

pyKNEEr: An image analysis workflow for open and reproducible research on femoral knee cartilage

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<https://sbonaretti.github.io/>

Open and reproducible research

- Openness and computational reproducibility are recent movements in research ([Woelfle 2011](#), [Bollen 2015](#))
- They are essential to researchers to assess value of scientific claims and confidently build on previous work ([Sandve 2013](#))
- Transparent *image-based* research is crucial to provide reliable answers to biological questions ([Poldrack 2017](#))
- Other communities, e.g. neuroimaging, largely benefit from open-access tools ([Gorgolewski 2011](#))

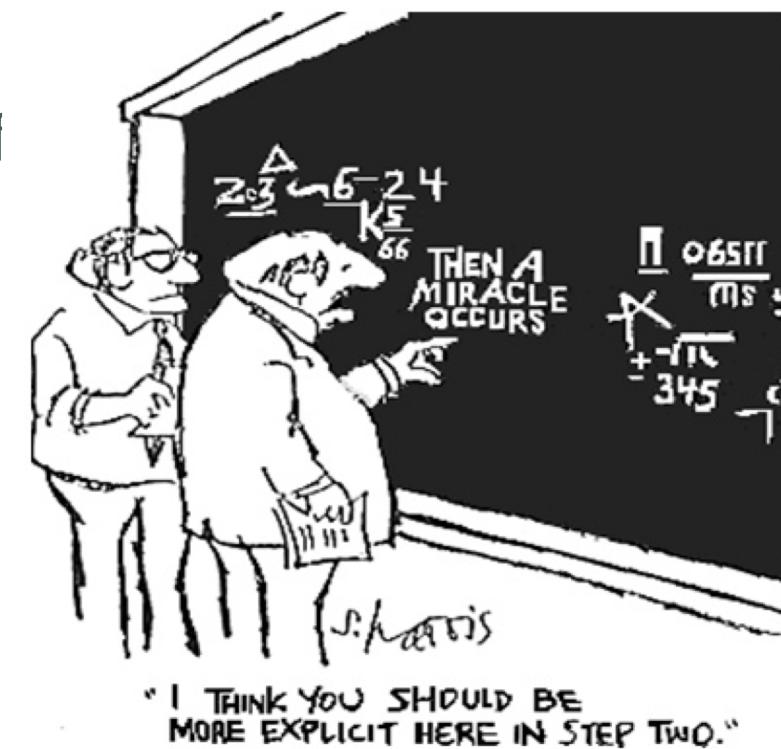
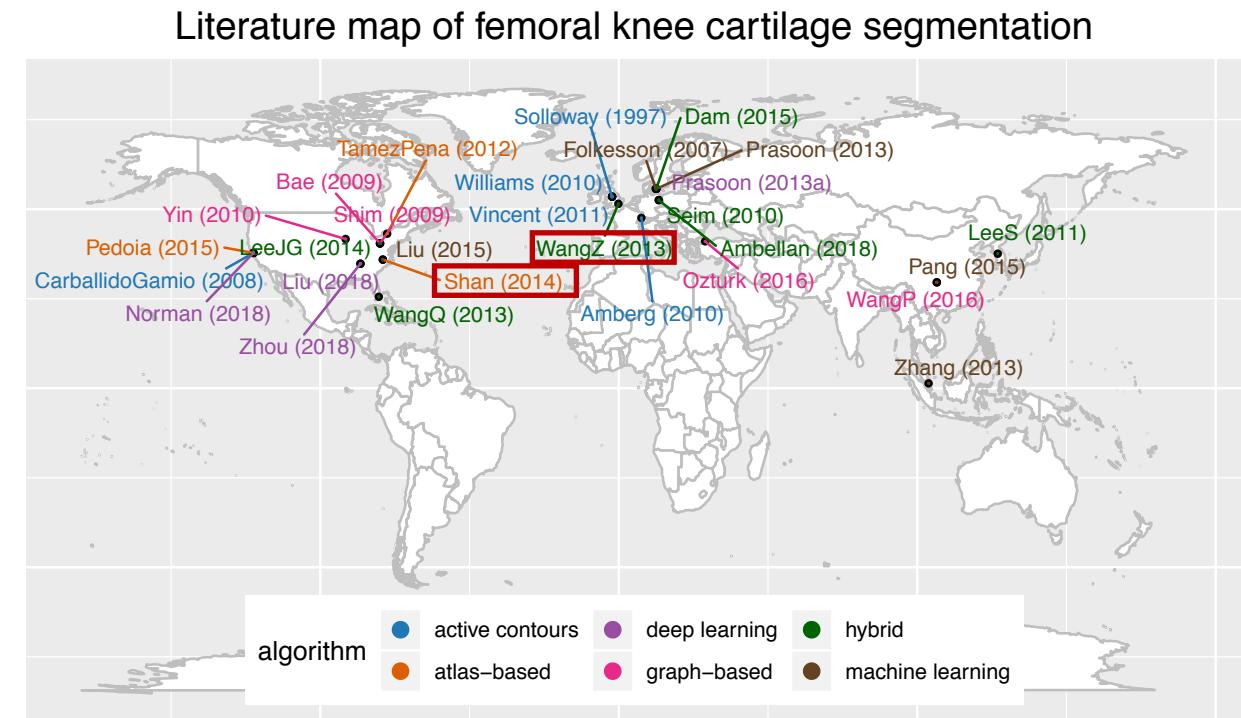


Image-based analysis of femoral knee cartilage

- In knee OA, analysis of femoral cartilage from MR images has assumed an important role ([Hafezi-Nejad 2018](#))
- Analysis workflows include:
 - Preprocessing
 - Segmentation
 - Morphology and Relaxometry
- Segmentation is a major challenge
 - Of 29 segmentation algorithms, only 2 have open source code ([Wang 2013](#), [Shan 2014](#))

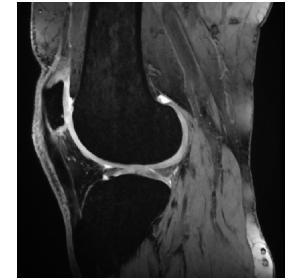


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Openness of pyKNEEr

- Code written in *python*
- Use of open file formats
 - *.mha, .txt, .csv*
- Modular structure to favor code expansion
- Reuse of developed code
 - Intensity preprocessing from Shan et al., *elastix*
- Available on GitHub
 - GNU GPLv3 license and DOI

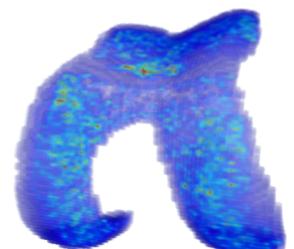
I. Preprocessing



II. Segmentation



III. Analysis



Reproducibility: Jupyter notebook

```
pyKNEEr
Relaxometry of Femoral Knee Cartilage
Exponential and linear fitting
• Exponential fitting is computationally expensive but more accurate
• Linear fitting is faster as data are transformed to their log and then linearly interpolated. However, linear fitting is less accurate because the nonlinear logarithmic transform provides larger weight to outliers

The fitting is computed:
• directly on the acquired images or after rigid registration of the following echo to the first echo
• voxel-wise, i.e. for each voxel the Echo Times (dcm tag: [0018,0081]) are the x-variable and the voxel intensities in each acquisition are the y-variable
• only in the mask volume to have short computation time

Image information
Inputs:
• input_file_name contains the list of the images used to calculate the relaxation maps
• method_flag is 0 if fitting is linear, 1 if fitting is exponential
• registration_flag is 0 for no registration, 1 for rigid registration
• n_of_cores = 4
• output_file_name = "exp_fit_aligned_OA1_T2.csv"

In [1]: input_file_name = "image_list_relaxometry_fitting_OA1_T2.txt"
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Read image data
• image_data is a dictionary (or struct), where each cell corresponds to an image. For each image, information such as paths and file names are stored

In [2]: image_data = io.load_image_data_fitting(input_file_name, method_flag, registration_flag)

Calculate fitting maps
Align acquisitions
Images are aligned rigidly to remove occasional subject motion among acquisitions
Note: This step is optional and can be skipped, given that:
• When images are aligned, the fitting is calculated on interpolated values obtained with rigid registration
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In [3]: if registration_flag == 1:
    rel.align_acquisitions(image_data, n_of_cores)

Compute the fitting
In [4]: rel.calculate_fitting_maps(image_data, n_of_cores)

Visualize fitting maps
2D MAP: For each image, fitting maps at medial and lateral compartments and flattened map
The flattened map is an average of neighboring voxels projected on the bone surface side of the femoral cartilage

In [5]: rel.show_fitting_maps(image_data)

3D MAP: Interactive rendering of fitting maps
(The error message "Error creating widget: could not find model" can appear when the notebook is moved to a different folder)

In [6]: # ID of the map to visualize (The ID is the one in the 2D visualization above)
image_ID = 1 - 1 # -1 because counting starts from 0

# read image
file_name = image_data[image_ID]["relaxometryFolder"] + image_data[image_ID]["mapFileName"]
image = itk.imread(file_name)

# view
viewer = view(image, gradient_opacity=0.0, ui_collapsed=False, shadow=False)
viewer

GRAPH: Dots represent the average value of fitting maps per image; bars represents the standard deviation
In [7]: rel.show_fitting_graph(image_data)

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- Open-source web application integrating
 - Live code
 - Narrative text with equations
 - Visualizations
- Versatile
- Easy to share among researchers

Reproducibility of pyKNEEr: Jupyter notebook

1. Link to GitHub repository

pyKNEEr

[Relaxometry of Femoral Knee Cartilage](#)

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<https://github.com/sbonaretti/pyKNEEr>

sbonaretti / pyKNEEr

[Code](#) [Issues 0](#) [Pull requests 0](#) [Projects 0](#) [Wiki](#) [Insights](#)

An image analysis workflow for open and reproducible research on femoral knee cartilage

[Manage topics](#)

132 commits

1 branch

0 releases

Branch: master [New pull request](#) [Create new](#)

 sbonaretti Update README.md

 code

 docs

 publication

 .DS_Store

 README.md

 environment.yml

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pyKNEEr

Introduction
Installation
Preprocessing
Segmentation
Morphology
Relaxometry
FAQ

MR acquisition Segmentation Cartilage thickness T₂ map

pyKNEEr is an image analysis workflow for **open** and **reproducible** research on **femoral knee cartilage**
It is implemented in **python** with **Jupyter notebooks**

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In [ ]: input_file_name = "image_list_relaxometry_fitting_0A11_T2.txt"
method_flag = 1 # 0 = linear, 1 = exponential
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n_of_cores = 4
output_file_name = "exp_fit_aligned_0A11_T2.csv"

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4. User inputs

input_file_name

```
./original/
001/BL
left
002/BL
left
003/BL
right
004/BL
left
005/BL
right
006/BL
left
007/BL
left
008/BL
left
009/BL
right
010/BL
left
011/BL
left
012/BL
right
```

n_of_cores

output_file_name

• • •

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watermark -v -m SimpleITK,matplotlib,numpy,pandas,scipy,itkwidgets,multiprocessing
```

- 1. Link to GitHub repository
- 2. Link to documentation

3. Introduction

4. User inputs

5. Commands with narrative

Cartilage Thickness

Separating subcondral surface and articular surface of cartilage

To calculate cartilage thickness, first the cartilage surface is extracted from the binary mask. Then subcondral surface and articular surface are divided in two separate point clouds

```
morph.separate_cartilage_surfaces(image_data, n_of_cores)
```

Visual check

Subcondral bone surface (yellow) and articular surface (blue) are visualized as flattened point clouds. The flattening is with respect to a cylinder interpolated into the cartilage surface [2]

```
morph.show_cartilage_surfaces(image_data)
```

Calculating cartilage thickness

Assign the chosen algorithm

```
morph.algorithm(image_data, thickness_algo)
```

Calculate thickness

```
morph.calculate_thickness(image_data, n_of_cores)
```

Reproducibility of pyKNEEr: Jupyter notebook

```
pyKNEEr
Relaxometry of Femoral Knee Cartilage

Exponential and linear fitting

- Exponential fitting is computationally expensive but more accurate
- Linear fitting is faster as data are transformed to their log and then linearly interpolated. However, linear fitting is less accurate because the nonlinear logarithmic transform provides larger weight to outliers


The fitting is computed:


- directly on the acquired images or after rigid registration of the following echo to the first echo
- voxel-wise, i.e. for each voxel the Echo Times (dicom tag: (0018,0081) are the x-variable and the voxel intensities in each acquisition are the y-variable
- only in the mask volume to have short computation time

Image information
Inputs:


- input_file_name = "image_list_relaxometry_fitting_OAII_T2.txt"
- method_flag = 0 = exponential
- registration_flag = 1 # = no rigid registration, 1 = execute rigid registration
- n_of_cores = 4
- output_file_name = "exp_fit_aligned_OAII_T2.csv"


In [ ]: input_file_name = "image_list_relaxometry_fitting_OAII_T2.txt"
method_flag = 0 = exponential
registration_flag = 1 # = no rigid registration, 1 = execute rigid registration
n_of_cores = 4
output_file_name = "exp_fit_aligned_OAII_T2.csv"

Read image data


- image_data is a dictionary (or struct), where each cell corresponds to an image. For each image, information such as paths and file names are stored


In [ ]: image_data = io.load_image_data_fitting(input_file_name, method_flag, registration_flag)

Calculate fitting maps
Align acquisitions
Images are aligned rigidly to remove occasional subject motion among acquisitions
Note: This step is optional and can be skipped, given that:


- When images are aligned, the fitting is calculated on interpolated values obtained with rigid registration
- When images are not aligned, the fitting is calculated on original intensities, but images might not be aligned


In [ ]: if registration_flag == 1:
    rel.align_acquisitions(image_data, n_of_cores)

Compute the fitting
In [ ]: rel.calculate_fitting_maps(image_data, n_of_cores)

Visualizing fitting maps
2D MAP: For each image, fitting maps at medial and lateral compartments and flattened map
The flattened map is an average of neighboring voxels projected on the bone surface side of the femoral cartilage
In [ ]: rel.show_fitting_maps(image_data)

3D MAP: Interactive rendering of fitting maps
(The error message "Error creating widget: could not find model" can appear when the notebook is moved to a different folder)
In [ ]: # ID of the map to visualize (The ID is the one in the 2D visualization above)
image_ID = 1 - 1 # because counting starts from 0

# read image
file_name = image_data[image_ID]['relaxometryFolder'] + image_data[image_ID]['mapFileName']
image = itk.imread(file_name)

# view
viewer = view(image, gradient_opacity=0.0, ui_collapsed=False, shadow=False)
viewer

GRAPH: Dots represent the average value of fitting maps per image; bars represents the standard deviation
In [ ]: rel.show_fitting_graph(image_data)

TABLE: Average and standard deviation of fitting maps per image
The table is saved as a csv file for subsequent analysis
In [ ]: rel.show_fitting_table(image_data, output_file_name)

References
[1] Borhani A., Wheaton A.J., Gopujadas A.J., Akella S.V., Regatte R.R., Charagundla S.R., Reddy R. In vivo measurement of T1 and T2 dispersion in the human brain at 1.5 Tesla. J Magn Reson Imaging. Apr;19(4):403-9. 2004.
[2] Li X, Benjamin Ma C, Link TM, Castillo D.D., Blumenkrantz G., Lozano J., Carballido-Gamio J., Ries M., Majumdar S. In vivo T1 and T2 mapping of articular cartilage in osteoarthritis of the knee using 3 T MRI. Osteoarthritis Cartilage. Jul;15(7):789-97. 2007.

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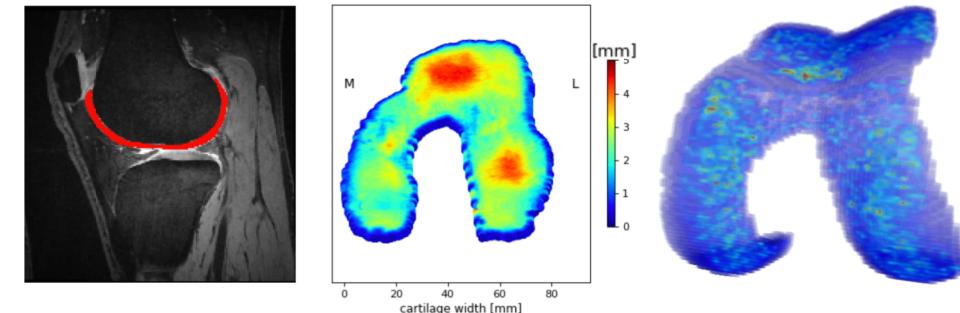
3. Introduction

4. User inputs

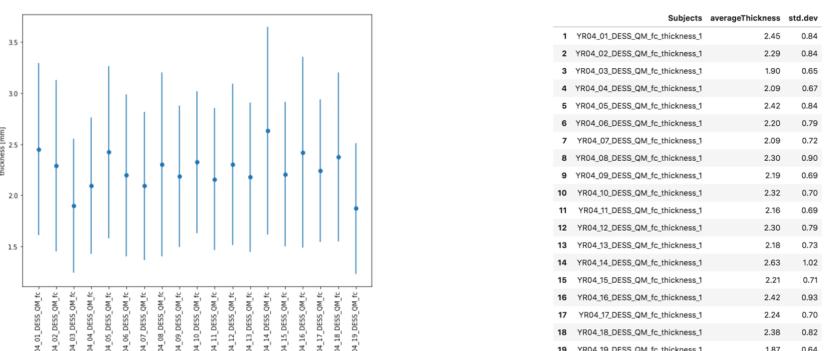
5. Commands with narrative

6. Visualization of outputs

Qualitative visualizations



Quantitative visualizations



Reproducibility of pyKNEEr: Jupyter notebook

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Relaxometry of Femoral Knee Cartilage

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- only in the mask volume to have short computation time

Image information
Inputs:


- input_file_name contains the list of the images used to calculate the relaxation maps
- method_flag is 0 if fitting is linear, 1 if fitting is exponential
- registration_flag is 0 for no registration, 1 for rigid registration
- output_file_name contains average and standard deviation of the fitting maps


In [1]: input_file_name = "image_list_relaxometry_fitting_0A11_T2.txt"
method_flag = 1 # 0 = linear, 1 = exponential
registration_flag = 1 # 0 = no rigid registration, 1 = execute rigid registration
n_of_cores = 4
output_file_name = "exp_fit_aligned_0A11_T2.csv"

Read image data

- image_data is a dictionary (or struct), where each cell corresponds to an image. For each image, information such as paths and file names are stored


In [2]: image_data = io.load_image_data_fitting(input_file_name, method_flag, registration_flag)

Calculate fitting maps
Align acquisitions

Images are aligned rigidly to remove occasional subject motion among acquisitions



Note: This step is optional and can be skipped, given that:



- When images are aligned, the fitting is calculated on interpolated values obtained with rigid registration
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In [3]: if registration_flag == 1:
    rel.align_acquisitions(image_data, n_of_cores)

Compute the fitting
In [4]: rel.calculate_fitting_maps(image_data, n_of_cores)

Visualize fitting maps

2D MAP: For each image, fitting maps at medial and lateral compartments and flattened map



The flattened map is an average of neighboring voxels projected on the bone surface side of the femoral cartilage


In [5]: rel.show_fitting_maps(image_data)



3D MAP: Interactive rendering of fitting maps



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In [6]: # ID of the map to visualize (the ID is the one in the 2D visualization above)
image_ID = 1 - 1 # -1 because counting starts from 0

# read image
file_name = image_data[image_ID]["relaxometryFolder"] + image_data[image_ID][“mapFileName”]
image = itk.imread(file_name)

# view
viewer = view(image, gradient_opacity=0.0, ui_collapsed=False, shadow=False)
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GRAPH: Dots represent the average value of fitting maps per image; bars represents the standard deviation


In [7]: rel.show_fitting_graph(image_data)



TABLE: Average and standard deviation of fitting maps per image



The table is saved as a csv file for subsequent analysis


In [8]: rel.show_fitting_table(image_data, output_file_name)

References
[1] Borhani A., Weston A.J., Gopaulas A.J., Akella S.V., Regatte R.R., Charagundla S.R., Reddy R. In vivo measurement of T1/T2 dispersion in the human brain at 1.5 Tesla. J Magn Reson Imaging. Apr;19(4):403-9. 2004.
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3. Introduction

4. User inputs

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```

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- 2. Link to documentation

3. Introduction

4. User inputs

5. Commands with narrative

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Dependencies

```
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%watermark -v -m -p matplotlib,numpy,pandas,scipy
```

CPython 3.7.1
IPython 7.2.0

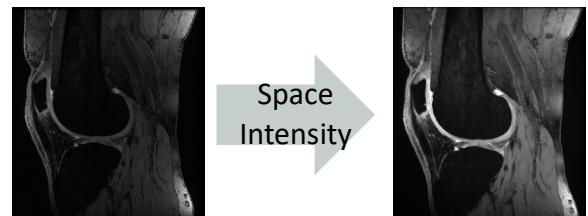
matplotlib 2.2.3
numpy 1.16.1
pandas 0.24.1
scipy 1.2.1

compiler : Clang 4.0.1 (tags/RELEASE_401/final)
system : Darwin
release : 17.7.0
machine : x86_64
processor : i386
CPU cores : 4
interpreter: 64bit

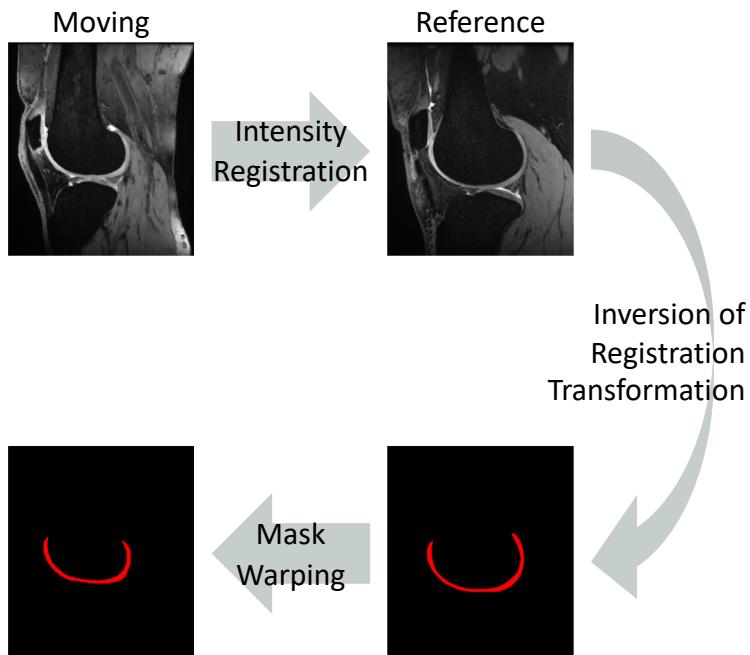
→ Reproducibility of computational environment

Algorithms in *pyKNEEr*

I. Preprocessing

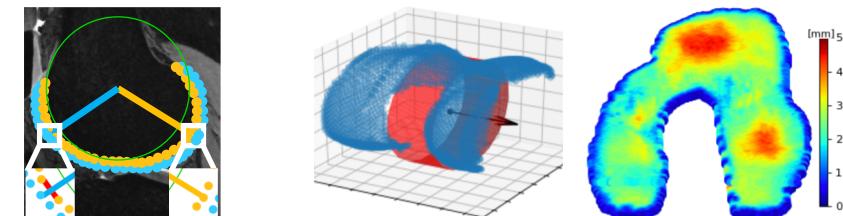


II. Segmentation

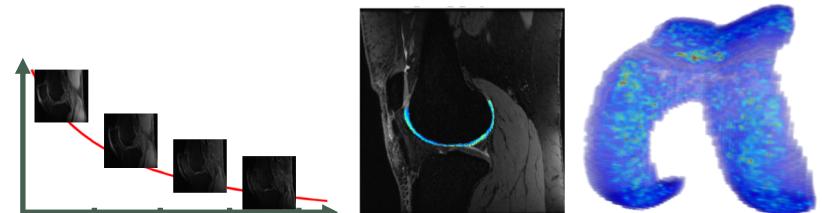


III. Analysis

Morphology



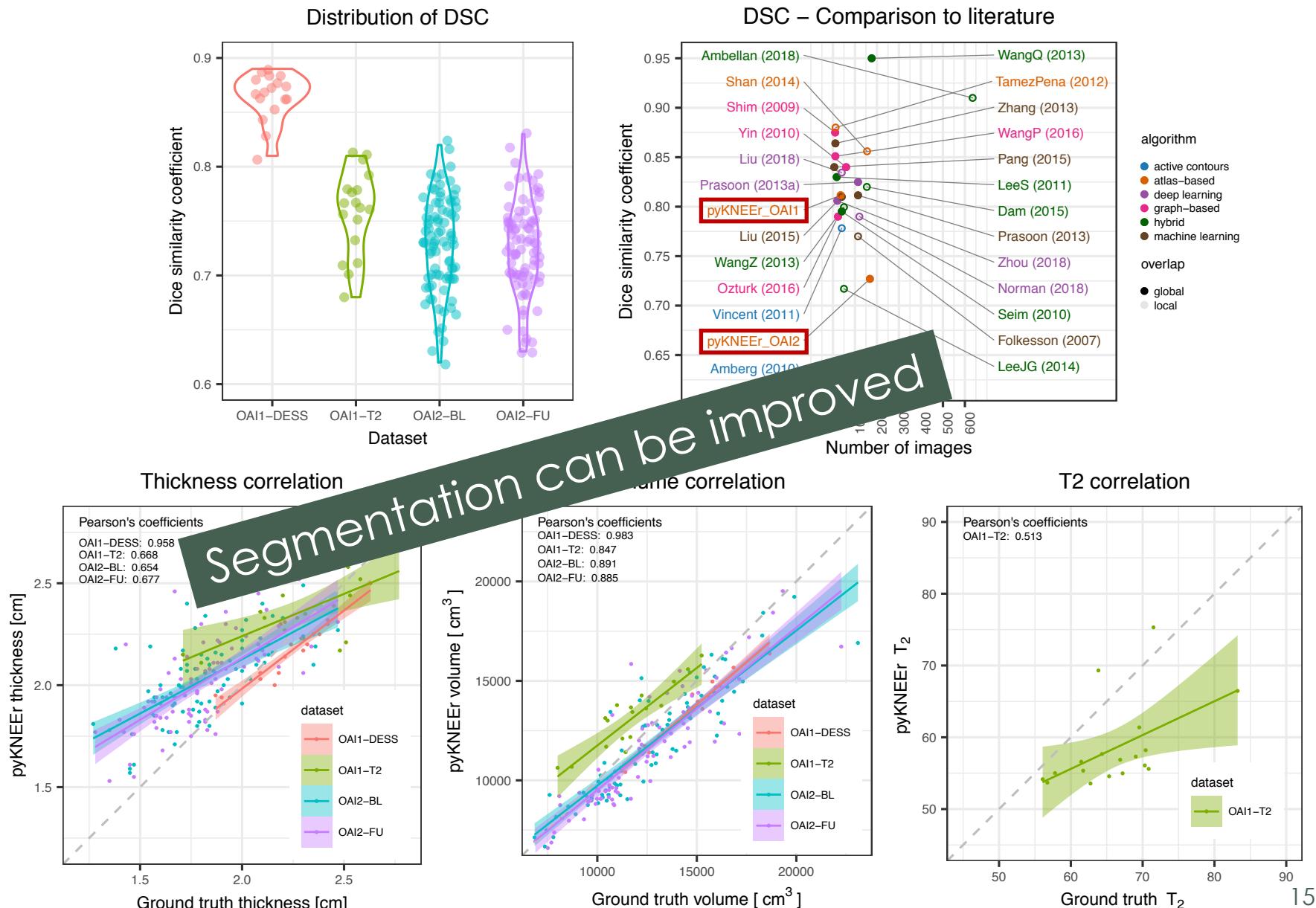
Relaxometry



Validation experiments

- OAI1: 19 subjects
 - OAI1-DESS
 - OAI1-T2
 - G. Truth: atlas-based
([TamezPena 2012](#))
- OAI2: 88 subjects
 - OAI2-DESS BL
 - OAI2-DESS FU
 - G. Truth: AAM
([Vincent 2011](#))

Data on [Zenodo](#)
Code on [GitHub](#)
Analysis on [binder](#)
Paper on [bioRxiv](#)



Discussion

- pyKNEEr can facilitate transparent research
 - Open source
 - Notebooks can be shared among researchers
 - Notebooks can be linked to publications
- Do you want to use pyKNEEr?
 - Go to terminal and type: `pip install pyKNEEr`
 - Download Jupyter notebooks from [GitHub](#)
 - Follow the instruction on the [documentation](#) (demo)
- Do you want to contribute to pyKNEEr?
 - Go to [GitHub](#) and create a branch
 - Implement your contribution
 - Segmentation: Machine and deep learning, hybrid, ...
 - Thickness algorithms: surface normal vectors, potential field lines, ...
 - Other knee tissues: tibial cartilage, patellar cartilage, menisci, ...
 - Send a pull request and your code will be merged to the main project

The screenshot shows the GitHub repository page for 'pyKNEEr'. At the top, there's a commit by 'sbonaretti' updating 'README.md' two days ago. Below the commit history is a list of files with their last update times: 'code' (4 days ago), 'docs' (3 days ago), 'publication' (3 days ago), '.DS_Store' (4 days ago), 'LICENSE' (4 days ago), 'README.md' (2 days ago), and 'environment.yml' (a month ago). The repository title is 'pyKNEEr: An image analysis workflow for open and reproducible research on femoral knee cartilage'. It includes links to 'Try pyKNEEr on Binder!', 'Documentation on website', 'Preprint paper on bioRxiv', and a DOI link 'DOI: 10.5281/zenodo.2574172'.

<https://github.com/sbonaretti/pyKNEEr>

Acknowledgments

- Uche Monu for her insights on cartilage segmentation
- Bragi Sveinsson for sharing his code on EPG modeling
- Valentina Mazzoli for her insight on relaxometry
- Christof Seiler for software engineering support
- Amy Silder, Julie Kolesar, and Scott Uhlrich for beta testing
- All the researchers that make their code open-source, on whose effort we could create pyKNEEr
- Merit Review Award Number I01 RX001811 from the United States Department of Veterans Affairs Rehabilitation R&D Service
- NIH EB002524 and NIH AR062068

pyKNEEr: An image analysis workflow for open and reproducible research on femoral knee cartilage

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²Musculoskeletal Research Laboratory, VA Palo Alto Health Care System, CA, USA

³Department of Bioengineering, Stanford University, Stanford, CA, USA