

# Winegrape Hardiness

## My final (I hope) dose response model

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## 1 Background

Dose response curves are regression models. The exact form of dose response models varies somewhat, encompassing a range of different statistical models including nonlinear regression models, generalized regression and parametric survival analyses Ritz *et al.* (2015). What connects these models is their application: to model the effect of some dose on a biological response. The model assesses the relationship between what is traditionally named the “dose” or “concentration” and the “response” or “effect”. This terminology comes from the model’s original use in pharmacology for modeling the effects of substances on physiology. The dose is usually some sort of biological stress that elicits a response from a biological organism. Dose values should be non-negative Rudemo *et al.* (1989) and the response values should change monotonically with increasing dose. Variations on the dose-response model have been applied to a wide variety of ecological questions, including species richness responses to nitrogen Jones *et al.* (2018), eco-toxicology Haanstra *et al.* (1985), and phenology (winegrape budburst dates) Kovaleski &

Londo (2019).

My analyses focus on sigmoid curve log-logistic models. These models are used to model relationships between variables with asymptotic minimum and maximum response values. I use the four parameter log-logistic model described in Ritz *et al.* (2015) and the below section, where the relationship between the dose and the response is a function of a maximum response, a minimum response, a response rate, and a dose giving a 50% response. A benefit of this model is that the parameters are easily interpretable and biologically meaningful Seefeldt *et al.* (1995)

## 2 Model Structure

### 2.1 Basic Model

As introduced above, I use a four parameter log-logistic model to model winegrape cold hardiness Equation (2.1). We used mean 2 day air temperature for our modeling because **Check Carl's reasoning** and because winegrape hardiness is closely linked to mean air temperatures Hubackova (1996). Air temperature readings were taken at **Penticton agCanada site???** **check**.

Some modifications to the original data are needed, though, before analysis. Firstly, to ensure the dose values are always positive, I added 30 to each air temperature value. Secondly, I multiplied the hardiness values with 1 so that larger values in the model meant higher winter hardiness. This was to make interpreting the minimum and maximum asymptotes of the model more intuitive.

$$\mu = f(x, (b, c, d, e)) = c + \frac{d - c}{1 + \exp^{b(\log(x) - \tilde{e})}}$$

$$\tilde{y}_i \sim normal(\mu_i, \sigma)$$

Where:

$x$  is the concentration of the dose (amount of winter cold)

$b$  is the response rate (slope)

$d$  is the upper asymptote of the response (maximum hardiness)

$c$  is the lower asymptote of the response (minimum hardiness)

$e$  is the effective dose ED50 (winter temperature where cold hardiness is half way between min and max)

$\tilde{e}$  is the log of the effective dose ED50

### 2.2 Full Model

The final model includes hierarchical variance for different varieties and sites on the  $d$  (maximum hardiness) parameter and for variety on the  $b$  (rate of change) parameter Equation ??

We expected different sites to vary in their maximum hardiness because the model uses temperature data for a single site but the weather conditions at sites in the Okanagan Valley can vary substantially. For example (**insert name of site**) is a site on a south facing slope close to the lake shore and so is warmer, whereas the colder site of (**insert name of site**) is further north and more inland. Such micro-climatic differences should cause maximum hardiness to be less in warmer sites and more in colder sites.

Winegrapes have been domesticated for many thousands of years, and over that time growers have cultivated a wide range of varieties (genetically unique variants) with different physiological and ecological characteristics. Winegrape varieties consequently vary a lot in many of their traits. Although the exact mechanisms behind winegrape winter hardiness are unknown, winter hardiness seems to vary across varieties Mills *et al.* (2006); Ferguson *et al.* (2014); Kovaleski *et al.* (2018). Winegrapes may have different rates of change of winter hardiness Kovaleski *et al.* (2018); Ferguson *et al.* (2014) and different maximum hardiness values Ferguson *et al.* (2014). We included a hierarchical effect of variety on both maximum hardiness and the rate of change of hardiness so we could assess how variable variety specific winter hardiness is.

We built our dose response model in a Bayesian framework using Stan **cite stan version** in R **cite rstan and r**. An essential part of modeling using Bayesian methods is the choice of prior expectations on each parameter value. Our priors are specified in Equation 2.2, and were generally chosen to encompass all possible parameter combinations according to our current physiological understanding of winegrape hardiness. The exception to this is parameter  $c$ , minimum winter hardiness. Our data did not span the full range of the sigmoid curve relationship of winter hardiness to air temperature; we lack data on minimum hardiness. This is a problem for model estimating. An estimation of minimum hardiness of winegrapes was taken from a selection of sources: -3°C Ferguson *et al.* (2011), -1.2°C Ferguson *et al.* (2014) **more sources**. We fed this estimation into the model as a prior constrained closely around -2°C.

$$\mu = f(x_i, (b, c, d, e)) = c + \frac{(d + d_{var,i} + d_{site,i}) - c}{1 + \exp^{b_{var}(\log(x_i) - \hat{e})}}$$

$$d_{var} = dr_{var} * \sigma_{dvar}$$

$$d_{site} = dr_{site} * \sigma_{dsite}$$

$$b_{var} = br_{var} * \sigma_{bvar}$$

$$\tilde{y}_i \sim normal(\mu_i, \sigma)$$

Where:

$x$  is the concentration of the dose (amount of winter cold)

$b$  is the response rate (slope)

$d$  is the grand upper asymptote of the response (maximum hardiness)  $d_{var}$  is the effect of each variety on the upper asymptote of the response (maximum hardiness)

$d_{site}$  is the effect of each site on the upper asymptote of the response (maximum hardiness)

$\sigma_{dvar}$  is the standard deviation of the effect of varieties on maximum winter hardiness

$\sigma_{d_{site}}$  is the standard deviation of the effect of sites on maximum winter hardiness  
 $dr_{var}$  is the non centred parameterization values for varieties effect on maximum hardiness d  
 $dr_{site}$  is the non centred parameterization values for sites effect on maximum hardiness d  
 $b$  is the lower asymptote of the response (minimum hardiness)  
 $\sigma_{bvar}$  is the standard deviation of the effect of varieties on rate of change of winter hardiness  
 $br_{var}$  is the non centred parameterization values for varieties effect on rate of change  
 $e$  is the effective dose ED50 (winter temperature where cold hardiness is half way between min and max)  
 $\tilde{e}$  is the log of the effective dose ED50

Priors (hardiness has been multiplied with -1 to be positive, and 30 has been added to air temp)  
 $b \sim \text{gamma}(7, 1)$   
 $\sigma_{bvar} \sim \text{normal}(0, 3)$   
 $br_{var} \sim \text{normal}(0, 1)$   
 $d \sim \text{Normal}(25, 10)$   
 $\sigma_{dvar} \sim \text{gamma}(2.5, 1.75)$   
 $dr_{var} \sim \text{normal}(0, 1)$   
 $\sigma_{d_{site}} \sim \text{gamma}(2.5, 1.75)$   
 $dr_{site} \sim \text{normal}(0, 1)$   
 $c \sim \text{normal}(2, 0.5)$   
 $\tilde{e} \sim \text{normal}(\log(30), 0.15)$   
 $\sigma \sim \text{normal}(0, 5)$

### 3 Model fit

#### 3.1 Prior Predictive Checks

Plot of predicted values

#### 3.2 Retrodictive Checks

Pairs plot Plot of predicted values against real data plot of predicted data against real data

#### 3.3 Estimated Parameter Values

Table of values for each parameter Panel of plots for each parameter Plot of effect of variety and site on d Plot of effect of varieties on d Plot of effect of sites on d Plot of effects of varieties on b

#### 3.4 Predictions

Description and citation of data from Washington Plot of model prediction x y

## 4 Discussion

Generally the model seems to do a good job of predicting values, even from new datasets/areas. Some uncertainty around predicted values though

Model overestimates hardiness in autumn if there is an especially cold snap - how can I work with this?

Site differences - how do these compare to our (Carl's) knowledge of the geography of the sites?

Variety differences - compare to results in Ferguson *et al.* (2014)'s table of winegrape variety max hardiness. Also contrast our results of no effect on rates of change to results by Ferguson *et al.* (2014); Kovaleski *et al.* (2018); Kovaleski & Londo (2019).

Compare relative importance of variety and site for growers adapting to climate change.

## References

- Ferguson, J.C., Moyer, M.M., Mills, L.J., Hoogenboom, G. & Keller, M. (2014) Modeling dormant bud cold hardiness and Budbreak in twenty-three *Vitis* genotypes reveals variation by region of origin. *American Journal of Enology and Viticulture* **65**, 59–71.
- Ferguson, J.C., Tarara, J.M., Mills, L.J., Grove, G.G. & Keller, M. (2011) Dynamic thermal time model of cold hardiness for dormant grapevine buds. *Annals of Botany* **107**, 389–396.
- Haanstra, L., Doelman, P. & Voshaar, J.H.O. (1985) The use of sigmoidal dose response curves in soil ecotoxicological research. *Plant and Soil* **84**, 293–297.
- Hubackova, M. (1996) Dependence of Grapevine Bud Cold Hardiness on Fluctuations in Winter Temperatures. *American Journal of Viticulture* **47**, 100–102.
- Jones, L., Milne, A., Hall, J., Mills, G., Provins, A. & Christie, M. (2018) Valuing Improvements in Biodiversity Due to Controls on Atmospheric Nitrogen Pollution. *Ecological Economics* **152**, 358–366.
- Kovaleski, A.P. & Londo, J.P. (2019) Tempo of gene regulation in wild and cultivated *Vitis* species shows coordination between cold deacclimation and budbreak. *Plant Science* **287**.
- Kovaleski, A.P., Reisch, B.I. & Londo, J.P. (2018) Deacclimation kinetics as a quantitative phenotype for delineating the dormancy transition and thermal efficiency for budbreak in *Vitis* species. *AoB PLANTS* **10**, 1–12.
- Mills, L.J., Ferguson, J.C. & Keller, M. (2006) Cold-Hardiness Evaluation of Grapevine Buds and Cane Tissues. *American Journal of Viticulture* **2**, 194–200.
- Ritz, C., Baty, F., Streibig, J.C. & Gerhard, D. (2015) Dose-Response Analysis Using R. *PLOS ONE* **10**, e0146021.
- Rudemo, M., Ruppert, D. & Streibig, J.C. (1989) Random-Effect Models in Nonlinear Regression with Applications to Bioassay. *Biometrics* **45**, 349–362.
- Seefeldt, S.S., Jensen, J.E. & Fuerst, E.P. (1995) Log-Logistic Analysis of Herbicide Dose-Response Relationships. *Weed Technology* **9**, 218–227.