

# Reconstructing transmission trees from genetic data: a Bayesian approach

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## The model, in a nutshell

We seek a simple probabilistic model allowing to reconstruct the transmission tree (who infected whom) of disease outbreaks based on RNA/DNA sequences sampled at given time points. This model is designed for densely sampled outbreaks of diseases with fairly short (epidemiological) generation times and moderate genetic diversity (typically, genomes should accumulate zero, one or maybe two mutations per generation of infection). For instance, the method should be relevant for influenza, but HIV is clearly out of the scope of the approach.

The model is inspired by *SeqTrack* in some of the key assumptions it makes:

- within-host evolution is considered negligible and mutations only occur during transmission events
- a single pathogen is considered for each patient (no multi-infection, no within-host diversity)
- reverse mutations are negligible

However, our model aims at improving *SeqTrack* in several respects:

- a Bayesian framework allowing parameter estimation and incorporating prior information
- the use of the generation time to compute the likelihood (cf Wallinga & Teunis)
- the ability to accommodate unobserved cases
- the incorporation of infection dates in the transmission model (as augmented data)
- the ability to incorporate multiple index cases

In a first approach, we assume that the generation time follows a known distribution. This could be relaxed in a more complex model where parameters of this distribution would be estimated. The elements we aim to infer are the transmission tree and the mutation rates.

## Data and parameters

### Data

For each patient  $i = 1, \dots, n$  we note the data:

- $s_i$ : the genetic sequence obtained for patient  $i$ .
- $t_i$ : the collection time for  $s_i$  (time is considered as a discrete variable).

### Augmented data

Augmented data are noted using capital latin letters:

- $T_i^{inf}$ : time at which patient  $i$  has been infected.

### Functions

We use the following functions of the data/augmented data:

- $d(s_i, s_j)$ : the number of transitions between  $s_i$  and  $s_j$ .
- $g(s_i, s_j)$ : the number of transversions between  $s_i$  and  $s_j$ .
- $l(s_i, s_j)$ : the number of nucleotide positions typed in both  $s_i$  and  $s_j$ .
- $w(\Delta_t)$ : generation time distribution (likelihood function for a secondary infection occurring  $\Delta_t$  unit times after the primary infection); we assume  $w(\Delta_t) = 0$  for  $\Delta_t \leq 0$ ; while not a requirement in theory, in practice this function will be truncated at a value  $\Delta_{max}$  so that  $w(\Delta_t) = 0$  if  $\Delta_t \geq \Delta_{max}$ .
- $f_w$ : a function of the generation time distribution ( $w$ ) indicating how likely it is to sequence an isolate at a given time after infection. By default, we set  $f_w = w$ , so that the probability of sequencing an isolate is proportional to the infectiousness of the host at the time of collection.

### Parameters

This model assumes that cases are ordered by increasing infection dates ( $T_i^{inf} \leq T_{i+1}^{inf}$ ). Parameters are indicated using greek letters:

- $\alpha_i$ : the closest observed ancestor of  $i$  in the infection tree;  $\alpha_i = j$  indicates that  $j$  has infected  $i$ , either directly, or with one or several intermediate generations, which were unobserved.  $\alpha_i = 0$  indicates that the infection was imported from the outside (by definition,  $\alpha_1 = 0$ ). We note the tree topology  $\alpha = \{\alpha_1, \dots, \alpha_n\}$ .
- $\kappa_i$ : an integer  $\geq 1$  indicating how many generations separate  $\alpha_i$  and  $i$ :  $\kappa_i = 1$  indicates that  $\alpha_i$  infected  $i$ ;  $\kappa_i = 2$  indicates that  $\alpha_i$  has infected an unobserved individual, who has in turn infected  $i$ . We note  $\kappa = \{\kappa_1, \dots, \kappa_n\}$ .
- $\mu_1$ : rates of transitions, given per site and per transmission event.
- $\mu_2$ : rate of transversions, parametrised as  $\mu_2 = \gamma\mu_1$  (with  $\gamma \in \mathbb{R}_+$ ) to account for the correlation between the two rates.
- $\pi$ : an hyperparameter corresponding to the proportion of observed cases, used to define the priors of  $\kappa_i$ .
- $\phi$ : an hyperparameter corresponding to the probability for an observed case to have been imported from the outside, used to define the priors on  $\alpha_i$ .

## Model

### Likelihood

The posterior distribution is proportional to the joint distribution:

$$p(\{s_i, t_i, T_i^{inf}\}_{(i=1, \dots, n)}, w, \alpha, \kappa, \mu_1, \gamma, \pi, \phi) \quad (1)$$

$$= p(\{s_i, t_i, T_i^{inf}\}_{(i=1, \dots, n)} | w, \alpha, \kappa, \mu_1, \gamma, \pi, \phi) \times p(w, \alpha, \kappa, \mu_1, \gamma, \pi, \phi) \quad (2)$$

where the first term is the likelihood of observed and augmented data, and the second, the prior. The likelihood can be decomposed as (the hyperparameters  $\pi$  and  $\phi$  are dropped as they only affect the priors on  $\kappa$  and  $\alpha$ ):

$$p(\{s_i, t_i, T_i^{inf}\}_{(i=1, \dots, n)} | w, \alpha, \kappa, \mu_1, \gamma) \quad (3)$$

$$= \prod_{i=2}^n p(s_i, t_i, T_i^{inf} | \{s_k, t_k, T_k^{inf}\}_{(k=1, \dots, i-1)}, w, \alpha, \kappa, \mu_1, \gamma) \times p(s_1, t_1, T_1^{inf} | w) \quad (4)$$

$$= \prod_{i=2}^n p(s_i, t_i, T_i^{inf} | s_{\alpha_i}, t_{\alpha_i}, T_{\alpha_i}^{inf}, w, \alpha_i, \kappa_i, \mu_1, \gamma) \times p(s_1, t_1, T_1^{inf} | w) \quad (5)$$

$$= \prod_{i=2}^n p(s_i, t_i, T_i^{inf} | s_{\alpha_i}, t_{\alpha_i}, T_{\alpha_i}^{inf}, w, \alpha_i, \kappa_i, \mu_1, \gamma) \times p(t_1 | T_1^{inf}, w) p(T_1^{inf}) p(s_1) \quad (6)$$

$p(t_1 | T_1^{inf}, w)$  is the probability of the first collection time given the first infection time. The term  $p(T_1^{inf}) p(s_1)$  is the probability of the first infection date and of the first sequence, treated as a constant. The term for case  $i$  ( $i = 2, \dots, n$ ) is:

$$p(s_i, t_i, T_i^{inf} | s_{\alpha_i}, t_{\alpha_i}, T_{\alpha_i}^{inf}, w, \alpha_i, \kappa_i, \mu_1, \gamma) \quad (7)$$

which can be decomposed into:

$$\begin{aligned} & p(s_i | t_i, T_i^{inf}, s_{\alpha_i}, t_{\alpha_i}, T_{\alpha_i}^{inf}, w, \alpha_i, \kappa_i, \mu_1, \gamma) \\ & \times p(t_i | T_i^{inf}, s_{\alpha_i}, t_{\alpha_i}, T_{\alpha_i}^{inf}, w, \alpha_i, \kappa_i, \mu_1, \gamma) \\ & \times p(T_i^{inf} | s_{\alpha_i}, t_{\alpha_i}, T_{\alpha_i}^{inf}, w, \alpha_i, \kappa_i, \mu_1, \gamma) \\ & = \underbrace{p(s_i | \alpha_i, s_{\alpha_i}, \kappa_i, \mu_1, \gamma)}_{\Omega_i^1} \times \underbrace{p(t_i | T_i^{inf}, w) p(T_i^{inf} | \alpha_i, T_{\alpha_i}^{inf}, \kappa_i, w)}_{\Omega_i^2} \end{aligned} \quad (8)$$

where  $\Omega_i^1$  is the genetic likelihood and  $\Omega_i^2$  if the epidemiological likelihood (derived from W&T).

As mutations only occur during transmission events, the expected divergence between two isolates is determined by the number of generations separating these two isolates, and  $\Omega_i^1$  is computed as (cf Kimura 1980):

$$\underbrace{\mathcal{B}(d(s_i, s_{\alpha_i}) | l(s_i, s_{\alpha_i}) \kappa_i, \mu_1)}_{\text{transitions}} \times \underbrace{\mathcal{B}(g(s_i, s_{\alpha_i}) | l(s_i, s_{\alpha_i}) \kappa_i, \gamma \mu_1)}_{\text{transversions}} \quad (9)$$

$\mathcal{B}(\cdot | n, p)$  is the probability mass function of a Binomial distribution with  $n$  draws and a probability  $p$ . This is approximated by:

$$\underbrace{\mathcal{P}(d(s_i, s_{\alpha_i}) | l(s_i, s_{\alpha_i}) \kappa_i \mu_1)}_{\text{transitions}} \times \underbrace{\mathcal{P}(g(s_i, s_{\alpha_i}) | l(s_i, s_{\alpha_i}) \kappa_i \gamma \mu_1)}_{\text{transversions}} \quad (10)$$

where  $\mathcal{P}(\cdot|\lambda)$  is the density of a Poisson distribution of parameter  $\lambda$ .

In the absence of genetic information (including external infections where  $\alpha_i = 0$ ),  $\Omega_i^1 = 1$ .

$\Omega_i^2$  is determined by the distribution of the generation time, and the dates of collection and infection:

$$\begin{aligned}\Omega_i^2 &= p(t_i|T_i^{inf}, w) \times p(T_i^{inf}|\alpha_i, T_{\alpha_i}^{inf}, \kappa_i, w) \\ &= f_w(t_i - T_i^{inf}) \times w^{(\kappa_i)}(T_i^{inf} - T_{\alpha_i}^{inf})\end{aligned}\tag{11}$$

where the first term is the likelihood of the collection date, and the second, the likelihood of the infection time.  $w^{(k)} = \underbrace{w * w * \dots * w}_{k \text{ times}}$ , where  $*$  denotes the convolution operator, defined, for two positive discrete

distributions  $a$  and  $b$ , by  $(a * b)(t) = \sum_{u=0}^t a(t-u)b(u)$ .

In the case of external infections ( $\alpha_i = 0$ ),  $\Omega_i^2$  is simply computed as:

$$\Omega_i^2 = p(t_i|T_i^{inf}, w) = f_w(t_i - T_i^{inf})\tag{12}$$

## Priors

For all model parameters, independent prior distributions have been chosen:

- $p(w) = \mathbf{1}_{\{w=w_0\}}$ : the distribution of the generation time will be fixed to a given distribution ( $w_0$ ) by default; this can be parameterized later in a more complex model.
- $p(\alpha_i) = \mathbf{1}_{\{k=0\}}\phi + \mathbf{1}_{\{k \neq i, k > 0\}}\frac{1-\phi}{n-1}$ , where  $\phi$  is the probability of cases imported from the outside.  $\phi$  will be assumed to follow a Beta distribution of fixed parameters (e.g.  $p(\pi) = \beta(1.5, 10)$  to allow for around 10% of external infections).
- $p(\kappa_i - 1) = \mathcal{NB}(1, 1 - \pi)$ , the probability mass function of a negative binomial distribution counting the number of unobserved cases before one observed case;  $1 - \pi$  is the proportion of unobserved (unsampled) cases in the outbreak. If we assume that the entire outbreak has been sampled,  $\pi = 1$  and  $p(\kappa_i) = \mathbf{1}_{\{\kappa_i=1\}}$ . Otherwise,  $\pi$  will be assumed to follow a Beta distribution of fixed parameters (e.g.  $p(\pi) = \beta(10, 1)$  for an a priori densely sampled outbreak).
- $p(\mu_1) = \text{Unif}(0, 1)$ .
- $p(\gamma) = \log\mathcal{N}(1, 1.25)$  where  $\log\mathcal{N}(\mu, \sigma)$  is the log-normal distribution with mean  $\mu$  and standard deviation  $\sigma$  (values are the default in BEAST).