## Class 11: AlphaFold

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Here we read the results from AlphaFold and try to interpret all the models and quality score metrics:

```
library(bio3d)

pth <- "dimer_23119/"

pdb.files <- list.files(path = pth, full.names = TRUE, pattern</pre>
```

Align and supperpose all these models

```
file.exists(pdb.files)
```

[1] TRUE TRUE TRUE TRUE TRUE

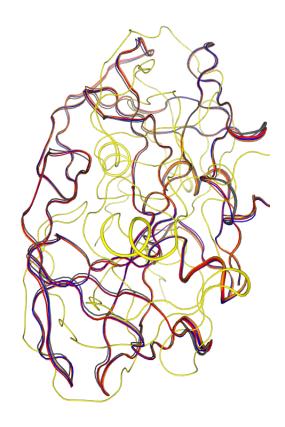
```
pdbs <- pdbaln(pdb.files, fit = TRUE, exefile="msa")</pre>
```

```
Reading PDB files:
dimer_23119//dimer_23119_unrelaxed_rank_001_alphafold2_multime
r_v3_model_2_seed_000.pdb
dimer_23119//dimer_23119_unrelaxed_rank_002_alphafold2_multime
r_v3_model_5_seed_000.pdb
dimer_23119//dimer_23119_unrelaxed_rank_003_alphafold2_multime
r_v3_model_4_seed_000.pdb
dimer_23119//dimer_23119_unrelaxed_rank_004_alphafold2_multime
r_v3_model_1_seed_000.pdb
dimer_23119//dimer_23119_unrelaxed_rank_005_alphafold2_multime
r_v3_model_3_seed_000.pdb
.....
```

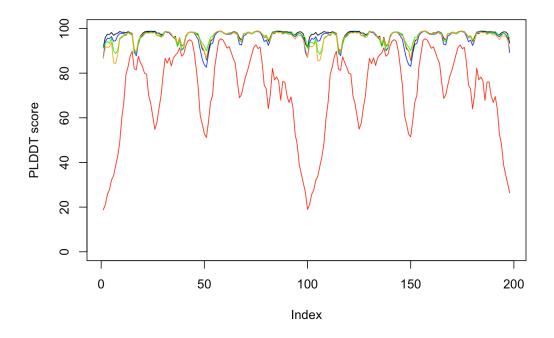
Extracting sequences

```
pdb/seq: 1    name:
dimer_23119//dimer_23119_unrelaxed_rank_001_alphafold2_multime
r_v3_model_2_seed_000.pdb
pdb/seq: 2    name:
dimer_23119//dimer_23119_unrelaxed_rank_002_alphafold2_multime
r_v3_model_5_seed_000.pdb
```

```
library(bio3dview)
view.pdbs(pdbs)
```



```
plot(pdbs$b[1,], typ="l", ylim=c(0,100), ylab="PLDDT score")
lines(pdbs$b[2,],typ="l", col="blue")
lines(pdbs$b[3,],typ="l", col="green")
lines(pdbs$b[4,],typ="l", col="orange")
lines(pdbs$b[5,],typ="l", col="red")
```

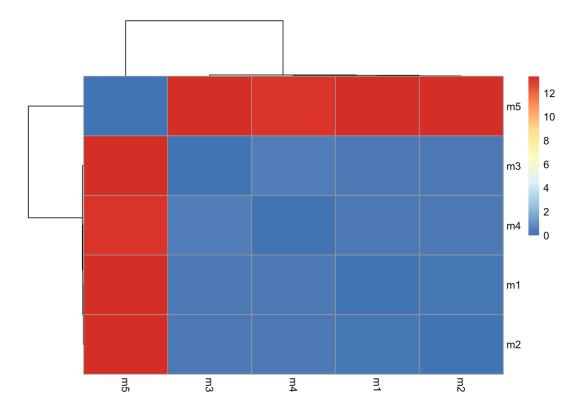


```
rd <- rmsd(pdbs)
```

Warning in rmsd(pdbs): No indices provided, using the 198 non NA positions

```
library(pheatmap)

colnames(rd) <- paste0("m",1:5)
rownames(rd) <- paste0("m",1:5)
pheatmap(rd)</pre>
```



```
pae1 <- read_json(pae_files[1],simplifyVector = TRUE)
pae2 <- read_json(pae_files[2],simplifyVector = TRUE)
pae3 <- read_json(pae_files[3],simplifyVector = TRUE)
pae4 <- read_json(pae_files[4],simplifyVector = TRUE)
pae5 <- read_json(pae_files[5],simplifyVector = TRUE)
attributes(pae1)</pre>
```

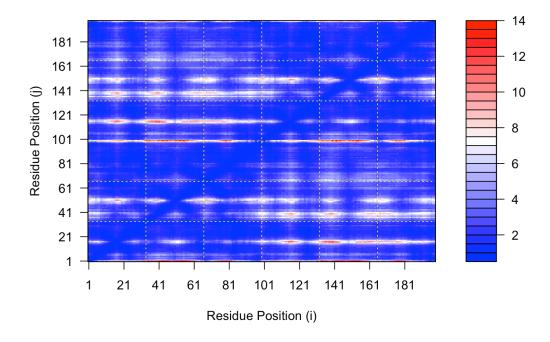
## \$names

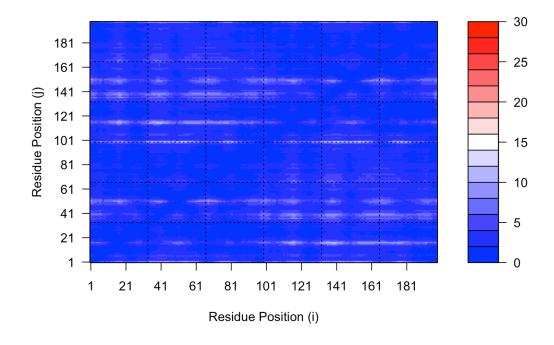
```
[1] "plddt" "max_pae" "pae" "ptm" "iptm"
```

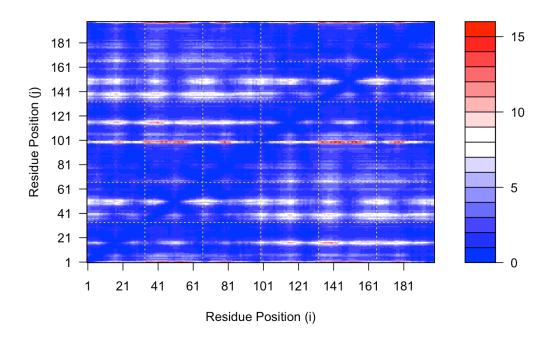
```
# Per-residue pLDDT scores
# same as B-factor of PDB..
head(pae1$plddt)
```

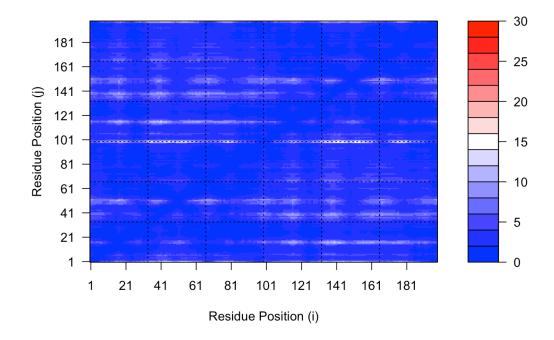
[1] 91.44 96.06 97.38 97.38 98.19 96.94

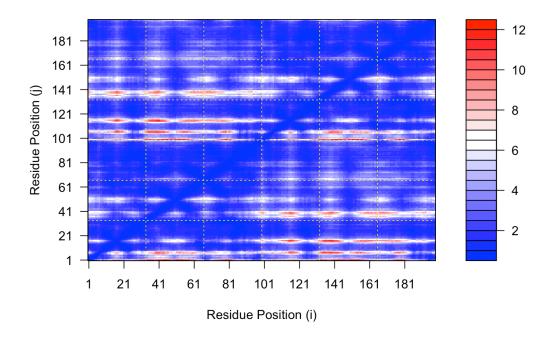
```
pae1$max_pae
[1] 13.57812
pae2$max_pae
[1] 15.71094
pae3$max_pae
[1] 12.41406
# Best (lowest) PAE score
pae4$max_pae
[1] 19.95312
pae5$max_pae
[1] 29.85938
plot.dmat(pae1$pae,
          xlab="Residue Position (i)",
          ylab="Residue Position (j)")
```

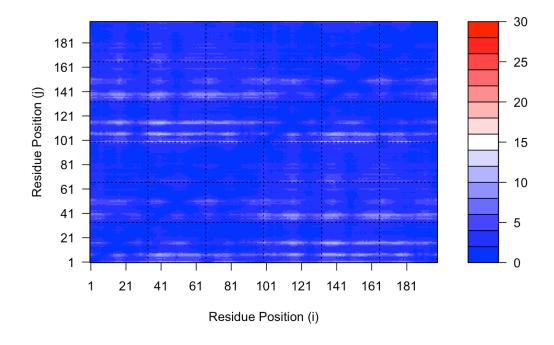


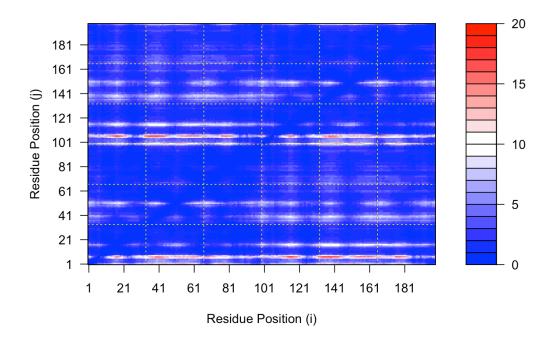


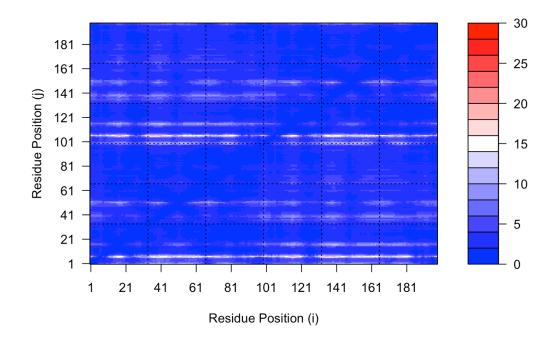


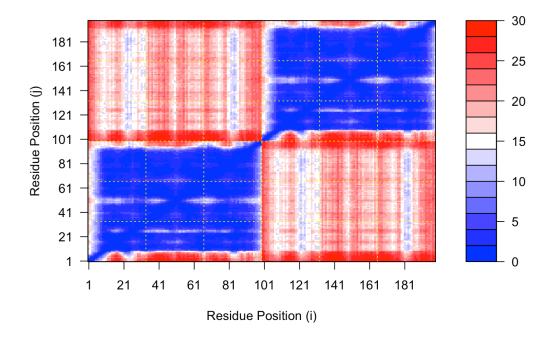


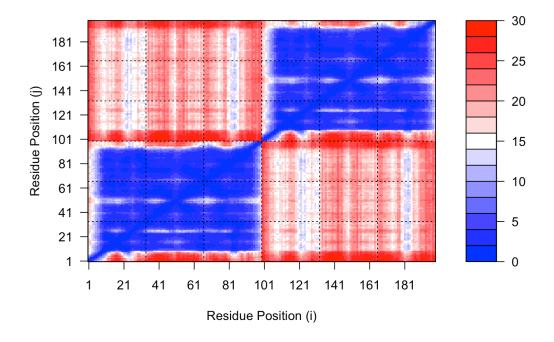












## Residue conservation from alignment file

[1] "dimer\_23119//dimer\_23119.a3m"

```
aln <- read.fasta(aln_file[1], to.upper = TRUE)</pre>
```

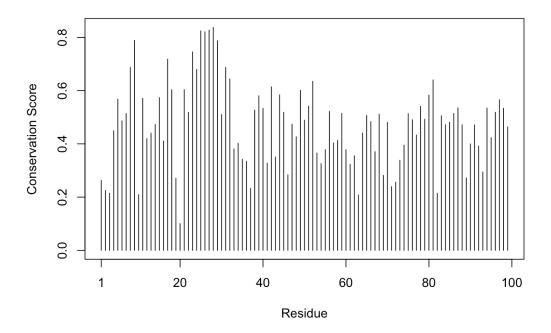
```
[1] " ** Duplicated sequence id's: 101 **"
[2] " ** Duplicated sequence id's: 101 **"
```

```
dim(aln$ali)
```

[1] 5378 132

```
sim <- conserv(aln)
```

```
plotb3(sim[1:99],
     ylab="Conservation Score")
```



```
con <- consensus(aln, cutoff = 0.9)
con$seq</pre>
```

- [19] "-" "-" "-" "-" "-" "D" "T" "G" "A" "-" "-" "-" "-"

[127] "-" "-" "-" "-" "-"

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```
m1.pdb <- read.pdb(pdb.files[1])
occ <- vec2resno(c(sim[1:99], sim[1:99]), m1.pdb$atom$resno)
write.pdb(m1.pdb, o=occ, file="m1_conserv.pdb")</pre>
```

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