

Homework Assignment

Protein Sequence Alignment

03-17-2023

Task A

```
choosebank("swissprot")
ac_1 <- "AOA1I9G1P5"
ac_2 <- "AOA1S0U2K5"

q1 <- query("q1", query = paste("AC=", ac_1, sep = ""))
q2 <- query("q2", query = paste("AC=", ac_2, sep = ""))

seq1 <- getSequence(q1$req[[1]])
seq2 <- getSequence(q2$req[[1]])

print(AAString(c2s(seq1)))

## 208-letter AAString object
## seq: MKLIVDSGHTGVNQLGGVFNAGRPLPDSTRQKIVDL...LAQLPPLTQANHLNKKDSGKLLWYSNYKGGWKELLI
print(AAString(c2s(seq2)))

## 133-letter AAString object
## seq: MLGDKKYSGHTGVNQLGGVFNAGRPLPDSTRQKIVD...EQPSIFAWEIRDKLLHEKVCSPDTIPSIHVGFISIHF
```

Task B

```
count_seq1 <- table(seq1)
count_seq2 <- table(seq2)

print(count_seq1)

## seq1
## A C D E F G H I K L M N P Q R S T V W Y
## 12 5 10 7 3 15 4 13 15 19 3 10 12 12 14 19 7 15 5 8
print(count_seq2)

## seq2
## A C D E F G H I K L M N P Q R S T V W Y
## 5 5 7 5 4 12 5 12 9 8 1 3 8 5 10 13 5 11 1 4

prop_seq1 <- proportions(count_seq1)
prop_seq2 <- proportions(count_seq2)

print(prop_seq1)
```

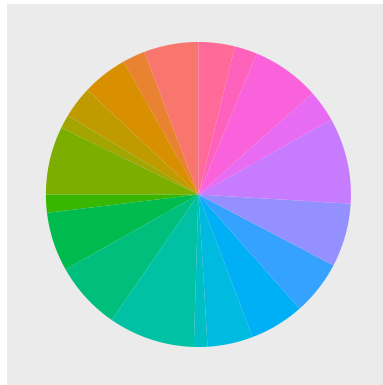
```
## seq1
##           A           C           D           E           F           G           H
## 0.05769231 0.02403846 0.04807692 0.03365385 0.01442308 0.07211538 0.01923077
##           I           K           L           M           N           P           Q
## 0.06250000 0.07211538 0.09134615 0.01442308 0.04807692 0.05769231 0.05769231
##           R           S           T           V           W           Y
## 0.06730769 0.09134615 0.03365385 0.07211538 0.02403846 0.03846154

print(prop_seq2)

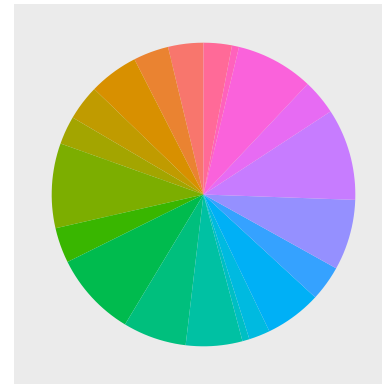
## seq2
##           A           C           D           E           F           G
## 0.037593985 0.037593985 0.052631579 0.037593985 0.030075188 0.090225564
##           H           I           K           L           M           N
## 0.037593985 0.090225564 0.067669173 0.060150376 0.007518797 0.022556391
##           P           Q           R           S           T           V
## 0.060150376 0.037593985 0.075187970 0.097744361 0.037593985 0.082706767
##           W           Y
## 0.007518797 0.030075188

theme_update(legend.key.size = unit(4, "mm"), legend.text = element_text(size = 6),
             axis.text = element_blank(), axis.ticks = element_blank(),
             panel.grid = element_blank())
df1 <- data.frame(slices = prop_seq1, amino_acids = paste(names(prop_seq1),
               "=", round(prop_seq1 * 100, 2), "%", sep = ""))
df2 <- data.frame(slices = prop_seq2, amino_acids = paste(names(prop_seq2),
               "=", round(prop_seq2 * 100, 2), "%", sep = ""))
plot1 <- ggplot(df1, aes(x = "", y = slices.Freq, fill = amino_acids)) +
  geom_bar(stat = "identity", width = 1) + coord_polar("y",
    start = 0) + ggtitle(ac_1) + xlab("") + ylab("") + guides(fill = guide_legend(title = NULL))
plot2 <- ggplot(df2, aes(x = "", y = slices.Freq, fill = amino_acids)) +
  geom_bar(stat = "identity", width = 1) + coord_polar("y",
    start = 0) + ggtitle(ac_2) + xlab("") + ylab("") + guides(fill = guide_legend(title = NULL))
grid.arrange(plot1, plot2, ncol = 2)
```

A0A1I9G1P5

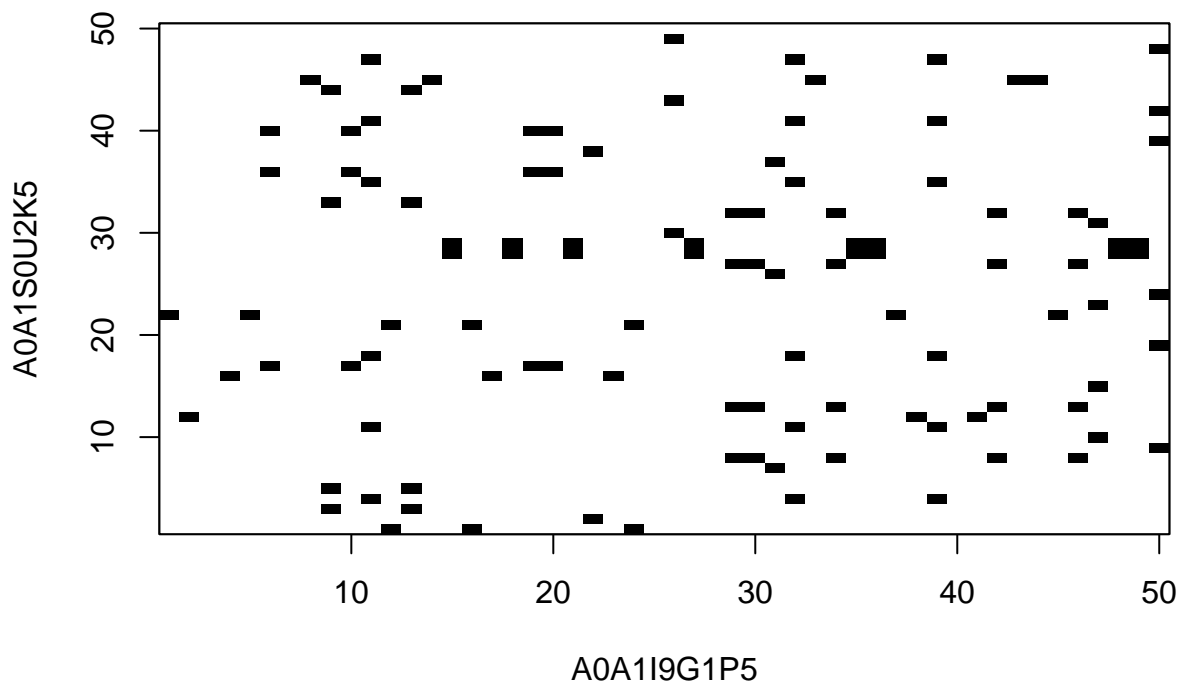


A0A1S0U2K5



Task C

```
dotPlot(tail(seq1, 50), tail(seq2, 50), xlab = ac_1, ylab = ac_2)
```



There is no similarity between the last 50 characters of the two sequences, evidenced by the lack of a diagonal line on the graph.

Task D

```
printPairwiseAlignment <- function(alignment, chunksize = 60,
  returnlist = FALSE) {
  require(Biostrings) # This function requires the Biostrings package
  seq1aln <- pattern(alignment) # Get the alignment for the first sequence
  seq2aln <- subject(alignment) # Get the alignment for the second sequence
  alnlen <- nchar(seq1aln) # Find the number of columns in the alignment
  starts <- seq(1, alnlen, by = chunksize)
  n <- length(starts)
  seq1alnresidues <- 0
  seq2alnresidues <- 0
  for (i in 1:n) {
    chunkseq1aln <- substring(seq1aln, starts[i], starts[i] +
      chunksize - 1)
    chunkseq2aln <- substring(seq2aln, starts[i], starts[i] +
      chunksize - 1)
    # Find out how many gaps there are in chunkseq1aln:
    gaps1 <- countPattern("-", chunkseq1aln) # countPattern() is from Biostrings package
    # Find out how many gaps there are in chunkseq2aln:
    gaps2 <- countPattern("-", chunkseq2aln) # countPattern() is from Biostrings package
    # Calculate how many residues of the first sequence
```

```

# we have printed so far in the alignment:
seq1alnresidues <- seq1alnresidues + chunksize - gaps1
# Calculate how many residues of the second
# sequence we have printed so far in the alignment:
seq2alnresidues <- seq2alnresidues + chunksize - gaps2
if (returnlist == "FALSE") {
  cat(paste0("\\textcolor{blue}{", chunkseq1aln, " ",
    seq1alnresidues, "}", "\\newline"))
  cat(paste0("\\textcolor{red}{", chunkseq2aln, " ",
    seq2alnresidues, "}", "\\newline"))
  cat(paste0(" \\newline"))
  # print(paste(chunkseq1aln, seq1alnresidues))
  # print(paste(chunkseq2aln, seq2alnresidues))
  # print(paste(' '))
}
}
if (returnlist == "TRUE") {
  vector1 <- s2c(substring(seq1aln, 1, nchar(seq1aln)))
  vector2 <- s2c(substring(seq2aln, 1, nchar(seq2aln)))
  mylist <- list(vector1, vector2)
  return(mylist)
}
}

```

```

ga <- pairwiseAlignment(AAString(c2s(seq1)), AAString(c2s(seq2)),
  substitutionMatrix = PAM250, type = "global", gapOpening = -10,
  gapExtension = -0.5)
print(ga)

```

```

## Global PairwiseAlignmentsSingleSubject (1 of 1)
## pattern: MKLIVD---SGHTGVNQLGGVFNVRPLPDSTRQ...QLPPLTQANHLNKKDSGKLLWVSNYKGGWKELLI
## subject: M--LGDKKYSGHTGVNQLGGVFNVRPLPDSTRQ...-----
## score: 534.5

```

```

printPairwiseAlignment(ga, chunksize = 30)

```

```

MKLIVD—SGHTGVNQLGGVFNVRPLPD 27
M-LGDKKYSGHTGVNQLGGVFNVRPLPD 28

```

```

STRQKIVDLAHQGARPCDISRILQVSNGCV 57
STRQKIVDLAHQGARPCDISRILQVSNGCV 58

```

```

SKILCRYYESGTIRPRAIGGSKPRVATVSV 87
SKILCRYYESGTIRPRAIGGSKPRVATVSV 88

```

```

CDKIESYKREQPSIFAWEIRDKLLHEKVCS 117
CDKIESYKREQPSIFAWEIRDKLLHEKVCS 118

```

```

PDTIPSVSSINRVLRLNLAkkeQQAMQNDF 147
PDTIPSI-----HVGf 129

```

```

YDRALRY 177
—SIHF 156

```

Task E

```
la <- pairwiseAlignment(AAString(c2s(seq1)), AAString(c2s(seq2)),
  substitutionMatrix = BLOSUM62, type = "local", gapOpening = -10,
  gapExtension = -0.5)
print(la)

## Local PairwiseAlignmentsSingleSubject (1 of 1)
## pattern: [7] SGHTGVNQLGGVFNNGRPLPDSTRQKIVDLAH...SYKREQPSIFAWEIRDKLLHEKVCSPDTIPSV
## subject: [8] SGHTGVNQLGGVFNNGRPLPDSTRQKIVDLAH...SYKREQPSIFAWEIRDKLLHEKVCSPDTIPSI
## score: 622

printPairwiseAlignment(la, chunksize = 30)
```

```
SGHTGVNQLGGVFNNGRPLPDSTRQKIVDL 30
SGHTGVNQLGGVFNNGRPLPDSTRQKIVDL 30
```

```
AHQGARPCDISRILQVSNNGCVSKILCRYYE 60
AHQGARPDISRILQVSNNGCVSKILCRYYE 60
```

```
SGTIRPRAIGGSKPRVATVSVCDKIESYKR 90
SGTIRPRAIGGSKPRVATVSVCDKIESYKR 90
```

```
EQPSIFAWEIRDKLLHEKVCSPDTIPSV 120
EQPSIFAWEIRDKLLHEKVCSPDTIPSI 120
```

The local alignment's score is higher than the global alignment. With the pairwise printed output we can see that there is just a single mismatch.

Task F

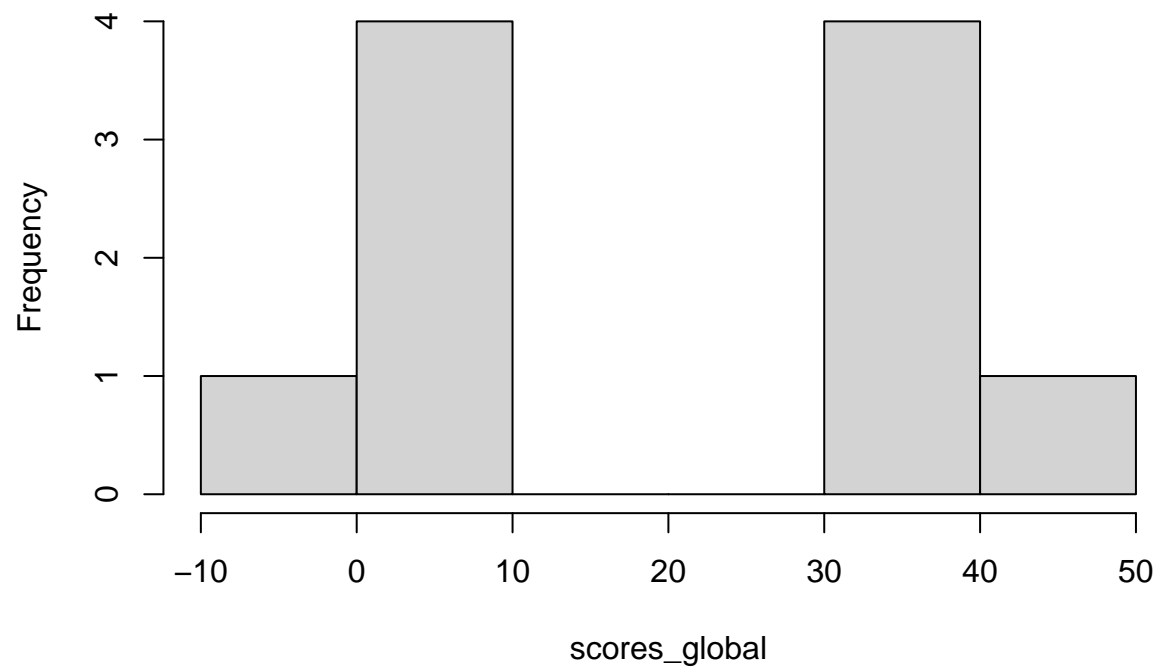
```
rand_seq1 <- generateSeqsWithMultinomialModel(c2s(seq1), 10)
rand_seq2 <- generateSeqsWithMultinomialModel(c2s(seq2), 10)

scores_global <- c()
scores_local <- c()

for (i in 1:length(rand_seq1)) {
  score_global <- pairwiseAlignment(pattern = AAString(c2s(rand_seq1[[i]])),
    subject = AAString(c2s(rand_seq2[[i]])), substitutionMatrix = PAM250,
    gapOpening = -10, gapExtension = -0.5, type = "global",
    scoreOnly = TRUE)
  score_local <- pairwiseAlignment(pattern = AAString(c2s(rand_seq1[[i]])),
    subject = AAString(c2s(rand_seq2[[i]])), substitutionMatrix = BLOSUM62,
    gapOpening = -10, gapExtension = -0.5, type = "local",
    scoreOnly = TRUE)
  scores_global <- append(scores_global, score_global)
  scores_local <- append(scores_local, score_local)
}

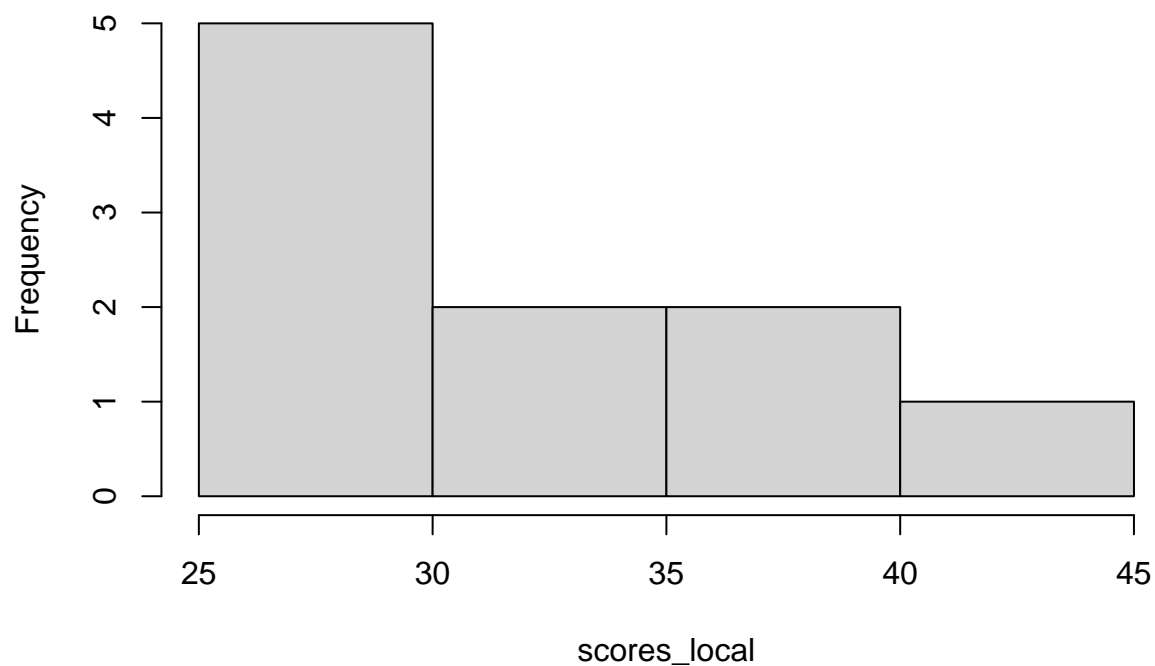
hist(scores_global)
```

Histogram of scores_global



```
hist(scores_local)
```

Histogram of scores_local



```
p_global <- sum(scores_global > score(ga))/length(scores_global)
p_local <- sum(scores_local > score(la))/length(scores_local)

print(p_global)
```

```
## [1] 0
```

```
print(p_local)
```

```
## [1] 0
```

Both the alignments are statistically significant as the 'p' values for both are less than 0.05.