

# **B. Tech. Project Report Phase II**

## **Deciphering Breast Cancer Dynamics: HOXB2 and MMP11 Insights**



Submitted in partial fulfilment of requirements for the award of the degree of  
Bachelor of Technology from IIT Guwahati

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## Certificate

This is to certify that the work presented in the report entitled “**Deciphering Breast Cancer Dynamics: HOXB2 and MMP11 Insights**” by Sarthak Ray (200106059), represents an original work under the guidance of **Prof. Anil Mukund Limaye, Department of Biosciences and Bioengineering**. This study has not been submitted elsewhere for a degree.

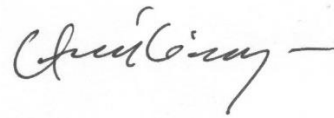
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Date: April 22, 2024

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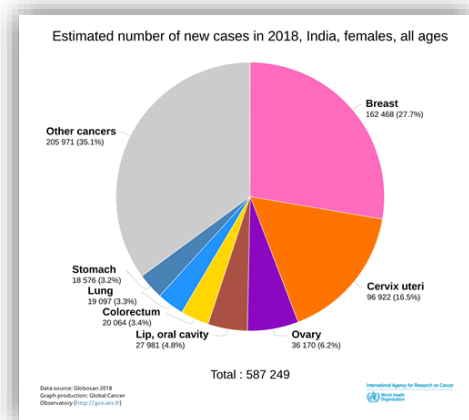
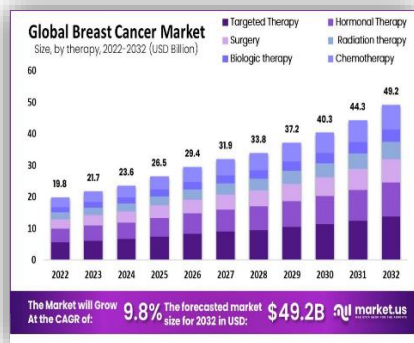
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# 1 Abstract

We delve deeper into the intricate landscape of breast cancer (BRCA) treatment by comprehensively analysing the HOXB2 and MMP11 genes' influence on ER alpha expressions. This study focuses on elucidating the specific impact of HOXB2 and MMP11 on the expression patterns of ER alpha. Through rigorous differential gene expression analysis utilising TCGA data, we aim to uncover novel insights that further refine our understanding of molecular signals in BRCA, ultimately enhancing the prospects for tailored therapeutic interventions and improved patient outcomes.

# 2 Introduction

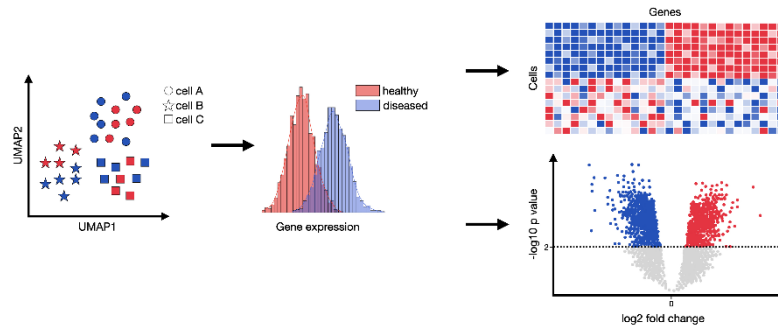
Breast cancer poses a serious threat to women's health around the world and is considered a powerful enemy in the field of global health. It is causing increasing concern due to its alarming 25% of all female malignancies. It is essential to understand the extent of breast cancer's impact on public health before diving into the disease's molecular complexity. Setting the stage for this investigation requires understanding the data and classifications.



Navigating the complexities of breast cancer involves understanding the intricate interplay between hormone expression and tumour behaviour, mainly influenced by estrogen and progesterone levels. These hormones, acting through their receptors, delineate distinct breast cancer subtypes, each presenting unique characteristics and therapeutic considerations. At the

molecular level, unravelling the intricate connections between hormone expression and tumour dynamics lays the groundwork for comprehending breast cancer pathogenesis.

Differential gene expression (DGE) analysis is a potent tool that sheds light on the molecular subtleties of the disease within this complex landscape. By closely examining the transcriptome landscape, DGE studies reveal minute changes in gene expression patterns, making it possible to identify unique genetic signatures linked to different breast cancer subtypes. Examining important genes like *HOXB2* and *MMP11* is essential to this effort because it provides information about how these genes affect the course of breast cancer and how well a treatment works. By carefully examining the dynamics of gene expression, we hope to expand our knowledge of the biology of breast cancer and open the door to more focused and efficient treatment approaches.



### 3. Literature Review:

A summary of the genes we are concerned with is as follows:

- **Homeobox (HOX) Genes:**

A family of transcription factors known as HOX genes is essential for tissue patterning, cell differentiation, and embryonic development. In breast cancer (BRCA), among other cancers, aberrant expression of HOX genes has been linked to tumour initiation, progression, and metastasis. *HOXB2* is one of the HOX genes that has become important in the pathophysiology of breast cancer. According to research, *HOXB2* expression is dysregulated in breast cancer tissues when compared to healthy breast tissue, and an aggressive tumour behaviour and unfavourable prognosis are linked to its overexpression. By modifying critical signalling pathways connected to cell proliferation, apoptosis

evasion, and the epithelial-mesenchymal transition (EMT), HOXB2 mechanistically stimulates tumour growth and invasion. Additionally, HOXB2 has been implicated in conferring resistance to endocrine therapy in estrogen receptor-positive (ER+) breast cancers, highlighting its clinical relevance as a therapeutic target.

- **Matrix Metalloproteinase (MMP) Genes:**

The zinc-dependent endopeptidases known as matrix metalloproteinases (MMPs) are essential modulators of the extracellular matrix (ECM) remodelling and are critical for the advancement and metastasis of cancer. MMP11, or stromelysin-3, is one of the MMP family members that has attracted much attention in breast cancer research because of its involvement in several tumour biology-related areas. When compared to normal breast tissue, MMP11 is frequently overexpressed in breast cancer tissues, and this overexpression is associated with advanced tumour stage, lymph node metastasis, and unfavourable patient outcomes. Through its ability to facilitate extracellular matrix degradation, enhance tumour cell migration and invasion, and modify the tumour microenvironment to support tumour growth and angiogenesis, MMP11 functionally promotes breast cancer invasion and metastasis. Moreover, MMP11 has been implicated in mediating resistance to chemotherapy and targeted therapies in breast cancer, underscoring its potential as a therapeutic target for overcoming treatment resistance and improving patient outcomes.

## **4. Objectives for Phase II:**

**To conduct Gene Expression analysis on BRCA data based on ER status and two specific genes, namely, HOXB2 and MMP11, elucidate the most correlated genes to the above, and document the entire analytical pipeline for comprehensive insights into targeted breast cancer treatments.**

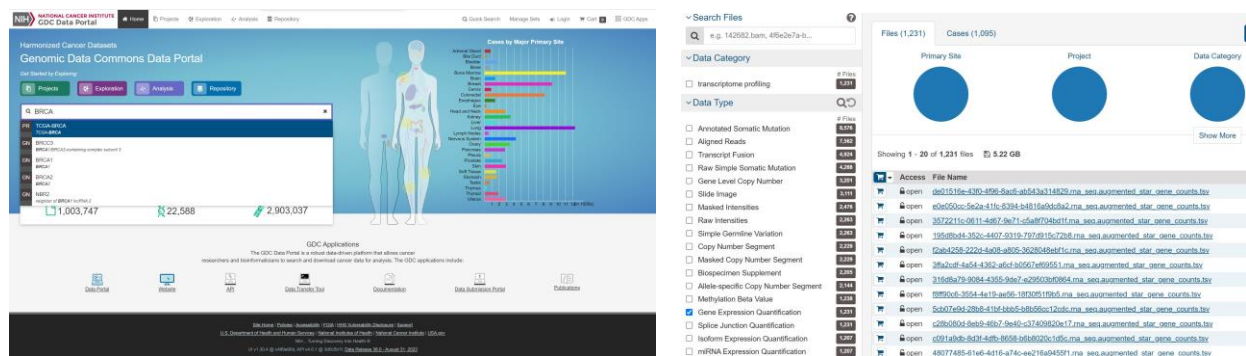
## **5. Materials and Methods**

The entire code for the following pipeline can be found [here](#).

### **5.1 Dataset for Analysis (TCGA)**

The Cancer Genome Atlas (TCGA) project catalogues the genetic mutations responsible for cancer using genome sequencing and bioinformatics. This joint effort between NCI and the National Human Genome Research Institute began in 2006, bringing together researchers from diverse disciplines and multiple institutions. The steps to obtain datasets are as follows:

- Visit <https://portal.gdc.cancer.gov/> and look for TCGA-BRCA in the search bar.
- Go to Explore Project Data.
- Select **Gene Expression Quantification** in the filter and download all the tsv files.



### 5.1.1 Count Data

The count data in the TCGA BRCA dataset represent the quantification of gene expression levels through RNA sequencing. This dataset is organised with genes as rows and samples as columns. The basis of the DGE analysis is these counts. This results from creating a data frame from all the tsv files' unstranded sequence counts. The resulting data frame looks as follows:

GeneIds	GeneNames	TCGA-A8-A086-01A	TCGA-D8-A	TCGA-AN-I
ENSG00000000003.15	TSPAN6	4263	4370	2443
ENSG00000000005.6	TNMD	9	7	144
ENSG000000000419.13	DPM1	2071	2625	2322
ENSG000000000457.14	SCYL3	1101	3005	1466
ENSG000000000460.17	C1orf112	717	1578	409
ENSG000000000938.13	FGR	312	599	1179
ENSG000000000971.16	CFH	2840	4864	11555
ENSG000000001036.14	FUCA2	2812	1944	2770
ENSG000000001084.13	GCLC	4188	1958	2260
ENSG000000001167.14	NFYA	2886	4597	2448
ENSG000000001460.18	STPG1	770	669	750
ENSG000000001461.17	NIPAL3	3410	3220	1922
ENSG000000001497.18	LAS1L	3809	3766	2862
ENSG000000001561.7	ENPP4	1029	3504	2457
ENSG000000001617.12	SEMA3F	7335	4516	3711
ENSG000000001626.16	CFTR	164	12	12

### 5.1.2 Clinical Data

The clinical data in TCGA BRCA complements genomic information by providing essential insights into patients' demographic, clinical, and pathological characteristics. This dataset encompasses a range of variables, including patient age, gender, tumour stage, hormone receptor status (ER/PR), HER2 status, survival status, and other relevant clinical annotations. Clinical data is indispensable in classifying patient samples into meaningful groups for DGE analysis, allowing researchers to investigate how gene expression patterns correlate with specific clinical features. The data frame looks as follows:

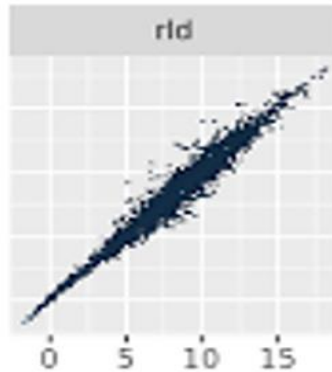
sampleID	AJCC_Stage	Age_at_Diag	CN_Cluster	Converted	Days_to_D	Days_to_d	ER_Status	Gender_nz	HER2_Fina	Integrated
TCGA-3C-AAAU-01										
TCGA-3C-AALI-01										
TCGA-3C-AALK-01										
TCGA-4H-AAAK-01										
TCGA-5L-AATO-01										
TCGA-5L-AAT1-01										
TCGA-ST-A9QA-01										
TCGA-A1-A Stage I		70	1 Stage I		259		Positive	FEMALE	Negative	
TCGA-A1-A Stage IIA		59	2 Stage IIA		437		Positive	FEMALE	Negative	
TCGA-A1-A Stage I		56	2 Stage I		1320		Positive	FEMALE	Negative	
TCGA-A1-A Stage IIA		54	3 Stage IIA		1463		Positive	FEMALE	Negative	
TCGA-A1-A Stage IIB		61	4 Stage IIB		433		Positive	FEMALE	Negative	
TCGA-A1-A Stage IIA		39	5 Stage IIA		1437		Negative	FEMALE	Negative	1
TCGA-A1-A Stage IIB		52	3 Stage IIB		634		Positive	FEMALE	Negative	
TCGA-A1-A Stage IIIA		39	3 Stage IIIA		426		Positive	FEMALE	Negative	1
TCGA-A1-A Stage IIA		54	1 Stage IIA		594	967	Negative	FEMALE	Negative	2
TCGA-A1-A Stage IIA		77	2 Stage IIA		242		Positive	MALE	Positive	
TCGA-A1-A Stage IIIA		50	5 Stage IIA		1196		Positive	FEMALE	Positive	
TCGA-A1-A Stage IIB		67	1 Stage IIB		852		Negative	FEMALE	Negative	2
TCGA-A1-A Stage IIA		40	Stage IIA		583		Negative	FEMALE	Negative	

## 5.2 Transformations on Variance

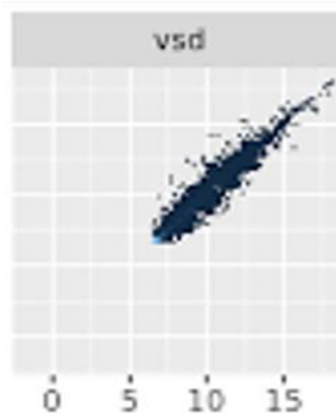
Following data preprocessing to address sequencing depth, gene length, and RNA composition biases, transformations on variance are applied to ensure robust and accurate evaluation of gene expression changes across samples. These transformations are essential because raw RNA-seq counts typically exhibit non-constant variance across the range of expression levels, violating the assumptions of many statistical methods. Three commonly used methods are:

- One commonly used method for variance stabilisation is the **regularised log transformation (rlog)** provided by the DESeq2 package. The rlog transformation stabilises the variance across the mean expression level, effectively normalising the data and making it more amenable to downstream analyses such as clustering and differential expression testing. It also has a regularised component that helps with small sample sizes.

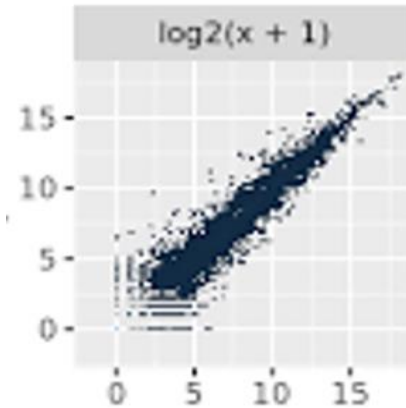




- Another widely used transformation is the **variance stabilising transformation (VST)**, also available in DESeq2. VST is based on a similar principle of stabilising the variance across the mean expression level but may be preferred in certain scenarios where the data exhibit specific characteristics. It models the relationship between the mean and variance of gene expression.

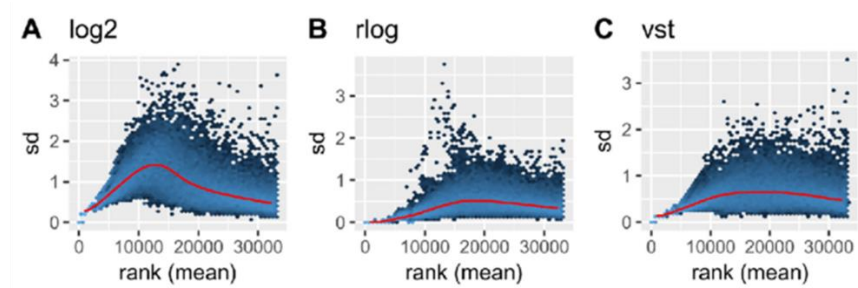


- Additionally, a common gene expression analysis practice is applying a **log2 transformation** to the normalised counts. This transformation helps in data visualisation and interpretation by compressing the dynamic range of expression values, making fold changes more intuitive and facilitating sample comparisons. The log2 transformation is beneficial for identifying differentially expressed genes and visualising expression patterns across experimental conditions.



Here, we use Variance Stabilising transformation (VST) using a simple line of code in R.

```
# Perform variance stabilizing transformation directly
vst_data <- varianceStabilizingTransformation(expression_matrix)
```



### 5.3 Data Analysis

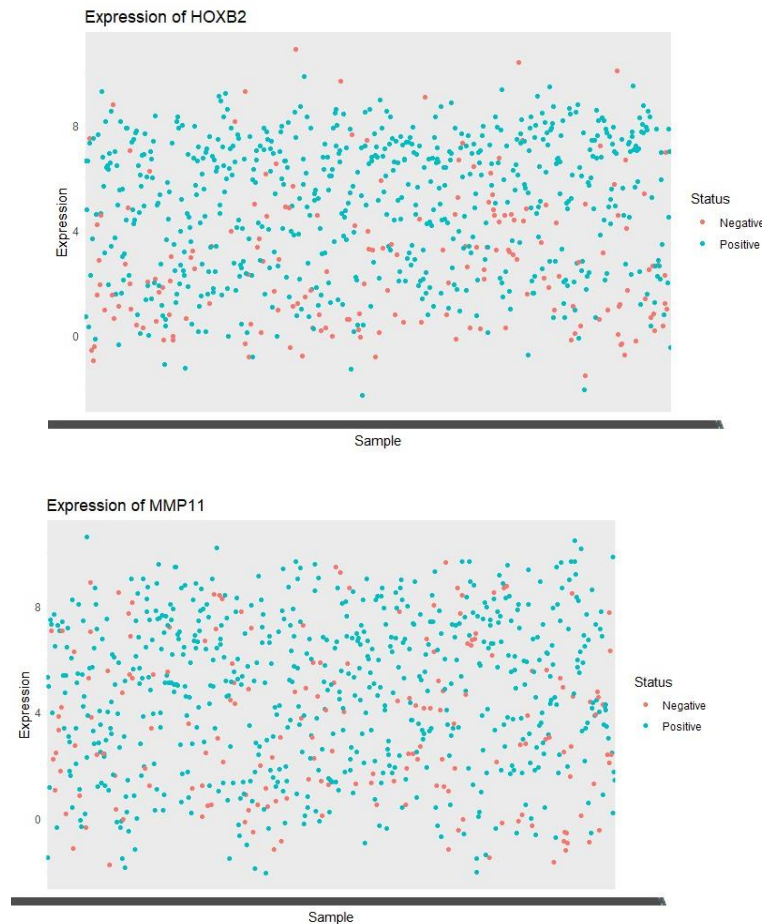
- Extract MMP and HOX gene expression data from the dataset. Split the data into two groups based on ER status: ER-positive (ER+) and ER-negative (ER-) samples.

```
hox_genes <- final_data_filtered[grepl("^HOX", final_data_filtered$GeneNames), ]
mmp_genes <- final_data_filtered[grepl("^MMP", final_data_filtered$GeneNames), ]
```

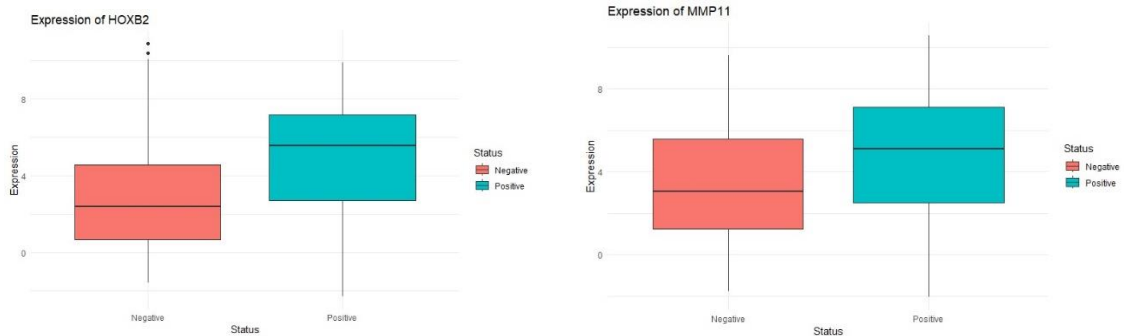
```
transposed_data <- t(hox_genes[, -1]) # Exclude the column with gene names
# Set the transposed gene names as column names
colnames(transposed_data) <- hox_genes$GeneNames

transposed_data <- data.frame(transposed_data)
transposed_data$status <- er_status_filtered$ER_status
```

- Generate dot plots to visualise the expression levels of MMP and HOX genes in ER+ versus ER- samples. Each dot represents the expression level of a gene in a specific sample, with ER+ and ER- samples plotted separately. This visualisation method quickly assesses gene expression patterns and differences between the two ER status groups.



- Draw box plots for each MMP and HOX gene, comparing expression levels between ER+ and ER- samples. Box plots provide a visual summary of the distribution of expression levels within each ER status group and facilitate the identification of potential differences.



- Calculate the correlation coefficients between each MMP and HOX gene and all other genes in the dataset. Correlation analysis provides insights into potential co-expression patterns and regulatory relationships between MMP, HOX, and other genes. Examining the correlation matrix makes it possible to identify genes highly correlated with MMP and HOX genes, which may indicate functional associations or shared regulatory mechanisms.

```
# Iterate over each gene and compute correlation with status
for (gene in colnames(all_gene_expression_data)) {
  # Create a dataframe for the current gene
  gene_data_b <- data.frame(Expression = all_gene_expression_data[, gene], MMP11 = mmp_data[, 4])

  # Compute correlation between status and gene expression
  correlation_value <- cor.test(gene_data_b$Expression, gene_data_b$MMP11)$estimate
  p_value <- cor.test(gene_data_b$Expression, gene_data_b$MMP11)$p.value

  # Append results to the data frame
  correlation_results_b2 <- rbind(correlation_results_b2, data.frame(Gene = gene,
                                                                    MMP11_correlation = correlation_value, p_value = p_value))
}
```

- Calculate the correlation coefficients between each MMP and HOX gene against the ER-alpha status. This will help us to find the relationship between the gene expression and whether or not the ER-alpha status is positive or negative.

```
# Iterate over each gene and compute correlation with status
for (gene in colnames(gene_expression_data_b)) {
  # Create a dataframe for the current gene
  gene_data_b <- data.frame(Expression = gene_expression_data_b[, gene], Status = status_column_b)

  # Convert 'Status' to a binary variable
  gene_data_b$Status_binary <- ifelse(gene_data_b$Status == "Positive", 1, 0)

  # Compute correlation between status and gene expression
  correlation_value <- cor.test(gene_data_b$Expression, gene_data_b$Status_binary)$estimate

  # Append results to the data frame
  correlation_results_b <- rbind(correlation_results_b, data.frame(Gene = gene, Correlation = correlation_value))
}
```

## 5.4 Additional analyses

Some other analyses can be further done on this data. They are as follows:

- **Pathway Enrichment Analysis:** Perform pathway enrichment analysis to identify biological pathways enriched among differentially expressed genes. This analysis can reveal the functional roles of MMP and HOX genes in breast cancer and provide insights into the underlying biological processes driving differential expression.
- **Machine Learning Classification:** Employ machine learning algorithms to build predictive models for breast cancer subtype classification based on MMP and HOX gene expression profiles. Evaluate the performance of these models using cross-validation techniques and assess the predictive power of MMP and HOX genes compared to other clinical and molecular features.
- **Network Analysis:** Employ machine learning algorithms to build predictive breast cancer subtype classification models based on MMP and HOX gene expression profiles. Evaluate the performance of these models using cross-validation techniques and assess the predictive power of MMP and HOX genes compared to other clinical and molecular features.

## 6.0 Results

We obtain lists of the most correlated genes with HOXB2 and MMP11. We also find the correlation between ER-alpha status and all the HOX and MMP genes.

- Most correlated genes with **MMP11**.

### Positive correlation

	Gene	MMP11_Correlation	p_value
cor2140	MMP11	1.0000000	0.000000e+00
cor3339	AEBP1	0.7566974	4.799297e-147
cor20275	CYS1	0.7526983	1.185813e-144
cor10077	MMP14	0.7448981	4.090054e-140
cor15298	NTM	0.7425146	9.233677e-139
cor5393	COL10A1	0.7357444	5.368370e-135
cor19660	PLPP4	0.7267734	3.488142e-130
cor12628	ANTXR1	0.7217749	1.385867e-127
cor7537	ITGA11	0.7191409	3.079714e-126
cor5315	PLAU	0.7047463	3.836250e-119
cor821	COL11A1	0.7035819	1.377453e-118

### Negative correlation

	Gene	MMP11_Correlation	p_value
cor60395	AL353135.2	-0.2066402	4.885291e-09
cor4969	BCL11A	-0.1941567	4.009122e-08
cor8826	PM20D2	-0.1923320	5.392412e-08
cor52193	AC006946.2	-0.1907413	6.966206e-08
cor7394	IL33	-0.1810804	3.150676e-07
cor32475	SOX9.AS1	-0.1784314	4.700337e-07
cor3264	MINDY4	-0.1780676	4.963530e-07
cor8094	TAF4B	-0.1774811	5.417902e-07
cor58760	AL356776.2	-0.1771041	5.730895e-07
cor26192	CADM3.AS1	-0.1759890	6.761626e-07
cor5643	SOX9	-0.1715314	1.296235e-06
cor52277	AC027449.1	-0.1695686	1.717372e-06

- Most correlated genes with **HOXB2**.

### Positive correlation

	Gene	HOXB2_Correlation	p_value
cor13605	HOXB2	1.0000000	0.000000e+00
cor5007	HOXB3	0.6955738	7.672181e-115
cor29231	HOXB.AS1	0.6852758	3.340457e-110
cor15312	HOXB4	0.5347363	2.065520e-59
cor47806	AC036222.1	0.4901554	8.414913e-49
cor35191	HOXB.AS2	0.4605725	1.406403e-42
cor39971	RNU6.863P	0.4522896	6.081057e-41
cor12038	PRR15L	0.3933254	1.612232e-30
cor42633	TRIM51DP	0.3864863	1.936574e-29
cor25017	AC092648.1	0.3826213	7.690841e-29
cor5005	HOXB5	0.3815634	1.118233e-28
cor7274	CALCOCO2	0.3575664	3.789903e-25

### Negative correlation

	Gene	HOXB2_Correlation	p_value
cor9098	FAM171A1	-0.2069920	4.594922e-09
cor15179	RGMA	-0.2049490	6.548909e-09
cor15619	PRKX	-0.1983764	1.997970e-08
cor730	FOXC1	-0.1967956	2.598209e-08
cor9274	CNKSR2	-0.1939109	4.173111e-08
cor9776	SRSF12	-0.1938799	4.194263e-08
cor28218	SNHG26	-0.1931562	4.718270e-08
cor2496	FERMT1	-0.1902332	7.556586e-08
cor9048	ZNF462	-0.1889767	9.231639e-08

- Most correlated **MMP** genes with ER-alpha.

	Gene	Correlation
cor21	MMP17	0.394946922
cor26	MMP28	0.317519449
cor15	MMP16	0.221780837
cor14	MMP21	0.217782440
cor8	MMP24OS	0.200604585
cor2	MMP11	0.179296855
cor17	MMP10	0.174744585
cor18	MMP26	0.108382471

- Most correlated **HOX** genes with ER-alpha.

	Gene	Correlation
cor36	HOXC4	0.257563687
cor39	HOXD.AS2	0.233971335
cor35	HOXC6	0.233523924
cor30	HOXD8	0.214376085
cor28	HOXC5	0.211013256
cor13	HOXB1	0.180793264
cor37	HOXB.AS1	0.163592588
cor33	HOXB4	0.146686814
cor12	HOXB3	0.143990653
cor32	HOXC10	0.128821505
cor40	HOXB.AS2	0.128064329
cor11	HOXB5	0.121584884
cor1	HOXC8	0.120477640
cor29	HOXB2	0.118674907

## 7.0 Conclusion & Future work

To sum up, our examination of the HOXB2 and MMP11 genes in breast cancer has revealed their complex relationships with other genes. We have also shed light on the relation between HOX and MMP genes to ER status, highlighting their importance in developing the disease and hormone receptor status. These results deepen our understanding of the biology of breast cancer and could lead to customised treatment plans depending on ER status. To optimise therapeutic interventions and enhance patient outcomes, these results must be validated in larger cohorts, mechanistic studies must be carried out to clarify underlying molecular mechanisms, and their prognostic and predictive value must be investigated.

## 8.0 References

- <https://portal.gdc.cancer.gov/projects/TCGA-BRCA>
- <https://bioconductor.org/packages/devel/bioc/vignettes/DESeq2/inst/doc/DESeq2.html>
- [https://lashlock.github.io/compbio/R\\_presentation.html](https://lashlock.github.io/compbio/R_presentation.html)
- <http://www.sthda.com/english/wiki/rna-seq-differential-expression-work-flow-using-deseq2>
- <https://www.blog.trainindata.com/variance-stabilizing-transformations-in-machine-learning/>
- <https://www.ncbi.nlm.nih.gov/gene/3212>
- <https://www.ncbi.nlm.nih.gov/gene/4320>