

#### Data construction for fusion v5

- MANE v1.4
  - 19, 226 transcripts (filtered via “filter\_mane\_gff.py”)  
Inclusion criteria:  
chr1-22, X, Y, protein\_coding, MANE\_Select (summary.txt => MANE Select; manually curated)
- GENCODE-r47
  - only use its FASTA source file => extract defined transcripts in MANE v1.4 gff via “bedtools getfasta”

The fusion v5 db is derived from the following downloaded files:

1. (MANE v1.4)  
MANE v1.4 DB  
([https://ftp.ncbi.nlm.nih.gov/refseq/MANE/MANE\\_human/release\\_1.4/](https://ftp.ncbi.nlm.nih.gov/refseq/MANE/MANE_human/release_1.4/))  
(/mnt/RD\_Develop/sandyteng/ACTFusionV5/db\_fusionv5/mane\_v1.4/OpenDB\_MANE\_human\_v1.4/release\_1.4/*MANE.GRCh38.v1.4.summary.txt.gz*)  
(/mnt/RD\_Develop/sandyteng/ACTFusionV5/db\_fusionv5/mane\_v1.4/OpenDB\_MANE\_human\_v1.4/release\_1.4/*MANE.GRCh38.v1.4.ensembl\_genomic.gff.gz*)
2. (Genome sequence, Grch38, GENCODE-r47)  
GRCh38.p14.genome.fa.gz  
Gencode V47  
([http://ftp.ebi.ac.uk/pub/databases/gencode/Gencode\\_human/release\\_47/](http://ftp.ebi.ac.uk/pub/databases/gencode/Gencode_human/release_47/))  
(/mnt/RD\_Develop/sandyteng/ACTFusionV5/db\_fusionv5/gencode\_v47/OpenDB\_GENCODE\_human\_r47/*GRCh38.p14.genome.fa.gz*)
3. (Probe information file provided by AD team)  
→ 1,039 probe regions bed file (genomic locations of the designed/target region  
  
(/mnt/RD\_Develop/sandyteng/ACTFusionV5/db\_fusionv5/captureprobe\_250401/*ACTFusionv5\_target-region\_PartAB\_individual\_1039.bed*) (obtained from AD team Lucy)

### <19,292-transcript sequence extraction>

The fasta sequence of the 19,292 transcripts are obtained via the following steps:

1. gff lines extraction (“filter\_manegff.py”)  
python3 /mnt/RD\_Develop/sandyteng/ACTFusionV5/code/filter\_manegff.py \  
-i MANE.GRCh38.v1.4.ensembl\_genomic.gff.gz \  
-o *MANE.GRCh38.v1.4.ensembl\_genomic.transcript.gff*
2. gff to bed file conversion (“/tools/Fusion/convert2bed”)  
/tools/Fusion/convert2bed -i gff -d  
< ./mane\_v1.4/OpenDB\_MANE\_human\_v1.4/derived/MANE.GRCh38.v1.4.ensembl\_genomic.transcript.gff  
> ./mane\_v1.4/OpenDB\_MANE\_human\_v1.4/derived/MANE.GRCh38.v1.4.ensembl\_genomic.transcript.bed
3. bed to fasta file conversion (“bedtools” in image actgenomics/fusion\_dev:v0.6)  
bedtools getfasta -name -s -  
fi ./gencode\_v47/OpenDB\_GENCODE\_human\_r47/derived/GRCh38.p14.genome.fa -  
bed ./mane\_v1.4/OpenDB\_MANE\_human\_v1.4/derived/MANE.GRCh38.v1.4.ensembl\_genomic.transcript.bed -  
fo ./mane\_v1.4/OpenDB\_MANE\_human\_v1.4/derived/MANE.GRCh38.v1.4.ensembl\_genomic.transcript.corrected.strand.fasta
4. Additional (-) or (+) strings (within the fasta file) removal  
sed -i  
's/([+-])//g' ./mane\_v1.4/OpenDB\_MANE\_human\_v1.4/derived/*MANE.GRCh38.v1.4.ensembl\_genomic.transcript.corrected.strand.fasta*

Note: The gff extraction program “filter\_manegff.py” extracts 19,292 protein\_coding transcripts located on chr1-22, X, Y from

“*MANE.GRCh38.v1.4.ensembl\_genomic.gff.gz*”

#### <Target transcript selection (ENST ID – RefSeq ID map)>

Among the 19,292 extracted transcripts, 66 transcripts are not labeled “MANE Select” in the “*MANE.GRCh38.v1.4.summary.txt.gz*” file (column: MANE\_status). The 66 transcripts are labeled “MANE Plus Clinical” instead. To avoid mapping ambiguity, we only include the 19,226 (=19,292 - 66) transcripts labeled “MANE Select”.

Note:

There are total 19,338 transcripts labeled “MANE Select” and 66 transcripts labeled “MANE Plus Clinical” in the \*summary.txt.gz file

The namemap files for the 19,404 (19,338 + 66) transcripts:

/mnt/RD\_Develop/sandyteng/ACTFusionV5/db\_fusionv5/mane\_v1.4/OpenDB\_MANE\_human\_v1.4/derived/*MANE.GRCh38.v1.4.select.and.plus.clinical.namemap*

#### <Empty pseudo intron annotation (loci) table + pseudo N (10N) fasta generation>

The pseudo intron sequences and the corresponding annotation (loci) tables (transcriptome + genome) for the 19,226 transcripts were generated via “RefFusion.v2.py”.

```
python3 /mnt/RD_Develop/sandyteng/ACTFusionV5/code/RefFusion.v2.py \
-g
/mnt/RD_Develop/sandyteng/ACTFusionV5/db_fusionv5/mane_v1.4/OpenDB_MANE_human_v1.4/derived/MANE.GRCh38.v1.4.ensembl_genomic.gff \
-m
/mnt/RD_Develop/sandyteng/ACTFusionV5/db_fusionv5/mane_v1.4/OpenDB_MANE_human_v1.4/derived/MANE.GRCh38.v1.4.summary.txt \
-f
/mnt/RD_Develop/sandyteng/ACTFusionV5/db_fusionv5/mane_v1.4/OpenDB_MANE_human_v1.4/derived/MANE.GRCh38.v1.4.ensembl_genomic.transcript.corrected.strand.fasta \
-p
/mnt/RD_Develop/sandyteng/ACTFusionV5/db_fusionv5/mane_v1.4/Output_Final/P.A053_ACTFusionV5_PseudoIntron_MANE-v1.4_GENCODE-r47_capture-v1.0_GRCh38.20250407.transcript.MANE.only.list \
-o
/mnt/RD_Develop/sandyteng/ACTFusionV5/db_fusionv5/Output_MANE_Select/20250407_MANE.r47
```

### <Probe anchored (mapping) exons extraction>

The 1,039 probe regions are converted to 533 exons located on the 19,226 extracted transcripts via “candidate\_exons\_mapping.sh”.

The information of the 533 extracted exons:

- *fusionv4.MANE.v1.4.GENCODE.r47.candidate.exons.transcript.bed*
- /mnt/RD\_Develop/sandyteng/ACTFusionV5/db\_fusionv5/InhouseDB\_Probe/captureprobe\_250407\_MANE\_Select/probeseq/*MANE.GRCh38.v1.4.0407.probe.r47.fasta*

For the fasta header for each probe sequence are converted via

“Probe\_faheader\_converter.py” and decompressed via “gunzip”:

- /mnt/RD\_Develop/sandyteng/ACTFusionV5/db\_fusionv5/InhouseDB\_Probe/captureprobe\_250407\_MANE\_Select/probeseq/*MANE.GRCh38.v1.4.0407.r47.probe.wtprimerlikeheader.fasta*

# sample command (prerequisite: “actgenomics/fusion\_dev:v0.6”)

docker run --rm -v /mnt:/mnt/ -it actgenomics/fusion\_dev:v0.6

```
bash ./candidate_exons_mapping.sh \  
    /path/to/genome.loci \  
    /path/to/transcript.loci \  
    /path/to/namemap \  
    /path/to/probe.bed \  
    fusionv4.MANE.v0.95.GENCODE.r38 \  
    /mnt/RD_Develop/sandyteng/workdir \  
    /tools/Fusion
```

# obtain mapping exons (pseudo locations on 10\*N transcriptome)

```
bash /mnt/RD_Develop/sandyteng/ACTFusionV5/code/candidate_exons_mapping.sh  
/mnt/RD_Develop/sandyteng/ACTFusionV5/db_fusionv5/Output_MANE_Select/20250407_MANE.r47.genome.loci  
/mnt/RD_Develop/sandyteng/ACTFusionV5/db_fusionv5/Output_MANE_Select/20250407_MANE.r47.transcript.loci  
/mnt/RD_Develop/sandyteng/ACTFusionV5/db_fusionv5/mane_v1.4/OpenDB_MANE_human_v1.4/derived/MANE.GRCh38.v1.4.select.and.plus.clinical.namemap  
/mnt/RD_Develop/sandyteng/ACTFusionV5/db_fusionv5/captureprobe_250401/ACTFusionv5_target-region_PartAB_individual_1039.bed  
fusionv4.MANE.v1.4.GENCODE.r47
```

```

/mnt/RD_Develop/sandyteng/ACTFusionV5/db_fusionv5/InhouseDB_Probe/capturep
robe_250407_MANE_Select/ /tools/Fusion
# extract mapped exons (candidate.exons.transcript.bed) sequences from gencode
fasta file (gencode.genome.fa)
bedtools getfasta -name -s -fi
/mnt/RD_Develop/sandyteng/ACTFusionV5/db_fusionv5/Output_MANE_Select/202
50407_MANE.r47.fasta -bed
/mnt/RD_Develop/sandyteng/ACTFusionV5/db_fusionv5/InhouseDB_Probe/capturep
robe_250407_MANE_Select/fusionv4.MANE.v1.4.GENCODE.r47.candidate.exons.
transcript.bed -fo
/mnt/RD_Develop/sandyteng/ACTFusionV5/db_fusionv5/InhouseDB_Probe/capturep
robe_250407_MANE_Select/probeseq/MANE.GRCh38.v1.4.0407.probe.r47.fasta

# probe fasta generation
python3
/mnt/RD_Develop/sandyteng/ACTFusionV5/code/Probe_faheader_converter.py \
-f
/mnt/RD_Develop/sandyteng/ACTFusionV5/db_fusionv5/InhouseDB_Probe/capturep
robe_250407_MANE_Select/probeseq/MANE.GRCh38.v1.4.0407.probe.r47.fasta \
-n
/mnt/RD_Develop/sandyteng/ACTFusionV5/db_fusionv5/mane_v1.4/OpenDB_MAN
E_human_v1.4/derived/MANE.GRCh38.v1.4.select.and.plus.clinical.namemap \
-o
/mnt/RD_Develop/sandyteng/ACTFusionV5/db_fusionv5/InhouseDB_Probe/capturep
robe_250407_MANE_Select/probeseq/MANE.GRCh38.v1.4.0407.r47.probe.wtprim
erlikeheader.fasta.gz
# unzip fasta.gz
gunzip
/mnt/RD_Develop/sandyteng/ACTFusionV5/db_fusionv5/InhouseDB_Probe/capturep
robe_250407_MANE_Select/probeseq/MANE.GRCh38.v1.4.0407.r47.probe.wtprim
erlikeheader.fasta.gz

```

< Probe/Exon (query) to Pseudo-intron Transcript (subject) alignment via blastn >

To annotate the 533 mapped exons to the selected transcriptome, the raw probe fasta file (*MANE.GRCh38.v1.4.0407.probe.r47.fasta*) is converted to a fasta file (*MANE.GRCh38.v1.4.0407.r47.probe.wtprimerlikeheader.fasta*) of the following format:

>Probe ID|Gene Name|RefSeq ID|ENST ID|exon number|F|probe length

Probe sequence (mapped exon sequence)

(e.g.,

>Probe-mane001|PSMB2|NM\_002794.5|ENST00000373237.4|2|F|123

ATCATGACAAGATGTTTAAGATGAGTGAAAAGATATTACTCCTGTGTGTTGG  
AGAGGCTGGAGACACTGTACAGTTTGCAGAATATATTCAGAAAAACGTGCA  
ACTTTATAAGATGCGAAATG)

The probe sequences (fasta file) were then aligned against the 19,226 pseudo-intron (10N) containing fast file (*20250407\_MANE.r47.fasta*) using the following command:

/tools/Fusion/ncbi-blast/bin/blastn -query

/mnt/RD\_Develop/sandyteng/ACTFusionV5/db\_fusionv5/InhouseDB\_Probe/capturep  
robe\_250407\_MANE\_Select/probeseq/*MANE.GRCh38.v1.4.0407.r47.probe.wtprim  
erlikeheader.fasta* -subject

/mnt/RD\_Develop/sandyteng/ACTFusionV5/db\_fusionv5/Output\_MANE\_Select/*202  
50407\_MANE.r47.fasta* -outfmt 6 -task blastn-short -dust no >

/mnt/RD\_Develop/sandyteng/ACTFusionV5/db\_fusionv5/InhouseDB\_Probe/capturep  
robe\_250407\_MANE\_Select/blastn/*20250407\_probe.r47.blastn*

After obtaining the “forward” probe alignment result, the “reverse” probe result is manually constructed and combined with the “forward” probe result via the following commands:

# create blastn result for “reverse probe” and concatenate all the alignments

cat *20250407\_probe.r47.blastn* > 20250407\_rprobe.r47.blastn

sed -i 's/|F|/|R|/' 20250407\_rprobe.r47.blastn

sed -i 's/mane/rmane/' 20250407\_rprobe.r47.blastn

cat 20250407\_probe.r38.blastn 20250407\_rprobe.r38.blastn >

*20250407\_probe.rprobe.r47.blastn*

```

# blastn parser (loci annotation)
python3 /mnt/RD_Develop/sandyteng/ACTFusionV5/code/blastnparser.py \
-if
/mnt/RD_Develop/sandyteng/ACTFusionV5/db_fusionv5/InhouseDB_Probe/capturep
robe_250407_MANE_Select/blastn/20250407_probe.rprobe.r47.blastn \
-mp
/mnt/RD_Develop/sandyteng/ACTFusionV5/db_fusionv5/mane_v1.4/OpenDB_MAN
E_human_v1.4/derived/MANE.GRCh38.v1.4.select.and.plus.clinical.namemap \
-lf
/mnt/RD_Develop/sandyteng/ACTFusionV5/db_fusionv5/Output_MANE_Select/202
50407_MANE.r47.transcript.loci >
/mnt/RD_Develop/sandyteng/ACTFusionV5/db_fusionv5/Output_Loci/250407/PA05
3_ACTFusionV5_PseudoIntron_MANE-v1.4_GENCODE-r47_capture-
v1.0_GRCh38.20250407.transcript.MANE.only.blastn.r47.loci

```