

# SBML Level 3: an extensible format for the exchange and reuse of biological models

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## Abstract

Systems biology has experienced dramatic growth in the number, size and complexity of computational models. To reproduce simulation results and reuse models, researchers must exchange unambiguous model descriptions. We review the latest edition of SBML (the Systems Biology Markup Language), a format designed for this purpose. A community of modelers and software authors developed SBML Level 3 over the past decade. Its modular form consists of a core suited to representing reaction-based models, and packages that extend the core with features suited to other model types including constraint-based models, reaction-diffusion models, logical network models, and rule-based models. The format leverages two decades of SBML and a rich software ecosystem that transformed how systems biologists build and interact with models. More recently, the rise of multiscale models of whole cells and organs, and new data sources such as single cell measurements and live imaging, have precipitated new ways of integrating data with models. We provide our perspectives on the challenges presented by these developments and how SBML Level 3 provides the foundation needed to support this evolution.

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# Introduction

Systems modeling and numerical simulations in biology can be traced to the mid-20<sup>th</sup> century. Though general theorizing about systems began earlier, the application of systems analysis to biology gained attention in the 1950's thanks to the work of biologists such as von Bertalanffy and Kacser (Kacser, 1957; Von Bertalanffy, 1950). The era of numerical simulation in biology truly began with the landmark works of Chance on enzyme kinetics (Chance *et al*, 1940), Hodgkin and Huxley on the molecular basis of neuronal transmission (Hodgkin and Huxley, 1952), and Turing on the chemical basis of morphogenesis (Turing, 1952). Since then, the number and variety of models have grown in all of the life sciences. As precise descriptions of phenomena that can be simulated, analyzed, and compared to experimental data, models provide unique insights that can confirm or refute hypotheses, suggest new experiments, and identify refinements to the models.

The availability of more data, more powerful modeling methods, and dramatically increased computing power led to the rise of systems biology as a compelling research theme around the turn of the millennium (Ideker *et al*, 2001; Kitano, 2000). Though computational models were at first published as printed equations in journal articles, the desire to reuse an ever-increasing number of models called for digital formats that were interoperable between software systems and could be easily exchanged between scientists (topics of interest as early as the 1960's; c.f. Garfinkel, 1969). This drove efforts to create tool-independent ways of representing models that could avoid the potential for human translation errors, be stored in databases, and provide a common starting point for simulations and analyses regardless of the software used (Goddard *et al*, 2001; Hucka *et al*, 2001; Lloyd *et al*, 2004). One such effort was SBML, the Systems Biology Markup Language. Its initial design was motivated by discussions to create a "metabolic model file format" following a 1999 workshop (recounted by Kell and Mendes, 2008). A distributed community thereafter discussed ideas that informed work at Caltech in late 1999/early 2000 and led (after a series of public drafts) to the specification of the official version of SBML Level 1 Version 1 being released in March 2001 (Hucka *et al*, 2003).

While SBML was initially developed to exchange compartmental models of biochemical reaction networks primarily formulated in terms of chemical kinetics (Hucka *et al*, 2001), it was always understood that there existed more types of models than the initial version of SBML could represent explicitly. However, seeking community consensus on a limited set of simpler features, which could be readily implemented in software at the time, was deemed a more pragmatic strategy. A deliberate decision was taken to delay the addition of more advanced capabilities to a later time. As a result, SBML has evolved in stages in a community-driven fashion that has benefited from the efforts of many researchers worldwide over two decades. As time passed, the need to support a broader range of model types, modeling frameworks, and research areas became apparent. SBML's success in serving as an interchange format for basic types of models led communities of modelers to ask whether it could be adapted or expanded to support more types. In addition to reaction-diffusion models, alternative modeling frameworks have risen in popularity in the past decade (Machado *et al*, 2011), and researchers have faced interoperability problems between software tools developed for their use. These needs drove a profound change in SBML's structure: a facility to permit layering the core of SBML with new features suited to more types of models, together with a way for individual models to identify which sets of extensions they need for proper interpretation. The release of SBML Level 3 (Hucka *et al*, 2010) has provided a new foundation to enable the exchange of a greater variety of models in various domains of biology (Figure 1).

In the rest of this article, we begin by summarizing SBML's general structure, then describe the modularity introduced in Level 3 and the wide range of modeling formalisms supported by Level 3 packages. We follow that by describing the community aspects of SBML development. We continue with a discussion of SBML's impact on both computational modeling and the modeling community, and finally, we close with a discussion of forthcoming challenges.

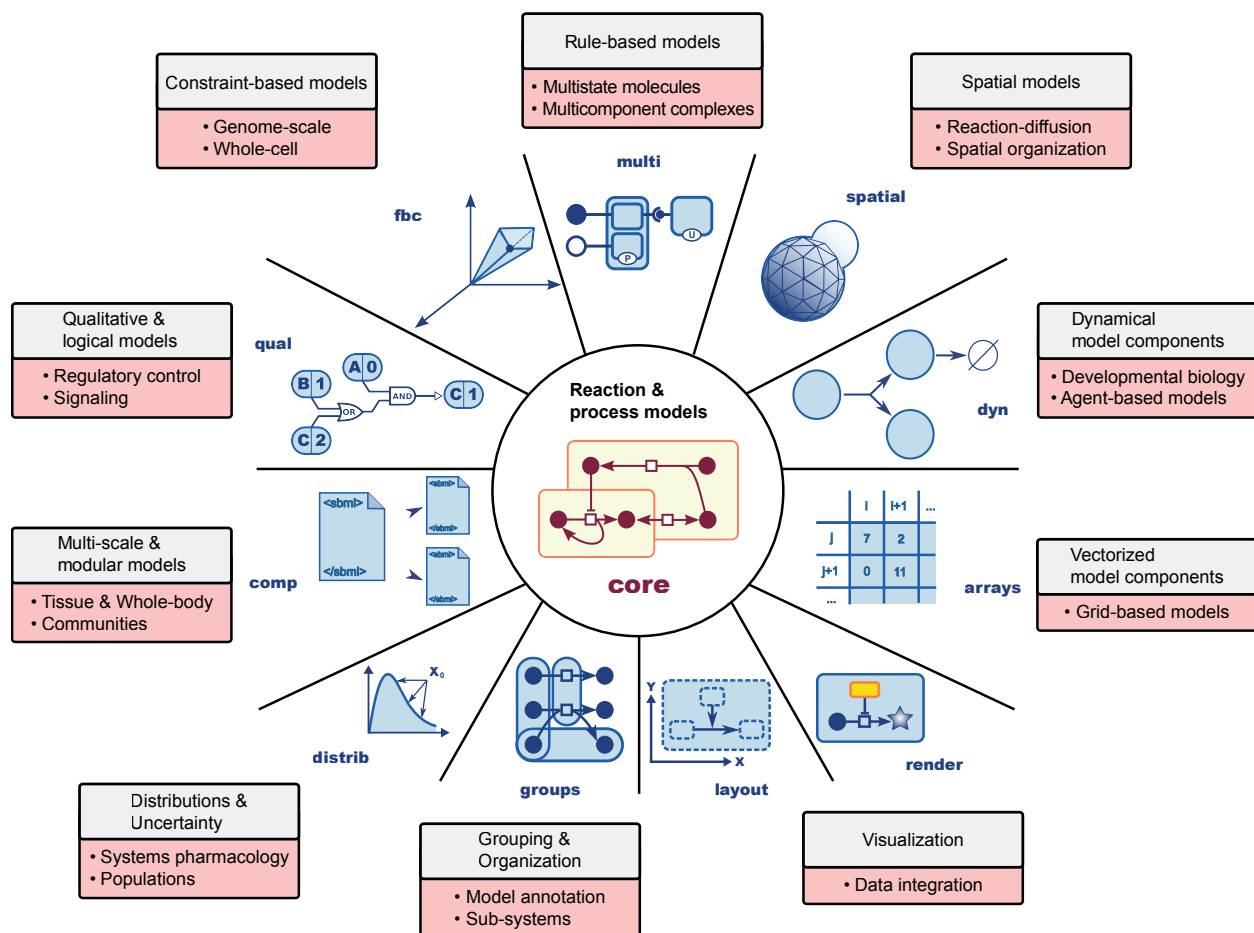


Fig 1: SBML Level 3 (Hucka et al, 2019) consists of a core (center) and specialized SBML Level 3 packages (in blue), which provide syntactical constructs to support additional modeling approaches. The packages support new types of modeling (in the gray boxes) needed for large and complex models such as those used in various domains and fields of biology (in the light red boxes). The meanings of SBML package labels such as “fbc” are given in Table 1, with additional package information in Box 1.

## The structure of SBML

The core of SBML is focused on encoding models in which entities are located in containers and are acted upon by processes that modify, create or destroy entities. The containers do not need to correspond to physical structures; they can be conceptual or abstract. Additional constructs allow parameters, initial conditions, other variables, and other mathematical relationships to be defined (Figure 2A). In the most common type of model, the “entities” are biochemical substances, the “containers” are well-mixed and spatially homogeneous, and the “processes” are biochemical reactions happening within or between the containers. This originally led to the SBML constructs being named *species*, *compartments*, and *reactions*, respectively (Figure 2B), but these names are historical artifacts and belie the generality of the underlying scheme. Software applications can map the names to other concepts to better suit their purposes. For instance, “species” could be mapped to populations of molecules, cells, or even organisms.

Modelers and software developers are encouraged to use SBML’s reaction construct to define a model’s behavior in preference to formulating the model explicitly as a system of equations. This gives users freedom to convert the model into the final format they prefer—a simpler operation than (for example) inferring a reaction network from a system of differential equations. More importantly, the approach also naturally handles models where reaction kinetics are unknown or unneeded, such as interaction maps, and supports the elaboration of the reaction construct using SBML packages (discussed below). That said, the use of reactions is optional, and SBML provides features sufficient for encoding a large diversity of purely mathematical models, too. Whether using reactions or not, values of model variables and their changes over time may be fixed or determined by mathematical expressions, either before or during simulation, continuously or in response to discrete events, with or without time delays. Units of measurement can be specified for all entities and values; in addition to adding a layer of essential physical knowledge (after all, how else could one interpret whether a time course is in milliseconds or years?), information about units can be used to verify the relationships expressed in a model. Units also facilitate reuse of models and components, interconnection of models, conversion of models between different frameworks, and integration of data with models.

SBML does not dictate which framework must be used to analyze or simulate a model; in fact, it purposefully lacks any explicit way to specify what is done with a model—whether to run simulations or other types of analyses, how to run them, or how to present the results—because externalizing this information enhances model reusability and permits independent innovation in separate but complementary formats. Two of the most popular methods for time-course simulation are commonly used: one is numerical integration of differential equations created from the reactions and other relationships affecting model variables, and the other is simulating the time evolution of the model as a stochastic system via algorithms such as the one developed by Gillespie (1977). Alternative approaches are also in use, particularly when a model is enhanced with SBML packages.

Any element of an SBML model can be elaborated using machine-readable metadata as well as human-readable notes. For metadata, two schemes are supported. The first is direct labeling of SBML elements with terms from the Systems Biology Ontology (SBO; Courtot *et al*, 2011), which allows the mathematical semantics of every element of a model to be precisely specified. The second scheme uses semantic web technologies and provides greater flexibility to capture additional metadata. For instance, a molecular species in a model can be linked to a UniProt entry (The UniProt Consortium, 2017) if it represents a protein, or to ChEBI entry (Hastings *et al*, 2013) if it represents a simple chemical. Gene Ontology terms (GO; Ashburner *et al*, 2000) can be attached to species, compartments, and mathematical elements representing biological processes and functions. Simple provenance data such as identities of creators can be added to facilitate attribution and versioning. To help standardize how annotations are stored, SBML encourages the use of guidelines and resources established for this purpose (Le Novère *et al*, 2005). Finally, software tools can also use annotations to encode tool-specific data in their own formats, thus providing a way to capture data that might otherwise be lost. Annotations thereby help enrich the meaning of model

components, facilitate the understanding and reuse of models, and help software work with SBML more flexibly (Neal *et al*, 2019).

The core features described above have been a backbone of SBML ever since Level 2, even as SBML continued to evolve. The development of the modular Level 3, discussed in the next section, provided an opportunity to rethink and redesign a few other rarely-used features. For example, the species *charge* attribute, designed to represent molecular charge, was removed in Level 3 in favor of letting an SBML package introduce more complete support for the relevant concepts.



Fig 2: A closer look at SBML. (A) Fragments of the global structure of an SBML file. In this example, the use of several SBML packages is declared in the file header. Model elements in the file include the descriptions of model variables, as well as their relationships. Elements of the same type are collected into “ListOf” elements; e.g., model parameters are in the ListOfParameters element. SBML package elements can refer to elements in the SBML Core as necessary. (B) Model elements are linked through unique identifiers used in the mathematical constructs and the elements describing the reactions, the molecular species, and their localization. The full model for this example is available in BioModels Database (Malik-Sheriff et al, 2020) as the model with identifier [MODEL1904090001](#).



## SBML Level 3's modularity and breadth

Constant evolution in scientific methods presents challenges for the creation of software tools and standards. One challenge arises because the creation of new standards requires labor, testing, and time. This often causes standardization efforts to lag behind the latest technical developments in a constantly-moving field. A second challenge is that users want support for new methods and standards in software tools, which pressures developers to implement support quickly. Combined with the first challenge, it means that sometimes problems with a standard's definition are not discovered until more developers attempt to use it in different situations, which in turn often means that revisions to a standard are needed after it is published. Finally, another challenge is that software development often takes place under resource constraints (e.g., funding and time), limiting the scope of work that software developers can undertake—including, sometimes, limiting how many features of a standard they can support in their software.

The SBML community sought to address these challenges by putting in place certain structural features in SBML's development process. The first is the notion of *Levels*. A Level in SBML is an attempt to provide a given set of features for describing models, with higher Levels providing more powerful features. For example, the ability to express discrete events was added to SBML Level 2 but does not exist in Level 1. SBML Levels are mostly upwardly compatible, in the sense that the vast majority of models encoded in Level  $n$  can be translated to Level  $n + 1$ . *Versions* are used to introduce refinements to a given Level to account for realizations that come from real-life use of SBML. Finally, SBML Level 3 introduced an extensible modular architecture consisting of a central set of fixed features (named *SBML Level 3 Core*), and a scheme for adding *packages* that can augment the *Core* by extending existing elements, adding new elements, and adjusting the meaning or scope of elements. A model declares which packages it uses in order to guide its interpretation by software applications. If a software tool detects the presence of packages that it does not support, it may inform users if it cannot work with the model. Together, these three features (Levels, Versions, packages) help address the challenges discussed above: they ease coping with evolution in methods by collecting significant changes into discrete stages (SBML Levels), they help deal with the inevitable need for revisions (Versions within Levels), and they allow developers to limit the feature set they implement (SBML Levels on the one hand, and SBML Level 3 packages on the other).

Packages allow SBML Level 3 (Hucka *et al*, 2019) to represent many model types and characteristics in a more natural way than if they had to be shoehorned into SBML Core constructs exclusively. Twelve packages have been proposed to date (Table 1); seven have been fully developed into consensus specifications and are each used by at least two software implementations (Box 1), and another three have draft specifications in use by software tools. New packages can be developed independently, within dedicated communities, at a pace that suits them. This was the case for logical modeling with the CoLoMoTo community (Naldi *et al*, 2015), constraint-based modeling within the COBRA community (Heirendt *et al*, 2019), and rule-based modeling with a community of like-minded software creators (Boutillier *et al*, 2018; Faeder *et al*, 2009; Palmisano *et al*, 2014; Zhang *et al*, 2013).

Several benefits accrue from leveraging SBML as a starting point rather than creating a new, independent format. One is it makes clear where common features overlap. Most computational modeling frameworks in the domain of biology share some common concepts—variables that represent characteristics of different kinds of entities, processes that represent interactions between entities, containers/locations, etc.—and reusing SBML Level 3 Core constructs makes the conceptual similarities explicit. This in turn makes interpretation of models easier (no need to learn new terminology) and reuse simpler (no need to translate between independent formats). Another benefit is that the creators of the format can leverage existing features developed for SBML, such as mechanisms for annotations, rather than spend time developing new approaches to achieving the same goals in a new format. This in turn leads to another benefit: the ability to reuse at least some parts of existing software libraries developed for SBML. It also means that a software application may be able to interpret at least *some* fundamental aspects of a model



even if the application is not designed to work with a particular SBML Level 3 package, by virtue of understanding SBML Core (and perhaps other packages used by the model). This improves the potential for model reuse, and benefits model creators and software developers alike. Finally, a common foundation simplifies the creation of multiframework models in which some parts of the model use one formalism and other parts use others (e.g., coupling kinetic models with flux-balance analysis; [Watanabe \*et al\*, 2018](#)).

Though this modular approach has benefits, it is not without potential pitfalls. The main risks are fragmentation of the community, and incompatibility of packages due to complex feature dependencies. The SBML community has addressed the former by maintaining communications between package developers; the community processes have such interactions built in. As for the latter, API libraries (see [Box 2](#)) can handle *some* combinations of packages and hide some of the complexity. Still, there remain some combinations of packages that are not fully understood, and it remains for future work to define how (if ever) they can be combined for use in a single model.

Table 1: Summary of SBML Level 3 Package statuses. Symbols: ● = released; ○ = not released; ⊕ = in progress; n/a = not applicable.

Package name	Label	Purpose	Specification	libSBML Support	JSBML Support	Test Suite	Reference
● Flux Balance Constraints	fbc	Define constraint-based models (a.k.a. steady-state models).	✓	✓	✓	✓	<a href="#">Olivier and Bergmann (2018)</a>
● Groups	groups	Collect elements together for annotation purposes. Groups have no mathematical meaning and do not affect simulations.	✓	✓	✓	n/a	<a href="#">Hucka and Smith (2016)</a>
● Hierarchical Model Composition	comp	Define models composed of other models. The “submodels” can be stored in the same file or as separate files.	✓	✓	✓	✓	<a href="#">Smith et al (2015)</a>
● Layout	layout	Store positions and sizes of model components in network diagrams of SBML models. (Cf. the Rendering package.)	✓	✓	✓	n/a	<a href="#">Gauges et al (2015)</a>
● Multistate, Multicomponent, & Multicompartment Species	multi	Define features such as states or binding sites on molecular species, optionally in combination with rule-based processes.	✓	✓	✓	⊕	<a href="#">Zhang and Meier-Schellersheim (2018)</a>
● Qualitative Models	qual	Allow model where SBML species’ values represent qualitative activity levels rather than amounts or concentrations.	✓	✓	✓	⊕	<a href="#">Chaouiya et al (2015)</a>
● Rendering	render	Extend the Layout package to enable storing graphical symbols and styles, curves, colors, and gradients in network diagrams.	✓	✓	✓	n/a	<a href="#">Bergmann et al (2018)</a>
○ Arrays	arrays	Define arrays of elements, such as arrays of compartments. (Core SBML Level 3 supports only scalar values.)	⊕	✓	✓	⊕	
○ Distributions	distrib	Define statistical distributions for quantitative values. (Core SBML does not enable indicating value ranges or distributions.)	⊕	✓	✓	⊕	
○ Dynamical Processes	dyn	Describe the creation, destruction, and movement of model elements during simulation.	⊕	✓	✓	⊕	
○ Extended math	math	Additional constructs not included in the subset of MathML used by SBML Level 3 Core for mathematical expressions.	⊕	⊕	⊕	⊕	
○ Spatial Processes	spatial	Define spatially-inhomogeneous compartment geometries and processes such as diffusion.	⊕	✓	✓	⊕	

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## Box 1. SBML Level 3 packages officially part of the standard

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**Hierarchical Model Composition** The “comp” package (Smith *et al*, 2015) allows users to build models from other complete models or from model fragments, as a way to manage complexity and construct composite models. “Submodels” can be described within the same SBML file or linked from external files. A submodel can act as a template, and the same definition can be reused multiple times in other models to avoid duplication and enable reuse of parts. The “comp” package also enables submodels to have explicit interfaces (known as *ports*) for optional black-box encapsulation. Finally, “comp” was designed so that a hierarchical model can be converted into a single SBML model that does not use any “comp” features, making it readable by software that does not directly support the package. The library libSBML (Bornstein *et al*, 2008) provides a facility to do this.

**Flux Balance Constraints** The “fbc” package (Olivier and Bergmann, 2018) provides a means of encoding constraint-based models and optimizations, such as is done in Flux Balance Analysis (Bordbar *et al*, 2014). Constructs in the “fbc” package allow for the definition of a list of objectives for minimization or maximization, as well as flux bounds on reactions and gene-reaction mappings. Additional information such as chemical formula and charge enable further model analyses, including calculation of reaction mass balances, electron leaks, or implausible sources of matter.

**Groups** The “groups” package (Hucka and Smith, 2016) provides constructs to describe conceptual relationships between model elements. Groupings can indicate classification, parthood, or merely a collection of things; a group’s meaning can be specified using semantic annotations. Groups have no semantic meaning and cannot influence the mathematical interpretation of an SBML model.

**Multistate, Multicomponent and Multicompartment Species** The “multi” package (Zhang and Meier-Schellersheim, 2018) manages the combinatorics produced by entities either composed of multiple components, such as molecular complexes, or that can exist in multiple states, such as proteins with post-translational modifications. With the “multi” package, rules can be defined for how reactions depend on the states of the entities and their locations. The package adds syntactic constructs for molecular species types, compartment types, features, binding sites, and bonds. Entire families of molecular complexes sharing certain properties can be defined using patterns created using these constructs.

**Qualitative Models** The “qual” package (Chaouiya *et al*, 2015) provides constructs to encode models whose dynamics can be represented by discrete, reachable states connected by state transitions denoting qualitative updates of model elements. Examples include logical regulatory networks (Boolean or multi-valued) and Petri nets. The “qual” package introduces SBML elements to allow the definition of qualitative species, which are used to associate discrete levels of activities with entity pools, as well as transitions, which define the possible changes between states in the transition graph.

**Layout and Rendering** The “layout” (Gauges *et al*, 2015) and “render” (Bergmann *et al*, 2018) packages extend SBML to allow graphical representations of networks or pathways to be stored within SBML files. The “layout” package enables the encoding of positions and sizes of graphical elements such as nodes and lines, while the information about colors, fonts, etc., are defined by the “render” package. This separation presents several advantages. For example, applications can offer multiple styles for visualizing the same layout of a network map. Most of the essential aspects of a network diagram can be expressed using just the “layout” package, and thus tools do not necessarily have to implement a full graphics environment if they do not need to support customizing a diagram’s look-and-feel.

## SBML as a community standard

SBML's success can be attributed largely to its community-based development and its consensus-oriented approach. SBML has always been developed through engagement with its user community to achieve goals expressed by that same community. To resolve occasionally conflicting technical demands, a guiding principle has been to seek consensus between different viewpoints and the needs of different groups, to find a middle ground that would be—while perhaps not a perfect solution—an *acceptable* and *usable* solution. This attracted the researchers and software developers who constitute SBML's foremost stakeholders. By using SBML in everything from software to textbooks, they helped drive further development to face the real needs expressed by the people who have those needs. This engagement allowed faster feedback from users to developers, and has helped produce a rich toolkit of software and other resources that facilitate SBML's incorporation into software (Box 2).

Over the years, the community has designed rules to organize its governance, develop and maintain the specifications, and facilitate collaboration among users. The development of SBML and its Level 3 packages is shepherded by the SBML Editors, a group of community-elected volunteers serving terms of three years who follow a written and public process detailed on the web portal [SBML.org](http://sbml.org)<sup>1</sup>. SBML Editors write or review SBML specification documents, organize discussions and vote on specific technical issues, and enact the decisions of the community. Major proposed changes to the specifications and packages are discussed by the community via the SBML mailing lists<sup>2</sup> as well as during annual face-to-face meetings.

The community currently comes together twice a year within the context of meetings organized by COMBINE (the Computational Modeling in Biology Network; Hucka *et al*, 2015). *HARMONY* (the Hackathon on Resources for Modeling in Biology) is a codefest that focuses on the development of software, in particular via the development of libraries, tools, and specifications; by contrast, the *COMBINE Forum* meetings focus on the presentation of novel tools and the discussion of proposed features. In addition to these general meetings, special SBML working groups are organized as needed to drive SBML package development. COMBINE's central activity is coordinating and harmonizing standardization in computational biology, and SBML is one of its core standards. FAIRsharing, a broader community network that covers life sciences more comprehensively (Sansone *et al*, 2019), maintains interconnected and organized collections of resources in many areas, including curated links between SBML and many associated funders, databases, and standards<sup>3</sup>.

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<sup>1</sup>[http://sbml.org/Documents/SBML\\_Development\\_Process](http://sbml.org/Documents/SBML_Development_Process)

<sup>2</sup><http://sbml.org/Forums/>

<sup>3</sup><https://fairsharing.org/FAIRsharing.9qv71f>

## Box 2. Software infrastructure for SBML

<p><b>Application Programming Interface (API)</b></p> <p>Open-source (LGPL) libraries and code generators help read, write, manipulate, validate, and transform SBML. They support all Levels and Versions of SBML, and all Level 3 packages.</p> <ol style="list-style-type: none"> <li>1. LibSBML (Bornstein <i>et al</i>, 2008) (<a href="http://sbml.org/Software/libSBML">http://sbml.org/Software/libSBML</a>), written in C++, offers interfaces for C, C++, C#, Java, JavaScript, MATLAB, Octave, Perl, PHP, Python, R, and Ruby</li> <li>2. JSBML (Rodriguez <i>et al</i>, 2015) (<a href="http://sbml.org/Software/JSBML">http://sbml.org/Software/JSBML</a>) offers a pure Java API</li> <li>3. Deviser (<a href="http://sbml.org/Software/Deviser">http://sbml.org/Software/Deviser</a>) generates libSBML code for rapid package prototyping</li> </ol>	<p><b>Test Suite</b></p> <p>The SBML Test Suite (<a href="http://sbml.org/Software/SBML_Test_Suite">http://sbml.org/Software/SBML_Test_Suite</a>) helps developers implement SBML compatibility and helps users check SBML features supported in software.</p> <ol style="list-style-type: none"> <li>1. Thousands of test cases for <ul style="list-style-type: none"> <li>• Semantic interpretation of models (for both deterministic and stochastic simulation)</li> <li>• Syntactic correctness</li> </ul> </li> <li>2. A graphical front end enables cases to be filtered by Level/Version and type of test</li> <li>3. An online database allows results to be uploaded and compared with results from other simulators</li> </ol>
<p><b>Validation Facilities</b></p> <p>Validation software can check files for compliance to the definition of SBML, good modeling practices, and consistency of units</p> <ol style="list-style-type: none"> <li>1. API libraries include built-in validation</li> <li>2. Online validator has simple user interface (<a href="http://sbml.org/Facilities/Validator">http://sbml.org/Facilities/Validator</a>)</li> <li>3. Web services support software access</li> </ol> <p>Validation ensures compliance with:</p> <ul style="list-style-type: none"> <li>• SBML syntax</li> <li>• SBML validation rules published as part of each accepted SBML specification</li> </ul>	<p><b>Conversion Facilities</b></p> <p>Converters (<a href="http://sbml.org/Software/Converters">http://sbml.org/Software/Converters</a>) can translate some other formats to/from SBML</p> <ol style="list-style-type: none"> <li>1. Conversion tools support format conversions from MATLAB, BioPAX, CellML, XPP, SBtab, and others</li> <li>2. Online services such as SBFC (Rodriguez <i>et al</i>, 2016) convert uploaded files to a variety of formats</li> <li>3. API libraries provide converters between different SBML Levels/Versions and different SBML constructs</li> </ol>

### Software Guide

A catalog ([http://sbml.org/SBML\\_Software\\_Guide](http://sbml.org/SBML_Software_Guide)) of software applications, libraries and online services known to support SBML—over 290 entries to date

1. A tabular interface highlights supported SBML features of each software system
2. A list interface displays human-readable summaries of software systems
3. Software can be added to the list upon request

## Impact of SBML

As contributors to developments in methods, software, and standards over the past two decades (Hucka *et al*, 2015), we can attest to SBML's profound impact on the field, both from our own first-hand experiences and from surveys (Klipp *et al*, 2007) that indicate SBML has become a *de facto* standard. The impact is a result of SBML's community-oriented development approach and its design.

The SBML development process has helped shape the field partly by directly involving software developers and modelers. Frequent workshops have provided essential feedback for developers to help them better serve modelers' needs (e.g., Waltemath *et al*, 2014). Workshops as well as resources such as the SBML Software Guide (see Box 2) helped raise awareness of existing tools, which in turn increased their use and the use of SBML. This helped create a culture of sharing models and building on existing work in systems biology (Stanford *et al*, 2015). It also led to new activities centered on the models themselves, including automatic model generation, analysis of model structures, model retrieval, and integration of models with experimental data (Dräger and Palsson, 2014). SBML's successful approach to community organization has led other standardization efforts (e.g., BioPAX, NeuroML, SBGN, SED-ML) to adopt some of the same approaches; SBML was also a founding member of COMBINE (Hucka *et al*, 2015), discussed above. Some of the primary standardization efforts in COMBINE, such as BioPAX (Demir *et al*, 2010) and NeuroML (Gleeson *et al*, 2010), are more domain-specific than SBML; others, such as CellML (Lloyd *et al*, 2004), overlap SBML's primary domains but offer alternative abstractions; and finally, still others such as SBGN (van Iersel *et al*, 2012), SBOL (Roehner *et al*, 2016), and SED-ML (Waltemath *et al*, 2011), are complementary formats.

Before the advent of SBML, it was challenging to exchange models because software tools used incompatible definition schemes. As models increased in size and complexity, manually rewriting them became more difficult, error-prone, and eventually, untenable. The development of SBML has enabled the use of a single model description throughout a project's life cycle even when projects involve heterogeneous software tools (Box 3). SBML-compatible software tools today allow researchers to use SBML in all aspects of a modeling project, including creation (manual or automated), annotation, comparison, merging, parametrization, simulation/analysis, results comparison, network motif discovery, system identification, omics data integration, visualization, and more. Such use of a standardized format, along with standard annotation schemes (Neal *et al*, 2019) and training in reproducible methods, improves research workflows and is generally recognized as promoting research reproducibility (Waltemath and Wolkenhauer, 2016).

The availability of a well-defined format has also facilitated the comparison of software tools to each other. Using SBML-encoded models has become the norm to assess the accuracy of modeling software: initially done manually using models from BioModels Database (Bergmann and Sauro, 2008), now it is more commonly done using the SBML Test Suite (Box 2). SBML's semantics are defined precisely enough that many simulation systems can produce equivalent results for over 1200 test cases, lending confidence that SBML-based simulations can be reproducible in different software environments.

While chemical kinetics models have been a staple of systems biology, other modeling frameworks exist. These have benefited from efforts to extend Level 3 to better suit their specific characteristics. Even when models could in principle be encoded using core SBML constructs, the use of features explicitly adapted to the needs of a domain can make model interpretation less error-prone and more natural. The former issue was demonstrated vividly when ad hoc methods of encoding genome-scale models led to incorrect interpretations, and a subsequent proposal to use SBML Level 3 "fbc" addressed representational inconsistencies that had hindered reproducibility (Ebrahim *et al*, 2015). The use of more domain-specific forms of encoding has been preferred by several communities, such as the qualitative and rule-based modeling communities. For example, the quickly adopted package SBML Level 3 "qual" (Chaouiya *et al*, 2015) supports software interoperability for qualitative modeling, illustrated by the use of CellNOpt (Terfve *et al*, 2012), which provides a set of optimal Boolean models that best explains the causal relationships between elements of a signal transduction network and associated data, and the subsequent use of GIN-



sim (Chaouiya *et al*, 2012) or Cell Collective (Helikar *et al*, 2012) to assess the dynamical properties of these models. Rule-based modeling can represent models that are impossible to express as reaction networks, such as polymerization (Faeder *et al*, 2009), or simply impractical to represent due to the combinatorial number of reactions implied by the rules (Hlavacek *et al*, 2003). Storing rule definitions in SBML is now feasible with the “multi” package, allowing rule-based modeling tools such as Simmune (Zhang *et al*, 2013) and BioNetGen (Faeder *et al*, 2009) to read and write the same model definitions.

SBML has also eased the automated processing of models to the point where they have become just another type of data in the life sciences. SBML is used today as an import/export format by many databases of mathematical models (Malik-Sheriff *et al*, 2020; Misirli *et al*, 2014; Norsigian *et al*, 2019), as well as by pathway databases (Caspi *et al*, 2015; Fabregat *et al*, 2017; Mi *et al*, 2016) and reaction databases (Ganter *et al*, 2013; Wittig *et al*, 2017). SBML is the preferred format for model curation in BioModels Database (Malik-Sheriff *et al*, 2020), not only because of its popularity but also because of its provisions to precisely encode and annotate models to support reproducible modeling. SBML is also used to share models by more generic data management platforms such as SEEK (Wolstencroft *et al*, 2016) and comprehensive online simulation environments (e.g., Lee *et al*, 2009; Moraru *et al*, 2008; Peters *et al*, 2017; Weidemann *et al*, 2008). Moreover, having an agreed-upon format has facilitated the introduction of better model management strategies. This includes support for tasks such as model storage and retrieval (Henkel *et al*, 2015), version control (Scharm *et al*, 2016b), and checking quality and validity (Liebermeister, 2008). The proliferation of derived models has led to the development of methods to compare model structure and semantic annotations (Lambusch *et al*, 2018), culminating in the development of several methods to quantify model similarities (Henkel *et al*, 2016) that can then be used to improve the relevance of model searches. Once model elements can be compared, one can align, combine, and merge different models (Krause *et al*, 2010).

A wider impact of SBML as a *de facto* standard has been the support of publishers and funding agencies. Many journals, aware of the challenges surrounding the reproducibility of scientific results, encourage authors not only to describe their models but also to make their models available in electronic form. Though journals avoid *requiring* a specific format, some journals, such as the BMC<sup>4</sup> and FEBS<sup>5</sup> journals, explicitly encourage authors to submit SBML files as supporting material for research where it is relevant; others, such as Biophysical Journal (Nickerson and Hunter, 2017), recommend authors deposit models in repositories such as BioModels Database (Malik-Sheriff *et al*, 2020), which encourages the use of common standard formats such as SBML. Many funding agencies also now have policies related to data sharing, and some program announcements suggested the use of SBML where appropriate<sup>6</sup>.

Finally, the continued development of SBML has stimulated collaborative work and the creation of consortia. This has led to better awareness and communication within groups interested in specific modeling frameworks. A good example is the CoLoMoTo effort mentioned above; it was launched by researchers who needed a format to exchange qualitative models between their software tools and developed the Qualitative Modeling package for SBML (Naldi *et al*, 2015) as the solution. Nevertheless, challenges remain, as discussed in the next section. These will need to be confronted to ensure the longevity of SBML as well as continued developments.

<sup>4</sup><https://www.biomedcentral.com/getpublished/writing-resources/additional-files>

<sup>5</sup><https://onlinelibrary.wiley.com/page/journal/17424658/homepage/ForAuthors.html>

<sup>6</sup>See, for example, <https://grants.nih.gov/grants/guide/pa-files/par-08-023.html>



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### Box 3. Examples of SBML use cases

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SBML's impact on computational systems biology includes its facilitation of collaborative work. In multiple instances, it has precipitated entirely new projects, as illustrated by the examples below.

**SBML throughout the model life-cycle** Encoding a model in a standard format such as SBML makes it easier to use different software tools for different purposes, and thus makes it easier to leverage the most suitable tools at different points in a workflow. The following is an example. A signaling pathway can be designed graphically using CellDesigner (Funahashi *et al*, 2003). The resulting model can then be semi-automatically annotated using the online tool semanticSBML (Krause *et al*, 2010). Experimental kinetic information can be retrieved in SBML format from the SABIO-Reaction Kinetics database (Wittig *et al*, 2017). Tools such as COPASI (Hoops *et al*, 2006) and PyBioNetFit (Mitra *et al*, 2019) provide facilities to estimate parameters and to simulate the model with various algorithms. Other SBML-enabled tools such as Tellurium (Medley *et al*, 2018) and PySCeS (Olivier *et al*, 2005) provide capabilities such as identifiability and bifurcation analysis. Each step of the process applied to a model from creation to publication of results—modeling, simulation and analysis—can be documented using notes attached to every model element. The model can even be turned into a publishable document using SBML2 $\LaTeX$  (Dräger *et al*, 2009). Finally, the model can be exported from selected modeling tools, together with data and other information all bundled together in COMBINE Archive format (Bergmann *et al*, 2014) and published in model repositories such as BioModels Database (Malik-Sheriff *et al*, 2020).

**Pipeline for automated model building** Being able to describe model elements precisely using semantic annotations facilitates the creation of automated pipelines (Dräger *et al*, 2010). Such pipelines can combine existing models with databases of molecular phenotypes or reaction kinetics (Li *et al*, 2010). They can also generate models *de novo* from data resources, as has been demonstrated by the Path2Models project (Büchel *et al*, 2013). Path2Models has produced 143,000 SBML models—all fully annotated—for over 2,600 organisms, by using pathway data. Metabolic pathways were encoded in SBML Level 3 Core while signaling pathways were encoded with the SBML “qual” package (Chaouiya *et al*, 2013). Moreover, constraint-based models of genome-scale reconstruction were provided for each organism. Other pipelines have now been built, including ones that can systematically generate alternative models for different tissue-types (Wang *et al*, 2012) and patient data (Uhlen *et al*, 2017), an important step towards personalized medicine.

**Development, sharing, and re-use of genome-scale models of human metabolism** Constraint-based modeling approaches such as Flux Balance Analysis and its variants permit the use of whole-genome reconstructions together with experimental molecular phenotypes, in order to predict how mutations or different environments affect metabolism as well as predict drug targets and biomarkers (O'Brien *et al*, 2015). With the availability of genome-scale metabolic reconstructions, the use of metabolic flux models at the same scale has been increasing (Bordbar *et al*, 2014). A recent development in the field has been the curation of consensus metabolic models, in particular for human metabolism (Brunk *et al*, 2018). Those community efforts rely on SBML for encoding and sharing the models, including annotations, which are crucial to being able to reuse the reconstructions later, and also for visual representation using the Layout (Gauges *et al*, 2015) and Rendering (Bergmann *et al*, 2018) packages. The Flux Balance Constraint package (Olivier and Bergmann, 2018) enables encoding of the information required for model optimization and flux calculation. Unambiguous encoding in SBML has been shown to be crucial for interpreting models and precisely computing fluxes (Ebrahim *et al*, 2015; Ravikrishnan and Raman, 2015).

## Forthcoming challenges

For nearly two decades, SBML has supported mathematical modeling in systems biology by helping to focus the efforts of the community and foster a culture of openness and sharing. The field is evolving rapidly, which presents challenges that the community and SBML must face.

The first challenge is to remain usable in the face of relentless growth in model sizes. One of the drivers of larger size is the rising popularity of genome-scale metabolic models (Bordbar *et al*, 2014), which can be produced semi-automatically (Henry *et al*, 2010). Modeling approaches have also been developed to combine the use of several such models (e.g., Bordbar *et al*, 2011). It is reasonable to expect models of ecosystems to be produced soon (e.g., microbiomes and their host). Model sizes will also increase as more models of tissues and organs are exchanged and reused, encouraged by the use of software packages that facilitate this approach, such as the open-source tools CHASTE (Mirams *et al*, 2013) and CompuCell3D (Swat *et al*, 2012). The challenge this presents is how to define, organize, and manage large models. Meeting the challenge will require a combination of novel approaches to model storage (e.g., Henkel *et al*, 2015) and comparison (e.g., Scharm *et al*, 2016a,b), as well as more effective use of SBML Level 3 features. For example, the SBML Hierarchical Model Composition (“comp”) package (Smith *et al*, 2015) provides a way to encode models in SBML out of separate building blocks or from preexisting models; this can make larger models easier to structure and maintain, and it is a natural way to construct multiscale models. Similarly, the Arrays package may help to define and structure larger models by allowing models to be defined in a more compact form.

A related challenge concerns human usability of SBML and similar XML-based formats. Though SBML is intended for software, not humans, to use directly, desire for a text-based or spreadsheet-based equivalent is often voiced (e.g., Kirouac *et al*, 2019). Various answers have been developed in the form of text-based notations (e.g., Gillespie *et al*, 2006; Smith *et al*, 2009) and spreadsheet conventions (e.g., Lubitz *et al*, 2016), with bidirectional translators for SBML. These formats have undeniable appeal for many users and use cases, despite that they do not capture the entirety of SBML (often having limited or missing facilities to express units, annotations, or SBML packages). Their chief drawback is that they become error-prone to use as model size increases. Graphical user interfaces (GUIs; e.g., Funahashi *et al*, 2003; Hoops *et al*, 2006; Moraru *et al*, 2008) can overcome this; software with GUIs can help with the cognitive burden of tracking large numbers of model elements. On the other hand, GUIs can be tedious to use when entering large models, performance of some software does not scale well with increasing model sizes, and some cannot be controlled programmatically for automation purposes. A middle ground may be domain-specific modeling languages layered on top of programming languages such as Python (e.g., Lopez *et al*, 2013; Olivier *et al*, 2005). However, these tend to appeal only to users who are comfortable with (or willing to take time to learn) the programming language used as a substrate. Overall, further innovation in this area would be welcome, both to help support SBML Level 3 packages and to help users cope with ever-increasing model sizes.

Because of the diversity of biological phenomena amenable to mathematical modeling, as well as their scales and properties, it is likely that a broad variety of modeling approaches will be added to every researcher’s essential toolbox (Cvijovic *et al*, 2014). Methods such as multiagent and lattice approaches are coming into broader use to represent evolving cell populations, cell migration, and deformation. Some researchers are experimenting with solutions using existing SBML packages (Varela *et al*, 2019; Watanabe and Myers, 2016). Modeling the development of tissues and organ function may also require combining these approaches with reaction-diffusion models, or multiphysics approaches (Nickerson *et al*, 2016). Population modeling will need to complement traditional instance-based systems if we want to take into account patient variability or information coming from single-cell measurements (Levin *et al*, 1997). The coupling of different approaches within the same simulation experiment is also becoming more frequent. Biomolecular reactions modeled using ODEs, Poisson processes, and Flux Balance Analyses have been coupled in the first whole-cell model (Karr *et al*, 2015). At the organ level, liver lobules have

been modeled using a combination of metabolism and multi-agent models (Schliess *et al*, 2014). Several approaches mixing modeling of cell mechanical properties and gene regulatory networks or signaling networks have been used to study morphogenesis (e.g., Tanaka *et al*, 2015). The coupling of different approaches can be done within a single hybrid model, or each model can be simulated using different software and with dynamic synchronization at run time (Mattioni and Le Novère, 2013). Once again, the SBML “comp” package can play a role in supporting these approaches, but other methods and software will be needed in the future, as well as better support for coupling models at run time using, for example, SED-ML (Waltemath *et al*, 2011).

These developments are arising in a landscape where structural models are sometimes not the central object of study, and instead function as collection of integrated information. An example of this is RECON3D, a comprehensive human metabolic network with metabolite and protein structure information (Brunk *et al*, 2018). SBML will continue to have a pivotal role here too. When SBML was introduced, the state of modeling workflows and software tools was more primitive and it was natural that a model was self-contained. SBML-encoded models often had predefined parameter values (e.g., as initial values for state variables or parameters for mathematical expressions), but today, modelers increasingly want to use the same model with different parameterizations, sometimes with parameter values expressed as distributions, lists, or ranges rather than unique values. A project may also use an ensemble of related models that differ in parameters or in turning some model elements on or off (Kuepfer *et al*, 2007). The semantic annotation of SBML elements also has become increasingly important, forming a bedrock for many of the analyses using SBML-encoded models. The growth in size and scope of annotations has recently led the modeling community to propose a standard way of storing annotations in separate linked files (Neal *et al*, 2019), relying on the COMBINE Archive format (Bergmann *et al*, 2014) to bundle everything together. Other formats that can complement SBML have been developed, and further coordination and evolution will undoubtedly happen in the future. As mentioned above, SED-ML is a format that provides a way to encode what to do with a model, which complements SBML and compensates for its lack of features to define procedures. Finally, experimentation in integrating SBML more directly with other formats and data also continues. For instance, preliminary work has shown that SBML can be enriched with SBOL (Voigt *et al*, 2018) to provide models of DNA components’ behavior (Roehner and Myers, 2014), and conversely, ongoing work in supporting genome-scale models of metabolism and gene expression (known as *ME-models*, Thiele *et al*, 2012) augments SBML with SBOL to more fully capture models for use with ME-modeling software. Future developments in modeling paradigms may require similar flexibility in how models are represented: some may be best served by implementing new SBML packages, others by extending existing packages, still others by combining SBML with other formats.

Besides the technical challenges, social and cultural challenges also exist for formats such as SBML. One is to continue raising awareness among researchers, software developers, and funders of the existence of SBML and related COMBINE standards. Some may not yet be using SBML simply because they are not aware of it, or its recent addition of support for many modeling formalisms (Figure 1). Raising awareness will require continual education and outreach, especially to students and early-career scientists. Awareness would be aided by greater promotion on the part of journals and reviewers of the use of SBML and related formats in paper submission guidelines. Despite some progress in this area (discussed in the previous section), the lack of stronger demands by journals and reviewers is surely one reason authors are either not aware or not motivated to publish their models in software-independent formats.

In addition, usability of standard formats depends crucially on their implementation in software tools, and motivating this work is another challenge for SBML. A pivotal factor for the success of SBML has been the extensive software ecosystem, which provides relatively easy import and export of SBML from popular software systems. However, implementing full SBML compatibility in software is not a simple matter, and problems with compatibility in the software ecosystem can be a significant source of frustration. Improving the software requires continuous investment in tool development.

That, in turn, is related to a final challenge: obtaining and maintaining funding. By virtue of not being a native format of any particular software tool, a format such as SBML may require extra work to define by consensus, and then again for developers to implement in software—and still, it will lag behind the leading edge of research because exchange formats only become important after more than one software system has something to exchange. Funders may wonder whether the resources, time and effort spent on standards development would not be better applied to other goals. However, these costs must be weighed against the costs to a whole research field of *not* having standards—and there are many such costs. To take one example, models in nonstandard formats are more difficult to review, verify, and reuse. Journal reviewers may not have access to the necessary software, or the software may not be well-tested, all of which increase the chances that the published model contains errors. Researchers can spend substantial time attempting to reproduce the results, only to fail. Worse, this is a repeating cost: failures to reproduce models are rarely published or publicized, which means an untold number of researchers may spend time (and research funding) on a futile effort. Funders recognize that too many research results are irreproducible, and have urged community action (e.g., [Collins and Tabak, 2014](#)). The continued development of exchange formats, such as SBML, is a crucial and cost-effective means to enable reproducible research.

## Conclusion

SBML and associated software libraries and tools have been instrumental in the growth of systems biology. As modeling and simulation grew in popularity, SBML allowed researchers to exchange and (re)use new models in an open, well-supported, interoperable format. SBML has made possible much of the research pursued by the authors of this article, and also helped us to structure our thoughts about our models and the biology they represent. Today, scientists can build, manipulate, annotate, store, reuse, publish, and connect models to each other and to basic data sources. In effect, SBML has turned models into a kind of data and transformed modeling in biology from an art to an exercise in engineering.

As the field of systems biology continues to grow and address emerging challenges, SBML will grow along with it. This evolution will (as it always has) depend on close cooperation between biologists and software developers. We hope that SBML will continue to be a source of inspiration for many researchers, especially those new to the field. In return, may they help develop the next generation of SBML to support more comprehensive, richer, and more diverse models, and expand the reach of systems modeling towards entire cells, organs, and organisms.

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## Author contributions

S.M.K., D.W., M.K., F.Z., A.D., C.C., and M.H. wrote the bulk of the manuscript. Together with F.B., A.M.F., C.G., T.H., S.H., R.M.-S., S.M., I.M., C.M., A.N., B.O., S.S., J.C.S., L.S., M.S., D.T., L.W., and D.J.W., they also wrote and/or edited specifications for SBML Level 3 Core and the Level 3 packages. M.L.B., K.B., J.F., H.G., T.M.H., Y.I., W.L., A.L., D.L., E.M., C.P., K.R., N.R., C.S., B.S., J.S., J.C.S., N.S., N.T., and J.W. contributed proposals for SBML Level 3 and/or are past or current members of the SBML Team. M.M.-S., H.M.S., B.P., H.B., H.K., A.F., H.H., J.C.D., and M.H. were principal investigators (or the equivalent, depending on the institution) for grants supporting SBML development.

The authors declare that they have no conflict of interest.

<sup>7</sup><https://summerofcode.withgoogle.com>

## References

- Ashburner M, Ball CA, Blake JA, Botstein D, Butler H, Cherry JM, Davis AP, Dolinski K, Dwight SS, Eppig JT, *et al* (2000) Gene Ontology: tool for the unification of biology. *Nature Genetics* 25: 25–29
- Bergmann FT, Adams R, Moodie S, Cooper J, Glont M, Golebiewski M, Hucka M, Laibe C, Miller AK, Nickerson DP, Olivier BG, Rodriguez N, Sauro HM, Scharm M, Soiland-Reyes S, Waltemath D, Yvon F, Le Novère N (2014) COMBINE archive and OMEX format: one file to share all information to reproduce a modeling project. *BMC Bioinformatics* 15: 369
- Bergmann FT, Keating SM, Gauges R, Sahle S, Wengler K (2018) SBML Level 3 Package: Render, Version 1, Release 1. *Journal of Integrative Bioinformatics* 15
- Bergmann FT, Sauro HM (2008) Comparing simulation results of SBML capable simulators. *Bioinformatics* 24: 1963–1965
- Bordbar A, Feist AM, Usaite-Black R, Woodcock J, Palsson BØ, Famili I (2011) A multi-tissue type genome-scale metabolic network for analysis of whole-body systems physiology. *BMC Systems Biology* 5: 180
- Bordbar A, Monk JM, King ZA, Palsson BØ (2014) Constraint-based models predict metabolic and associated cellular functions. *Nature Reviews Genetics* 15: 107–20
- Bornstein BJ, Keating SM, Jouraku A, Hucka M (2008) LibSBML: an API library for SBML. *Bioinformatics* 24: 880–881
- Boutillier P, Maasha M, Li X, Medina-Abarca HF, Krivine J, Feret J, Cristescu I, Forbes AG, Fontana W (2018) The Kappa platform for rule-based modeling. *Bioinformatics* 34: i583–i592
- Brunk E, Sahoo S, Zielinski DC, Altunkaya A, Dräger A, Mih N, Gatto F, Nilsson A, Gonzalez GAP, Aurich MK, Prlić A, Sastry A, Danielsdottir AD, Heinken A, Noronha A, Rose PW, Burley SK, Fleming RMT, Nielsen J, Thiele I, *et al* (2018) Recon3D enables a three-dimensional view of gene variation in human metabolism. *Nature Biotechnology* 36: 272–281
- Büchel F, Rodriguez N, Swainston N, Wrzodek C, Czauderna T, Keller R, Mittag F, Schubert M, Glont M, Golebiewski M, van Iersel M, Keating S, Rall M, Wybrow M, Hermjakob H, Hucka M, Kell DB, Müller W, Mendes P, Zell A, *et al* (2013) Path2Models: large-scale generation of computational models from biochemical pathway maps. *BMC Systems Biology* 7: 116
- Caspi R, Billington R, Ferrer L, Foerster H, Fulcher CA, Keseler IM, Kothari A, Krummenacker M, Latendresse M, Mueller LA, Ong MQ, Paley S, Subhraveti P, Weaver DS, Karp PD (2015) The MetaCyc database of metabolic pathways and enzymes and the BioCyc collection of pathway/genome databases. *Nucleic Acids Research* 44: D471–D480
- Chance B, Brainerd J, Cajori F, Millikan G (1940) The kinetics of the enzyme-substrate compound of peroxidase and their relation to the Michaelis theory. *Science* 92: 455
- Chaouiya C, Bérenguier D, Keating SM, Naldi A, van Iersel MP, Rodriguez N, Dräger A, Büchel F, Cokelaer T, Kowal B, Wicks B, Gonçalves E, Dorier J, Page M, Monteiro PT, von Kamp A, Xenarios I, de Jong H, Hucka M, Klamt S, *et al* (2013) SBML qualitative models: a model representation format and infrastructure to foster interactions between qualitative modelling formalisms and tools. *BMC Systems Biology* 7: 135
- Chaouiya C, Keating SM, Berenguier D, Naldi A, Thieffry D, Iersel MPv, Le Novère N, Helikar T (2015) SBML Level 3 package: Qualitative Models, Version 1, Release 1. *Journal of Integrative Bioinformatics* 12: 691–730

- Chaouiya C, Naldi A, Thieffry D (2012) Logical modelling of gene regulatory networks with GINsim, In *Bacterial Molecular Networks*, Springer, pp. 463–479
- Collins FS, Tabak LA (2014) Policy: NIH Plans to Enhance Reproducibility. *Nature* 505: 612–613
- Courtot M, Juty NJ, Knüpfer C, Waltemath D, Zhukova A, Dräger A, Dumontier M, Finney A, Golebiewski M, Hastings J, Hoops S, Keating S, Kell DB, Kerrien S, Lawson J, Lister A, Lu J, Machne R, Mendes P, Pocock M, *et al* (2011) Controlled vocabularies and semantics in systems biology. *Molecular Systems Biology* 7: 1
- Cvijovic M, Almquist J, Hagmar J, Hohmann S, Kaltenbach HM, Klipp E, Krantz M, Mendes P, Nelander S, Nielsen J, Pagnani A, Przulj N, Raue A, Stelling J, Stoma S, Tobin F, Wodke JAH, Zecchina R, Jirstrand M (2014) Bridging the gaps in systems biology. *Molecular Genetics and Genomics* 289: 727–734
- Demir E, Cary MP, Paley S, Fukuda K, Lemer C, Vastrik I, Wu G, D’Eustachio P, Schaefer C, Luciano J, Schacherer F, Martinez-Flores I, Hu Z, Jimenez-Jacinto V, Joshi-Tope G, Kandasamy K, Lopez-Fuentes AC, Mi H, Pichler E, Rodchenkov I, *et al* (2010) The BioPAX community standard for pathway data sharing. *Nature Biotechnology* 28: 935
- Dräger A, Palsson BØ (2014) Improving collaboration by standardization efforts in systems biology. *Frontiers in Bioengineering* 2: 61
- Dräger A, Planatscher H, Wouamba DM, Schröder A, Hucka M, Endler L, Golebiewski M, Müller W, Zell A (2009) SBML2 $\LaTeX$ : Conversion of SBML files into human-readable reports. *Bioinformatics* 25: 1455–1456
- Dräger A, Schröder A, Zell A (2010) Automating Mathematical Modeling of Biochemical Reaction Networks, In *Systems Biology for Signaling Networks*, Choi S (ed), New York, NY: Springer, pp. 159–205
- Ebrahim A, Almaas E, Bauer E, Bordbar A, Burgard AP, Chang RL, Dräger A, Famili I, Feist AM, Fleming RMT, Fong SS, Hatzimanikatis V, Herrgård MJ, Holder A, Hucka M, Hyduke D, Jamshidi N, Lee SY, Le Novère N, Lerman JA, *et al* (2015) Do Genome-scale Models Need Exact Solvers or Clearer Standards? *Molecular Systems Biology* 11: 831
- Fabregat A, Jupe S, Matthews L, Sidiropoulos K, Gillespie M, Garapati P, Haw R, Jassal B, K€orninger F, May B, Milacic M, Roca C, Rothfels K, Sevilla C, Shamovsky V, Shorser S, Varusai T, Viteri G, Weiser J, Wu G, *et al* (2017) The Reactome pathway knowledgebase. *Nucleic Acids Research* 46: D649–D655
- Faeder JR, Blinov ML, Hlavacek WS (2009) Rule-based modeling of biochemical systems with BioNetGen, In *Systems biology*, Maly IV (ed), Methods in Molecular Biology, Totowa, NJ: Humana Press, pp. 113–167
- Funahashi A, Tanimura N, Morohashi M, Kitano H (2003) CellDesigner: a process diagram editor for gene-regulatory and biochemical networks. *BIOSILICO* 1: 159–162
- Ganter M, Bernard T, Moretti S, Stelling J, Pagni M (2013) MetaNetX.org: a website and repository for accessing, analysing and manipulating metabolic networks. *Bioinformatics* 29: 815–816
- Garfinkel D (1969) Construction of biochemical computer models. *FEBS Letters* 2: S9–S13
- Gauges R, Rost U, Sahle S, Wengler K, Bergmann FT (2015) The Systems Biology Markup Language (SBML) Level 3 Package: Layout, Version 1 Core. *Journal of Integrative Bioinformatics* 12: 267
- Gillespie CS, Wilkinson DJ, Proctor CJ, Shanley DP, Boys RJ, Kirkwood TBL (2006) Tools for the SBML Community. *Bioinformatics* 22: 628–629
- Gillespie DT (1977) Exact stochastic simulation of coupled chemical reactions. *The Journal of Physical Chemistry* 81: 2340–2361



- Gleeson P, Crook S, Cannon RC, Hines ML, Billings GO, Farinella M, Morse TM, Davison AP, Ray S, Bhalla US, Barnes SR, Dimitrova YD, Silver RA (2010) NeuroML: A language for describing data driven models of neurons and networks with a high degree of biological detail. *PLoS Computational Biology* 6: 1–19
- Goddard NH, Hucka M, Howell F, Cornelis H, Shankar K, Beeman D (2001) Towards NeuroML: Model Description Methods for Collaborative Modelling in Neuroscience. *Philosophical transactions of the Royal Society of London Series B Biological sciences* 356: 12091228
- Hastings J, de Matos P, Dekker A, Ennis M, Harsha B, Kale N, Muthukrishnan V, Owen G, Turner S, Williams M, Steinbeck C (2013) The ChEBI reference database and ontology for biologically relevant chemistry: enhancements for 2013. *Nucleic Acids Research* 41: D456–D463
- Heirendt L, Arreckx S, Pfau T, Mendoza SN, Richelle A, Heinken A, Haraldsdóttir HS, Wachowiak J, Keating SM, Vlasov V, Magnúsdóttir S, Ng CY, Preciat G, Žagare A, Chan SHJ, Aurich MK, Clancy CM, Modamio J, Sauls JT, Noronha A, *et al* (2019) Creation and Analysis of Biochemical Constraint-Based Models Using the COBRA Toolbox v.3.0. *Nature Protocols* 14: 639–702
- Helikar T, Kowal B, McClenathan S, Bruckner M, Rowley T, Madrahimov A, Wicks B, Shrestha M, Limbu K, Rogers JA (2012) The Cell Collective: toward an open and collaborative approach to systems biology. *BMC Systems Biology* 6: 96
- Henkel R, Hoehndorf R, Kacprowski T, Knüpfer C, Liebermeister W, Waltemath D (2016) Notions of similarity for systems biology models. *Briefings in Bioinformatics* 19: 77–88
- Henkel R, Wolkenhauer O, Waltemath D (2015) Combining computational models, semantic annotations and simulation experiments in a graph database. *Database* 2015
- Henry CS, DeJongh M, Best AA, Frybarger PM, Lindsay B, Stevens RL (2010) High-throughput generation, optimization and analysis of genome-scale metabolic models. *Nature Biotechnology* 28: 977
- Hlavacek WS, Faeder JR, Blinov ML, Perelson AS, Goldstein B (2003) The complexity of complexes in signal transduction. *Biotechnology and Bioengineering* 84: 783–794
- Hodgkin AL, Huxley AF (1952) A quantitative description of membrane current and its application to conduction and excitation in nerve. *The Journal of Physiology* 117: 500–544
- Hoops S, Sahle S, Gauges R, Lee C, Pahle J, Simus N, Singhal M, Xu L, Mendes P, Kummer U (2006) COPASI – a complex pathway simulator. *Bioinformatics* 22: 3067–3074
- Hucka M, Bergmann FT, Chaouiya C, Dräger A, Hoops S, Keating SM, König M, Novère NL, Myers CJ, Olivier BG, Sahle S, Schaff JC, Sheriff R, Smith LP, Waltemath D, Wilkinson DJ, Zhang F (2019) The Systems Biology Markup Language (SBML): Language Specification for Level 3 Version 2 Core Release 2. *Journal of Integrative Bioinformatics* 16: 20190021
- Hucka M, Bergmann FT, Hoops S, Keating SM, Sahle S, Schaff JC, Smith L, Wilkinson DJ (2010) The Systems Biology Markup Language (SBML): Language Specification for Level 3 Version 1 Core. *Nature Precedings*
- Hucka M, Finney A, Sauro HM, Bolouri H, Doyle JC, Kitano H, Arkin AP, Bornstein BJ, Bray D, Cornish-Bowden A, Cuellar AA, Dronov S, Gilles ED, Ginkel M, Gor V, Goryanin II, Hedley WJ, Hodgman TC, Hofmeyr JHHS, Hunter PJ, *et al* (2003) The systems biology markup language (SBML): a medium for representation and exchange of biochemical network models. *Bioinformatics* 19: 524–531
- Hucka M, Nickerson DP, Bader GD, Bergmann FT, Cooper J, Demir E, Garny A, Golebiewski M, Myers CJ, Schreiber F, Waltemath D, Le Novère N (2015) Promoting Coordinated Development of Community-Based Information Standards for Modeling in Biology: The COMBINE Initiative. *Frontiers in Bioengineering and Biotechnology* 3

- Hucka M, Sauro HM, Finney A, Bolouri H, Doyle J, Kitano H (2001) The ERATO Systems Biology Workbench: An integrated environment for multiscale and multitheoretic simulations in systems biology, In *Foundations of Systems Biology*, Kitano H (ed), chapter 6, MIT Press, pp. 125–143
- Hucka M, Smith LP (2016) SBML Level 3 package: Groups, Version 1 Release 1. *Journal of Integrative Bioinformatics* 13: 8–29
- Ideker T, Galitski T, Hood L (2001) A new approach to decoding life: systems biology. *Annual Review of Genomics and Human Genetics* 2: 343–372
- Kacser H (1957) Some Physico-Chemical Aspects of Biological Organisation, In *The Strategy of the Genes: A Discussion of Some Aspects of Theoretical Biology*, Waddington CH (ed), London: George Allen & Unwin, Ltd., pp. 191–249
- Karr JR, Takahashi K, Funahashi A (2015) The principles of whole-cell modeling. *Current Opinion in Microbiology* 27: 18–24
- Kell DB, Mendes P (2008) The markup is the model: reasoning about systems biology models in the Semantic Web era. *Journal of Theoretical Biology* 252: 538–543
- Kirouac DC, Cicali B, Schmidt S (2019) Reproducibility of Quantitative Systems Pharmacology Models: Current Challenges and Future Opportunities. *CPT Pharmacometrics Systems Pharmacology* 8: 205–210
- Kitano H (2000) Perspectives on systems biology. *New Generation Computing* 18: 199–216
- Klipp E, Liebermeister W, Helbig A, Kowald A, Schaber J (2007) Systems biology standards—the community speaks. *Nature Biotechnology* 25: 390–391
- Krause F, Uhlenendorf J, Lubitz T, Schulz M, Klipp E, Liebermeister W (2010) Annotation and merging of SBML models with semanticSBML. *Bioinformatics* 26: 421–422
- Kuepfer L, Peter M, Sauer U, Stelling J (2007) Ensemble modeling for analysis of cell signaling dynamics. *Nature Biotechnology* 25: 1001
- Lambusch F, Waltemath D, Wolkenhauer O, Sandkuhl K, Rosenke C, Henkel R (2018) Identifying frequent patterns in biochemical reaction networks: a workflow. *Database* 2018
- Le Novère N, Finney A, Hucka M, Bhalla US, Campagne F, Collado-Vides J, Crampin EJ, Halstead M, Klipp E, Mendes P, Nielsen P, Sauro H, Shapiro BE, Snoep JL, Spence HD, Wanner BL (2005) Minimum information requested in the annotation of biochemical models (MIRIAM). *Nature Biotechnology* 23: 1509–1015
- Lee DY, Saha R, KhanYusufi FN, Park W, Karimi IA (2009) Web-based applications for building, managing and analysing kinetic models of biological systems. *Briefings in Bioinformatics* 10: 65–74
- Levin SA, Grenfell B, Hastings A, Perelson AS (1997) Mathematical and Computational Challenges in Population Biology and Ecosystems Science. *Science* 275: 334–343
- Li P, Dada JO, Jameson D, Spasic I, Swainston N, Carroll K, Dunn W, Khan F, Malys N, Messiha HL, Simeonidis E, Weichart D, Winder C, Wishart J, Broomhead DS, Goble CA, Gaskell SJ, Kell DB, Westerhoff HV, Mendes P, *et al* (2010) Systematic integration of experimental data and models in systems biology. *BMC Bioinformatics* 11: 582
- Liebermeister W (2008) Validity and combination of biochemical models. In *Proceedings of 3rd International ESCEC Workshop on Experimental Standard Conditions on Enzyme Characterizations*, Kettner C, Hicks MG (eds). Beilstein-Institut

- Lloyd CM, Halstead MDB, Nielsen PF (2004) CellML: its future, present and past. *Progress in Biophysics and Molecular Biology* 85: 433–450 1 2
- Lopez CF, Muhlich JL, Bachman JA, Sorger PK (2013) Programming Biological Models in Python Using PySB. *Molecular Systems Biology* 9: 646 3 4
- Lubitz T, Hahn J, Bergmann FT, Noor E, Klipp E, Liebermeister W (2016) SBtab: A Flexible Table Format for Data Exchange in Systems Biology. *Bioinformatics* 32: 2559–2561 5 6
- Machado D, Costa RS, Rocha M, Ferreira EC, Tidor B, Rocha I (2011) Modeling formalisms in Systems Biology. *AMB Express* 1: 45 7 8
- Malik-Sheriff RS, Glont M, Nguyen TVN, Tiwari K, Roberts MG, Xavier A, Vu MT, Men J, Maire M, Kananathan S, Fairbanks EL, Meyer JP, Arankalle C, Varusai TM, Knight-Schrijver V, Li L, Dueñas-Roca C, Dass G, Keating SM, Park YM, *et al* (2020) BioModels—15 Years of Sharing Computational Models in Life Science. *Nucleic Acids Research* : gkz1055 9 10 11 12
- Mattioni M, Le Novère N (2013) Integration of biochemical and electrical signaling-multiscale model of the medium spiny neuron of the striatum. *PLoS One* 8: e66811 13 14
- Medley JK, Choi K, König M, Smith L, Gu S, Hellerstein J, Sealfon SC, Sauro HM (2018) Tellurium notebooks—An environment for reproducible dynamical modeling in systems biology. *PLoS Computational Biology* 14: e1006220 15 16 17
- Mi H, Huang X, Muruganujan A, Tang H, Mills C, Kang D, Thomas PD (2016) PANTHER version 11: expanded annotation data from Gene Ontology and Reactome pathways, and data analysis tool enhancements. *Nucleic Acids Research* 45: D183–D189 18 19 20
- Mirams GR, Arthurs CJ, Bernabeu MO, Bordas R, Cooper J, Corrias A, Davit Y, Dunn SJ, Fletcher AG, Harvey DG, Marsh ME, Osborne JM, Pathmanathan P, Pitt-Francis J, Southern J, Zemzemi N, Gavaghan DJ (2013) Chaste: an open source C++ library for computational physiology and biology. *PLoS Computational Biology* 9: e1002970 21 22 23 24
- Misirli G, Hallinan J, Wipat A (2014) Composable modular models for synthetic biology. *ACM Journal on Emerging Technologies in Computing Systems JETC* 11: 22 25 26
- Mitra ED, Suderman R, Colvin J, Ionkov A, Hu A, Sauro HM, Posner RG, Hlavacek WS (2019) PyBioNetFit and the Biological Property Specification Language. *iScience* 19: 1012–1036, 00005 27 28
- Moraru II, Morgan F, Li Y, Loew LM, Schaff JC, Lakshminarayana A, Slepchenko BM, Gao F, Blinov ML (2008) Virtual Cell modelling and simulation software environment. *IET Systems Biology* 2: 352–362 29 30
- Naldi A, Monteiro PT, Müssel C, Consortium for Logical Models and Tools, Kestler HA, Thieffry D, Xenarios I, Saez-Rodriguez J, Helikar T, Chaouiya C (2015) Cooperative development of logical modelling standards and tools with CoLoMoTo. *Bioinformatics* 31: 1154–1159 31 32 33
- Neal ML, König M, Nickerson D, Mısırlı G, Kalbasi R, Dräger A, Atalag K, Chelliah V, Cooling MT, Cook DL, Crook S, de Alba M, Friedman SH, Garny A, Gennari JH, Gleeson P, Golebiewski M, Hucka M, Juty N, Myers C, *et al* (2019) Harmonizing semantic annotations for computational models in biology. *Briefings in Bioinformatics* 20: 540–550 34 35 36 37
- Nickerson D, Atalag K, de Bono B, Geiger J, Goble C, Hollmann S, Lonien J, Müller W, Regierer B, Stanford NJ, Golebiewski M, Hunter P (2016) The Human Physiome: how standards, software and innovative service infrastructures are providing the building blocks to make it achievable. *Interface Focus* 6: 20150103 38 39 40

- Nickerson DP, Hunter PJ (2017) Introducing the physiome journal: improving reproducibility, reuse, and discovery of computational models. In *2017 IEEE 13th International Conference on e-Science (e-Science)*. IEEE 1  
2  
3
- Norsigian CJ, Pusarla N, McConn JL, Yurkovich JT, Dräger A, Palsson BO, King Z (2019) BiGG Models 2020: multi-strain genome-scale models and expansion across the phylogenetic tree. *Nucleic Acids Research* 4  
5
- O'Brien EJ, Monk JM, Palsson BØ (2015) Using genome-scale models to predict biological capabilities. *Cell* 161: 971–987 6  
7
- Olivier BG, Bergmann FT (2018) SBML Level 3 Package: Flux Balance Constraints version 2. *Journal of Integrative Bioinformatics* 15: 20170082 8  
9
- Olivier BG, Rohwer JM, Hofmeyr JHS (2005) Modelling cellular systems with PySCeS. *Bioinformatics* 21: 560–561 10  
11
- Palmisano A, Hoops S, Watson LT, Jones Jr TC, Tyson JJ, Shaffer CA (2014) Multistate Model Builder (MSMB): a flexible editor for compact biochemical models. *BMC Systems Biology* 8: 42 12  
13
- Peters M, Eicher JJ, van Niekerk DD, Waltemath D, Snoep JL (2017) The JWS Online simulation database. *Bioinformatics* 33: 1589–1590 14  
15
- Ravikrishnan A, Raman K (2015) Critical assessment of genome-scale metabolic networks: the need for a unified standard. *Briefings in Bioinformatics* 16: 1057–1068 16  
17
- Rodriguez N, Pettit JB, Dalle Pezze P, Li L, Henry A, van Iersel MP, Jalowicki G, Kutmon M, Natarajan KN, Tolnay D, Stefan MI, Evelo CT, Le Novère N (2016) The systems biology format converter. *BMC Bioinformatics* 17 18  
19  
20
- Rodriguez N, Thomas A, Watanabe L, Vazirabad IY, Kofia V, Gómez HF, Mittag F, Matthes J, Rudolph JD, Wrzodek F, Netz E, Diamantikos A, Eichner J, Keller R, Wrzodek C, Fröhlich S, Lewis NE, Myers CJ, Le Novère N, Palsson BØ, *et al* (2015) JSBML 1.0: providing a smorgasbord of options to encode systems biology models. *Bioinformatics* 31: 3383–3386 21  
22  
23  
24
- Roehner N, Beal J, Clancy K, Bartley B, Misirli G, Grünberg R, Oberortner E, Pocock M, Bissell M, Madsen C, Nguyen T, Zhang M, Zhang Z, Zundel Z, Densmore D, Gennari JH, Wipat A, Sauro HM, Myers CJ (2016) Sharing Structure and Function in Biological Design with SBOL 2.0. *ACS Synthetic Biology* 5: 498–506 25  
26  
27
- Roehner N, Myers CJ (2014) A methodology to annotate systems biology markup language models with the synthetic biology open language. *ACS Synthetic Biology* 3: 57–66 28  
29
- Sansone SA, McQuilton P, Rocca-Serra P, Gonzalez-Beltran A, Izzo M, Lister A, Thurston M, Batista D, Granell R, Adekale M, Dauga D, Ganley E, Hodson S, Lawrence R, Khodiyar V, Tenenbaum J, Axton JM, Ball M, Besson S, Bloom T, *et al* (2019) FAIRsharing as a community approach to standards, repositories and policies. *Nature Biotechnology* 37: 358–367 30  
31  
32  
33
- Scharm M, Waltemath D, Mendes P, Wolkenhauer O (2016a) COMODI: an ontology to characterise differences in versions of computational models in biology. *Journal of Biomedical Semantics* 7: 46 34  
35
- Scharm M, Wolkenhauer O, Waltemath D (2016b) An algorithm to detect and communicate the differences in computational models describing biological systems. *Bioinformatics* 32: 563–570 36  
37
- Schliess F, Hoehme S, Henkel SG, Ghallab A, Driesch D, Böttger J, Guthke R, Pfaff M, Hengstler JG, Gebhardt R, Häussinger D, Drasdo D, Zellmer S (2014) Integrated metabolic spatial-temporal model for the prediction of ammonia detoxification during liver damage and regeneration. *Hepatology* 60: 2040–2051 38  
39  
40  
41

- Smith LP, Bergmann FT, Chandran D, Sauro HM (2009) Antimony: A Modular Model Definition Language. *Bioinformatics* 25: 2452–2454 1
- Smith LP, Hucka M, Hoops S, Finney A, Ginkel M, Myers CJ, Moraru I, Liebermeister W (2015) SBML Level 3 package: Hierarchical Model Composition, Version 1 Release 3. *Journal of Integrative Bioinformatics* 12: 268 3
- Stanford NJ, Wolstencroft K, Golebiewski M, Kania R, Juty N, Tomlinson C, Owen S, Butcher S, Hermjakob H, Le Novère N, Mueller W, Snoep J, Goble C (2015) The evolution of standards and data management practices in systems biology. *Molecular Systems Biology* 11: 851 6
- Swat MH, Thomas GL, Belmonte JM, Shirinifard A, Hmeljak D, Glazier JA (2012) Multi-scale modeling of tissues using CompuCell3D, In *Methods in Cell Biology*, volume 110, Elsevier, pp. 325–366 9
- Tanaka S, Sichau D, Iber D (2015) LBIBCell: a cell-based simulation environment for morphogenetic problems. *Bioinformatics* 31: 2340–2347 11
- Terfve C, Cokelaer T, Henriques D, MacNamara A, Goncalves E, Morris MK, van Iersel M, Lauffenburger DA, Saez-Rodriguez J (2012) CellNOptR: a flexible toolkit to train protein signaling networks to data using multiple logic formalisms. *BMC Systems Biology* 6: 133 13
- The UniProt Consortium (2017) UniProt: the universal protein knowledgebase. *Nucleic Acids Research* 45: D158–D169 16
- Thiele I, Fleming RMT, Que R, Bordbar A, Diep D, Palsson BO (2012) Multiscale Modeling of Metabolism and Macromolecular Synthesis in *E. coli* and Its Application to the Evolution of Codon Usage. *PLOS ONE* 7: e45635 18
- Turing AM (1952) The Chemical Basis of Morphogenesis. *Philosophical Transactions of the Royal Society of London Series B Biological Sciences* 237: 5–72 21
- Uhlen M, Zhang C, Lee S, Sjöstedt E, Fagerberg L, Bidkhori G, Benfeitas R, Arif M, Liu Z, Edfors F, Sanli K, von Feilitzen K, Oksvold P, Lundberg E, Hober S, Nilsson P, Mattsson J, Schwenk JM, Brunnström H, Glimelius B, *et al* (2017) A pathology atlas of the human cancer transcriptome. *Science* 357: eaan2507 23
- van Iersel MP, Villéger AC, Czauderna T, Boyd SE, Bergmann FT, Luna A, Demir E, Sorokin A, Dogrusoz U, Matsuoka Y, Funahashi A, Aladjem MI, Mi H, Moodie SL, Kitano H, Le Novère N, Schreiber F (2012) Software support for SBGN maps: SBGN-ML and LibSBGN. *Bioinformatics* 28: 2016–2021 26
- Varela PL, Ramos CV, Monteiro PT, Chaouiya C (2019) EpiLog: A software for the logical modelling of epithelial dynamics [version 2; peer review: 3 approved]. *F1000Research* 7: 1145 29
- Voigt MA, Dräger A, Lloyd C, King ZA, Yang L (2018) draeger-lab/SBMLme: SBMLme converter (Version 0.0.6). Available from Zenodo at <https://doi.org/10.5281/zenodo.1238905> 31
- Von Bertalanffy L (1950) An outline of general system theory. *The British Journal for the Philosophy of Science* 1: 134–165 33
- Waltemath D, Adams R, Bergmann FT, Hucka M, Kolpakov F, Miller AK, Moraru II, Nickerson D, Sahle S, Snoep JL, Le Novère N (2011) Reproducible computational biology experiments with SED-ML—the Simulation Experiment Description Markup Language. *BMC Systems Biology* 5: 198 35
- Waltemath D, Bergmann FT, Chaouiya C, Czauderna T, Gleeson P, Goble C, Golebiewski M, Hucka M, Juty N, Krebs O, Le Novère N, Mi H, Moraru II, Myers CJ, Nickerson D, Olivier BG, Rodriguez N, Schreiber F, Smith L, Zhang F, *et al* (2014) Meeting report from the fourth meeting of the Computational Modeling in Biology Network (COMBINE). *Standards in Genomic Sciences* 9: 1285–1301 38

Waltemath D, Wolkenhauer O (2016) How modeling standards, software, and initiatives support reproducibility in systems biology and systems medicine. <i>IEEE Transactions on Biomedical Engineering</i> 63: 1999–2006	1 2 3
Wang Y, Eddy JA, Price ND (2012) Reconstruction of genome-scale metabolic models for 126 human tissues using mCADRE. <i>BMC Systems Biology</i> 6: 153	4 5
Watanabe LH, König M, Myers CJ (2018) Dynamic Flux Balance Analysis Models in SBML. Technical report	6
Watanabe LH, Myers CJ (2016) Efficient Analysis of Systems Biology Markup Language Models of Cellular Populations Using Arrays. <i>ACS Synthetic Biology</i> 5: 835–841	7 8
Weidemann A, Richter S, Stein M, Sahle S, Gauges R, Gabdoulline R, Surovtsova I, Semmelrock N, Besson B, Rojas I, Wade R, Kummer U (2008) SYCAMORE—a systems biology computational analysis and modeling research environment. <i>Bioinformatics</i> 24: 1463–1464	9 10 11
Wittig U, Rey M, Weidemann A, Kania R, Müller W (2017) SABIO-RK: an updated resource for manually curated biochemical reaction kinetics. <i>Nucleic Acids Research</i> 46: D656–D660	12 13
Wolstencroft K, Krebs O, Snoep JL, Stanford NJ, Bacall F, Golebiewski M, Kuzyakiv R, Nguyen Q, Owen S, Soiland-Reyes S, Straszewski J, van Niekerk D, Williams A, Malmström L, Rinn B, Müller W, Goble C (2016) FAIRDOMHub: a repository and collaboration environment for sharing systems biology research. <i>Nucleic Acids Research</i> 45: D404–D407	14 15 16 17
Zhang F, Angermann BR, Meier-Schellersheim M (2013) The Simmune Modeler visual interface for creating signaling networks based on bi-molecular interactions. <i>Bioinformatics</i> 29: 1229–1230	18 19
Zhang F, Meier-Schellersheim M (2018) SBML Level 3 package: Multistate, Multicomponent and Multi-compartment Species, Version 1, Release 1. <i>Journal of Integrative Bioinformatics</i> 15: 20170077	20 21