# Review Letter

# How good are predictions of protein secondary structure?

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The three most widely used methods for the prediction of protein secondary structure from the amino acid sequence are tested on 62 proteins of known structure using a program package and data collection not previously available. None of these methods predicts better than 56% of the residues correctly, for a three state model (helix, sheet and loop). The algorithms of Robson et al. [J. Mol. Biol. (1978) 120, 97-120] and Lim [J. Mol. Biol. (1974) 88, 873-894] are the best of those tested. New methods, now under development, can be tested against this benchmark.

Protein structure

Secondary structure prediction

Amino acid sequence

#### 1. INTRODUCTION

The explosive increase in our knowledge of DNA sequences (currently at about 1 Megabase) has led to increased use of protein secondary structure predictions from the amino acid sequence. Typically, one wants to know what structural type of protein the DNA codes for and whether the protein is related to one of known function or structure. The most interesting practical use has been the prediction of antigenic oligopeptides as potential vaccines [3,4]. For any of these uses it is important to know how well secondary structure prediction methods work.

Assessment of available prediction methods is best made by comparing predictions with the crystallographically determined structure. Such comparisons have been made [5], but have been hindered by two facts:

(i) There are ambiguities in two of the best known methods, those of Chou [6] and Lim [2], in that they often give different results in the hands of different people and are therefore not programmable without extension or modification; (ii) There is considerable variation in the definitions of secondary structure given by crystallographers.

We have now solved both of these difficulties and report the results of a completely objective, up-to-date assessment of the most widely used prediction methods on 62 proteins with more than 10000 residues. For a three-state definition of secondary structure (helix, sheet, loop/turn) the overall prediction accuracy for new protein structures does not exceed 56% for the best of these methods and is only 50% for the most widely used (Chou) method. We caution against the overinterpretation of predictions made by presently available methods and provide a benchmark against which new methods, now under development, can be tested.

#### 2. METHODS

Ambiguities in the method of Chou [6] were overcome by selecting possible secondary structure segments such that the sum of preference parameters over *all* chosen segments is maximal; technically, this is a difficult optimization problem

but was achieved by a recursive algorithm which was added to a program written by C. Oefner [7]. Turn prediction, done separately by Chou [8], was not included. Conceivably the overall success of Chou's method can be improved by rules for eliminating overlaps of predicted turns with predicted helix/sheet residues. Ambiguities in the method of Lim [2] were overcome by a simplified iterative procedure for segment selection which was added to a program written by J.A. Lenstra [9]. The (unambiguous) method of Robson was used as programmed by the authors [1].

Known methods not compared here include: Nagano [10] (bad beta prediction in our hands); Maxfield and Scheraga [11] (similar to Robson's, reportedly 57% accurate for five states); Ptitsyn and Finkelstein [12] (new version just published [13]); Palau and Argos [14] (reportedly 56% accurate for four states).

Objective and accurate assignment of secondary structure was achieved by a pattern recognition algorithm [15] which extracts hydrogen-bonded features from the full atomic coordinates as deposited with the Protein Data Bank [16].

# 3. RESULTS AND DISCUSSION

Predictive success is given in table 1 for each

Table 1

Predictive success of the three most widely used secondary structure prediction methods: details for each protein, averaged over the three structure states

a)			24 pr	utein	structures pre-1974
fraction correct (%)					
	Robson				
[6]	[1]	[2]	IDEN	KE5	NAHE
54	59	72	1CPV	108	CALCIUM-BINDING PARVALBUMIN B
60	55	51	285C	85	CYTOCHROME B5 (UXIDIZED)
		55		103	CYTUCHROME ( (UXIDIZED).
43	4.3	70	1020	112	CYTUCHROME C2 (FERRI)
35		50			FERREDOXIN (PEPTOCOCCUS ALROGENES)
ь5		69			RUBREDOXIN (OXIDIZED, FE(III))
40		71	11NS	51	INSULIN (A AND B CHAIN)
58		64		129	LYSUZYME (HEN EGG WHITE, TRICEINIC)
58		68		142	STAPHYLUCUCCAL NUCLEASE (COMPLEX)
57	66	66	18145	124	RIBONUCLEASE-5
47	61	59 59	1CPA	308	CARBOXYPEPTIDASE A
50	52	59	2TLN	316	THERMOLYSIN
53	54	74	2GCH	236	GAMMA CHYMOTRYPSIN A
57	53	66	15 LN	223	
	57	67	1SBT	275	SUBTILISIN BPN'
55	59	58 70	1EST	240	TOSYL-ELASTASE
53	55	70	8PAP	212	PAPAIN
59	62	71	3CNA	231	
46		51		329	
51	72	71	1MBN	153	MYOGLOBIN (FERRIC IRON - METMYOGLOBIN)
30	54	ь3	1ECD	136	HEMOGLOBIN (ERYTHROCRUORIN DEOXY)
47	57	63 62	2MHB	287	HEMOGLOBIN (HORSE, AQUO MET)
37	55	12	1LHB		
		12		58	TRYPSIN INHUBITOR
51	57	65			SUBTOTAL PRE-1974

)			۳۰ دد	ocein	structures jour-1974
actio	n corr	ect (%)			protein structure
	Rob son				
[6]	[1]	[2]	IDEN	rL5	NAME
51		411	1 24 11		T Z MOTERNE PERCE MARKET
53	53	49 54	1,467	0 ل ز	E-ARADIADSE-SERVERS PROTECT OXIDIZED HIGH POTERTIAL I COL PROTECT
33	40	04	THILL	05	ONTO THE MENT OF THE TOTAL TOTAL TO
46	44	51 66	1209	103	CATOCHARINE OBSIZ (E. COLI, CAID'E D
		00	1000	1.54	CYTOCHROLE C550
49 36	06 43	61 72	251C 1FXC 3FXH 1AZU 1PU/	54	effochRows up51 (Oxidiate) Flaggodxi4 (Swisdulfor Plot Note)
50 50	59	7.2	30.41	120	FLAVOURIN (DIPLIZ P)
59	47	0.7	1.670	150	AZBRIA
56	52	5.7	1017	125	PLAST JUYANIN
5 <sub>6</sub>	44	10	1007	32	AVIAN PANCPENTIC PULYPEPTION
45	41	60	1F PT 1GUN	-10	GLUCAGON (PH t=/)
50	47	50	1802	123	PHOSPHOLIPASE A2
32	56	51 57 39 59 50 52	18P2 1LZM	164	LYSOZYME (BACTERIOPHNG T4)
54	59	1,2	1 100	324	ACID PROTEAST (SHIPPOND BIT TOLD)
46	53	62 50	1APR 1APP	253	MCID PROTLINASE (PENT INC. FOR A)
48	52	5.3	1.31 P	133	ALPHA LYTIC PROTLANC
56	60	52 52	IALP ISGA	131	PROTEINASE A FROM SET OFFICE A PILLUS
51	56	51	CACT	218	ACTINIDIN
54	60	51 60 60	1FAB		
57	54	60	IREI	107	BENCE-JONES IMMUNO OBULIN
44	55	61	DEGM	230	PHOSOHUGI YCEDATE MULASE IDE-110 0001
50	65	59	1TIM	246	PHOSPHUGLYCEPATE MUTASE (DE-110 000) TRIOSE PHOSPHATE ISOMEPASE
50	53	61 59 61 50	1CAC	25n	CAPBONIC ANHYDRASE FORM C
46	51	50	1CAC 1DFR	152	DIHYDROFOLATE REDUCTASE (UNIPLEX)
47	25	58 52	1GPD	333	D=GYCLRALDEHYDE=3=PHOS MATE DESCRIBING NAME
39	44	5,2	4 ADH	374	D-GYCERALDEHYDE-3-PHOS MATE DEMYOR NEW APO-LIVER ALCOHOL DEHYDROGENAS!
45	49	43	2685	4n !	GERTATRIONE REDUCTASI
_	12				
52	69	50	1HBI	150	LEGRETOGLOGIA CYLLEGA LDE'AD
37	33	44	LURN	4p	CRAMSIN
48	54	77	1000	56	OVEMBOUTD THERE DOMAIN
51	63	54	255T	107	STREPTONICES SUBTILISED LABOURE
68	65	69	1CTx	71	ALPHA LOGRATOXIN
42	42	46	1MLT	26	MELITTIN
50	61	60	1N/3	02	NEURUTUKIN B
52	73	55	2AUK	194	ADENYLATE KINASE
55	54	54	1PHD	293	RHUDANLUL
46	4.2	40	2PAB	114	OU ZA SUPERIXITO DISTURBANCE LEGALIMOCODI (MILLEGI IN TO LWANDIA UWONDODI THI RY DOTALY STREPTORTHES SUBTLISTY I CHISTOR ALPHA CUBRATOXIN MELITTIM MERROTOCIN B ADENYLATE KIRASE RHODANES, PREALBOMIN (MITAN TLASAT)
49	55*	56*		LL30	SUBTOTAL POST-1074
50	E.		1	0.256	TOTAL FOR . 1 PROTEINS

All methods are compared according to how well they predict the three states  $\alpha$ -helix,  $\beta$ -sheet and loop (everything else) or older (a) and newer (b) protein structures. Fraction correct is the number of residues predicted correctly in any state divided by the total number of residues. The protein name is preceded by the protein data bank [16] identifier IDEN and the number of residues RES. The \* indicates the percentage of correctly predicted residues one can expect in applying the methods of Robson and Lim to newly determined sequences

protein and each method as the percentage of residues predicted correctly in a three state description of secondary structure. The result of the comparison is similar to that of Busetta and Hospital [17] who have 47% success for Chou and 57% success for Robson on 34 proteins. The method of Lim has a surprising 65% success rate for protein structures known in 1974 when his method was published, but this drops to 56% for proteins elucidated after 1974. The difference can be understood to be due to special rules tailored to particular proteins in Lim's method.

Structure predictions can be evaluated in more

detail by calculation of assorted quality indices [5] which indicate how well a particular state is predicted, whether there is over- or underprediction etc. All of these indices can be calculated from the predicted/observed matrix in table 2 which indicates, say, how many of the 2295 observed helical residues are correctly predicted as helical (H) and how many are wrongly predicted as loop/turn (L) or sheet (E, for extended); or, how many of the 2684 residues predicted as helical by Lim are sheet, loop or helical in the crystallographic structure. One such quality index for each state is the 'fraction correct of observed' in table 2. For example, we see that 74% of the observed loop residues are correctly predicted by Lim, while only 36% of the observed sheet residues are correct; this imbalance is related to an overall underprediction (1690/2295) of sheet and an over-

Table 2

Predictive success of the three most widely used secondary structure prediction methods: details of sheet (E), loop/turn (L) and helix (H) prediction averaged over all proteins

	Ch	[e] world		Robson [1]			L	lət L	2]				
	observed/predicted matrix (number of residues)												
observed p observed L observed H	µr E 1195 1361 ∂94	2900	417	E 1244 1161	689	H 362 1203	820 575	redic L 1151 4027 1211	813 813				
	observed/predicted total (number of residues)												
observed predicted	2795	5421	3047	E 2235 2975	5421	3047	E 2295 1590	5421					
	fraction correct (%)												
of observed of predicted	E 52 35	53 65	H 42 45	E 54 42	50		E 36 49	74	H 51 57	(a) (b)			
	rms error in "redicting secondary Structure content (residues per 100 residues)												
	E 17	L 14	н 1ь	E 12	L 16	н 16	E 13	L 14	н 13				

- (a) Number of residues correctly predicted in state S divided by number of residues observed in state S = percentage of correct predictions when state S is observed
- (b) Number of residues correctly predicted in state S divided by number of residues predicted in state S = percent probability of correct prediction when state S is predicted. The latter 'probability of correct prediction', PC(S), is practically useful in predicting unknown secondary structure

prediction (6389/5421) of loop residues in Lim's method.

Suppose you have predicted a residue as helical and want to know the chances of being right. For a particular method, the average 'fraction correct of predicted' (table 2) defined as:

$$PC(S) = \frac{N \text{ (correctly predicted in state S)}}{N \text{ (predicted in state S)}} \times 100$$

is a direct measure of the probability of correct prediction having predicted a residue to be in state S. Curiously, PC(S) does not appear among the quality indices commonly used [5], but is perhaps the most useful in prediction practice (after all, in a truly unknown protein structure no reference can be made to observed states). For example, when Lim's method predicts a sheet strand, we can estimate from table 2 that there is a 49% chance of correct prediction. Note the high probability of correct loop prediction of 63–68% which is related to the high fraction (50%) of observed loop residues.

Suppose you do not care about the details of secondary structure assignments but merely want to use a secondary structure prediction method to predict the helix/sheet *content* of a protein; for example, for comparison with spectroscopic determinations (such as circular dichroism). The rootmean-square average difference between predicted and observed secondary structure content for the 62 proteins is 12-17 residues/100 residues (table 2). For example, a prediction by Robson of sheet content has a typical uncertainty of  $\pm 12\%$ . An uncertainty of this size renders present comparisons of predicted secondary structure content with circular dichroism experiments useless in all but extreme cases.

We conclude that one may expect a success rate, for three states, of about 50% with Chou's method and of 55-56% with either Robson's or Lim's method. In any event, an error rate of 44% is unacceptable for many purposes and newly developing methods must do better. We estimate that empirical-statistical prediction of secondary structure alone may eventually reach 70% accuracy for three states; higher accuracy will, in our opinion, only come with a protein-folding theory aiming at prediction of the complete three-dimensional structure.

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#### **REFERENCES**

- [1] Garnier, J., Osguthorpe, D.J. and Robson, B. (1978) J. Mol. Biol. 120, 97-120.
- [2] Lim, V.I. (1974) J. Mol. Biol. 88, 873-894.
- [3] Mueller, G.M., Shapira, M. and Arnon, J. (1981) Proc. Natl. Acad. Sci. USA 79, 569-573.
- [4] Pfaff, E., Mussgay, M., Boehm, H.O., Schulz, G.E. and Schaller, H. (1982) EMBO J. 1, 869-874.
- [5] Schulz, G.E. and Schirmer, R.H. (1979) Principles of Protein Structure, Ch.6, Springer-Verlag, New York.

- [6] Chou, P.Y. and Fasman, G.D. (1978) Adv. Enzymol. 47, 45-148.
- [7] Oefner, C. (1982) Thesis (Diplomarbeit), University of Heidelberg.
- [8] Chou, P.Y. and Fasman, G.D. (1979) Biophys. J. 25, 367-383.
- [9] Lenstra, J.A. (1977) Biochim. Biophys. Acta 491, 333-338.
- [10] Nagano, K. (1977) J. Mol. Biol. 109, 251-274.
- [11] Maxfield, F.R. and Scheraga, H.A. (1979) Biochemistry 18, 697-704.
- [12] Finkelstein, A.V. and Ptitsyn, O.B. (1971) J. Mol. Biol. 62, 613-624.
- [13] Ptitsyn, O.B. and Finkelstein, A.V. (1983) Biopolymers 22, 15-25.
- [14] Palau, J., Argos, P. and Puigdomenech, P. (1982) Int. J. Pept. Prot. Res. 19, 394-401.
- [15] Kabsch, W. and Sander, C. (1983) Biopolymers, in press.
- [16] Bernstein, F.C., Koetzle, T.F., Williams, G.J.B., Meyer, E.F., Brice, M.D., Rodgers, J.R., Kennard, O., Shimanouchi, T. and Tasumi, M. (1977) J. Mol. Biol. 112, 535-542.
- [17] Busetta, B. and Hospital, M. (1982) Biochim. Biophys. Acta 701, 111-118.