept (Phi)	3.07	0.38	3.05	2.36	3.85	-	2268	1.004
	Plant size (Phi)	0.22	0.12	0.22	-0.01	0.44	-	2538	1.003
Seeds per pod	Intercept	25.38	2.29	25.48	20.64	29.38	<0.0001	1651	1.001
	Pollen count	1.21	0.32	1.20	0.61	1.87	0.0001	1046	1.002
	Plant size	4.76	0.78	4.76	3.24	6.23	<0.0001	1752	1.001
	Bay centre	-0.50	0.45	-0.50	-1.39	0.38	0.2639	5315	0.999
	HB distance	-0.02	0.12	-0.01	-0.25	0.21	0.881	3007	1.000
	Flower density	-0.11	0.06	-0.11	-0.23	0.00	0.0451	2596	1.000
	Pods per plant	1.65	0.35	1.65	0.94	2.34	<0.0001	2485	1.001
	Flowers per plant	-4.26	0.74	-4.27	-5.60	-2.73	<0.0001	1571	1.001
	Sigma	4.25	0.16	4.25	3.92	4.56	-	2951	1.000
	Sigma (field)	1.97	0.32	1.95	1.41	2.68	-	1767	1.002
	Lambda	1.67	1.00	1.39	0.60	4.28	-	2448	0.999
Seed size	Intercept	1.17	0.50	1.18	0.21	2.16	0.0189	2004	1.001
	Pollen count	-0.02	0.05	-0.02	-0.12	0.08	0.7002	2345	1.000
	Seeds per pod	-0.01	0.01	-0.01	-0.03	0.00	0.0765	3719	0.999
	Plant size	0.23	0.06	0.23	0.12	0.34	<0.0001	3607	1.000
	Plant density	0.41	0.11	0.41	0.20	0.63	0.0002	1988	1.000
	LCB distance	0.07	0.07	0.07	-0.06	0.22	0.3146	4466	1.000
	Sigma	0.75	0.03	0.75	0.69	0.81	-	3533	1.000
	Sigma (field)	0.36	0.06	0.35	0.25	0.49	-	2060	1.000
	Lambda	4.44	1.22	4.23	2.70	7.42	-	2125	1.000

Total yield models

Formulas for total yield models are using `lmer`-style R formulas. Predicted yield for each plant was calculated as: pods per plant $\times$ seeds per pod $\times$ weight per seed. Terms on right side of $\sim$ indicate fixed effects, while terms in brackets indicate random intercepts (and slopes).

$$\text{Total Yield} \sim log(\text{Predicted Yield}) + (log(\text{Predicted Yield})|\text{Field})+$$
$$(log(\text{Predicted Yield})|\text{Plot}), \text{distribution} = \text{log-normal}$$

Table S6: Summary of parameters for total yield models

Field Type	Parameter	Mean	SD	Z	median	Min	Max	p-value	N_{eff}	$\hat{R}$
	Intercept	-0.318	0.042	-7.62	-0.317	-0.397	-0.236	0.000	982	1.000
	Predicted Yield	1.007	0.018	55.64	1.007	0.972	1.044	0.000	1044	1.000
	Sigma (field intercept)	0.119	0.041	2.89	0.112	0.051	0.216	0.004	292	1.020
	Sigma (field slope)	0.026	0.019	1.37	0.023	0.001	0.072	0.170	295	1.013
Commodity	Sigma (plot intercept)	0.389	0.037	10.47	0.388	0.318	0.465	0.000	506	1.005
	Sigma (plot slope)	0.169	0.017	9.86	0.168	0.135	0.204	0.000	556	1.006
	Sigma	0.253	0.008	33.30	0.253	0.239	0.268	0.000	2002	1.000
	Correlation (Int:Slope field)	-0.400	0.448	-0.89	-0.525	-0.944	0.626	0.372	660	1.005
	Correlation (Int:Slope plot)	-0.989	0.007	-140.44	-0.990	-0.998	-0.972	0.000	936	1.005
	Intercept	-0.210	0.072	-2.90	-0.210	-0.348	-0.066	0.004	738	1.007
	Predicted Yield	0.856	0.025	34.62	0.856	0.806	0.904	0.000	672	1.008
	Sigma (field intercept)	0.040	0.018	2.28	0.038	0.011	0.079	0.022	226	1.005
	Sigma (field slope)	0.010	0.007	1.56	0.009	0.001	0.023	0.118	217	1.005
Seed	Sigma (plot intercept)	0.097	0.013	7.26	0.097	0.072	0.124	0.000	470	1.005
	Sigma (plot slope)	0.032	0.005	5.83	0.032	0.021	0.043	0.000	379	1.006
	Sigma	0.289	0.012	23.59	0.289	0.266	0.314	0.000	1218	1.001
	Correlation (Int:Slope field)	-0.470	0.436	-1.08	-0.616	-0.946	0.603	0.281	457	1.005
	Correlation (Int:Slope plot)	-0.902	0.038	-23.81	-0.909	-0.954	-0.808	0.000	486	1.007