

Methods

Input format and preprocessing

We analyze a codon alignment in PHYLIP sequential format. The first line declares the number of taxa and sites; subsequent blocks consist of a taxon name line followed by wrapped sequence lines. Sequences are uppercased on read. Characters outside {A,C,G,T} (e.g., N, ?, -) are treated as missing and excluded from per-site calculations. The script verifies that all sequences have equal length and that the total length is divisible by three.

Per-site and pooled composition statistics

For each alignment column (nucleotide site), we count observed A/C/G/T across taxa after discarding missing characters. Sites with fewer than two observed bases are ignored for entropy computations. We also compute pooled base frequencies across the entire matrix (all taxa × sites), again ignoring missing values; these frequencies (p_A , p_C , p_G , p_T) are used to parameterize expectations under full substitution saturation (FSS).

Entropy and the Index of Substitution Saturation (Iss)

Per-site Shannon entropy is computed as:

$$H = - \sum_{b \in \{A, C, G, T\}} p_b \log p_b$$

where p_b is the observed frequency of base b at the site (natural log units). We average over usable sites to obtain H . Following Xia's formulation, the script defines:

$$Iss = \frac{\tilde{H}}{H_{FSS}}$$

where H_{FSS} is the expected per-site entropy under full substitution saturation given the number of taxa and the pooled base frequencies.

Exact expectation under full substitution saturation

To obtain H_{FSS} and its variance, the script evaluates the exact multinomial expectation over all possible site compositions for N taxa. Specifically, for all non-negative integer quadruples (n_A, n_C, n_G, n_T) with $\sum n_b = N$, it weights the entropy of the composition:

$$-\sum_b \binom{n_b}{N} \log \left(\frac{n_b}{N} \right)$$

by the multinomial probability:

$$\frac{N!}{\prod_b n_b!} \prod_b p_b^{n_b}$$

It also accumulates $E[H^2]$ to report $\text{Var}(H_{FSS}) = E[H^2] - E[H]^2$. This yields the theoretical mean and variance of per-site entropy under FSS without simulation. Computationally this is $O(N^3)$ states; practical for typical N .

Position-specific Iss

To characterize heterogeneity across codon positions, sites are partitioned by position (1st, 2nd, 3rd) and their mean entropies are normalized by the same H_{FSS} to yield $Iss_{pos1/2/3}$.

Codon-level contributions

To localize saturation-like behavior, the script computes a codon contribution score for each codon k . It averages the entropies of its three nucleotide sites, then normalizes by H_{FSS} :

$$\text{Iss_contrib}_k = \frac{\frac{1}{3} \sum_{i \in \{3k-2, 3k-1, 3k\}} H_i}{H_{\text{FSS}}}$$

Higher values indicate codons whose within-column variability is closer to the FSS expectation.

Species (taxon) contributions via leave-one-out

To identify taxa that inflate saturation, the script performs leave-one-taxon-out (LOTO) analyses.

For each taxon t , it recomputes Iss on the reduced alignment and reports:

$$\Delta \text{Iss}_t = \text{Iss}_{(-t)} - \text{Iss}_{\text{full}}$$

Large positive ΔIss indicates that removing t reduces saturation (i.e., t had been pushing Iss upward).

Monte Carlo FSS check (optional)

As an empirical complement to the exact expectation, the script can run a Monte Carlo test by simulating independent sites under FSS: for each site, it draws N nucleotides i.i.d. with the pooled base frequencies, computes the site entropy, and averages over the number of usable sites to obtain H_{sim} . Repetition yields an empirical distribution of H under FSS; the script reports $p_{\text{hi}} = P(H_{\text{sim}} \geq H_{\text{obs}})$ and $p_{\text{lo}} = P(H_{\text{sim}} \leq H_{\text{obs}})$. Small p_{lo} indicates the observed mean entropy is well below FSS (i.e., unsaturated).

Thresholding and filtered alignment output

Two user-supplied thresholds enable targeted cleaning: Codon filter: drop codons with $\text{Iss_contrib} \geq \tau_{\text{codon}}$. Taxon filter: drop taxa with $\Delta \text{Iss} \geq \tau_{\text{taxon}}$. Filters can be applied separately or jointly.

When filtering, entire codons (all three sites) are removed to preserve the reading frame. The script writes PHYLIP-sequential alignments for the codon-filtered, taxon-filtered, and combined-filtered datasets, using the same wrapping width as the input. Manifests listing removed codons and taxa are also written.

Outputs

A summary table reporting N, total and usable sites, H, H_FSS, Var(H_FSS), I_{ss}, I_{ss_pos1/2/3}, and pooled base frequencies. A codon-wise table of I_{ss_contrib} (and flags, if thresholded). A taxon-wise table of ΔI_{ss} (and flags, if thresholded). Optional Monte Carlo FSS results (p_{hi}, p_{lo}). Optional filtered PHYLIP alignments ready for downstream inference.

How this differs from DAMBE—and why the results remain valid

DAMBE implements Xia's I_{ss} test together with critical thresholds I_{ss_c} (for symmetric vs. asymmetric trees and varying numbers of taxa), derived from extensive simulations, and uses those I_{ss_c} tables to make a formal decision ("significantly saturated or not"). This script (i) computes I_{ss} with the same core definition H/H_FSS; (ii) obtains H_FSS and Var(H_FSS) by exact multinomial expectation given the observed base composition and taxon count; and (iii) provides an empirical FSS Monte Carlo check rather than consulting I_{ss_c} tables tied to tree-shape assumptions. Thus, while it does not label results using DAMBE's I_{ss_c} decision framework, its I_{ss} values are directly comparable and its "distance from FSS" assessments are grounded in the same entropy logic that underpins I_{ss}. Moreover, the script adds diagnostically useful localization (codon contributions) and influence (leave-one-out taxa) analyses that aid targeted data curation prior to phylogenetic inference.