

# Package ‘CNVseq’

December 6, 2024

**Version** 1.0.0  
**Date** 2024-12-05  
**Type** Package  
**Title** CNVseq: A tool for CNV-seq data calling variants  
**Author** Zhan-Ni Chen  
**Maintainer** Zhan-Ni Chen <sanadamakomi@gmail.com>  
**Depends** R (>= 3.4.0)

**Imports** utils,  
S4Vectors,  
IRanges,  
GenomicRanges,  
Biostrings,  
bitops,  
GenomeInfoDb,  
Rsamtools,  
GenomicAlignments,  
DNACopy,  
data.table

**Description** Detect CNVs in CNV-seq data. By default, it identifies DUP and DEL larger than 100 kb with a mosaicism level above 0.5, and CNVs larger than 5 Mb with a mosaicism level above 0.1. The mosaicism level is adjustable.

**License** Artistic-2.0  
**Encoding** UTF-8

**URL** <https://github.com/sanadamakomi/CNVseq>  
**BugReports** <https://github.com/sanadamakomi/CNVseq/issues>  
**LazyData** true  
**RoxygenNote** 7.3.2

## Contents

batch_call_cnv . . . . .	2
batch_cov . . . . .	3
batch_gender . . . . .	3
batch_merge . . . . .	4

batch_split . . . . .	4
bed_to_gr . . . . .	5
call_cnv . . . . .	5
check_bam . . . . .	5
count_reads_of_region . . . . .	6
count_reads_of_region_multicore . . . . .	6
creat_chunk . . . . .	7
do_cbs . . . . .	7
get_hg19_seqinfo . . . . .	8
get_id . . . . .	8
get_wgs_bin . . . . .	8
make_vcf_format . . . . .	9
make_vcf_header . . . . .	9
make_vcf_info . . . . .	9
make_vcf_matrix . . . . .	10
merger_cov_files . . . . .	10
merge_bin_df . . . . .	11
merge_result_files . . . . .	11
merge_segment . . . . .	11
output_vcf . . . . .	12
ploidy_to_cn . . . . .	12
read_db_file . . . . .	12
write_cov_file . . . . .	13

## Index 14

---

batch_call_cnv	<i>Call CNVs from read count files</i>
----------------	--

---

### Description

Call CNVs from read count files

### Usage

```
batch_call_cnv(
  cnm_path,
  cnn_paths,
  cnn_dirs,
  out_dir,
  test_id,
  ref_ids,
  by_gender = FALSE,
  fraction = c(0.5, 0.5)
)
```

### Arguments

cnm_path	Path of merged CNN file.
cnn_paths	Path of CNN files, separated by comma(,).
cnn_dirs	Path of CNN directory, separated by comma(,).

out_dir	Output directory path.
test_id	Sample id to call CNV.
ref_ids	Reference ids to create a baseline, separated by comma(,).
by_gender	A bool value to call by gender.
fraction	Fraction of CNV >=100Kb and >=5Mb, default:c(0.5, 0.5).

---

batch_cov	<i>Calculate read count in bams</i>
-----------	-------------------------------------

---

### Description

Calculate read count in bams

### Usage

```
batch_cov(bam_dirs, bam_paths, bed_path, out_dir, thread = 4, mapq.filter = 0)
```

### Arguments

bam_dirs	Path of BAM directory, separated by comma(,).
bam_paths	Path of BAM file, separated by comma(,).
bed_path	Path of BED file.
out_dir	A character string of directory to output coverage files.
thread	An integer providing the number of thread, default: 4.
mapq.filter	A non-negative integer specifying the minimum mapping quality to include. BAM reads with mapping qualities less than mapqFilter are discarded, default: 0.

---

batch_gender	<i>Calculate gender and total read from a coverage file.</i>
--------------	--

---

### Description

Calculate gender and total read from a coverage file.

### Usage

```
batch_gender(cnn_paths, cnn_dirs, out_dir)
```

### Arguments

cnn_paths	Path of CNN files, separated by comma(,).
cnn_dirs	Path of CNN directory, separated by comma(,).
out_dir	Output directory path.

---

batch_merge	<i>Merge CNN, VCF, CNR, CNS file.</i>
-------------	---------------------------------------

---

### Description

Input `cnn_paths` or `cnn_dirs`, it will merge multiple samples' read count result files(.cnn) and create a merged file. Input `auto_file` and `sex_file`, it will merge autosome and sex chromosome result, the VCF, CNR, CNS file can be input.

### Usage

```
batch_merge(cnn_paths, cnn_dirs, auto_file, sex_file, out_path)
```

### Arguments

<code>cnn_paths</code>	Path of CNN files, separated by comma(,).
<code>cnn_dirs</code>	Path of CNN directory, separated by comma(,).
<code>auto_file</code>	path of VCF, CNR, CNS file.
<code>sex_file</code>	path of VCF, CNR, CNS file.
<code>out_path</code>	Output file path.

---

batch_split	<i>Split wgs region into bins</i>
-------------	-----------------------------------

---

### Description

Split wgs region into bins

### Usage

```
batch_split(path, bin = 10000, access_bed = "")
```

### Arguments

<code>path</code>	Path of output, a BED file.
<code>bin</code>	An integer of bin size, default: 1E4.
<code>access_bed</code>	BED file include sequence-accessible region. If NULL it will use whole genome region.

---

bed_to_gr	<i>Read BED file and output grange</i>
-----------	--

---

**Description**

Read BED file and output grange

**Usage**

```
bed_to_gr(file)
```

**Arguments**

file	Path of BED file.
------	-------------------

---

call_cnv	<i>Calling CNV</i>
----------	--------------------

---

**Description**

Calling CNV

**Usage**

```
call_cnv(data, s_id, is_male = TRUE, fraction = c(0.5, 0.5))
```

**Arguments**

data	A data frame of log2ratio to do CBS. It must has three column: chromosome, end, log2.
s_id	A charactor string of sample id.
is_male	A bool value, TRUE when the gender is male. change-points, default: 0.05.
fraction	Fraction of CNV >=100Kb and >=5Mb, default:c(0.5, 0.5).

---

check_bam	<i>Check BAM file.</i>
-----------	------------------------

---

**Description**

It will stop if BAM file is illegal.

**Usage**

```
check_bam(x)
```

**Arguments**

x	A character string or vector of BAM File path.
---	--

---

count\_reads\_of\_region    *Calculating read count in region*

---

### Description

Calculating read count in region

### Usage

```
count_reads_of_region(region, bamPath, mapq.filter = 0, minoverlap = 75L)
```

### Arguments

region	A grange object of region to extract reads in BAM file.
bamPath	A character string of the BAM path.
mapq.filter	A non-negative integer specifying the minimum mapping quality to include. BAM reads with mapping qualities less than mapqFilter are discarded.
minoverlap	Minimum overlap size for region and reads, default: 75L.

---

count\_reads\_of\_region\_multicore  
                                   *Calculating read count in multiple thread*

---

### Description

Calculating read count in multiple thread

### Usage

```
count_reads_of_region_multicore(
  region,
  bamPath,
  thread,
  batch,
  tmpDir = NULL,
  mapq.filter = 0,
  minoverlap = 75L
)
```

### Arguments

region	A grange object of region to extract reads in BAM file.
bamPath	A character string of the BAM path.
thread	An integer providing the number of thread.
batch	An integer giving how many GRanges are performed in a batch.
tmpDir	A character string of directory to output coverage files (<sampleid>.cnn). Default is the current folder.

mapq.filter	A non-negative integer specifying the minimum mapping quality to include. BAM reads with mapping qualities less than mapqFilter are discarded.
minoverlap	Minimum overlap size for region and reads, default: 75L.

---

creat_chunk	<i>Create grange into chunk</i>
-------------	---------------------------------

---

## Description

Create grange into chunk

## Usage

```
creat_chunk(gr, bin = 10000)
```

## Arguments

gr	A grange object.
bin	Bin size, default: 1E4.

---

do_cbs	<i>Circular Binary Segmentation</i>
--------	-------------------------------------

---

## Description

Circular Binary Segmentation

## Usage

```
do_cbs(data, s_id, alpha = 0.05, min.width = 2)
```

## Arguments

data	A data frame of log2ratio to do CBS. It must has three column: chromosome, end, log2.
s_id	A charactor string of sample id.
alpha	A numeric value of significance levels for the test to accept change-points, default: 0.05.
min.width	An integer value of the minimum number of markers for a changed segment, default: 2.

---

get_hg19_seqinfo	<i>Hg19 seqinfo</i>
------------------	---------------------

---

**Description**

Hg19 seqinfo

**Usage**

```
get_hg19_seqinfo()
```

---



---

get_id	<i>Extract sample id from file path</i>
--------	---

---

**Description**

Extract sample id from file path

**Usage**

```
get_id(file, ptn = NULL)
```

**Arguments**

file	Path of file.
ptn	A charactor for spliting string. By default NULL, it will split string by '.'

---



---

get_wgs_bin	<i>Split wgs region into bins</i>
-------------	-----------------------------------

---

**Description**

Split wgs region into bins

**Usage**

```
get_wgs_bin(bin = 10000, access_bed = "")
```

**Arguments**

bin	An integer of bin size, default: 1E4.
access_bed	BED file include sequence-accessible region. If NULL it will use whole genome region.



---

make_vcf_format	Create VCF FORMAT column
-----------------	--------------------------

---

**Description**

Create VCF FORMAT column

**Usage**

```
make_vcf_format(x)
```

**Arguments**

x	A data frame of CNV result.
---	-----------------------------

---

make_vcf_header	Create VCF header
-----------------	-------------------

---

**Description**

Create VCF header

**Usage**

```
make_vcf_header()
```

---

make_vcf_info	Create VCF INFO column
---------------	------------------------

---

**Description**

Create VCF INFO column

**Usage**

```
make_vcf_info(x)
```

**Arguments**

x	A data frame of CNV result.
---	-----------------------------

---

make_vcf_matrix	<i>Create VCF matrix</i>
-----------------	--------------------------

---

**Description**

Create VCF matrix

**Usage**

```
make_vcf_matrix(x)
```

**Arguments**

x	A data frame of CNV result.
---	-----------------------------

---

merger_cov_files	<i>Merge coverage files.</i>
------------------	------------------------------

---

**Description**

Align and merge coverage files (<filename>.cnn) with chromosome, start and end position. Four required fields in a coverage file are chromosome name, start and end position, depth of coverage.

**Usage**

```
merger_cov_files(files, path = NULL)
```

**Arguments**

files	A character vector contains several coverage files path.
path	Path to write to.

**Value**

A data frame, of which columns are chromosome, start position, end position, and depths in input coverage files.

---

merge_bin_df	<i>Merge log2ratio with a specific interval of bins</i>
--------------	---

---

**Description**

Merge log2ratio with a specific interval of bins

**Usage**

```
merge_bin_df(data, group_size = 50)
```

**Arguments**

data	A data frame of cnr file.
group_size	A interval to merge, default: 50.

---

merge_result_files	<i>Merge autosome and sex chromosome result</i>
--------------------	---

---

**Description**

Merge autosome and sex chromosome result

**Usage**

```
merge_result_files(auto_file, sex_file, out_path)
```

**Arguments**

auto_file	path of VCF, CNR, CNS file.
sex_file	path of VCF, CNR, CNS file.
out_path	path of output file.

---

merge_segment	<i>Filtering transcripts</i>
---------------	------------------------------

---

**Description**

Filtering transcripts

**Usage**

```
merge_segment(x)
```

**Arguments**

x	A data.frame of CNV calling result.
---	-------------------------------------

---

output_vcf	<i>Output CNV calling result to VCF file</i>
------------	--

---

**Description**

Output CNV calling result to VCF file

**Usage**

```
output_vcf(dat, path)
```

**Arguments**

dat	A data.frame of CNV calling result.
path	Path of VCF file.

---

ploidy_to_cn	<i>Count copy numer with different mosaicism level</i>
--------------	--

---

**Description**

Count copy numer with different mosaicism level

**Usage**

```
ploidy_to_cn(f, ploidy, cn)
```

**Arguments**

f	Fraction of variants.
ploidy	the ploidy of chromosome.
cn	Copy number.

---

read_db_file	<i>Read annovation database file and return granges</i>
--------------	---

---

**Description**

Read annovation database file and return granges

**Usage**

```
read_db_file(prefix, db_path, p = 1, header = FALSE, file_encode = "UTF-8")
```

**Arguments**

prefix	A prefix of database file, e.g. centromere_telomere for hg19_centromere_telomere.txt.
db_path	Path of database directory.
p	A integer for the column index of chromosome, default: 1.
header	A bool for the header of database file, default: FALSE.
file_encode	The file encode of database file, default: UTF-8.

---

write_cov_file	<i>Write read count result file</i>
----------------	-------------------------------------

---

**Description**

Write read count result file

**Usage**

```
write_cov_file(gr, path)
```

**Arguments**

gr	A GRange with a column named rc.
path	Path to write to.

# Index

[batch\\_call\\_cnv](#), [2](#)  
[batch\\_cov](#), [3](#)  
[batch\\_gender](#), [3](#)  
[batch\\_merge](#), [4](#)  
[batch\\_split](#), [4](#)  
[bed\\_to\\_gr](#), [5](#)  
  
[call\\_cnv](#), [5](#)  
[check\\_bam](#), [5](#)  
[count\\_reads\\_of\\_region](#), [6](#)  
[count\\_reads\\_of\\_region\\_multicore](#), [6](#)  
[creat\\_chunk](#), [7](#)  
  
[do\\_cbs](#), [7](#)  
  
[get\\_hg19\\_seqinfo](#), [8](#)  
[get\\_id](#), [8](#)  
[get\\_wgs\\_bin](#), [8](#)  
  
[make\\_vcf\\_format](#), [9](#)  
[make\\_vcf\\_header](#), [9](#)  
[make\\_vcf\\_info](#), [9](#)  
[make\\_vcf\\_matrix](#), [10](#)  
[merge\\_bin\\_df](#), [11](#)  
[merge\\_result\\_files](#), [11](#)  
[merge\\_segment](#), [11](#)  
[merger\\_cov\\_files](#), [10](#)  
  
[output\\_vcf](#), [12](#)  
  
[ploidy\\_to\\_cn](#), [12](#)  
  
[read\\_db\\_file](#), [12](#)  
  
[write\\_cov\\_file](#), [13](#)