

Prediction of the Effect of Single Amino Acid Protein Variants Using Deep Mutational Scanning Data

University of Bologna — Master Thesis in Bioinformatics

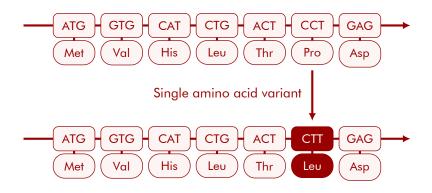
Pierotti Saul

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External Advisor: Prof. Arne Elofsson (Stockholm University)

Single Amino Acid Variant

A mutation that replaces exactly one amino acid in a protein



Deep Mutational Scanning

A technique for obtaining fitness information on a large number of mutations

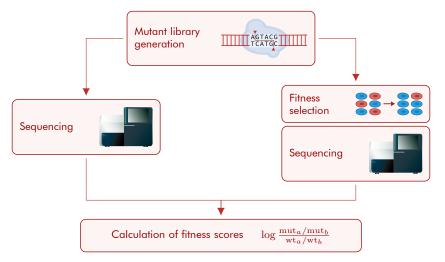


Image sources: restriction_enzyme icon by Simon Dürr; Illumina_miseq icon by DBCLS; modification of stem_cell_colony icon by Marcel Tisch. Full credits in appendix.

Predicting the Effect of Mutations

Why is it useful?

- Targeted medical treatments
- ► Protein engineering



Why is it needed?

- Experiments are insufficient
- ► Experiments are expensive





How can it be done?

- ▶ Machine learning
- ► Statistical models





Image sources: modification of patient icon by Marcel Tisch; zincfinger icon by Simon Dürr; Illumina_miseq icon by DBCLS; sand-clock icon by Servier; ai icon by Simon Dürr; sigmoid icon by Simon Dürr. Full credits in appendix.

Previous Work in the Field

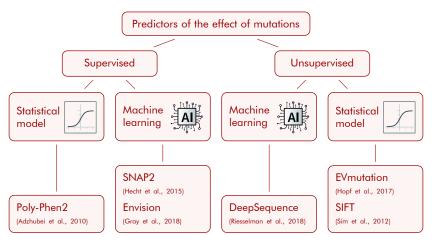
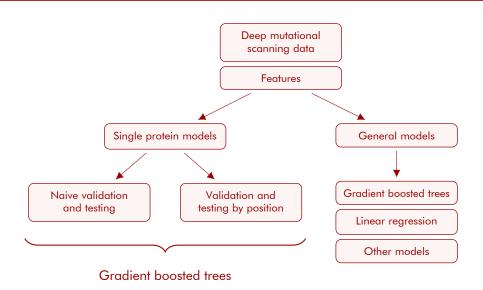


Image sources: ai icon by Simon Dürr; sigmoid icon by Simon Dürr. Full credits in appendix.

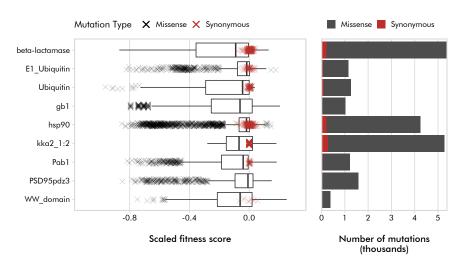
Among these predictors only Envision was trained on deep mutational scanning data

My Approach



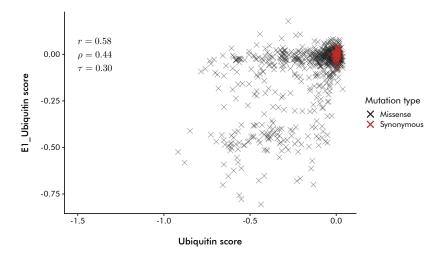
Training Data

I borrowed the training dataset of the predictor Envision (Gray et al., 2018)

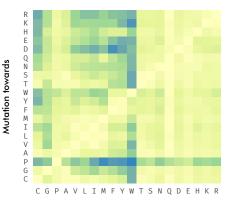


Experiments Do Not Agree Much with Each Other

Two independent deep mutational scanning experiments on Ubiquitin are present in the training dataset. Their correlation is low.

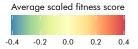


Interesting Patterns in Mutation Sensitivity



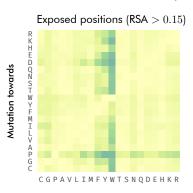
- ➤ Polar residues seem more tolerant to mutations than hydrophobic residues
- Proline (P) is the most disruptive residue
- ► Tryptophan (W) is hard to replace

Mutation from

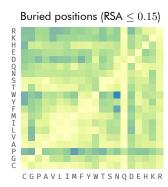


Exposure Explains the Mutability of Polar Residues

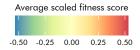
When filtering by Relative Solvent Accessibility (RSA) apolar residues are **not** more sensitive to mutations than polar residues



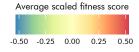




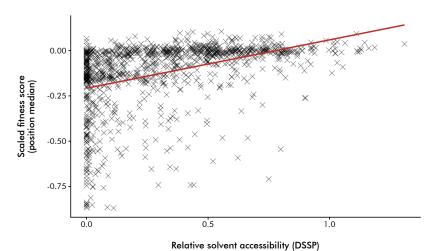






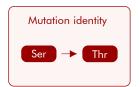


Buried Residues Are More Conserved



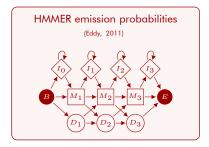
Features Used by the Predictors

All the features are derived from the sequences: I did **not** use any structural information



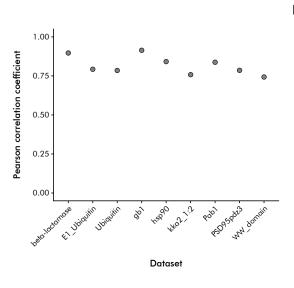








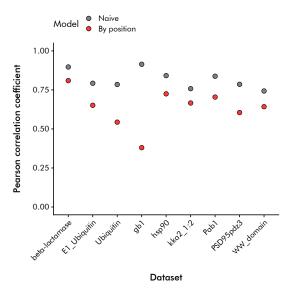
Single Protein Models



Naive approach

- A different model trained for each protein
- ► Half of the mutations used for testing and half for cross-validation
- ▶ Too good to be true

Single Protein Models



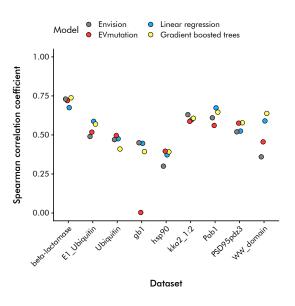
Naive approach

- ► A different model trained for each protein
- Half of the mutations used for testing and half for cross-validation
- ► Too good to be true

Segregating protein positions

- Same as above but mutations in the same position segregated in the training or testing sets
- ► Performances are more realistic

Leave-One-Protein-Out (LOPO) Models



- Models trained on the whole dataset while leaving one protein out
- For the left-out protein, half of the mutations used for testing and half for validation
- Spearman correlation coefficient used for evaluation

Discussion and Future Directions

What I learned

- ▶ The testing strategy is crucial
- Good performances without structural features
- Strong variability between datasets
- ► Complex models not necessarily better

Ideas for the future

- Using residue contacts in a graph convolutional neural network
- ➤ Training on more deep mutational scanning studies
- ► Finding a better normalization strategy





Image sources: personal creation; hell-birne-karriere-klettern-licht by mohamed_hassan. Full credits in appendix.

Questions?



Image sources: hombre-pensamiento-silueta-4484226 by mohamed hassan. Full credits in appendix.

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Images Credits

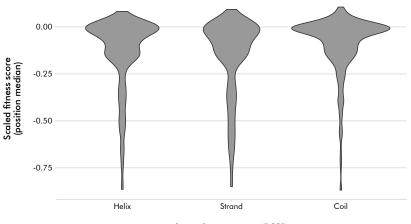
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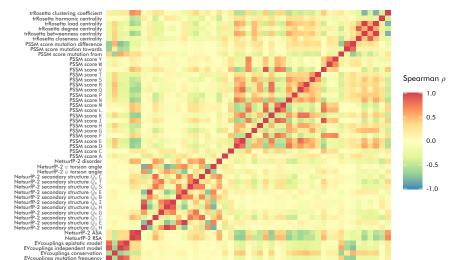
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Secondary structure is of limited importance in the discrimination of damaging mutations



Secondary structure (DSSP)

The correlation among features follows predictable patterns



Precision of trRosetta (Yang et al., 2020) in predicting residue contacts

Dataset	Medium-range ($s \geq 12$)			Long-range ($s \geq 24$)		
	Top $L/5$	Top $L/2$	Top L	Top $L/5$	Top $L/2$	Top L
beta-lactamase	1.00	0.92	0.86	0.96	0.93	0.76
WW domain	0.95	0.90	0.83	0.90	0.87	0.75
PSD95pdz3	0.96	0.92	0.80	0.92	0.81	0.70
kka2 1:2	1.00	1.00	0.96	1.00	1.00	0.89
hsp90	1.00	1.00	0.96	1.00	1.00	0.89
Ubiquitin	0.98	0.92	0.82	1.00	0.90	0.70
Pab 1	0.80	0.72	0.67	0.87	0.74	0.60
E1 Ubiquitin	0.82	0.86	0.77	0.91	0.75	0.54
gb1	1.00	0.85	0.46	0.63	0.40	0.22

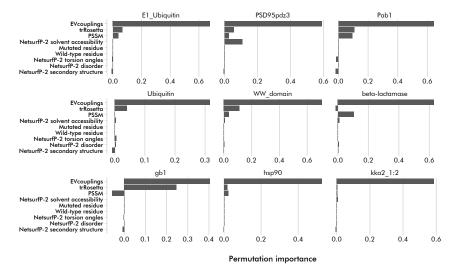
Quality of the predicted structural features from NetsurfP-2 (Klausen et al., 2019)

Feature	Evaluation metric	Score
Relative solvent accessibility	Pearson r	0.79
Accessible surface area	Pearson r	0.80
Q ₃ secondary structure	Q_3 accuracy	0.85
Q_8 secondary structure	Q_{8} accuracy	0.72
φ torsion angle	Circular correlation	0.73
ψ torsion angle	Circular correlation	0.87

Relationship between the features used in the models and the fitness scores

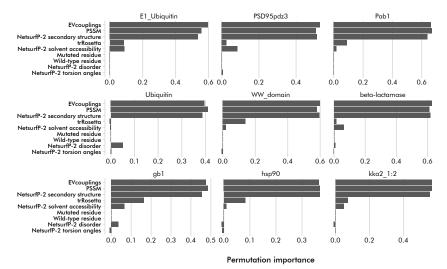
Feature	Pearson r	Spearman $ ho$	Kendall τ
PSSM mutation score	-0.29	-0.28	-0.19
Netsurf predicted RSA	0.34	0.37	0.25
Netsurf predicted ASA	0.32	0.35	0.24
Netsurf predicted disorder	0.06	0.18	0.12
EVcouplings epistatic model	0.46	0.50	0.34
EVcouplings independent model	0.44	0.44	0.30
EVcouplings frequency	0.19	0.35	0.24
EVcouplings conservation	-0.32	-0.33	-0.23
Closeness centrality (trRosetta predicted contacts)	-0.16	-0.17	-0.11
Betweenness centrality (trRosetta predicted contacts)	-0.20	-0.29	-0.19
Degree centrality (trRosetta predicted contacts)	-0.12	-0.13	-0.09
Load centrality (trRosetta predicted contacts)	-0.20	-0.29	-0.19
Harmonic centrality (trRosetta predicted contacts)	-0.19	-0.20	-0.14
Clustering coefficient (trRosetta predicted contacts)	0.23	0.25	0.17
			Linear-circular correlation
Netsurf predicted ϕ torsion angle			0.01
Netsurf predicted ψ torsion angle			0.02
	Kruskal-Wallis χ^2		p-value
Wild-type residue	1482.40		< 2.20 · 10 ⁻¹⁶
Mutated residue	708.53		< 2.20 · 10 - 16
Netsurf predicted Q_3 secondary structure	215.33		< 2.20 · 10 - 16
Netsurf predicted Q_8 secondary structure	351.97		< 2.20 · 10 ⁻¹⁶

Feature importances for the gradient boosted tree general models



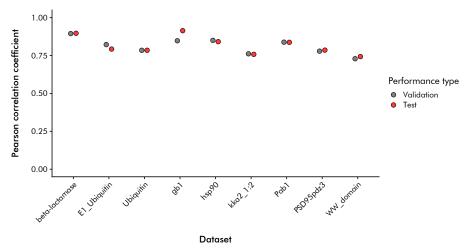
Feature group

Feature importances for the linear regression general models

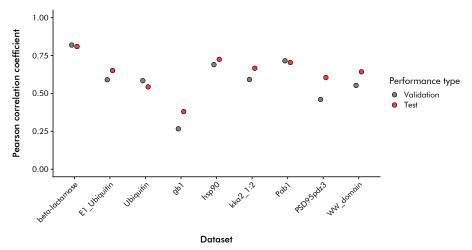


Feature group

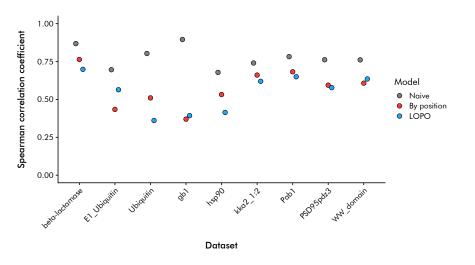
Validation and testing performances for the single protein models trained with the naive approach



Validation and testing performances for the single protein models trained by segregating protein positions



Comparison of the performances of single protein models and general models



Confidence intervals in prediction performances

Dataset name	Model	95 % C.I. (Pearson)	95 % C.I. (Spearman)
beta-lactamase	Naive	0.89 to 0.91	0.86 to 0.88
beta-lactamase	By position	0.79 to 0.83	0.75 to 0.78
beta-lactamase	LOPO	_	0.68 to 0.72
WW domain	Naive	0.67 to 0.82	0.70 to 0.84
WW domain	By position	0.57 to 0.73	0.52 to 0.72
WW domain	LÓPO	_	0.56 to 0.73
PSD95pdz3	Naive	0.74 to 0.83	0.73 to 0.80
PSD95pdz3	By position	0.55 to 0.67	0.54 to 0.65
PSD95pdz3	LÓPO	_	0.53 to 0.63
kka2 1:2	Naive	0.74 to 0.78	0.72 to 0.76
kka2 1:2	By position	0.65 to 0.69	0.64 to 0.68
kka2 1:2	LÓPO	_	0.60 to 0.64
hsp90	Naive	0.82 to 0.87	0.65 to 0.71
hsp90	By position	0.69 to 0.76	0.50 to 0.57
hsp90	LÓPO	_	0.38 to 0.45
Ubiquitin	Naive	0.75 to 0.83	0.78 to 0.83
Ubiquitin	By position	0.49 to 0.60	0.46 to 0.57
Ubiquitin	LÓPO	_	0.30 to 0.43
Pab 1	Naive	0.80 to 0.87	0.75 to 0.82
Pab1	By position	0.65 to 0.76	0.64 to 0.73
Pab1	LÓPO	_	0.60 to 0.70
E1 Ubiquitin	Naive	0.75 to 0.85	0.65 to 0.75
E1 Ubiquitin	By position	0.59 to 0.72	0.36 to 0.51
E1 Ubiquitin	LOPO	_	0.50 to 0.63
gb1	Naive	0.90 to 0.93	0.88 to 0.92
gb1	By position	0.31 to 0.46	0.29 to 0.45
gb1	LOPO	_	0.32 to 0.47

Statistical significance of performance differences. Starred values are significant with Bonferroni correction.

$$\alpha = \frac{0.05}{27} = 0.00185185$$

Dataset name	Model 1	Model 2	$p ext{-value}$
beta-lactamase	Linear regression	Gradient boosted trees	1 · 10-4 *
beta-lactamase	Linear regression	EVmutation	1 · 10 - 4 *
beta-lactamase	EVmutation	Gradient boosted trees	1 · 10 - 4 *
WW domain	Linear regression	Gradient boosted trees	1 · 10 - 4 *
WW_domain	Linear regression	EVmutation	1 · 10 - 4 *
WW domain	EVmutation	Gradient boosted trees	1 · 10-4 *
PSD95pdz3	Linear regression	Gradient boosted trees	1 · 10 - 4 *
PSD95pdz3	Linear regression	EVmutation	1 · 10-4 *
PSD95pdz3	EVmutation	Gradient boosted trees	0.51
kka2_1:2	Linear regression	Gradient boosted trees	0.00
kka2_1:2	Linear regression	EVmutation	0.01
kka2_1:2	EVmutation	Gradient boosted trees	1 · 10 - 4 *
hsp90	Linear regression	Gradient boosted trees	1 · 10 - 4 *
hsp90	Linear regression	EVmutation	1 · 10 - 4 *
hsp90	EVmutation	Gradient boosted trees	0.24

The table continues on the next slide

Statistical significance of performance differences. Starred values are significant with Bonferroni correction.

$$\alpha = \frac{0.05}{27} = 0.00185185$$

The table continues from the previous slide

Dataset name	Model 1	Model 2	$p ext{-value}$
Ubiquitin	Linear regression	Gradient boosted trees	1 · 10-4 *
Ubiquitin	Linear regression	EVmutation	1 · 10 - 4 *
Ubiquitin	EVmutation	Gradient boosted trees	1 · 10-4 *
Pab1	Linear regression	Gradient boosted trees	1 · 10 - 4 *
Pab1	Linear regression	EVmutation	1 · 10 - 4 *
Pab1	EVmutation	Gradient boosted trees	1 · 10 - 4 *
E1 Ubiquitin	Linear regression	Gradient boosted trees	1 · 10-4 *
E1 Ubiquitin	Linear regression	EVmutation	1 · 10 - 4 *
E1 Ubiquitin	EVmutation	Gradient boosted trees	1 · 10-4 *
gb1	Linear regression	Gradient boosted trees	1 · 10-4 *
gb1	Linear regression	EVmutation	1 · 10-4 *
gb1	EVmutation	Gradient boosted trees	1 · 10 - 4 *