

# DRAFT

## Diagnostic Protocol *Conotrachelus nenuphar*

### Pest Information

The plum curculio, *Conotrachelus nenuphar* Herbst (Coleoptera: Curculionidae), is a native fruit pest in the United States and Canada. Its geographic range extends from the eastern Rockies to the Atlantic and from southern Canada to the Gulf Coast. The plum curculio is an agricultural pest of orchard fruit—especially plums, peaches, nectarines, apples, cherries, and blueberries (Quaintance & Jenne 1912; Chapman 1938).

The adults feed on, and the larvae develop within, the fruit of these plants. Crop damage comes from oviposition sites and from adult and larval feeding on fruits. The adult feeding punctures often deform the fruit and open up the skin to further damage by other insect pests or fungal attacks. The developing larvae consume the flesh of the fruit and cause the fruit to drop from the tree before ripening. Both forms of damage are problems for fresh market fruits, and premature drop prevents the fruit from being used as a processed food item.

The plum curculio is endemic and native to North America. Native hosts include hawthorn trees (*Crataegus spp.*), crabapple trees (*Malus spp.*), and wild plum (*Prunus spp.*). The adult beetles will feed on the fruits of a great many kinds of rosaceous and ericaceous plants: plums, apples, peaches, nectarines, cherries, apricots, pears, strawberries, quince, blueberries, haws, huckleberry, as well as grape (Vitaceae), gooseberry and currant (both Grossulariaceae), persimmon (Ebenaceae), and if given the opportunity will even feed on tropical fruits not available within its current range (Quaintance & Jenne 1912; Chapman 1938; Hallman & Gould 2004). The beetle discriminates among these potential food sources and prefers stone and pome fruits—especially plums, peaches, cherries, apricots, apples, and pears (Jenkins, Cottrell, et al. 2006b; Leskey & Wright 2007). Females will oviposit in these fruits, and larvae can successfully develop in any of them. Larvae have been known to develop in fungal black knot (*Plowrightia morbosa*) on cherry trees (Quaintance & Jenne 1912; Jenkins, Cottrell, et al. 2006b).

The geographic range of the plum curculio is limited to the United States and Canada east of the Rocky Mountains. There are no established populations of plum curculio in the western United States, except for an infestation in Box Elder County, Utah dating to the 1980's, primarily of fruit trees in home yards and wild plums (Alston et al. 2005). There are no known established populations of the plum curculio outside of North America.

There are two phenological strains of plum curculio, a northern strain and a southern strain. The number of generations per year is a defining characteristic

of the strains. The northern strain plum curculio must diapause to become reproductively mature (obligate diapause) and has a single brood per year, with adults entering diapause in the late summer and early fall before female reproductive features have developed. The southern strain plum curculio often has only one brood per year but has the ability to develop reproductively and have a second or even in rare cases a third generation in a single season (facultative diapause) (Smith & Salkeld 1964). For this reason, summer and fall harvested fruit may have viable larvae in them in the southeastern United States, though this is rare.



Figure 1: Plum curculio adult

## Taxonomic Information

Name:

*Conotrachelus nenuphar* Herbst, 1797

Synonyms:

*Curculio nenuphar* Herbst, 1797

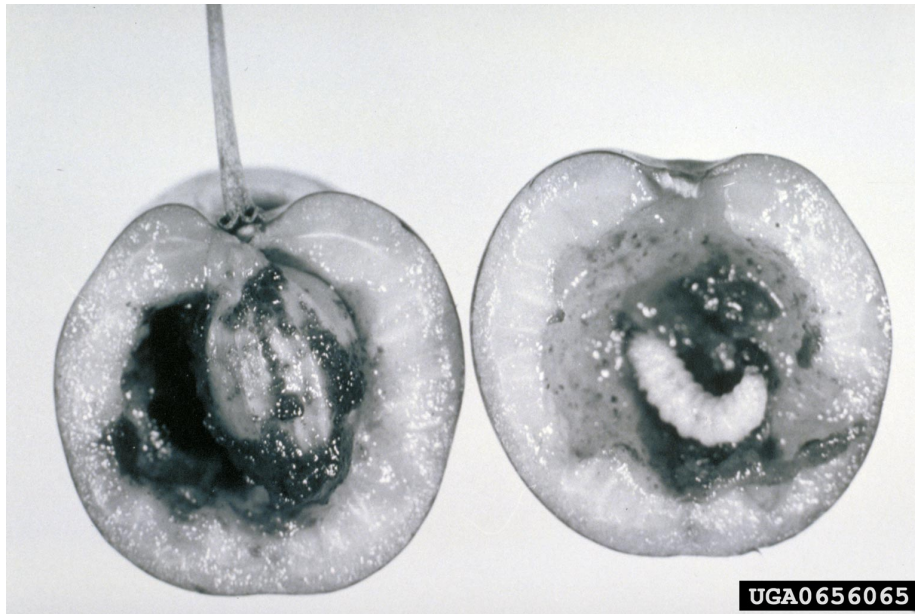


Figure 2: Plum curculio larva in cherry fruit

*Conotrachelus nenuphar* (Hbst.) Leconte and Horn, 1876

*Rhynchaenus argula* Fabricius, 1801

*Cryptorhynchus argula* (Fab.) Say, 1831

*Rhynchaenus cerasi* Peck, 1819

Taxonomic position:

Insecta, Coleoptera, Curculionidae, Molytinae

Common name:

Plum curculio

## Detection

The larvae are the life stage most likely to be transported in late season fruit, especially from the southern extent of its range. Pupae may be transported in soil along with tree seedlings or transplants. Adults may be transported in nursery material, rootstocks, branches, flowers, and fresh packed fruit.

***C. nenuphar* is commonly found on the follow plant parts and plant-associated media depending on life stage:**

- Eggs: immature fruit tissue, mature fruit tissue (if from southern range)
- Larvae: immature fruit tissue, mature fruit tissue
- Pupae: in the soil
- Adults: on the leaves, branches, flowers, and fruits

### **Signs of eggs and larvae:**

In immature fruit, a small crescent-shaped cut and scar is indicative of oviposit. To lay an egg, a female must puncture the skin of the developing fruit with her mandible (on the distal end of the rostrum) and excavate a small, shallow cavity. A single egg is deposited in the center of this cavity. A single female may lay multiple eggs on a single fruit.

In mature fruit, the oviposition scar becomes more diffuse and takes on a corky appearance. These can look like mottled fans with a small scar at the base of the fan.

### **Signs of adult action:**

Adults feed on fruit. In immature fruit, punctures look circular (not crescent shaped) and extend up to 3mm into the fruit. On mature fruit, punctures also appear circular and can tend to cluster around the calyx of the fruit.

### **Methods of insect recovery from plants and plant products:**

Eggs and Larvae: Eggs can be detected by observing fresh fruit for signs of oviposition and inspection of plant tissue beneath the scar. Larvae can be recovered from fruit by splitting the fruit and looking for signs of larval feeding and for larvae. Larvae will only exit the fruit after fruit drop, and so any fruit still on the stem may yield live larvae.

Pupae: Pupae can be recovered from soil by sifting and hand searching with visual inspection of the soil associated with any plant product. Pupae tend to be found within 20mm of the soil surface. Pupae can be reared also from soil, following established guidelines (citation needed).

Adults: Adults can be found by visual inspection of any plant part, including flowers, leaves, branches, and trunks. Adults are well camouflaged and will

tend to appear as a small piece of bark. Plum curculio exhibit thanatosis, or tonic immobility, when disturbed. Collection in the field is done by jarring or disturbing the medium on which the adults are found (branches and trunks) over a white sheet, followed by visual inspection for immobile adults. Immobile adults fold their legs under their body and bend the rostrum under the prothorax, and appear as an oblong shape.



Figure 3: An adult plum curculio exhibiting mimicry via tonic immobility

### Similar signs due to other insect sources:

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## Identification

Identification of *Conotrachelus nenuphar* by morphological examination is restricted to adult specimens because there are no adequate keys for the identification of eggs, larvae or pupae. A guide to identification of adult plum curculio is given below.

Larval and pupal life stages are especially a risk for misidentification because of the lack of reliable identification diagnostics or keys for them. Molecular methods can be applied to all life stages including the immature stages for which morphological identification to species is not possible and to specimen fragments. Molecular approaches can also be used to narrow the identification of the phenological strain and the geographic region of origin for the sample.

## Preparation of adult beetles for microscopic examination

Adult weevils in the genus *Conotrachelus* are generally less than 9mm and may be examined for morphological identification under 50x to 600x magnification. Most diagnostic characters can be observed at this level of magnification. For routine identification, dissection of genitalia is usually not necessary.

The typical size for adult *Conotrachelus nenuphar* specimens is between 4mm and 6mm, allowing for pin mounting directly through the right elytron. There are several important diagnostic characters on the legs, so spreading the legs while mounting is recommended.

## Morphological identification of adult weevils

The weevil family, Curculionidae, is very large with more than 60,000 described species (Marvaldi et al, Oberprieler). The best external morphological characters for the recognition of the weevils are associated with their rostrum (snout or beak) although some weevils have a very short rostrum and some have none (especially in the Scolytinae and Platypodinae). The length of the rostrum, its curvature, or lack of curvature, and the degree of punctation or sculpturing, and/or the type and density of vestiture all are used in classification. Another set of diagnostic characters are those of the antennae. The first article (the scape) is elongate and inserted away from the base, usually near the middle and at times near the apex. Often it rests in a lateral groove (scrobe), and is directed in many ways, e.g., dorsally, ventrally, and has various lengths and shapes. The number of funicular articles varies from 4 to 8 articles and the last three antennal articles normally form a compact club.

[Table to come...]

## Morphological identification of adult *Conotrachelus*

*Conotrachelus* is a New World beetle genus with approximately 1,200 named species (Dejean 1837; O'Brien & Wibmer 1982). The majority of species diversity is concentrated in South America and there are many species found only in South America. The *Conotrachelus* diversity found in the United States and Canada (where *Conotrachelus nenuphar* is endemic) is limited to approximately 63 of

1,200 described species. Of these, roughly 46 *Conotrachelus* species are broadly sympatric with *C. nenuphar* being found in the eastern portion of North America, here defined as north of Mexico and east of the Rocky Mountains. Of those *Conotrachelus* species found in the same geographic regions as *C. nenuphar*, none are known to use commercial fruit trees as hosts. The potential for confusion with other fruit pests is greater than with congeners. Of critical importance in diagnosing *Conotrachelus* species is the postmedian elytral band. This is a region in the middle of the elytra and there are diagnostic differences between groups of species.

Table of Diagnostic Characters for the genus:

Body Part	Characteristic
Antennae	Length of scape, number of funicular articles, club.
Rostrum	Length, shape, sculpture, vestiture.

### **Morphological identification of adult *Conotrachelus nenuphar***

Table of Diagnostic Characters for the species:

Body Part	Characteristic
Appendages	Tarsal claws divergent, not close together.
Prothorax	No median ridge or furrow.
Elytra	Two distinct costae (or crests), one on each elytron on interval 3. Region between and around
Thorax	The mesoscutellum is gently sloped, depressed and flat on the basal side and not prominent all
Elytra	costae, and patterns and color of vestiture
Venter	punctuation and vestiture
Legs	Femoral tooth and metaunci

### **Morphological identification of voltinic strains of adult *Conotrachelus nenuphar***

Smith and Salkeld (1964) dissected the maturing ovaries of northern and southern strain plum curculio adult females and laid out how the breeding behavior of the two strains was related to their diapause behavior. They established that northern strain females do not develop mature oocytes prior to diapause, and that southern strain females do. This was the first demonstration that the diapause behavior between the strains was different, with the northern strain required to diapause to develop oocytes. To this day, this is the only reliable method to distinguish the strains and their diapause behavior.

## Molecular assays for identifying *Conotrachelus nenuphar*

There are several methods available for molecular diagnostic identification of the plum curculio. The *COI* gene is a useful marker for diagnosing *Conotrachelus nenuphar* from its congeners as well as identifying certain regional variants within the species. The northern and southern regions are readily diagnosable from each other, and within the southern populations, genetic variants found west of the Mississippi and in the mid-Atlantic are also diagnosable from the broader southern distribution.

### DNA isolation

Nucleic acids can be isolated using any standard technique. In the case of adults, a fragment of the whole adult specimen can be used in lysis to preserve morphological characters for independent diagnosis. A single hind leg from fresh tissue yields sufficient DNA concentration for PCRs. To semi-destructively sample a specimen for DNA, the head and prothorax of each individual can be separated from the body and subjected DNA extraction. This protocol allows for digestion of the soft internal tissue of the head and prothorax for isolation of nucleic acids while preserving the hard sclerotized external anatomy. After lysis of internal soft tissue, the head and prothorax is reattached to each specimen.

The Qiagen DNeasy Blood & Tissue Kit (Qiagen, Inc) nucleic acid extraction technique has been used successfully on fresh whole specimens, single legs from fresh specimens, and head and prothorax from museum specimens (with modification, see Crane 2013).

### 3' *COI* sequence-based identification of *Conotrachelus nenuphar* and its geographic strains

Using the 3' *COI* gene fragment, one is able to diagnose regional variation within *Conotrachelus nenuphar*. Two geographically distinct mitochondrial groups are resolvable with purely diagnostic single nucleotide variants and enable the diagnosis to identify regional source populations in the case of mid-Atlantic (Sb haplotypes) and Midwestern (Sa haplotypes).

Target Locus: 3' end of the *COI* mitochondrial gene

Expected Amplicon Size: 826bp

Primers:

Primer Name	Marker	Sequence (5' - 3')	Direction
C1-J-2183 Jerry	COI	CAACATTTATTTTGATTTTTTGG	F
TL2-N3014 Pat	COI	TCCAATGCACTAATCTGCCATATTA	R



PCR Conditions: Setup for a 25 µl PCR reaction: Molecular-grade water (volume to 25 µl), 10mM dNTPs (0.5 µl), 10µM Forward primer (0.5 µl), 10µM Reverse primer (0.5 µl), isolated DNA (0.5 µl), *Taq* DNA polymerase (0.1 µl).

Thermocycler program: initial denaturation at 95°C (30 seconds); 45 cycles of amplification of 95°C (15 seconds), 56°C (30 seconds), 68°C (60 seconds); final extension at 72°C (5 minutes).

DNA sequencing was done using the BigDye® Terminator v3.1 Cycle Sequencing kit (Applied Biosystems, Inc.) on an ABI Prism™ 3730 DNA Analyzer.

The DNA sequences produced by this method can be analyzed via multiple sequence alignment and subsequent character analysis. Figure XX provides species-level molecular diagnostic characters for identification of closely-related *Conotrachelus* species. Reference sequences can be obtained from GenBank.

Taxa/Position	1	7	22	55	59	94	124	127	169	172	220	232	253	271	286	292	301	304	328	581	607	627	628	639	659	667	668	670	671	718	724	733	741	760	772	
affinis/hicoriae	C	A	A	C	G	A	T	T	A	A	T	C	A	T	C	A	T	A	T	T	A	T	A	T	T	C	C	A	A	G	T	A	A	T	T	A
albicinctus	T	A	A	C	G	A	T	T	A	A	T	C	A	T	T	A	T	A	T	T	A	T	A	C	C	C	C	A	A	T	A	A	T	A	T	A
aratus/schoofi	T	A	A	C	G	T	T	T	A	A	T	C	A	T	T	A	T	A	T	T	A	T	A	T	T	T	C	T	A	T	A	A	T	T	C	A
buchanani	T	G	A	T	G	A	T	T	A	T	T	A	A	Y	T	A	T	A	T	T	A	T	A	T	T	C	C	A	A	T	A	A	T	T	C	A
corni	T	A	A	C	G	A	T	T	A	T	-	-	-	-	-	-	-	-	-	-	T	A	C	A	T	T	C	C	A	A	T	G	G	T	A	
downiei	T	G	A	T	G	A	T	T	A	T	T	A	A	C	T	A	T	A	T	T	A	T	A	T	T	C	C	A	A	T	A	A	T	T	C	A
elegans	T	A	A	C	G	A	T	T	A	A	T	C	A	T	T	T	A	T	A	T	A	T	A	T	T	C	T	A	A	T	A	A	T	T	T	A
iowensis	T	A	A	C	G	A	T	T	A	A	T	C	G	G	T	T	G	C	A	T	T	A	T	A	T	T	C	C	A	A	T	A	A	T	T	A
juglandis	T	A	A	T	G	A	T	T	A	A	T	T	A	T	T	A	T	A	T	G	T	T	A	T	G	T	T	C	C	A	A	T	A	A	T	A
nenuphar_N	T	A	A	C	A	A	T	T	C	A	T	C	A	T	T	A	T	A	T	T	A	T	A	T	T	C	C	A	A	A	A	T	A	T	T	A
nenuphar_S	T	A	A	G	G	A	T	T	A	C	T	C	A	A	T	A	T	A	T	T	G	T	A	T	T	Y	C	A	A	T	A	A	T	A	T	A
retentus	T	A	G	T	G	A	T	T	A	T	C	A	T	T	A	A	A	C	C	A	T	A	T	T	C	C	A	A	T	A	A	T	A	T	A	A
seniculus	A	A	A	T	G	A	C	A	T	T	T	A	A	C	T	A	A	A	A	T	T	T	T	A	T	T	T	T	A	T	T	A	A	T	A	A

Figure 4: 3' COI *Conotrachelus nenuphar* species-level diagnostic characters

## 5' COI sequence-based identification of *Conotrachelus nenuphar* and some congeneric species

Target Locus: 5' end of the COI mitochondrial gene

Expected Amplicon Size: 658bp

Primers: (Folmer et al. 1994).

Primer Name	Marker	Sequence (5' - 3')	Direction
LCO1490	COI	GGTCAACAAATCATAAAGATATTGG	F
HCO2198	COI	TAAACTTCAGGGTGACCAAAAAATCA	R

PCR Conditions: Setup for a 25 µl PCR reaction: Molecular-grade water (volume to 25 µl), 10mM dNTPs (0.5 µl), 10µM Forward primer (0.5 µl), 10µM Reverse primer (0.5 µl), isolated DNA (0.5 µl), *Taq* DNA polymerase (0.1 µl).

Thermocycler program: initial denaturation at 95°C (30 seconds); 45 cycles of amplification of 95°C (15 seconds), 56°C (30 seconds), 68°C (60 seconds); final extension at 72°C (5 minutes).

DNA sequencing was done using the BigDye® Terminator v3.1 Cycle Sequencing kit (Applied Biosystems, Inc.) on an ABI Prism™ 3730 DNA Analyzer.

The DNA sequences produced by this method can be analyzed and a species designation made using the Barcode of Life database (Project code NAPCB).

## **RFLP**

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