

# Nanoq: fast quality control for nanopore reads

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## Software

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## Summary

Nanopore sequencing is now routinely integrated in a variety of genomics applications, including whole genome assembly ([Jain et al., 2018](#)) and real-time infectious disease surveillance ([Meredith et al., 2020](#)). As a consequence, the amount of nanopore sequence data in the public domain has increased rapidly in the last few years. One of the first steps in any workflow is to assess the quality of reads and obtain basic summary statistics after basecalling raw nanopore signal, and to filter low quality reads. [NanoPack](#) (biopython parser) ([De Coster et al., 2018](#)), [Filtlong](#) ([Klib](#) parser) and [MinIONQC](#) (summary file parser) ([Lanfear et al., 2018](#)) are common tools used to filter and obtain summary statistics from nanopore reads. However, these tools can be relatively slow due to bottlenecks in read parsing ([NanoPack](#), [Filtlong](#)), are not immediately usable due to reliance on summary files ([MinIONQC](#)), or focus on data exploration and visualization. We therefore implement [nanoq](#), a command line tool to accelerate summary and quality control for nanopore reads in Rust.

## Statement of need

A common practice for quality control and filtering of reads for length and quality is to use a sequencing summary file as index to speed up iteration and computation over millions of individual reads and their precomputed metrics from the basecalling process (e.g. the main access mode for [MinIONQC](#)), which requires access to signal level data or shared summary files. With increasing throughput on scalable nanopore platforms like [GridION](#) or [PromethION](#), fast quality control of sequence reads and the ability to generate summary statistics on-the-fly are required. [Nanoq](#) is highly competitive in processing speed (see benchmarks) and can be effectively applied to nanopore data from the public domain, where sequencing summaries are unavailable, as part of automated pipelines, in streaming applications, or directly from the command line to check on the progress of active sequencing runs.

## Applications

[Nanoq](#) is implemented in Rust using the read parsers from [needletail](#) and [Rust-Bio](#) ([Köster, 2015](#)).

Tests can be run within the [nanoq](#) repository:

```
cargo test
```

[Nanoq](#) accepts a file or stream of sequence reads in `fast{a/q}` and compressed formats on `stdin`:

```
cat test.fq | nanoq
```

35 Basic summary statistics are output to stderr:

```
100000 400398234 5154 44888 5 4003 3256 8.90 9.49
```

- 36     ▪ number of reads
- 37     ▪ number of base pairs
- 38     ▪ N50 read length
- 39     ▪ longest and shorted reads
- 40     ▪ mean and median read length
- 41     ▪ mean and median read quality

42 Extended output analogous to NanoStat can be obtained using multiple `--detail` flags:

```
cat test.fq | nanoq -d -d -d
```

43 Reads filtered by minimum read length (`--length`) and mean read quality (`--quality`) are  
44 output to stdout:

```
cat test.fq | nanoq -l 1000 -q 10 > reads.fq
```

45 Advanced two-pass filtering analogous to `Filtlong` removes the worst 20% of bases using  
46 sorted reads by quality (`--keep_percent`) or the worst quality reads until approximately 500  
47 Mbp remain (`--keep_bases`):

```
nanoq -f test.fq -p 80 -b 500000000 > reads.fq
```

48 Live sequencing run data directory:

```
RUN=/data/nanopore/run
```

49 Check total run statistics of active run:

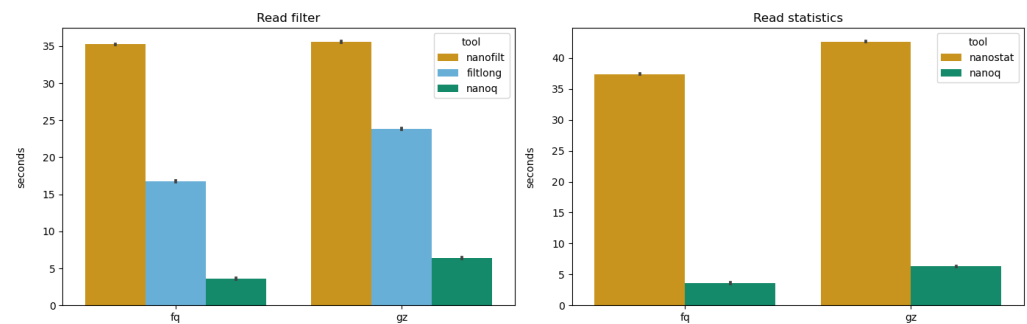
```
find $RUN -name *.fastq -print0 | xargs -0 cat | nanoq
```

50 Check per-barcode statistics of active run:

```
for i in {01..12}; do
  find $RUN -name barcode${i}.fastq -print0 | xargs -0 cat | nanoq
done
```

## 51 Benchmarks

52 Benchmarks evaluate processing speed of a long-read filter and computation of summary  
53 statistics on the first 100,000 reads (`test.fq.gz` in Docker container) of the even [Zymo](#)  
54 [mock community](#) ([Nicholls et al., 2019](#)) (GridION) using the `nanoq:v0.2.0` [Benchmark](#)  
55 image with comparison to [NanoFilt](#), [NanoStat](#) and [Filtlong](#)



**Figure 1:** Nanoq benchmarks compared to Filtlong and Nanopack on 100,000 reads of the Zymo mock community

program	ftype	task	mean sec (+/- sd)	~ reads / sec	speedup
nanofilt	fq	filter	35.25 (0.35)	2,836	1.00 x
filtlong	fq	filter	16.71 (0.47)	5,984	2.11 x
nanoq	fq	filter	03.63 (0.45)	27,548	9.71 x
nanostat	fq	stats	37.39 (0.50)	2,674	1.00 x
nanoq	fq	stats	03.57 (0.57)	28,011	10.4 x
nanofilt	fq.gz	filter	35.58 (0.36)	2,810	1.00 x
filtlong	fq.gz	filter	23.84 (0.60)	4,195	1.49 x
nanoq	fq.gz	filter	06.37 (0.41)	14,858	5.28 x
nanostat	fq.gz	stats	42.21 (0.37)	2,369	1.00 x
nanoq	fq.gz	stats	06.30 (0.28)	15,873	6.70 x

## Availability

Nanoq is open-source on GitHub (<https://github.com/esteinig/nanoq>) and available through:

- Cargo: `cargo install nanoq`
- Docker: `docker pull esteinig/nanoq`
- BioConda: `conda install -c bioconda nanoq`
- Singularity: `singularity pull docker://esteinig/nanoq`

Nanoq is integrated with [pipelines servicing research projects](#) at [Queensland Genomics](#) using nanopore sequencing to detect infectious agents in septic patients, reconstruct transmission dynamics of bacterial pathogens, and conduct outbreak sequencing at the Townsville University Hospital (QLD, Australia).

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