Antibody binding experiment

- 1. Prepare 8ml fluorescein stock solution of concentration 1.2nM.
- 2. Prepare 1ml antibody 4-4-20 solution of concentration 12nM.
- 3. Take 0.5ml solution 2 into a new test tube. Add 0.5 ml buffer into the new test tube. Mix thoroughly.
- 4. Repeat step 3 for 10 times. Then we have 12 antibody solutions with concentration of a factor 2 less successively.
- 5. Put 0.5 ml fluorescein stock solution 1 into each of the 12 test tubes and mix.
- 6. Select an excitation wavelength of 470nm and an emission wavelength range of 500-600nm.
- 7. Measure the buffer and 0.6nM fluorescein emission.
- 8. Start with lowest concentration of antibody samples. Measure the emission spectrum.
- 9. For each emission spectrum, correct the background by subtracting the background spectrum. Then integrate the spectrum to get the total fluorescent intensity. Record the value of total intensity.
- 10. Analyze data:
 - a. Calculate the fluorescence quenching percentage of each fluoresceinantibody sample with respect to pure fluorescein

$$Q = \left(1 - \frac{I}{I}_{\text{Pure fluorescein}}\right) \times 100\% .$$

- b. Calculate the fluorescence ratio r between the completely bound fluorescein and free fluorescein $r = I_{\text{Completely Bound}} / I_{\text{Pure fluorescein}}$.
- c. Use the ratio r to calculate the free ligand concentration with the formular $(I I_{\text{completely bound}}) / I_{\text{Pure fluorescein}}.$ $[L] = [L]_T \frac{1 r}{1 r}.$
- d. Calculate the free antibody binding sites concentration by subtracting the total enzyme concentration by the bound antibody concentration $[ABS]_{Free} = [ABS]_{Total} ([L]_T [L]) \times 2$.
- e. Plot quenching Q as a function of $[ABS]_{\mathrm{Free}}$ and fit to the function

$$Q = \frac{a \times [ABS]_{Free}}{K_d + [ABS]_{Free}} + b \text{ to obtain } K_d.$$

Polarization Experiment

1. Measure the anisotropy of free fluorescein, partly bound fluorescein, and completely bound fluorescein.