# Modeling with k-means, Part 4

In Part 4 I continue to apply the methods of Parts 1, 2, and 3. The difference is that here we are looking for 3 clusters rather than just 2.

As in Part 2, since there are so few records in the *wine* dataset (178 records, or samples), I will include all records in the training set data and compare model performance based on cross-validation scores.

The wine samples represent three varietals.

\* \* \* \* \*

## **Preliminaries**

```
In []: require(rattle)
    require(car)
    require(gpplot2)
    require(stringr)
    require(parallel)
    require(faraway)
    require(randomForest)
    require(plyr)
    require(e1071)
    require(xgboost)
In [2]: options(digits= 5, show.signif.stars= FALSE)
```

#### Load the data

```
In [3]: # Load the data. The wine dataset is found in
# package rattle.

data(wine, package="rattle")
dim(wine)
colnames(wine)
```

178 14

'Type' 'Alcohol' 'Malic' 'Ash' 'Alcalinity' 'Magnesium' 'Phenols' 'Flavanoids' 'Nonflavanoids' 'Proanthocyanins' 'Color' 'Hue' 'Dilution' 'Proline'

```
In [4]: # Note that all of the predictors are numeric (continuous).
# Note that Type is a factor.
head(wine[, 1:14])
```

A data.frame: 6 × 14

	Type	Alcohol	Malic	Ash	Alcalinity	Magnesium	Phenols	Flavanoids	Nonflavanoids	Proanthocyanins	Color	Hue
	<fct></fct>	<dbl></dbl>	<dbl></dbl>	<dbl></dbl>	<dbl></dbl>	<int></int>	<dbl></dbl>	<dbl></dbl>	<dbl></dbl>	<dbl></dbl>	<dbl></dbl>	<dbl></dbl>
1	1	14.23	1.71	2.43	15.6	127	2.80	3.06	0.28	2.29	5.64	1.04
2	1	13.20	1.78	2.14	11.2	100	2.65	2.76	0.26	1.28	4.38	1.05
3	1	13.16	2.36	2.67	18.6	101	2.80	3.24	0.30	2.81	5.68	1.03

```
Malic
               Type Alcohol
                                        Ash Alcalinity Magnesium Phenols Flavanoids Nonflavanoids Proanthocyanins Color
                                                                                                                                      Hue
                                                                                     <dbl>
               <fct>
                        <dbl>
                               <dbl>
                                      <dbl>
                                                  <dbl>
                                                               <int>
                                                                        <dbl>
                                                                                                    <dbl>
                                                                                                                      <dbl> <dbl>
                                                                                                                                     <dbl>
                        14 37
                                1 95
                                        2 50
                                                   16.8
                                                                113
                                                                         3 25
                                                                                      3 49
                                                                                                     n 24
                                                                                                                        2 12
                                                                                                                              7 80
                                                                                                                                      በ ጸ6
In [5]: tail(wine)
           A data.frame: 6 x 14
                                  Malic
                                          Ash Alcalinity Magnesium Phenols Flavanoids Nonflavanoids Proanthocyanins Color
                                                                                                                                        Нι
                  Type Alcohol
                  <fct>
                          <dbl>
                                 <dbl>
                                         <dbl>
                                                    <dbl>
                                                                 <int>
                                                                           <dbl>
                                                                                       <dbl>
                                                                                                      <dbl>
                                                                                                                        <dbl> <dbl>
                                                                                                                                       <db
            173
                     3
                           14.16
                                   2.51
                                          2.48
                                                     20.0
                                                                   91
                                                                            1.68
                                                                                        0.70
                                                                                                        0.44
                                                                                                                          1.24
                                                                                                                                  9.7
                                                                                                                                        0.6
            174
                     3
                           13.71
                                   5.65
                                          2.45
                                                     20.5
                                                                   95
                                                                            1.68
                                                                                        0.61
                                                                                                        0.52
                                                                                                                          1.06
                                                                                                                                  7.7
                                                                                                                                        9.0
            175
                     3
                           13 40
                                   3 91
                                          2 48
                                                     23.0
                                                                   102
                                                                            1 80
                                                                                        0.75
                                                                                                        0.43
                                                                                                                          1 41
                                                                                                                                  7.3
                                                                                                                                        0.7
            176
                     3
                           13.27
                                   4.28
                                          2.26
                                                     20.0
                                                                   120
                                                                            1.59
                                                                                        0.69
                                                                                                        0.43
                                                                                                                          1.35
                                                                                                                                 10.2
                                                                                                                                        0.5
            177
                     3
                           13.17
                                   2.59
                                          2.37
                                                     20.0
                                                                   120
                                                                            1.65
                                                                                        0.68
                                                                                                        0.53
                                                                                                                          1.46
                                                                                                                                  9.3
                                                                                                                                        9.0
            178
                     3
                           14.13
                                   4.10
                                          2.74
                                                     24.5
                                                                    96
                                                                            2.05
                                                                                        0.76
                                                                                                        0.56
                                                                                                                          1.35
                                                                                                                                  9.2
                                                                                                                                        0.6
```

```
In [4]: # Shuffle the data.

set.seed(1234)
smp <- sample(rownames(wine), nrow(wine), replace=FALSE)
wine <- wine[smp,]
smp02 <- sample(rownames(wine), nrow(wine), replace=FALSE)
wine <- wine[smp02,]
head(rownames(wine))
tail(rownames(wine))</pre>
```

'113' '159' '21' '131' '137' '133'

'104' '144' '32' '140' '28' '168'

```
In [5]: train <- wine
In [6]: # The wine samples are from 3 grape varietals.</pre>
```

1 2 3 59 71 48

## **Basic functions**

table(wine\$Type)

```
In [6]: # Function to output a confusion matrix and the
    # accuracy score.

get_confusion <- function(preds, df_actual) {

    # df_actual is a one-column dataframe;
    # preds is a named vector of predictions;
    # preds is of type factor; it is assumed there are
    # at least 2 factor levels

levs <- levels(preds)
    n_levs <- length(levs)
    if(n_levs== 1) { levs <- c('0', '1') }
    n_levs <- max(n_levs, 2)
    actual <- as.vector(df_actual[, 1])
    names(actual) <- rownames(df_actual)</pre>
```

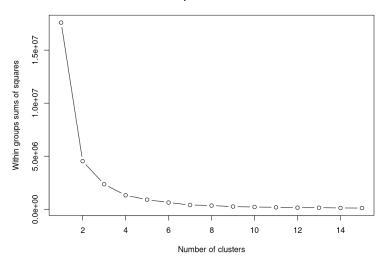
```
datout \leftarrow rep(0, n_levs * (n_levs + 1))
             dim(datout) <- c(n_levs, n_levs + 1)</pre>
             datout <- as.data.frame(datout)</pre>
             colnames(datout) <- c(levs, "class.error")</pre>
             rownames(datout) <- levs
             result <- vector("list", length= 2)</pre>
             names(result) <- c("matrix", "acc")</pre>
             # for each factor level, identify the rcd names
             # which should be classed as such
             for(rowlev in levs) {
                 actlev names <- names(actual[actual == rowlev])</pre>
                 # columns are for the predicted values:
                 for(collev in levs) {
                     predlev names <- names(preds[preds == collev])</pre>
                     if(length(predlev_names > 0)) {
                         datout[rowlev, collev] <- sum(predlev_names %in% actlev_names)</pre>
                 nonrow cols <- levs[!(levs %in% rowlev)]</pre>
                 datout[rowlev, "class.error"] <- round(sum(as.vector(datout[rowlev, nonrow_cols]))/</pre>
                                                           sum(as.vector(datout[rowlev, levs])), 4)
             }
             result$matrix <- datout
             mat <- as.matrix(datout[, 1:nrow(datout)])</pre>
             result[[2]] <- round(sum(diag(mat))/floor(sum(mat)), 4)</pre>
             return(result)
In [7]: # The following function is from Robert Kabacoff's "R in Action", pp.379-380.
        wssplot <- function(data, title="wss plot", nc=15, seed=1233) {</pre>
             # wss[1] is the total sum of squares when there is only
             # one cluster. In R's kmeans help this is called 'totss'.
             # Here is another way to compute totss:
             # ss <- function(x) sum(scale(x, scale = FALSE)^2)
             wss <- (nrow(data) - 1)*sum(apply(data, 2, var))</pre>
             for(i in 2:nc) {
                 set.seed(seed)
                 km_model <- suppressWarnings(kmeans(data, centers=i, iter.max=50,</pre>
                                                       nstart=15))
                 wss[i] <- sum(km model$withinss)</pre>
             plot(1:nc, wss, type='b', xlab="Number of clusters",
                 ylab="Within groups sums of squares",
                 main= title)
In [8]: # Function for identifying which cluster each record
        # belongs to.
        getCluster <- function(x, centers) {</pre>
             # x is a row of a dataframe; its columns need
             # to be in the same order as centers (a matrix'
             # constructed from kmeans)
             cl_dist <- apply(centers, 1, function(y) sqrt(sum((x-y)^2)))</pre>
             return(which.min(cl dist)[1])
In [9]: # Function to constrain range of data between 0 and 1.
        range01 <- function(x) {(x - min(x))/(max(x) - min(x))}
```

```
In [10]: # Function to constrain range of data between min_x and max_x.
         # This function is used to transform validation data.
         range02 <- function(x, min_x, max_x) \{(x - min_x)/(max_x - min_x)\}
In [11]: # Function to generate combination of parameters for gridSearch;
         # each combination must add to a number ~1. Returns a dataframe,
         # each row of which is a valid combination.
         # I re-factored this ftn using R's expand.grid ftn. expand.grid
         # actually takes more time to run. This is probably due to
         # type-checking. It appears that we also run out of memory more
         # quickly when using expand.grid. So at the moment I am
         # reverting to the deprecated section.
         generate_combs <- function(arglist, tol=0.0001) {</pre>
             # arglist is a named list; each name is a column
             # name of the dataframe which goes to k-means
             # this next section is an alternative to expand.grid
             # if(FALSE) {
             n_args <- length(arglist)</pre>
             param_vlens <- rep(NA, n_args)</pre>
             for(i in 1:n_args) {
                 param vlens[i] <- length(arglist[[i]])</pre>
             n_rows <- prod(param_vlens)</pre>
             datout <- rep(NA, n_args*n_rows)</pre>
             dim(datout) <- c(n_rows, n_args)</pre>
             datout <- as.data.frame(datout)</pre>
             colnames(datout) <- names(arglist)</pre>
             cprod <- 1
             for(j in 1:n_args) {
                 vect <- arglist[[j]]</pre>
                 val <- rep(vect, rep(cprod, length(vect)))</pre>
                 datout[, j] <- rep(val, n_rows/length(val))</pre>
                 cprod <- cprod*length(vect)</pre>
             # } ## end of 'if(FALSE)'
             # datout <- expand.grid(arglist, KEEP.OUT.ATTRS= FALSE)</pre>
             # colnames(datout) <- names(arglist)</pre>
             row sums <- round(rowSums(datout), 4)</pre>
             names(row_sums) <- rownames(datout)</pre>
             row_sums <- row_sums[which((as.numeric(row_sums) <= (1 + tol)) & (as.numeric(row_sums)</pre>
             datout <- datout[names(row_sums),]</pre>
             return(datout)
 In [ ]:
```

## Section 1: Get best models for train data

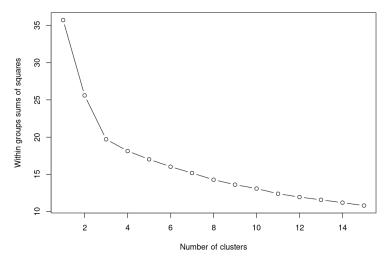
```
wssplot(train[, -1], title="wss plot of wine data")
```

#### wss plot of wine data



```
In [134]: # Scale the data; then plot.
    train_scaled <- scale(train[, -1], center=TRUE, scale=TRUE)
    train_scaled <- rangeO1(train_scaled)
    options(repr.plot.width= 8, repr.plot.height= 6)
    wssplot(train_scaled, title="wss plot of wine data, scaled")</pre>
```

#### wss plot of wine data, scaled



```
In [ ]: ### COMMENT:

# We see that after scaling the data, it is easier to
# "identify" 3 clusters. Without the scaling, the wss
# plot makes it appear that the data is best partitioned
# into 2 classes rather than 3.
```

### Find best random forest model

```
In [56]: # Initial model.
```

```
set.seed(123)
         rfclf <- randomForest(Type ~ ., data= train,</pre>
                                ntree= 100, mtry= 3, nodesize= 1,
                                importance=TRUE)
         print(rfclf)
         Call:
          randomForest(formula = Type ~ ., data = train, ntree = 100, mtry = 3,
                                                                                       nodesize = 1,
         importance = TRUE)
                        Type of random forest: classification
                               Number of trees: 100
         No. of variables tried at each split: 3
                 00B estimate of error rate: 1.69%
         Confusion matrix:
            1 2 3 class.error
         1 59 0 0
                        0.000000
         2 1 68 2
                       0.042254
         3 0 0 48
                        0.000000
In [57]: print(round(rfclf$importance, 3))
                                           3 MeanDecreaseAccuracy MeanDecreaseGini
                              1
         Alcohol
                          0.172 0.074 0.008
                                                             0.087
                                                                             16.068
         Malic
                          0.013 0.008 0.016
                                                             0.012
                                                                              2.767
                          0.003 0.003 0.014
         Ash
                                                             0.006
                                                                              1.803
                          0.012 0.003 0.030
                                                             0.012
         Alcalinity
                                                                              2.804
                          0.063 0.011 0.002
         Magnesium
                                                             0.025
                                                                              3.517
                          0.093 0.000 0.049
         Phenols
                                                             0.044
                                                                              7.943
         Flavanoids
                          0.128 0.027 0.238
                                                             0.114
                                                                             17.190
         Nonflavanoids 0.012 -0.002 0.004
                                                             0.005
                                                                              1.364
         Proanthocyanins 0.012 0.001 0.014
                                                             0.008
                                                                              1.770
                          0.107 0.091 0.135
                                                             0.109
                                                                             17.491
                                                                              9.597
         Hue
                          0.052 0.015 0.130
                                                             0.057
         Dilution
                         0.114 0.015 0.214
                                                             0.100
                                                                             16.171
         Proline
                          0.263 0.055 0.027
                                                             0.115
                                                                             18.078
 In [ ]: | ### COMMENTS:
         # If the models that we want to boost using k-means are too
         # powerful, the k-means algorithm will probably have nothing
         # to add. So we need to reduce the number of predictors.
         # In a previous notebook I had selected only Magnesium, Phenols,
         # and Flavanoids. Using only these 3 predictors, the typical
         # cross-val accuracy score was around 87%. The k-means
         # algorithm was still not able to improve upon the best of
         # the models (an xgboost model) using only these 3 predictors.
         # In the xgboost model, Phenols was insignificant relative
         # to the other predictors, with a gain under 3%. For Magnesium
         # the gain was just under 20%. For Flavanoids, the gain was # almost 78%. It may be that in order for k-means to boost
         # a model's performance, we also need more balance in the
         # importance of the predictors. We saw greater balance between
         # the 3 predictors I used in Parts 1 and 2 for the cow data.
         # It is worth trying Hue, Phenols, and Alcalinity. (See
         # the above importance output.) This choice should give us
         # more balance and lower the predictive power of the models
         # to a point where the k-means algorithm can have something
         # to contribute.
In [ ]: #&* Bookmark
In [12]: train <- wine[, c("Type","Hue","Phenols","Alcalinity")]</pre>
In [13]: set.seed(123)
         rfclf <- randomForest(Type ~ ., data= train,</pre>
```

```
ntree= 100, mtry= 3, nodesize= 1,
                                 importance=TRUE)
         print(rfclf)
         Call:
           randomForest(formula = Type ~ ., data = train, ntree = 100, mtry = 3,
                                                                                         nodesize = 1,
         importance = TRUE)
                         Type of random forest: classification
                                Number of trees: 100
         No. of variables tried at each split: 3
                  00B estimate of error rate: 20.22%
         Confusion matrix:
             1 2 3 class.error
         1 51 8 0
                         0.13559
          2 11 52 8
                         0.26761
         3 0 9 39
                         0.18750
In [14]: print(round(rfclf$importance, 3))
                                      3 MeanDecreaseAccuracy MeanDecreaseGini
                     0.188 0.104 0.496
                                                        0.235
                                                                         50.687
         Phenols
                     0.342 0.041 0.229
                                                        0.192
                                                                         43.940
                                                        0.087
                                                                         21.824
         Alcalinity 0.148 0.021 0.105
         Tune the random forest model
In [15]: # This function is called from get_cvScore_rf.
         get Acc rf <- function(traindat, valdat, ntrees, mtry, nodesize) {</pre>
              rfmod <- randomForest(Type ~ ., data= traindat, ntree= ntrees,</pre>
                                     mtry= mtry, nodesize= nodesize)
              preds <- predict(rfmod, newdata= valdat, type="response")</pre>
              ans <- get_confusion(preds, valdat[, "Type", drop=FALSE])</pre>
              return(as.numeric(ans[[2]]))
         }
In [16]: # Function to obtain a cross-validation score, averaging the
          # accuracy scores of the folds. This function is called from
         # avg_seedScores_rf.
         get_cvScore_rf <- function(seed, dat, ntrees, mtry,</pre>
                                      nodesize, folds= 5) {
              # divide dat by the number of folds
              segment_size <- round(nrow(dat)/folds)</pre>
              diff <- nrow(dat) - folds * segment_size</pre>
              last_seg_size <- segment_size + diff</pre>
              segmentsv <- c(rep(segment_size, (folds - 1)), last_seg_size)</pre>
              stopifnot(sum(segmentsv) == nrow(dat))
              # shuffle dat
              set.seed(seed)
              smp <- sample(rownames(dat), nrow(dat), replace= FALSE)</pre>
              dat <- dat[smp,]</pre>
              # split the data into the folds
              row_list <- vector("list", length= folds)</pre>
              names(row_list) <- as.character(1:folds)</pre>
              startpt <- 1
              for(i in 1:folds) {
                  endpt <- startpt + segmentsv[i] - 1</pre>
```

stopifnot(endpt <= dim(dat)[1])</pre>

```
In [18]: # This grid search is specific to finding the best random forest
          # classifier for train.
          gridSearch02 <- function(seed_vector, traindat, ntree_vector,</pre>
                                       mtry_vector, nodesizes, folds=5) {
               tree_len <- length(ntree_vector)</pre>
               mtry_len <- length(mtry_vector)</pre>
               node len <- length(nodesizes)</pre>
               # We need to capture the gridSearch parameters as well as
               # the cross-val scores.
               datout <- rep(NA, 2 * tree_len * mtry_len * node_len)</pre>
               dim(datout) <- c((tree_len * mtry_len * node_len), 2)</pre>
               datout <- as.data.frame(datout)</pre>
               colnames(datout) <- c("params", "acc")</pre>
               datout$params <- ""
               index <- 0
               for(i in 1:tree_len) {
                   n_trees <- ntree_vector[i]</pre>
                   for(j in 1:mtry_len) {
                        mtry <- mtry_vector[j]</pre>
                        for(k in 1:node_len) {
                            index \leftarrow index + 1
                             nodesize <- nodesizes[k]</pre>
                             param_string <- paste(as.character(n_trees),</pre>
                                                     as.character(mtry),
                                                     as.character(nodesize), sep= "--")
                             datout$params[index] <- param_string</pre>
                            \label{lem:condition} datout\$acc[index] <- avg\_seed\overline{S}cores\_rf(seed\_vector, traindat, n\_trees,
                                                                            folds=folds, mtry=mtry,
```

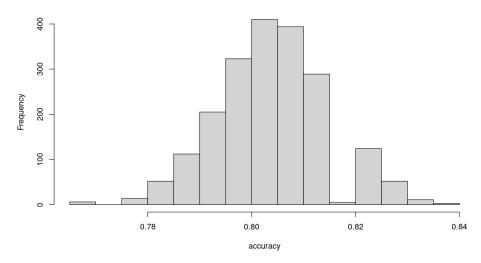
```
nodesize=nodesize)
                       }
                   }
              }
              return(datout)
In [19]: # Run grid search to get better parameters for the
          # random forest model. Test with 200 seeds. For each
          # seed, an average is taken over 5 folds.
          set.seed(7541)
          seed_smp <- sample(1:9999, 200, replace=FALSE)</pre>
          tree_vector <- c(60, 80, 100, 120, 150)
          mtry_vector <- c(1, 2)
          node_vector <- c(1, 2)</pre>
          start <- Sys.time()</pre>
          paste("Start time: ", start, sep="")
          ans <- gridSearch02(seed_smp, train, tree_vector, mtry_vector, node_vector)</pre>
          stop <- Sys.time()</pre>
          round(stop - start, 2)
          # Time difference of 2.21 mins
          (best params <- ans[which(ans$acc == max(ans$acc)),]$params)</pre>
          # '150--1--2'
          (best rf acc <- ans[which(ans$acc == max(ans$acc)),]$acc)</pre>
          # 0.80765
          'Start time: 2021-05-30 06:56:33'
          Time difference of 2.21 mins
          '150--1--2'
          0.80765
In [20]: # Refine the search.
          set.seed(7543)
          seed_smp <- sample(1:9999, 200, replace=FALSE)</pre>
          tree_vector <- c(150, 180)
          mtry vector \leftarrow c(1)
          node_vector <- c(1, 2)</pre>
          start <- Sys.time()</pre>
          paste("Start time: ", start, sep="")
          ans <- gridSearch02(seed_smp, train, tree_vector, mtry_vector, node_vector)</pre>
          stop <- Sys.time()</pre>
          round(stop - start, 2)
          # Time difference of 30.33 secs
          (best_params <- ans[which(ans$acc == max(ans$acc)),]$params)</pre>
          # '150--1--2'
          (best rf acc <- ans[which(ans$acc == max(ans$acc)),]$acc)
          # 0.8087
          'Start time: 2021-05-30 07:00:40'
          Time difference of 30.33 secs
          '150--1--2'
          0.80868
```

Get an average accuracy score for rfclf\_best on training set data

```
In [21]: # Get stable scores for the best random forest model. I will
          # refer to this model as rfclf_best. Note that 2000 seeds
          # are being used.
          set.seed(1433)
          seed_smp <- sample(1:9999, 2000, replace=FALSE)</pre>
          datout <- rep(NA, 2 * length(seed_smp))</pre>
          dim(datout) <- c(length(seed_smp), 2)</pre>
          datout <- as.data.frame(datout)</pre>
          colnames(datout) <- c("seed", "Acc")</pre>
          datout$seed <- seed_smp</pre>
          start <- Sys.time()</pre>
          for(i in 1:length(seed_smp)) {
              set.seed(seed_smp[i])
               rfmod <- randomForest(Type ~ ., data= train, ntree=150,</pre>
                                           mtry= 1, nodesize= 2)
              # preds <- predict(rfmod, newdata= dat, type="response")</pre>
              # ans <- get_confusion(preds, dat[, "Type", drop=FALSE])</pre>
              # mat <- as.matrix(ans[[1]])</pre>
              mat <- rfmod$confusion</pre>
              # percent correct <- sum(diag(mat))/floor(sum(mat))</pre>
              # datout[i, c("Acc")] <- round(percent_correct, 4)</pre>
              datout[i, c("Acc")] <- round(1-median(rfmod$err.rate[,1]), 4)</pre>
          stop <- Sys.time()</pre>
          round(stop - start, 2)
          # Time difference of 21.38 secs
```

Time difference of 21.38 secs

#### Distribution of accuracy scores for rfclf\_best on train data



```
In [23]: # rfclf_best's average accuracy score on the trainset data.
round(mean(datout$Acc), 4)
# 0.8040
```

0.804

```
In [36]: # Identify seeds with an accuracy score between 0.8039
         # and 0.8041. When constructing rfclf_best, I want to
         # use a seed which has an accuracy in the center of the
         # above distribution of accuracy scores.
         rf_candidate_seeds <- datout[which((datout$Acc > 0.8033) & (datout$Acc < 0.8046)),]$seed
         length(rf_candidate_seeds)
         # 394
         head(rf_candidate_seeds)
         # 2781
         394
         5605 8920 8072 2781 644 7123
In [37]: # Best random forest model, for the purposes of
         # showing average performance on the training set.
         set.seed(2781)
         rfclf_best <- randomForest(Type ~ ., data= train, ntree=150,
                                       mtry= 1, nodesize= 2)
         print(rfclf best)
         # 00B estimate of error rate: 17.98%; 32 records misclassified
         # Again, this is the AVERAGE PERFORMANCE of our best rf model.
         Call:
          randomForest(formula = Type \sim ., data = train, ntree = 150, mtry = 1,
                                                                                    nodesize = 2)
                        Type of random forest: classification
                              Number of trees: 150
         No. of variables tried at each split: 1
                 00B estimate of error rate: 17.98%
         Confusion matrix:
            1 2 3 class.error
         1 48 11 0
                       0.18644
         2 8 58 5
                        0.18310
         3 0 8 40
                        0.16667
```

### Get comparative cross-val score for best random forest model

For model performance comparisons, we want a cross-val score over many folds for our best random forest model.

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get cvScore rfBest <- function(seed, dat, folds= 5) {</pre>

```
# divide dat by the number of folds
segment_size <- round(nrow(dat)/folds)</pre>
diff <- nrow(dat) - folds * segment_size</pre>
last_seg_size <- segment_size + diff</pre>
segmentsv <- c(rep(segment_size, (folds - 1)), last_seg_size)</pre>
stopifnot(sum(segmentsv) == nrow(dat))
# shuffle dat
set.seed(seed)
smp <- sample(rownames(dat), nrow(dat), replace= FALSE)</pre>
dat <- dat[smp,]</pre>
# split the data into the folds
row_list <- vector("list", length= folds)</pre>
names(row_list) <- as.character(1:folds)</pre>
startpt <- 1
for(i in 1:folds) {
    endpt <- startpt + segmentsv[i] - 1</pre>
    stopifnot(endpt <= nrow(dat))</pre>
    row_list[[i]] <- rownames(dat)[startpt:endpt]</pre>
    startpt <- endpt + 1
}
train_list <- test_list <- vector("list", length= folds)</pre>
for(j in 1:folds) {
    testdat <- dat[row_list[[j]],]</pre>
    traindat <- dat[which(!(rownames(dat) %in% rownames(testdat))),]</pre>
    stopifnot((length(rownames(traindat)) + length(rownames(testdat))) == dim(dat)[1])
    test_list[[j]] <- testdat</pre>
    train_list[[j]] <- traindat</pre>
}
scores <- mcmapply(get_Acc_rfBest, train_list, test_list,</pre>
                     SIMPLIFY= TRUE, mc.cores=5)
# The following mean is over 5 accuracy scores, one for each
# of the folds.
return(round(mean(scores), 5))
```

```
In [40]: # Function to get a cross-val accuracy score over many
# folds for the best random forest model.

compute_cvScore_rf <- function(seedv, dat) {
    seedv_len <- length(seedv)
    result <- rep(NA, length=seedv_len)
    names(result) <- as.character(seedv)

for(i in 1:seedv_len) {
    cur.seed <- seedv[i]
    # For each seed in seedv, compute a cross-val
    # accuracy score.
    result[i] <- get_cvScore_rfBest(cur.seed, dat)
}
return(result)
}</pre>
```

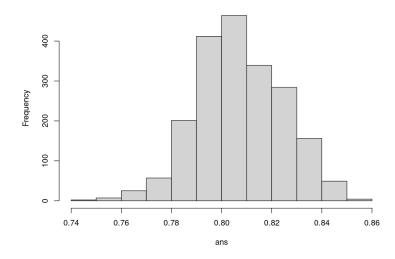
```
In [41]: # Use 2000 seeds. This equates to getting a
    # comparative cross-val score over 10,000 folds.
# Since train has 178 records in it, each fold
# will consist of 35 or 36 records. There are
# 1.56e37 ways of selecting 35 from 178.

set.seed(1931)
seedv <- sample(1:9999, 2000, replace=FALSE)

start <- Sys.time()</pre>
```

```
paste("Start time: ", start, sep="")
         ans <- compute_cvScore_rf(seedv, train)</pre>
         stop <- Sys.time()</pre>
         round(stop - start, 2)
         # Time difference of 1.23 mins
         'Start time: 2021-05-30 07:09:24'
         Time difference of 1.23 mins
In [42]: summary(ans)
            Min. 1st Qu.
                           Median
                                      Mean 3rd Qu.
                                                       Max.
           0.747
                    0.797
                            0.808
                                             0.820
                                                      0.859
                                     0.807
In [43]: # This is our comparative cross-val accuracy score for
         # the current best random forest model.
         round(mean(ans), 4)
         # 0.8653
         0.8074
In [44]: round(median(ans), 4)
          round(sd(ans), 6)
         # median: 0.8085
         # sd: 0.017064
         0.8085
         0.017064
In [45]: options(repr.plot.width= 8, repr.plot.height= 6)
         hist(ans, breaks=16, main="Distribution of cross-val accuracy scores for rfclf_best")
```

### Distribution of cross-val accuracy scores for rfclf\_best

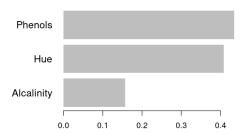


# Find best gradient boosting model

```
# packageDate("xgboost")
           ans <- unlist(packageDescription("xgboost")[1:5])</pre>
           (df <- data.frame(ans))</pre>
           A data.frame: 5 x 1
                                      ans
                                     <chr>
            Package
                                   xgboost
               Type
                                   Package
               Title Extreme Gradient Boosting
             Version
                                    1.4.1.1
               Date
                                2021-04-22
In [46]: table(train$Type)
            1 2 3
           59 71 48
In [47]: # We need to transform Type for xgboost to work. The
           # classes must be numeric, starting at 0.
           gbtrain <- train
           gbtrain$label <- NA</pre>
           for(i in 1:nrow(gbtrain)) {
                if(gbtrain$Type[i]== "1") { gbtrain$label[i] <- 0 }
if(gbtrain$Type[i]== "2") { gbtrain$label[i] <- 1 }</pre>
                if(gbtrain$Type[i]== "3") { gbtrain$label[i] <- 2 }</pre>
           table(as.factor(gbtrain$label))
            0 1 2
           59 71 48
In [48]: # There are 3 predictors.
           colnames(gbtrain)
           'Type' 'Hue' 'Phenols' 'Alcalinity' 'label'
In [49]: # Remove Type column from gbtrain and make label
           # the first column.
           gbtrain <- gbtrain[, -1]</pre>
           cols <- colnames(gbtrain)[1:3]</pre>
           gbtrain <- gbtrain[, c("label", cols)]</pre>
           head(gbtrain)
           A data.frame: 6 × 4
                 label
                        Hue Phenols Alcalinity
                      <dbl>
                 <dbl>
                                <dbl>
                                         <dbl>
            113
                        1.23
                                1.75
                                           20
                    1
            159
                    2
                        0.57
                                2.80
                                           25
             21
                        1.09
                                3.00
                                            16
                    0
            131
                    2
                        0.76
                                1.51
                                            18
```

```
label
                      Hue Phenols Alcalinity
               <dbl> <dbl>
                            <dbl>
                                     <dbl>
In [50]: dtrain <- xgb.DMatrix(data = as.matrix(gbtrain[, -1]), label = gbtrain$label)</pre>
In [52]: set.seed(123)
          gbmod <- xgboost(data= dtrain, booster="gbtree",</pre>
                             objective="multi:softmax", num_class=3,
                             eta= 0.25, max_depth=5, gamma=3,
                             subsample=0.5, nrounds=8, nthread=10,
                            eval_metric="merror")
          [1]
                   train-merror:0.191011
          [2]
                   train-merror:0.196629
          [3]
                   train-merror:0.179775
          [4]
                   train-merror:0.174157
                   train-merror:0.174157
          [5]
          [6]
                   train-merror:0.174157
          [7]
                   train-merror:0.157303
          [8]
                   train-merror:0.157303
In [53]: preds <- predict(gbmod, as.matrix(gbtrain[, -1]))</pre>
          print(length(preds))
          table(as.factor(preds))
          [1] 178
           0 1 2
          64 69 45
In [54]: # Get accuracy score for the xgboost model.
          preds02 <- as.factor(preds)</pre>
          names(preds02) <- rownames(gbtrain)</pre>
          ans <- get_confusion(preds02, gbtrain[, "label", drop=FALSE])</pre>
          ans$matrix
          ans$acc
          A data.frame: 3 × 4
                0
                      1
                            2 class.error
                  <dbl>
             <dbl>
                        <dbl>
                                  <dbl>
                                  0.1017
           0
               53
                      6
                            0
           1
               11
                     56
                            4
                                  0.2113
           2
                0
                      7
                           41
                                  0.1458
          0.8427
In [55]: # Get feature importances for the gbmod model.
          options(repr.plot.width= 6, repr.plot.height= 4)
          importance matrix <- xgb.importance(model = gbmod)</pre>
          print(importance_matrix)
          xgb.plot.importance(importance_matrix = importance_matrix)
```

Feature Gain Cover Frequency



### Tune the gradient boost model

```
In [57]: # Function to obtain a cross-validation score, averaging the
          # accuracy scores of the folds. This function is called from
          # avg_seedScores_xgb.
          get_cvScore_xgb <- function(seed, dat, eta, maxDepth,</pre>
                                         gamma, folds= 5) {
              # divide dat by the number of folds
              segment_size <- round(nrow(dat)/folds)</pre>
              diff <- nrow(dat) - folds * segment_size</pre>
              last_seg_size <- segment_size + diff</pre>
              segmentsv <- c(rep(segment_size, (folds - 1)), last_seg_size)</pre>
              stopifnot(sum(segmentsv) == nrow(dat))
              # shuffle dat
              set.seed(seed)
              smp <- sample(rownames(dat), nrow(dat), replace= FALSE)</pre>
              dat <- dat[smp,]</pre>
              # split the data into the folds
              row list <- vector("list", length= folds)</pre>
              names(row_list) <- as.character(1:folds)</pre>
              startpt <- 1
              for(i in 1:folds) {
                   endpt <- startpt + segmentsv[i] - 1</pre>
                   stopifnot(endpt <= nrow(dat))</pre>
                   row_list[[i]] <- rownames(dat)[startpt:endpt]</pre>
                   startpt <- endpt + 1</pre>
```

```
}
              train_list <- test_list <- vector("list", length= folds)</pre>
              for(j in 1:folds) \overline{\{}
                  testdat <- dat[row_list[[j]],]</pre>
                  traindat <- dat[which(!(rownames(dat) %in% rownames(testdat))),]</pre>
                  stopifnot((length(rownames(traindat)) + length(rownames(testdat))) == dim(dat)[1])
                  test_list[[j]] <- testdat</pre>
                  train_list[[j]] <- traindat</pre>
              }
              # Do NOT use mcmapply with xgboost. The function is
              # already using 10 cores, or threads.
              scores <- mapply(get_Acc_xgb, train_list, test_list,</pre>
                                 MoreArgs= list(eta= eta, maxDepth=maxDepth,
                                                 gamma=gamma), SIMPLIFY= TRUE)
              return(round(mean(scores), 5))
In [58]: # Since the seed value can affect the results, I take
          # the average over a number of seeds. This ftn is
          # called from gridSearch02_xgb.
          avg seedScores xgb <- function(seed vector, traindat, eta, maxDepth,
                                            gamma, folds= 5) {
              seed_len <- length(seed_vector)</pre>
              outv <- rep(NA, seed_len)</pre>
              for(i in 1:seed_len) {
                  seed <- seed_vector[i]</pre>
                  outv[i] <- get_cvScore_xgb(seed, traindat, eta, maxDepth,</pre>
                                                gamma, folds=folds)
              return(round(mean(outv), 5))
In [59]: # This grid search is specific to finding the best xgboost
          # classifier for traindat.
          gridSearch02 xgb <- function(seed vector, traindat, eta vector,</pre>
                                         maxDepth_vector, gamma_vector, folds=5) {
              eta_len <- length(eta_vector)</pre>
              maxDepth_len <- length(maxDepth_vector)</pre>
              gamma_len <- length(gamma_vector)</pre>
              # We need to capture the gridSearch parameters as well as
              # the cross-val scores.
              datout <- rep(NA, 2 * eta len * maxDepth len * gamma len)
              dim(datout) <- c((eta len * maxDepth len * gamma len), 2)</pre>
              datout <- as.data.frame(datout)</pre>
              colnames(datout) <- c("params", "acc")</pre>
              datout$params <- ""
              index <- 0
              for(i in 1:eta_len) {
                  eta <- eta_vector[i]</pre>
                   for(j in 1:maxDepth_len) {
                       maxDepth <- maxDepth_vector[j]</pre>
                       for(k in 1:gamma_len) {
                           index \leftarrow index + 1
                           gamma <- gamma_vector[k]</pre>
                           param string <- paste(as.character(eta),</pre>
                                                   as.character(maxDepth),
                                                   as.character(gamma), sep= "--")
```

datout\$acc[index] <- avg\_seedScores\_xgb(seed\_vector, traindat, eta=eta,</pre>

folds=folds, maxDepth=maxDepth,

gamma=gamma)

datout\$params[index] <- param\_string</pre>

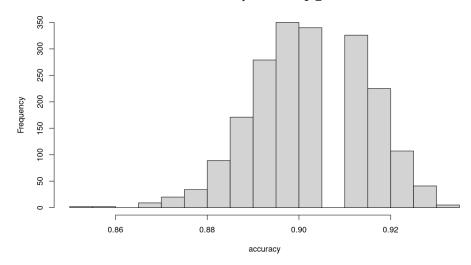
```
}
              }
              return(datout)
          # Run grid search to get better parameters for the
In [60]:
          # xgboost model. Test with 350 seeds. For each
          # seed, an average is taken over 5 folds.
          set.seed(7541)
          seed smp <- sample(1:9999, 350, replace=FALSE)</pre>
          eta_vector \leftarrow c(0.20, 0.23, 0.25)
          maxDepth_vector <- 3:5</pre>
          gamma_vector <- 1:2</pre>
          start <- Sys.time()</pre>
          paste("Start time: ", start, sep="")
          ans <- gridSearch02_xgb(seed_smp, gbtrain, eta_vector, maxDepth_vector, gamma_vector)</pre>
          stop <- Sys.time()</pre>
          round(stop - start, 2)
          # Time difference of 3.67 mins
          (best_params <- ans[which(ans$acc == max(ans$acc)),]$params)</pre>
          # '0.23--5--1'
          (best xgb acc <- ans[which(ans$acc == max(ans$acc)),]$acc)</pre>
          # 0.7940
          'Start time: 2021-05-30 07:16:47'
          Time difference of 3.67 mins
          '0.23--5--1'
          0.79396
In [61]: # Refine the search.
          set.seed(1933)
          seed_smp <- sample(1:9999, 350, replace=FALSE)</pre>
          eta_vector <- c(0.22, 0.23, 0.24)
          maxDepth_vector <- 5:6</pre>
          gamma_vector <- 1:2</pre>
          start <- Sys.time()</pre>
          paste("Start time: ", start, sep="")
          ans <- gridSearch02_xgb(seed_smp, gbtrain, eta_vector, maxDepth_vector, gamma_vector)</pre>
          stop <- Sys.time()</pre>
          round(stop - start, 2)
          # Time difference of 2.55 mins
          (best params <- ans[which(ans$acc == max(ans$acc)),]$params)</pre>
          # '0.24--6--1'
          (best_xgb_acc <- ans[which(ans$acc == max(ans$acc)),]$acc)</pre>
          # 0.7936
          'Start time: 2021-05-30 07:21:34'
          Time difference of 2.55 mins
          '0.24--6--1'
          0.7936
```

Get an average accuracy score for xgb best on the training set

```
In [62]: # Get stable scores for the best xgboost model. I will
          # refer to this model as xgb_best. Note that 2000 seeds
          # are being used.
          set.seed(1433)
          seed_smp <- sample(1:9999, 2000, replace=FALSE)</pre>
          datout <- rep(NA, 2 * length(seed_smp))</pre>
          dim(datout) <- c(length(seed_smp), 2)</pre>
          datout <- as.data.frame(datout)</pre>
          colnames(datout) <- c("seed", "Acc")</pre>
          datout$seed <- seed_smp</pre>
          start <- Sys.time()</pre>
          for(i in 1:length(seed_smp)) {
              set.seed(seed_smp[i])
              xgbmod <- xgboost(data= dtrain, booster="gbtree",</pre>
                             objective="multi:softmax", num_class=3,
                             eta=0.24, max_depth=6, gamma=1,
                             subsample=0.5, nrounds=8, nthread=10,
                             eval_metric="merror", verbose=0)
              preds <- predict(xgbmod, as.matrix(gbtrain[, -1]))</pre>
              preds <- as.factor(preds)</pre>
              names(preds) <- rownames(gbtrain)</pre>
              ans <- get_confusion(preds, gbtrain[, "label", drop=FALSE])</pre>
              mat <- as.matrix(ans[[1]])</pre>
              percent_correct <- sum(diag(mat))/floor(sum(mat))</pre>
              datout[i, c("Acc")] <- round(percent_correct, 4)</pre>
          stop <- Sys.time()</pre>
          round(stop - start, 2)
          # Time difference of 16 secs
```

Time difference of 16.06 secs

#### Distribution of accuracy scores for xgb\_best on trainset



```
In [64]: # xgb_best's average accuracy score on the trainset data.
round(mean(datout$Acc), 4)
```

```
# For rfclf_best we had an accuracy score of 0.8040
         # 0.902
         0.902
In [76]: # Identify seeds with an accuracy score near 0.9020.
         xgb candidate seeds <- datout[which((datout$Acc > 0.8990) & (datout$Acc < 0.9050)),]$seed</pre>
         length(xgb candidate seeds)
         # 340
         head(xgb_candidate_seeds)
         # 4998
         340
         3837 947 381 4998 2141 4887
In [77]: # Best xgb boost model for the purposes of showing
         # average performance on the training set. The seed
         # value is part of the model.
         set.seed(4998)
         xgb_best <- xgboost(data= dtrain, booster="gbtree",</pre>
                           objective="multi:softmax", num_class=3,
                            eta=0.24, max_depth=6, gamma=1,
                            subsample=0.5, nrounds=8, nthread=10,
                            eval_metric="merror", verbose=0)
In [78]: # Get accuracy score for xgb_best.
         preds <- predict(xgb_best, as.matrix(gbtrain[, -1]))</pre>
         preds <- as.factor(preds)</pre>
         names(preds) <- rownames(gbtrain)</pre>
         ans <- get_confusion(preds, gbtrain[, "label", drop=FALSE])</pre>
         acc <- round(as.numeric(ans[[2]]), 4)</pre>
         print(acc)
         # 0.9045
         # This is the AVERAGE PERFORMANCE of our best xgboost model on
         # the training set.
          [1] 0.9045
In [79]: # xqb best misclassifies, on average, 17 of the 178
         # records. rfclf best misclassified, on average, 32
         # of the 178 records. These are scores on the training set.
         ans[[1]]
         A data.frame: 3 × 4
                           2 class.error
                0
                     1
                       <dbl>
             <dbl> <dbl>
                                 <dbl>
                                 0.0678
               55
          1
                6
                    61
                           4
                                0.1408
                0
                     3
                          45
                                 0.0625
```

# Get comparative cross-val score for best xgboost model

```
In [80]: # This function is called from get_cvScore_xgbBest.
# It returns an accuracy score on the validation set.
```

```
get Acc xgbBest <- function(traindat, valdat) {</pre>
    dtrain <- xgb.DMatrix(data = as.matrix(traindat[, -1]),</pre>
                            label = traindat$label)
    # This is our current best xgboost model.
    set.seed(4998)
    xgbmod <- xgboost(data= dtrain, booster="gbtree",</pre>
                        objective="multi:softmax", num_class=3,
                        eta=0.24, max_depth=6, gamma=1,
                        subsample=0.5, nrounds=8, nthread=10,
                        eval_metric="merror", verbose=0)
    preds <- predict(xgbmod, as.matrix(valdat[, -1]))</pre>
    preds <- as.factor(preds)</pre>
    names(preds) <- rownames(valdat)</pre>
    ans <- get_confusion(preds, valdat[, "label", drop=FALSE])</pre>
    return(ans[[2]])
}
```

```
In [81]: # Function to obtain a cross-validation score, averaging the
          # accuracy scores of the folds. This function is called from
          # compute_cvScore_xgb.
          get_cvScore_xgbBest <- function(seed, dat, folds= 5) {</pre>
              # divide dat by the number of folds
              segment_size <- round(nrow(dat)/folds)</pre>
              diff <- nrow(dat) - folds * segment_size</pre>
              last_seg_size <- segment_size + diff</pre>
              segmentsv <- c(rep(segment_size, (folds - 1)), last_seg_size)</pre>
              stopifnot(sum(segmentsv) == nrow(dat))
              # shuffle dat
              set.seed(seed)
              smp <- sample(rownames(dat), nrow(dat), replace= FALSE)</pre>
              dat <- dat[smp,]</pre>
              # split the data into the folds
              row_list <- vector("list", length= folds)</pre>
              names(row_list) <- as.character(1:folds)</pre>
              startpt <- 1
              for(i in 1:folds) {
                  endpt <- startpt + segmentsv[i] - 1</pre>
                  stopifnot(endpt <= nrow(dat))</pre>
                  row_list[[i]] <- rownames(dat)[startpt:endpt]</pre>
                  startpt <- endpt + 1</pre>
              train_list <- test_list <- vector("list", length= folds)</pre>
              for(j in 1:folds) {
                  testdat <- dat[row_list[[j]],]</pre>
                  traindat <- dat[which(!(rownames(dat) %in% rownames(testdat))),]</pre>
                  stopifnot((length(rownames(traindat)) + length(rownames(testdat))) == dim(dat)[1])
                  test_list[[j]] <- testdat</pre>
                  train_list[[j]] <- traindat</pre>
              # Do NOT use mcmapply with xgboost. The function is
              # already using 10 cores, or threads.
              scores <- mapply(get_Acc_xgbBest, train_list, test_list,</pre>
                                 SIMPLIFY= TRUE)
              # The following average is over the 5 folds created using
              # the current seed.
              return(round(mean(scores), 5))
          }
```

## In [83]: head(gbtrain)

A data.frame: 6 × 4

	label	Hue	Phenols	Alcalinity
	<dbl></dbl>	<dbl></dbl>	<dbl></dbl>	<dbl></dbl>
113	1	1.23	1.75	20
159	2	0.57	2.80	25
21	0	1.09	3.00	16
131	2	0.76	1.51	18
137	2	0.75	1.38	21
133	2	0.66	1.15	24

```
In [84]: # Use the same initial seed as we did to get the
    # corresponding score for the random forest model.

set.seed(1931)
seedv <- sample(1:9999, 2000, replace=FALSE)

start <- Sys.time()
paste("Start time: ", start, sep="")
ans <- compute_cvScore_xgb(seedv, gbtrain)
stop <- Sys.time()
round(stop - start, 2)
# Time difference of 1.24 mins</pre>
```

'Start time: 2021-05-30 07:35:28'

Time difference of 1.23 mins

```
In [85]: summary(ans)
```

Min. 1st Qu. Median Mean 3rd Qu. Max. 0.719 0.780 0.792 0.792 0.804 0.849

```
In [86]: # This is our comparative cross-val accuracy score for the
# current best gradient boosting model.

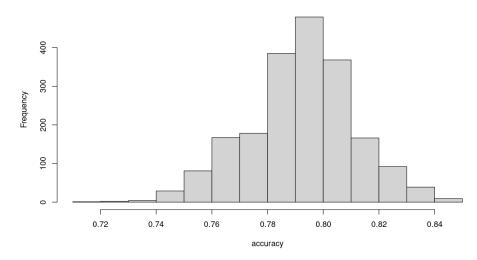
round(mean(ans), 4)
# 0.7920
```

0.792

```
In [87]: round(median(ans), 4)
    round(sd(ans), 6)
# 0.7925
# 0.019219

In [88]: options(repr.plot.width= 10, repr.plot.height= 6)
    hist(ans, breaks=16, xlab="accuracy",
        main="Distribution of cross-val accuracy scores for xgb_best")
```

#### Distribution of cross-val accuracy scores for xgb\_best



### **SVM** classifier

```
In [13]: summary(train[, -1])
# All variables are numeric and positive.
```

Hue Phenols Alcalinity :0.480 Min. :0.98 Min. :10.6 1st Qu.:0.782 1st Qu.:1.74 1st Qu.:17.2 Median :0.965 Median :2.35 Median :19.5 Mean :0.957 Mean :2.30 Mean :19.5 3rd Qu.:1.120 3rd Qu.:2.80 3rd Qu.:21.5 Max. :1.710 Max. :3.88 Max. :30.0

```
In [89]: # For SVM modeling, we need to scale the data. In
# Parts 1 and 2 we got a better model if we scaled
# the data without applying any prior transformations.
# I will take the same approach with this data set.

svm_scaled <- scale(svmtrain[, -1])
summary(svm_scaled)</pre>
```

```
Hue
                   Phenols
                                    Alcalinity
                                       :-2.66350
Min.
      :-2.089
                Min.
                     :-2.1013
                                  Min.
1st Qu.:-0.765
                1st Qu.:-0.8830
                                  1st Qu.:-0.68720
Median : 0.033
                Median : 0.0957
                                  Median : 0.00151
Mean : 0.000
                Mean : 0.0000
                                  Mean : 0.00000
                3rd Qu.: 0.8067
3rd Qu.: 0.711
                                  3rd Qu.: 0.60039
      : 3.292
                Max. : 2.5324
                                        : 3.14564
Max.
                                  Max.
```

```
In [90]: # Attach response variable.
         svm_scaled <- as.data.frame(cbind(svmtrain$Type, svm_scaled),</pre>
                                       row.names=rownames(svmtrain))
         colnames(svm scaled) <- colnames(svmtrain)</pre>
In [91]: # Try with kernel = radial basis function.
         svm01 <- svm(I(as.factor(Type)) ~ ., data=svm_scaled, kernel="radial",</pre>
                       gamma= 0.01, cost= 30, scale=FALSE)
         preds <- as.factor(fitted(svm01))</pre>
         ans <- get confusion(preds, svm scaled[, "Type", drop=FALSE])</pre>
         print(ans[[1]])
         paste0("Accuracy score for svm model: ", as.character(ans[[2]]))
             1 2 3 class.error
         1 48 11 0
                          0.1864
          2 10 56 5
                           0.2113
         3 0 3 45
                           0.0625
         'Accuracy score for svm model: 0.8371'
```

#### Tune the svm model

```
In [92]: # Function to compute an accuracy score for an svm cv-fold.
          get_Acc_svm <- function(traindat, valdat, gamma, cost) {</pre>
              # Scale traindat.
              train_scaled <- scale(traindat[, -1])</pre>
              train_centers <- attr(train_scaled, "scaled:center")</pre>
              train_scales <- attr(train_scaled, "scaled:scale")</pre>
              train_scaled <- as.data.frame(cbind(traindat$Type, train_scaled),</pre>
                                               row.names=rownames(traindat))
              colnames(train_scaled) <- colnames(traindat)</pre>
              svmmod < - svm(I(as.factor(Type)) \sim ., data= train scaled, gamma=gamma,
                               cost=cost, scale=FALSE, kernel="radial")
              # Scale valdat.
              test_scaled <- scale(valdat[, -1], center=train_centers,</pre>
                                     scale=train scales)
              test_scaled <- as.data.frame(test_scaled, row.names=rownames(valdat))</pre>
              preds <- as.factor(predict(svmmod, newdata= test_scaled))</pre>
              ans <- get_confusion(preds, valdat[, "Type", drop=FALSE])</pre>
              return(ans[[2]])
In [93]: # This grid search searches for the best parameters for svm
          # modeling of the data.
          gridSearch_svm <- function(seedv, dat, gammav, costv, folds=5) {</pre>
              gamma_len <- length(gammav)</pre>
              cost_len <- length(costv)</pre>
              # We need to capture the gridSearch parameters as well as
              # the cross-val scores.
```

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datout <- rep(NA, 2 \* gamma\_len \* cost\_len)
dim(datout) <- c((gamma\_len \* cost\_len), 2)</pre>

colnames(datout) <- c("params", "Acc")</pre>

datout <- as.data.frame(datout)</pre>

```
datout$params <- ""
# Divide dat by the number of folds to get a
# size for each fold.
segment_size <- round(nrow(dat)/folds)</pre>
diff <- nrow(dat) - folds * segment_size</pre>
last seg size <- segment size + diff</pre>
segmentsv <- c(rep(segment_size, (folds - 1)), last_seg_size)</pre>
stopifnot(sum(segmentsv) == nrow(dat))
index <- 0
for(i in 1:gamma_len) {
    gamma <- gammav[i]</pre>
    for(j in 1:cost len) {
        index \leftarrow index + 1
        cost <- costv[j]</pre>
        param_string <- paste(as.character(gamma),</pre>
                                as.character(cost), sep= "--")
        datout$params[index] <- param_string</pre>
        # Each set of parameters gets tested over many folds.
        # The different folds are created using different seeds.
        # Create a vector to store the Acc score for each seed.
        seedv_len <- length(seedv)</pre>
        seed_scores <- rep(NA, seedv_len)</pre>
        for(h in 1:seedv len) {
             # shuffle dat
             cur_seed <- seedv[h]</pre>
             set.seed(cur_seed)
             smp <- sample(rownames(dat), nrow(dat), replace= FALSE)</pre>
             dat <- dat[smp,]</pre>
             # Each element of row list will be the rows we pick
             # out for one of the folds. E.g., the first element
             # of row list will contain the rows we want for the
             # first fold, the second element of row_list will
             # contain the rows we want for the second fold, and
             # so forth.
             row_list <- vector("list", length=folds)</pre>
             names(row_list) <- as.character(1:folds)</pre>
             startpt <- 1
             for(k in 1:folds) {
                 endpt <- startpt + segmentsv[k] - 1</pre>
                 stopifnot(endpt <= nrow(dat))</pre>
                 row_list[[k]] <- rownames(dat)[startpt:endpt]</pre>
                 startpt <- endpt + 1
             }
             train list <- test list <- vector("list", length= folds)
             for(k in 1:folds) {
                 testdat <- dat[row_list[[k]],]</pre>
                 traindat <- dat[which(!(rownames(dat) %in% rownames(testdat))),]</pre>
                 stopifnot((length(rownames(traindat)) + length(rownames(testdat))) == n
                 test_list[[k]] <- testdat</pre>
                 train_list[[k]] <- traindat</pre>
             # When there are only 5 folds, only 5 cores get used.
             scores <- mcmapply(get_Acc_svm, train_list, test_list,</pre>
                                 MoreArgs= list(gamma=gamma, cost=cost),
                                 SIMPLIFY= TRUE, mc.cores=5)
             # For the current seed, store the average of the accuracy
             # scores, the average taken over the folds.
             seed_scores[h] <- round(mean(scores), 5)</pre>
        } ## end of for-loop, index h
        # Here I am taking an average of average scores. This
```

```
# could be improved by simply taking a single average.
                        datout$Acc[index] <- round(mean(seed scores), 5)</pre>
                   } ## end of for-loop, index j
               } ## end of for-loop, index i
               return(datout)
In [94]: # Find the best set of parameters.
          set.seed(7543)
          seed vector <- sample(1:9999, 200, replace=FALSE)</pre>
          gamma_v \leftarrow seq(0.01, 0.1, by=0.01)
          cost_v \leftarrow seq(10, 50, by=10)
          start <- Sys.time()</pre>
          paste("Start time: ", start, sep="")
          ans <- gridSearch_svm(seed_vector, svmtrain, gamma_v, cost_v)</pre>
          stop <- Sys.time()</pre>
          round(stop - start, 2)
          # Time difference of 5.19 mins
          (best_params <- ans[which(ans$Acc == max(ans$Acc)),]$params)</pre>
          # '0.03--30'
          (best Acc <- ans[which(ans$Acc == max(ans$Acc)),]$Acc)</pre>
          # 0.8281
          'Start time: 2021-05-30 07:41:47'
          Time difference of 5.19 mins
          '0.03--30'
          0.82814
In [95]: # Refine the search.
          set.seed(1981)
          seed_vector <- sample(1:9999, 250, replace=FALSE)</pre>
          gamma_v \leftarrow seq(0.02, 0.04, by=0.01)
          cost_v \leftarrow c(25, 30, 35)
          start <- Sys.time()</pre>
          paste("Start time: ", start, sep="")
          ans <- gridSearch_svm(seed_vector, svmtrain, gamma_v, cost_v)</pre>
          stop <- Sys.time()</pre>
          round(stop - start, 2)
          # Time difference of 1.17 mins
          (best_params <- ans[which(ans$Acc == max(ans$Acc)),]$params)</pre>
          # '0.03--25'
          (best_Acc <- ans[which(ans$Acc == max(ans$Acc)),]$Acc)</pre>
          'Start time: 2021-05-30 07:48:50'
          Time difference of 1.17 mins
          '0.03--25'
          0.82811
```

### Get scores for best svm (svm02)

```
In [96]: # Construct an svm with the identified parameters.
         # Set probability=TRUE in order to get probability estimates
         # from the output.
         svm02 <- svm(I(as.factor(Type)) ~ ., data=svm scaled, kernel="radial",</pre>
                      gamma= 0.03, cost= 25, scale=FALSE, probability=TRUE)
         preds <- as.factor(fitted(svm02))</pre>
         ans <- get_confusion(preds, svm_scaled[, "Type", drop=FALSE])</pre>
         print(ans[[1]])
         paste0("Accuracy score for svm02 (best svm): ", as.character(ans[[2]]))
         # 0.8596
         # 25 of the 178 records are misclassified.
            1 2 3 class.error
         1 55 4 0
                        0.0678
         2 12 53 6
                         0.2535
         3 0 3 45
                         0.0625
```

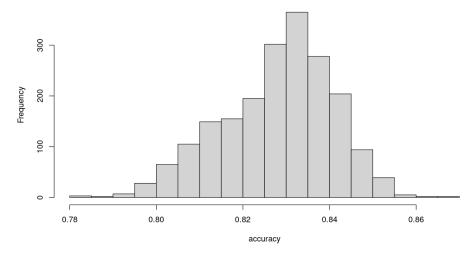
'Accuracy score for svm02 (best svm): 0.8596'

## Get comparative cross-val score for best svm model

```
In [97]: # Function to obtain a cross-validation score, averaging the
          # accuracy scores of the folds. This function is called from
          # compute_cvScore_svm.
          get_cvScore_svm02 <- function(seed, dat, folds= 5) {</pre>
              # divide dat by the number of folds
              segment size <- round(nrow(dat)/folds)</pre>
              diff <- nrow(dat) - folds * segment_size</pre>
              last_seg_size <- segment_size + diff</pre>
              segmentsv <- c(rep(segment_size, (folds - 1)), last_seg_size)</pre>
              stopifnot(sum(segmentsv) == nrow(dat))
              # shuffle dat
              set.seed(seed)
              smp <- sample(rownames(dat), nrow(dat), replace= FALSE)</pre>
              dat <- dat[smp,]</pre>
              # split the data into the folds
              row_list <- vector("list", length= folds)</pre>
              names(row_list) <- as.character(1:folds)</pre>
              startpt <- 1
              for(i in 1:folds) {
                   endpt <- startpt + segmentsv[i] - 1</pre>
                   stopifnot(endpt <= nrow(dat))</pre>
                   row_list[[i]] <- rownames(dat)[startpt:endpt]</pre>
                   startpt <- endpt + 1</pre>
              }
              train_list <- test_list <- vector("list", length= folds)</pre>
              for(j in 1:folds) {
                   testdat <- dat[row_list[[j]],]</pre>
                   traindat <- dat[which(!(rownames(dat) %in% rownames(testdat))),]</pre>
                   stopifnot((length(rownames(traindat)) + length(rownames(testdat))) == dim(dat)[1])
                   test_list[[j]] <- testdat</pre>
                   train_list[[j]] <- traindat</pre>
              scores <- mcmapply(get_Acc_svm, train_list, test_list,</pre>
                                   MoreArgs= list(gamma=0.03, cost=25),
                                   SIMPLIFY= TRUE, mc.cores=5)
```

```
# The following average is over the 5 folds created using
               # the current seed.
               return(round(mean(scores), 5))
          # Function to get a cross-val accuracy score over many
          # folds for the svm02 model.
          compute_cvScore_svm <- function(seedv, dat) {</pre>
               seedv_len <- length(seedv)</pre>
               result <- rep(NA, length=seedv_len)</pre>
               names(result) <- as.character(seedv)</pre>
               for(i in 1:seedv_len) {
                   cur.seed <- seedv[i]</pre>
                   # For each seed in seedv, compute a cross-val
                   # accuracy score.
                   result[i] <- get_cvScore_svm02(cur.seed, dat)</pre>
               return(result)
In [99]: # Again, use the same initial seed we used for
          # these scores above. The result is over 10000 folds.
          set.seed(1931)
          seed vector <- sample(1:9999, 2000, replace=FALSE)</pre>
          start <- Sys.time()</pre>
          paste("Start time: ", start, sep="")
          ans <- compute_cvScore_svm(seed_vector, svmtrain)</pre>
          stop <- Sys.time()</pre>
          round(stop - start, 2)
          # Time difference of 1.04 mins
           'Start time: 2021-05-30 07:53:25'
          Time difference of 1.04 mins
In [100]: summary(ans)
             Min. 1st Qu. Median
                                       Mean 3rd Qu.
                                                        Max.
             0.782 0.820
                              0.827
                                      0.828 0.837
                                                       0.866
In [101]: # This is our comparative cross-val accuracy score for the
          # current best svm model.
          round(mean(ans), 4)
          # 0.8278
          0.8278
In [102]: round(median(ans), 4)
          round(sd(ans), 6)
          # median: 0.8271
          # sd: 0.012713
          0.8271
          0.012713
```

#### Distribution of cross-val accuracy scores for svm02



```
In []: ### COMMENTS (RESULTS THUS FAR):

# Thus far, svm02 is our best model. It has a cross-val score
# of 0.8278. This equates to an average of 30.65 misclassified
# for every 178 records.
# The next best model is rfclf_best. It has a cross-val score
# of 0.8074; this equates to an average of 34.28 misclassified
# for every 178 records.
```

# Section 2: k-means models, base and initial hybrid

#### Base k-means model

In Parts 1 and 2 we were able to find a k-means hybrid model which outperformed all competing models. One thing that likely made this possible was that the base k-means model itself was nearly as good classifying the data as any of the other best models. With the current example, the base k-means model does not perform nearly as well as the models we have already looked at.

Among other things, this means that it will likely be more difficult to construct a hybrid k-means model which outperforms svm02. It is much more difficult for the k-means algorithm to contribute something additional to the classification task when it has low performance on that task relative to the other models. Another disadvantage for our hybrid model, relative to what we saw in Parts 1 and 2, is that here we have to beat an accuracy score of almost 83% on 178 records, whereas in Part 2 (for example) we had to beat an accuracy score of around 70.5% on 400 records. Thus, with the wine dataset, the bar is much higher. I could lower it a bit by choosing a different set of 3 predictors, but let's see what happens.

\* \* \* \* \*

```
In [104]: # First scale the data. Two kinds of scaling are
    # being applied here. In Section 5 below, I employ
    # a different approach to scaling.

df <- train

df02 <- scale(df[, -1], center=TRUE, scale=TRUE)</pre>
```

```
# Apply min-max scaling, moving all values between 0 and 1.
          df_scaled <- apply(as.matrix(df02), MARGIN=2, range01)</pre>
          df_scaled <- as.data.frame(cbind(df$Type, df_scaled),</pre>
                                       row.names=rownames(df))
          colnames(df_scaled) <- colnames(df)</pre>
          summary(df_scaled[, -1])
                 Hue
                               Phenols
                                               Alcalinity
           Min.
                  :0.000
                            Min. :0.000
                                             Min. :0.000
            1st Qu.:0.246
                            1st Qu.:0.263
                                             1st Qu.:0.340
                                             Median :0.459
                            Median :0.474
           Median :0.394
           Mean :0.388
                            Mean :0.453
                                             Mean :0.459
            3rd Qu.:0.520
                            3rd Qu.:0.628
                                             3rd Qu.:0.562
           Max.
                 :1.000
                            Max.
                                  :1.000
                                             Max.
                                                   :1.000
In [105]: # Run k-means with number of clusters set to 3.
          set.seed(1233)
          fit_km \leftarrow kmeans(df_scaled[, -1], 3, iter.max = 50, nstart = 30)
          print(fit_km$size)
           [1] 74 44 60
In [106]: datout <- as.data.frame(cbind(df scaled$Type, fit km$cluster))</pre>
          colnames(datout) <- c("Type", "cluster")</pre>
          rownames(datout) <- rownames(df_scaled)</pre>
          head(datout)
          A data.frame: 6 x 2
                Type cluster
               <dbl>
                      <dbl>
           113
                  2
                         2
           159
                  3
                         2
            21
                         1
           131
           137
                         3
In [107]: # We need to map cluster to Type level.
          table(datout$Type, as.factor(datout$cluster))
                1 2 3
            1 50 9 0
            2 24 32 15
            3 0 3 45
In [108]: ans <- as.matrix(table(datout$Type, as.factor(datout$cluster)))</pre>
          print(apply(ans, MARGIN=2, which.max))
          1 2 3
          1 2 3
In [109]: # Code snippet from get mapping function (see below).
          # (The point of this cell is to show output for a
          # section of the code in get_mapping.)
          ans <- table(datout$Type, as.factor(datout$cluster))</pre>
               # For each cluster, pick out the row (i.e., Type level) which
               # the cluster will almost certainly not map to.
```

```
min_levs <- apply(ans, MARGIN=2, which.min)</pre>
               cl_minlev <- as.numeric(min_levs["1"])</pre>
               c2_minlev <- as.numeric(min_levs["2"])</pre>
               c3_minlev <- as.numeric(min_levs["3"])</pre>
               # For each cluster, identify the remaining Type levels to
               # which it could map.
               cllevs <- c(1:3)[which(!(1:3 %in% c1_minlev))]</pre>
               c2levs <- c(1:3)[which(!(1:3 %in% c2_minlev))]
               c3levs <- c(1:3)[which(!(1:3 %in% c3_minlev))]
               tmpdat <- rbind(c1levs, c2levs, c3levs)</pre>
               colnames(tmpdat) \leftarrow 1:(dim(ans)[1] - 1)
           tmpdat
           A matrix: 3 x 2
           of type int
                  1 2
            c1levs 1 2
            c2levs 1 2
            c3levs 2 3
  In [ ]: #&* Bookmark
 In [13]: # Helper function for function valid_mappings.
           all_levels <- function(rowvals, levs) {</pre>
               n_levels <- length(levs)</pre>
               result <- FALSE
               if(sum(levs %in% rowvals) == n_levels) result <- TRUE</pre>
               return(result)
  In [ ]:
In [111]: # Apply mapping resulting from fit_km above.
           tmpdat <- datout
           tmpdat[which(tmpdat$cluster== 1),]$Type <- 1</pre>
           tmpdat[which(tmpdat$cluster== 2),]$Type <- 2</pre>
           tmpdat[which(tmpdat$cluster== 3),]$Type <- 3</pre>
           dim(tmpdat)
           178 2
```

```
In [112]: # Generate confusion matrix for the k-means clusters.
# Output accuracy for this confusion matrix.

preds <- as.factor(tmpdat$Type)
    names(preds) <- rownames(tmpdat)
    ans <- get_confusion(preds, df_scaled[, "Type", drop=FALSE])
    print(ans$matrix)
    ''

print(paste("Accuracy score for the base k-means model: ", as.character(ans[[2]]), sep=""))
# 0.7135

1 2 3 class.error
1 50 9 0 0.1525
2 24 32 15 0.5493
3 0 3 45 0.0625
"

[1] "Accuracy score for the base k-means model: 0.7135"</pre>
```

### Get comparative cross-val score for the base k-means model

```
In [113]: ans <- as.vector(table(train$Type))</pre>
          names(ans) <- levels(train$Type); print(ans)</pre>
           1 2 3
          59 71 48
In [14]: # Function returning a list of possible mappings
          # between the k-means clusters and the Type levels.
          valid_mappings <- function(df, levs) {</pre>
              # Row 1 of df is for cluster 1, row 2 for
              # cluster 2, and so forth. Each row of df
              # lists the Type levels the given cluster
              # might be assigned to.
              df_asList <- split(df, seq(nrow(df)))</pre>
              ans <- expand.grid(df_asList, KEEP.OUT.ATTRS=FALSE)</pre>
              valid_rows <- apply(as.matrix(ans), MARGIN=1, FUN=all_levels, levs=levs)</pre>
              return(ans[valid_rows,])
In [15]: # Function returning mapping between clusters and
          # Type levels. As currently written, this function
          # only works for 3-cluster solutions. We choose the
          # mapping that yields the best accuracy score.
          ### NOTE: Choosing the mapping that yields the best
          ### accuracy score does not give the k-means algorithm
          ### an advantage over other algorithms. This is
          ### because we are comparing models based on how they
          ### perform on new data, and the mapping we settle upon
          ### is based on the training set data, not the validation
          ### set.
          get_mapping <- function(dat, type_levels=c(1,2,3)) {</pre>
              # Returns a named vector on which a lookup can be done
              # by cluster name. Values of the vector are the Type
              # levels.
              # dat is a dataframe with 2 columns, c("Type", "cluster");
              # nrow(dat) = number of predictions from the model;
              # dat$Type = traindat$Type (from the calling function)
```

```
# We find the correct mapping between cluster number and
               # Type level by computing accuracy scores for the different
                                   We choose the mapping with the best
               # valid mappings.
               # accuracy score.
               tbl <- as.matrix(table(dat$Type, as.factor(dat$cluster)))</pre>
               # The colnames of tbl refer to the names of the clusters.
               # initial_map <- apply(tbl, MARGIN=2, which.max)</pre>
               # The names for initial map are the cluster names; the
               # values are the Type levels.
               # c1_init <- as.numeric(initial_map["1"])</pre>
               # c2_init <- as.numeric(initial_map["2"])</pre>
               # c3_init <- as.numeric(initial_map["3"])</pre>
               # Ideally we have c1_init <> c2_init, c1_init <> c3_init,
               # and c2_init <> c3_init
               # For each cluster, pick out the row (i.e., Type level) which
               # the cluster will almost certainly not map to.
               min_levs <- apply(tbl, MARGIN=2, which.min)</pre>
               c1_minlev <- as.numeric(min_levs["1"])</pre>
               c2 minlev <- as.numeric(min levs["2"])</pre>
               c3 minlev <- as.numeric(min levs["3"])</pre>
               # For each cluster, identify the remaining Type levels to
               # which it could map.
               cllevs <- c(1:3)[which(!(1:3 %in% c1_minlev))]</pre>
               c2levs <- c(1:3)[which(!(1:3 %in% c2_minlev))]</pre>
               c3levs <- c(1:3)[which(!(1:3 %in% c3_minlev))]
               tmpdat01 <- rbind(c1levs, c2levs, c3levs)</pre>
               colnames(tmpdat01) \leftarrow 1:(dim(tbl)[1] - 1)
               # Identify valid candidate mappings.
               df_maps <- valid_mappings(tmpdat01, type_levels)</pre>
               rownames(df_maps) <- 1:nrow(df_maps)</pre>
               scores <- rep(NA, nrow(df maps))</pre>
               for(i in 1:nrow(df_maps)) {
                   cand_levs <- df_maps[i,]</pre>
                   # colnames(df_maps) are the cluster names:
                   names(cand_levs) <- colnames(df_maps)</pre>
                   tmpdat <- dat
                   tmpdat[which(tmpdat$cluster== 1),]$Type <- as.numeric(cand_levs["1"])</pre>
                   tmpdat[which(tmpdat$cluster== 2),]$Type <- as.numeric(cand_levs["2"])</pre>
                   tmpdat[which(tmpdat$cluster== 3),]$Type <- as.numeric(cand_levs["3"])</pre>
                   preds <- as.factor(tmpdat$Type)</pre>
                   names(preds) <- rownames(tmpdat)</pre>
                   ans <- get confusion(preds, dat[, "Type", drop=FALSE])</pre>
                   scores[i] <- ans[[2]]
               }
               # I am transposing the extracted row in order to convert
               # object to a vector.
               vals <- t(df_maps[which.max(scores)[1],])[, 1]</pre>
               vals <- as.vector(vals)</pre>
               names(vals) <- colnames(df_maps)</pre>
               return(vals)
           }
In [116]: # Function for obtaining average of confusion matrix
           # accuracy score. This function is called from compute_cvScore_km.
```

```
get_cvScore_km <- function(traindat, valdat) {

# Transform and scale training set data
    df <- scale(traindat[, -1], center=TRUE, scale=TRUE)
    centers <- attr(df, "scaled:center")</pre>
```

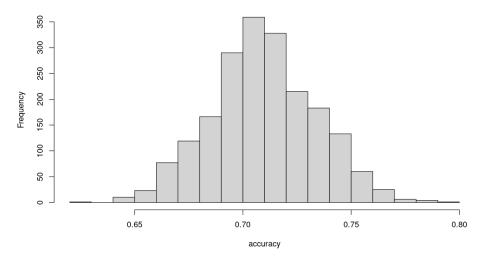
```
scales <- attr(df, "scaled:scale")</pre>
df <- as.matrix(df)</pre>
traindat_scaled <- apply(df, MARGIN=2, range01)</pre>
colnames(traindat_scaled) <- colnames(traindat)[-1]</pre>
rownames(traindat_scaled) <- rownames(traindat)</pre>
# Get mins and maxs for scaling of valdat.
traindat_mins <- as.numeric(apply(df, MARGIN=2, min))</pre>
traindat maxs <- as.numeric(apply(df, MARGIN=2, max))</pre>
# Transform and scale valdat.
df02 <- scale(valdat[, -1], center=centers, scale=scales)</pre>
df02_t \leftarrow t(as.matrix(df02))
df02_asList <- split(df02_t, seq(nrow(df02_t)))</pre>
names(df02 asList) <- colnames(valdat)[-1]</pre>
valdat_scaled <- mapply(range02, df02_asList, traindat_mins,</pre>
                          traindat maxs)
# The next step is crucial.
valdat scaled <- as.data.frame(valdat scaled, row.names=rownames(valdat))</pre>
colnames(valdat_scaled) <- colnames(valdat)[-1]</pre>
###########################
# Construct k-means model.
kmod <- suppressWarnings(kmeans(traindat scaled, 3, iter.max = 50, nstart=30))</pre>
# See how the clusters are associated with Type level.
dfout <- as.data.frame(cbind(traindat$Type, kmod$cluster))</pre>
colnames(dfout) <- c("Type", "cluster")</pre>
rownames(dfout) <- rownames(traindat)</pre>
mapping <- get_mapping(dfout)</pre>
###################################
# Apply the k-means model to valdat_scaled.
# Each element of the following list is a row of valdat_wghts.
valdat_asList <- split(valdat_scaled[, colnames(kmod$centers)],</pre>
                        seq(nrow(valdat scaled)))
ctr list <- vector("list", length= nrow(valdat))</pre>
for(i in 1:nrow(valdat)) {
    ctr_list[[i]] <- kmod$centers</pre>
names(ctr list) <- rownames(valdat)</pre>
# Get the predictions for the validation set.
preds <- mcmapply(getCluster, valdat_asList, ctr_list,</pre>
                   SIMPLIFY=TRUE, USE.NAMES= FALSE, mc.cores=6)
valdat_scaled$cluster <- as.numeric(preds)</pre>
valdat_scaled$pred_Type <- NA</pre>
# Apply mapping to the assigned clusters. Since valdat is
# quite small (around 35 records if folds = 5), and there
# are 3 clusters, it is possible that one of the LHS expressions
# below is NA. I will make the changes if the program fails.
valdat_scaled[which(valdat_scaled$cluster==1), c("pred_Type")] <- as.numeric(mapping["1</pre>
valdat_scaled[which(valdat_scaled$cluster==2), c("pred_Type")] <- as.numeric(mapping["2</pre>
valdat_scaled[which(valdat_scaled$cluster==3), c("pred_Type")] <- as.numeric(mapping["3</pre>
# Generate confusion matrix for the k-means clusters and
# the corresponding f-score.
preds <- as.factor(valdat_scaled$pred_Type)</pre>
```

```
names(preds) <- rownames(valdat)</pre>
              ans <- get confusion(preds, valdat[, "Type", drop=FALSE])</pre>
              return(ans[[2]])
         }
In [29]: # Function for computing the cross-val score for the base
         # k-means model, the hybrid k-means model, and the hybrid
         # k-means model with weights. The mapply line in the
         # following function needs to be altered to call the
         # appropriate function for the model we are working with.
         compute_cvScore_km <- function(seed_vector, dat, folds=5) {</pre>
              n seeds <- length(seed vector)</pre>
              datout <- rep(NA, 2*n seeds)
              dim(datout) <- c(n_seeds, 2)</pre>
              datout <- as.data.frame(datout)</pre>
              colnames(datout) <- c("seed", "Acc")</pre>
              datout$seed <- seed_vector</pre>
              ###############################
              # Partition the data into folds.
              # divide dat by the number of folds
              segment_size <- round(nrow(dat)/folds)</pre>
              diff <- nrow(dat) - folds * segment_size</pre>
              last seg size <- segment size + diff
              segmentsv <- c(rep(segment_size, (folds - 1)), last_seg_size)</pre>
              stopifnot(sum(segmentsv) == nrow(dat))
              for(h in 1:n_seeds) {
                  # shuffle dat
                  cur seed <- seed vector[h]</pre>
                  set.seed(cur seed)
                  smp <- sample(rownames(dat), nrow(dat), replace= FALSE)</pre>
                  dat <- dat[smp,]</pre>
                  # Each element of row_list will be the rows we pick
                  # out for one of the folds. E.g., the first element
                  # of row_list will contain the rows we want for the
                  # first fold, the second element of row_list will
                  # contain the rows we want for the second fold, and
                  row list <- vector("list", length=folds)</pre>
                  names(row_list) <- as.character(1:folds)</pre>
                  startpt <- 1
                  for(i in 1:folds) {
                      endpt <- startpt + segmentsv[i] - 1</pre>
                      stopifnot(endpt <= nrow(dat))</pre>
                      row_list[[i]] <- rownames(dat)[startpt:endpt]</pre>
                      startpt <- endpt + 1</pre>
                  }
                  train list <- test list <- vector("list", length= folds)</pre>
                  for(j in 1:folds) {
                      testdat <- dat[row_list[[j]],]</pre>
                      traindat <- dat[which(!(rownames(dat) %in% rownames(testdat))),]</pre>
                      stopifnot((length(rownames(traindat)) + length(rownames(testdat))) == nrow(dat)
                      test list[[j]] <- testdat
                      train_list[[j]] <- traindat</pre>
                  # When there are only 5 folds, only 5 cores get used.
                  # The next line varies based on which model we are trying to
                  # evaluate. [*** TURN OFF mcmapply WHEN USING xgboost ***]
                  scores <- mcmapply(get_cvScore_pcaHybrid_wghts, train_list, test_list,</pre>
                                       SIMPLIFY= TRUE, mc.cores=5)
                  # For the current seed, store the average of the accuracy
                  # scores, the average taken over the folds.
```

0.024145

```
datout$Acc[h] <- round(mean(scores), 5)</pre>
               } ## end of for-loop, index h
               return(datout)
           }
In [118]: # Again, use the same initial seed that we have
           # been using for this score.
           set.seed(1931)
           seed_vector <- sample(1:9999, 2000, replace=FALSE)</pre>
           start <- Sys.time()
paste("Start time: ", start, sep="")</pre>
           ans <- compute_cvScore_km(seed_vector, train)</pre>
           stop <- Sys.time()</pre>
           round(stop - start, 2)
           # Time difference of 5.48 mins
           'Start time: 2021-05-30 08:02:33'
           Time difference of 5.48 mins
In [119]: summary(ans$Acc)
              Min. 1st Qu. Median
                                        Mean 3rd Qu.
                                                         Max.
             0.624 0.692 0.708
                                       0.710 0.725
                                                        0.792
In [120]: round(mean(ans$Acc), 4)
           # 0.7095
           # svm02 has an accuracy score that is more than 11 percentage
           # points greater.
           0.7095
In [121]: round(median(ans$Acc), 4)
           round(sd(ans$Acc), 6)
           # median: 0.7085
           # sd: 0.024145
           0.7085
```

### Distribution of cross-val accuracy scores for base k-means model



# First hybrid k-means model

```
In [128]: # For the probability columns, use the output from svm02.
           preds <- predict(svm02, newdata=svm scaled, scale=FALSE, probability=TRUE)</pre>
           prob01 <- as.numeric(attr(preds, "probabilities")[, 2])</pre>
           prob02 <- as.numeric(attr(preds, "probabilities")[, 3])</pre>
           round(head(prob01), 3)
           round(head(prob02), 3)
           0.014 0.721 0.001 0.752 0.863 0.965
           0.02 0.051 0.919 0.011 0.002 0
In [127]: str(prob01)
            num [1:178] 0.0143 0.7209 0.0013 0.7522 0.8629 ...
In [129]: df <- train</pre>
           df$prob01 <- prob01
           df$prob02 <- prob02
           df02 <- scale(df[, -1], center=TRUE, scale=TRUE)</pre>
           df_scaled <- apply(as.matrix(df02), MARGIN=2, range01)</pre>
           df_scaled <- as.data.frame(cbind(df$Type, df_scaled),</pre>
                                        row.names=rownames(df))
           colnames(df_scaled) <- colnames(df)</pre>
           summary(df_scaled[, -1])
```

178 2

```
prob01
                 Hue
                                Phenols
                                                Alcalinity
                   :0.000
                             Min. :0.000
                                              Min. :0.000
                                                                      :0.00000
            Min.
                                                               Min.
                             1st Qu.:0.263
                                              1st Qu.:0.340
            1st Qu.:0.246
                                                                1st Qu.:0.00468
                             Median :0.474
                                              Median :0.459
            Median :0.394
                                                               Median :0.02751
                   :0.388
                                     :0.453
                                                      :0.459
                                                                       :0.27161
            Mean
                             Mean
                                              Mean
                                                               Mean
In [130]: # Run k-means with number of clusters set to 3.
           set.seed(1233)
           fit km < -kmeans(df scaled[, -1], 3, iter.max = 50, nstart = 30)
           print(fit_km$size)
           [1] 68 59 51
In [131]: datout <- as.data.frame(cbind(df_scaled$Type, fit_km$cluster))</pre>
           colnames(datout) <- c("Type", "cluster")</pre>
           rownames(datout) <- rownames(df_scaled)</pre>
           head(datout)
           A data.frame: 6 × 2
                Type cluster
                <dbl>
                      <dbl>
           113
                   2
                         2
                   3
                         3
           159
            21
                   1
                         1
           131
                   3
                         3
           137
                   3
                         3
           133
                         3
                   3
In [132]: # Map cluster to Type level.
           table(datout$Type, as.factor(datout$cluster))
                1
                   2
             1 55 4
                      0
             2 13 52
                      6
                0
                  3 45
In [133]: tmpdat <- datout</pre>
           tmpdat[which(tmpdat$cluster== 1),]$Type <- 1</pre>
           tmpdat[which(tmpdat$cluster== 2),]$Type <- 2</pre>
           tmpdat[which(tmpdat$cluster== 3),]$Type <- 3</pre>
           dim(tmpdat)
```

```
In [134]: # Generate confusion matrix for the k-means clusters.

preds <- as.factor(tmpdat$Type)
names(preds) <- rownames(tmpdat)
ans <- get_confusion(preds, df_scaled[, "Type", drop=FALSE])
print(ans$matrix)
"

print(paste("Accuracy score for the hybrid k-means model: ", as.character(ans[[2]]), sep=""
# 0.8539

1 2 3 class.error
1 55 4 0 0.0678
2 13 52 6 0.2676
3 0 3 45 0.0625
"

[1] "Accuracy score for the hybrid k-means model: 0.8539"</pre>
```

## Get comparative cross-val score for the hybrid model

```
In [135]: # Function for obtaining average of confusion matrix accuracy
           # score. This function is called from compute_cvScore_km.
           get_cvScore_hybrid <- function(traindat, valdat) {</pre>
                # Scale traindat for purpose of an svm model.
                svm_scaled <- scale(traindat[, -1])</pre>
                svm_centers <- attr(svm_scaled, "scaled:center")
svm_scales <- attr(svm_scaled, "scaled:scale")</pre>
                svm_scaled <- as.data.frame(cbind(traindat$Type, svm_scaled),</pre>
                                                  row.names=rownames(traindat))
                colnames(svm_scaled) <- colnames(traindat)</pre>
                # This is our current best svm model for the trainset data
                svmod <- svm(I(as.factor(Type)) ~ ., data=svm_scaled, kernel="radial",</pre>
                                gamma= 0.03, cost= 25, scale=FALSE, probability=TRUE)
                preds <- predict(svmod, newdata=svm_scaled, scale=FALSE, probability=TRUE)</pre>
                traindat$prob01 <- as.numeric(attr(preds, "probabilities")[, 2])
traindat$prob02 <- as.numeric(attr(preds, "probabilities")[, 3])</pre>
                #################################
                # Scale training set data for the k-means model.
                df <- scale(traindat[, -1], center=TRUE, scale=TRUE)</pre>
                centers <- attr(df, "scaled:center")
scales <- attr(df, "scaled:scale")</pre>
                df <- as.matrix(df)</pre>
                # Move the scaled data between 0 and 1.
                traindat_scaled <- apply(df, MARGIN=2, range01)</pre>
                colnames(traindat_scaled) <- colnames(traindat)[-1]</pre>
                rownames(traindat_scaled) <- rownames(traindat)</pre>
                # Get mins and maxs for scaling of valdat.
                traindat_mins <- as.numeric(apply(df, MARGIN=2, min))</pre>
                traindat_maxs <- as.numeric(apply(df, MARGIN=2, max))</pre>
                ##############################
                # Prepare valdat for svm modeling.
                svmval_scaled <- scale(valdat[, -1], center=svm_centers, scale=svm_scales)</pre>
                svmval_scaled <- as.data.frame(svmval_scaled, row.names=rownames(valdat))</pre>
                # Compute prob01 and prob02 columns.
```

}

```
preds01_b <- predict(svmod, newdata=svmval_scaled, scale=FALSE, probability=TRUE)</pre>
valdat$prob01 <- as.numeric(attr(preds01_b, "probabilities")[, 2])
valdat$prob02 <- as.numeric(attr(preds01_b, "probabilities")[, 3])</pre>
# Scale valdat.
df02 <- scale(valdat[, -1], center=centers, scale=scales)</pre>
df02_t <- t(as.matrix(df02))</pre>
df02_asList <- split(df02_t, seq(nrow(df02_t)))</pre>
names(df02_asList) <- colnames(valdat)[-1]</pre>
# Applying min-max scaling here:
valdat_scaled <- mapply(range02, df02_asList, traindat_mins,</pre>
                             traindat maxs)
# The next step is crucial.
valdat_scaled <- as.data.frame(valdat_scaled, row.names=rownames(valdat))</pre>
colnames(valdat_scaled) <- colnames(valdat)[-1]</pre>
###############################
# Construct k-means model.
kmod <- kmeans(traindat scaled, 3, iter.max = 50, nstart=30)</pre>
# See how the clusters are associated with Type level.
dfout <- as.data.frame(cbind(traindat$Type, kmod$cluster))</pre>
colnames(dfout) <- c("Type", "cluster")</pre>
rownames(dfout) <- rownames(traindat)</pre>
mapping <- get_mapping(dfout)</pre>
###############################
# Apply the k-means model to valdat_scaled.
# Each element of the following list is a row of valdat_scaled.
valdat_asList <- split(valdat_scaled[, colnames(kmod$centers)],</pre>
                           seq(nrow(valdat_scaled)))
ctr list <- vector("list", length= nrow(valdat))</pre>
for(i in 1:nrow(valdat)) {
    ctr_list[[i]] <- kmod$centers</pre>
names(ctr list) <- rownames(valdat)</pre>
# Get the predictions for the validation set.
cluster_assgns <- mcmapply(getCluster, valdat_asList, ctr_list,</pre>
                                SIMPLIFY=TRUE, USE.NAMES= FALSE, mc.cores=6)
valdat scaled$cluster <- as.numeric(cluster assgns)</pre>
valdat scaled$pred Type <- NA
# Apply mapping to the assigned clusters. Since valdat is
# quite small (around 35 records if folds = 5), and there
# are 3 clusters, it is possible that one of the LHS expressions
# below is NA. I will make the changes if the program fails.
valdat_scaled[which(valdat_scaled$cluster==1), c("pred_Type")] <- as.numeric(mapping["1
valdat_scaled[which(valdat_scaled$cluster==2), c("pred_Type")] <- as.numeric(mapping["2
valdat_scaled[which(valdat_scaled$cluster==3), c("pred_Type")] <- as.numeric(mapping["3</pre>
# Generate confusion matrix for the k-means clusters and
# the corresponding f-score.
preds <- as.factor(valdat_scaled$pred_Type)</pre>
names(preds) <- rownames(valdat)</pre>
ans <- get_confusion(preds, valdat[, "Type", drop=FALSE])</pre>
return(ans[[2]])
```

In [141]: # Use the same initial seed as we have been using for

```
# this score.
          set.seed(1931)
          seed_vector <- sample(1:9999, 2000, replace=FALSE)</pre>
          start <- Sys.time()</pre>
          paste("Start time: ", start, sep="")
          ans <- compute_cvScore_km(seed_vector, train)</pre>
          stop <- Sys.time()</pre>
          round(stop - start, 2)
          # Time difference of 6.83 mins
          'Start time: 2021-05-30 08:43:33'
          Time difference of 6.83 mins
In [142]: | summary(ans$Acc)
              Min. 1st Qu. Median
                                       Mean 3rd Qu.
                                                        Max.
             0.775
                     0.821
                              0.831
                                      0.830
                                                        0.860
                                               0.837
In [143]: round(mean(ans$Acc), 4)
          # 0.8295
          # For svm02, this score was 0.8278.
          0.8295
In [144]: round(median(ans$Acc), 4)
          round(sd(ans$Acc), 6)
          # median: 0.8314
          # sd: 0.012021
          0.8314
          0.012021
          options(repr.plot.width= 10, repr.plot.height= 6)
          hist(ans$Acc, breaks=14, xlab="accuracy",
                main="Distribution of cross-val accuracy scores for hybrid k-means model")
```

Distribution of cross-val accuracy scores for hybrid k-means model 400 300 -requency 200 100 0.78 0.80 0.82 0.84 0.86 accuracy

```
In [ ]: ### COMMENT:
        # At this point we already have a better model than our
        # previous best model, svm02. The difference in means
        # is 0.0017. This yields an F-statistic of 4.34, giving
```

```
# us a two-tailed p-value of 1.42e-05.
# The difference in median cross-val accuracy scores is
# 0.0043.

# On average, the hybrid model would misclassify 30.35 of
# 178 new records. svm02 would misclassify 30.65 of the
# 178 new records. While the improvement in the accuracy
# score is small, it is real and shows that the k-means
# algorithm does have something to add to the modeling.
# In Parts 1 and 2 we saw around a 3% improvement in the
# accuracy score, but there the number we had to beat was
# around 70%, not the nearly 83% score we are trying to
# beat here. Also, in this example we are working with
# less than half of the records of the cow data.
```

# Section 3: Hybrid k-means model with weights

### Find the best weights for the above hybrid model

We find the best weights using cross-validation. We then get the comparative cross-val score that we need. We expect to get a slightly better score than what we have already seen.

```
In [43]: # Function for obtaining average of confusion matrix
           # accuracy score. This function is called from gridSearch03.
          get_cvScore_kmWghts <- function(traindat, valdat, wghts) {</pre>
               # Scale traindat for purpose of an svm model.
               svm_scaled <- scale(traindat[, -1])</pre>
               svm_centers <- attr(svm_scaled, "scaled:center")
svm_scales <- attr(svm_scaled, "scaled:scale")</pre>
               svm_scaled <- as.data.frame(cbind(traindat$Type, svm_scaled),</pre>
                                                 row.names=rownames(traindat))
               colnames(svm_scaled) <- colnames(traindat)</pre>
               # This is our current best svm model for the trainset data
               svmod <- svm(I(as.factor(Type)) ~ ., data=svm_scaled, kernel="radial",</pre>
                               gamma= 0.03, cost= 25, scale=FALSE, probability=TRUE)
               preds <- predict(svmod, newdata=svm_scaled, scale=FALSE, probability=TRUE)</pre>
               traindat$prob01 <- as.numeric(attr(preds, "probabilities")[, 2])
traindat$prob02 <- as.numeric(attr(preds, "probabilities")[, 3])</pre>
               ################################
               # Scale training set data for the k-means model.
               df <- scale(traindat[, -1], center=TRUE, scale=TRUE)</pre>
               centers <- attr(df, "scaled:center")
scales <- attr(df, "scaled:scale")</pre>
               df <- as.matrix(df)</pre>
               traindat_scaled <- apply(df, MARGIN=2, range01)</pre>
               colnames(traindat_scaled) <- colnames(traindat)[-1]</pre>
               rownames(traindat_scaled) <- rownames(traindat)</pre>
               # Get mins and maxs for scaling of valdat.
               traindat_mins <- as.numeric(apply(df, MARGIN=2, min))</pre>
               traindat_maxs <- as.numeric(apply(df, MARGIN=2, max))</pre>
               ##################################
               # Apply weights to traindat. The sqrt should have
               # been taken in the calling function.
               cols <- names(wghts)</pre>
               df2 <- t(t(traindat_scaled[, cols]) * as.numeric(wghts[cols]))</pre>
               traindat wghts <- as.data.frame(df2, row.names=rownames(traindat))</pre>
```

```
# Prepare valdat.
svmval_scaled <- scale(valdat[, -1], center=svm_centers, scale=svm_scales)</pre>
svmval_scaled <- as.data.frame(svmval_scaled, row.names=rownames(valdat))</pre>
# Compute prob01 and prob02 columns.
preds01_b <- predict(svmod, newdata=svmval_scaled, scale=FALSE, probability=TRUE)</pre>
valdat$prob01 <- as.numeric(attr(preds01_b, "probabilities")[, 2])</pre>
valdat$prob02 <- as.numeric(attr(preds01_b, "probabilities")[, 3])</pre>
# Scale valdat.
df02 <- scale(valdat[, -1], center=centers, scale=scales)</pre>
df02_t \leftarrow t(as.matrix(df02))
df02_asList <- split(df02_t, seq(nrow(df02_t)))</pre>
names(df02_asList) <- colnames(valdat)[-1]</pre>
valdat_scaled <- mapply(range02, df02_asList, traindat_mins,</pre>
                        traindat maxs)
# The next step is crucial.
valdat scaled <- as.data.frame(valdat scaled, row.names=rownames(valdat))</pre>
colnames(valdat_scaled) <- colnames(valdat)[-1]</pre>
# Apply weights to valdat_scaled.
df4 <- t(t(valdat_scaled[, cols]) * as.numeric(wghts[cols]))</pre>
valdat_wghts <- as.data.frame(df4, row.names=rownames(valdat))</pre>
# Construct k-means model.
kmod <- kmeans(traindat_wghts, 3, iter.max = 50, nstart=30)</pre>
# See how the clusters are associated with Type level.
dfout <- as.data.frame(cbind(traindat$Type, kmod$cluster),</pre>
                       row.names=rownames(traindat))
colnames(dfout) <- c("Type", "cluster")</pre>
mapping <- get_mapping(dfout)</pre>
################################
# Apply the k-means model to valdat wghts.
# Each element of the following list is a row of valdat_wghts.
valdat_asList <- split(valdat_wghts[, colnames(kmod$centers)],</pre>
                       seq(nrow(valdat_wghts)))
ctr list <- vector("list", length= nrow(valdat))</pre>
for(i in 1:nrow(valdat)) {
    ctr_list[[i]] <- kmod$centers</pre>
names(ctr_list) <- rownames(valdat)</pre>
# Get the predictions for the validation set.
cluster_assgns <- mcmapply(getCluster, valdat_asList, ctr_list,</pre>
                           SIMPLIFY=TRUE, USE.NAMES= FALSE, mc.cores=6)
valdat_wghts$cluster <- as.numeric(cluster_assgns)</pre>
valdat_wghts$pred_Type <- NA</pre>
# Apply mapping to the assigned clusters. Since valdat is
# quite small (around 35 records if folds = 5), and there
# are 3 clusters, it is possible that one of the LHS expressions
# below is NA. I will make the changes if the program fails.
valdat_wghts[which(valdat_wghts$cluster==1), c("pred_Type")] <- as.numeric(mapping["1"]</pre>
valdat_wghts[which(valdat_wghts$cluster==2), c("pred_Type")] <- as.numeric(mapping["2"]</pre>
```

```
valdat_wghts[which(valdat_wghts$cluster==3), c("pred_Type")] <- as.numeric(mapping["3"]</pre>
              # Generate confusion matrix for the k-means clusters and
              # the corresponding f-score.
              preds <- as.factor(valdat_wghts$pred_Type)</pre>
              names(preds) <- rownames(valdat)</pre>
              ans <- get confusion(preds, valdat[, "Type", drop=FALSE])</pre>
              return(ans[[2]])
In [17]: # This grid search searches for the best set of weights to use
         # in our k-means hybrid model. The best weights are those
         # which generalize best to the validation set. So we look for
         # the best cross-validation accuracy score.
         # Because our training set is so small---only 178 records---we
         # need to run the gridSearch over many seeds. Otherwise, we
         # will not get a meaningful result.
         gridSearch03 <- function(seed_vector, dat, df_params, folds=5) {</pre>
              datout <- rep(NA, 2*nrow(df_params))</pre>
              dim(datout) <- c(nrow(df_params), 2)</pre>
              datout <- as.data.frame(datout)</pre>
              colnames(datout) <- c("row", "Acc")</pre>
              datout$row <- rownames(df_params)</pre>
              # We want the sqrt of the weights.
              df params <- df params^0.5</pre>
              params_rows <- rownames(df_params)</pre>
              ###############################
              # Partition the data into folds.
              # divide dat by the number of folds
              segment_size <- round(nrow(dat)/folds)</pre>
              diff <- nrow(dat) - folds * segment_size</pre>
              last_seg_size <- segment_size + diff</pre>
              segmentsv <- c(rep(segment_size, (folds - 1)), last_seg_size)</pre>
              stopifnot(sum(segmentsv) == nrow(dat))
              # Create a dataframe, each row for a distinct seed.
              # Each column of the dataframe is for a distinct set
              # of weights. The entries in the cells are accuracy
              # scores.
              seedv len <- length(seed vector)</pre>
              df scores <- rep(NA, seedv len*nrow(df params))</pre>
              dim(df scores) <- c(seedv len, nrow(df params))</pre>
              df_scores <- as.data.frame(df_scores)</pre>
              colnames(df_scores) <- rownames(df_params)</pre>
              rownames(df_scores) <- as.character(seed_vector)</pre>
              for(h in 1:seedv_len) {
                  # shuffle dat
                  cur seed <- seed vector[h]</pre>
                  set.seed(cur_seed)
                  smp <- sample(rownames(dat), nrow(dat), replace= FALSE)</pre>
                  dat <- dat[smp,]</pre>
                  # Each element of row list will be the rows we pick
                  # out for one of the folds. E.g., the first element
                  # of row list will contain the rows we want for the
                  # first fold, the second element of row list will
                  # contain the rows we want for the second fold, and
                  # so forth.
                  row_list <- vector("list", length=folds)</pre>
                  names(row_list) <- as.character(1:folds)</pre>
                  startpt <- 1
                  for(i in 1:folds) {
                       endpt <- startpt + segmentsv[i] - 1</pre>
```

```
stopifnot(endpt <= nrow(dat))</pre>
                        row list[[i]] <- rownames(dat)[startpt:endpt]</pre>
                        startpt <- endpt + 1</pre>
                    }
                    for(i in 1:nrow(df_params)) {
                        cur_row <- params_rows[i]</pre>
                        wghts <- as.numeric(df params[i,])</pre>
                        names(wghts) <- colnames(df_params)</pre>
                        train_list <- test_list <- vector("list", length= folds)</pre>
                        for(j in 1:folds) {
                             testdat <- dat[row list[[j]],]</pre>
                             traindat <- dat[which(!(rownames(dat) %in% rownames(testdat))),]</pre>
                             stopifnot((length(rownames(traindat)) + length(rownames(testdat))) == nrow(
                             test list[[j]] <- testdat</pre>
                             train_list[[j]] <- traindat</pre>
                        ### Do NOT use multiple cores if xgboost is used.
                        scores <- mcmapply(get cvScore hybrid pcaWqhts, train list, test list,</pre>
                                             MoreArgs= list(wghts=wghts),
                                             SIMPLIFY= TRUE, mc.cores=5)
                        # For the current seed, store the average of the accuracy
                        # scores, the average taken over the folds.
                        df_scores[as.character(cur_seed), cur_row] <- round(mean(scores), 5)</pre>
                    } # end of for-loop, index i
               } ## end of for-loop, index h
               # Compute the average over the seeds of the accuracy scores
               # obtained for each set of parameters in df_params.
               datout$Acc <- round(apply(df_scores, MARGIN=2, mean), 5)</pre>
               return(datout)
In [148]: # There are 5 parameter lists to work with. Start by
           # exploring the region around the space where all
           # parameters have an equal weight---in this case, a
           # weight of 0.20.
           lst <- vector("list", length= 5)</pre>
           names(lst) <- c(colnames(train)[-1], "prob01", "prob02")</pre>
           lst[[1]] \leftarrow lst[[2]] \leftarrow lst[[3]] \leftarrow lst[[4]] \leftarrow lst[[5]] \leftarrow seq(0.10, 0.30, by=0.02)
           start <- Sys.time()</pre>
           dfc01 <- generate_combs(lst)</pre>
           stop <- Sys.time()</pre>
           # round(stop - start, 2)
           dim(dfc01)
           # 8,801
           8801 5
```

A data.frame: 6 × 5

			Hue	Phenois	Alcalinity	prob01	prob02
			<dbl></dbl>	<dbl></dbl>	<dbl></dbl>	<dbl></dbl>	<dbl></dbl>
		47656	0.16	0.28	0.26	0.14	0.16
		94566	0.28	0.20	0.10	0.20	0.22
		26846	0.20	0.28	0.12	0.28	0.12
		10006	0.22	0.24	0.20	0.24	0.10
		136376	0.26	0.10	0.20	0.16	0.28
In	[153]:	# Use is set.set seed_ve start < dat_res stop < round(s # Time	ed(123 ector - Syssult - Systop - diffe	eds.  33) <- samples.time() <- grids <- start erence of	ole(1:99 ) Gearch03	99, 11 (seed_ <i>mins</i>	, repla
Tn	[154]:	summary	/(dat	result	Acc)		
111	[134].			_	e range	is aho	u+ 0 06
		Min 0.81		Qu. Mo .823 (		Mean 0.825	3rd Qu. 0.827
In	[155]:	# result # not w # this # It to # 0.002 # in th # 2000 round(s	lting worth point urns c 235 in he cro folds	accurac the tro t is who but I wi n order pss-val	a small cy score buble. at is "s ill need for wei accurac	s, the [* Wha mall" d much i ghts to y score	n weigh t I don and wha more va o make
		0.00235					
In	[156]:		<- dat	$t\overline{0}1[ordet$	t er(dat01	\$Acc,	decreas
		A data.fra	me: 15 ×	¢ 2			

	row	Acc	
	<chr></chr>	<dbl></dbl>	
917	41936	0.83262	
510	83046	0.83167	
704	68186	0.83165	
524	72166	0.83108	
430	73916	0.83057	
819	155896	0.83054	

```
row
                             Acc
                           <dbl>
                   <chr>
             726
                   65866 0.83010
                   28266 0.83006
             749
             995 150556 0.83004
             216
                   74986 0.83001
             667
                 117226 0.82960
             238
                   83896 0.82959
                   47656 0.82956
In [157]:
            bestrows <- as.character(dat01$row)[1:10]</pre>
            acc <- dat01[1:10, c("Acc")]
            dat02 <- cbind(dfc01[bestrows, ], acc)</pre>
            dat02
            A data.frame: 10 × 6
                      Hue Phenols Alcalinity prob01 prob02
                                                                 acc
                     <dbl>
                              <dbl>
                                        <dbl>
                                               <dbl>
                                                       <dbl>
                                                               <dbl>
              41936
                                                        0.14 0.83262
                      0.16
                               0.22
                                        0.20
                                                0.28
              83046
                      0.22
                                        0.18
                                                0.24
                                                        0.20 0.83167
                               0.16
                               0.20
              68186
                      0.24
                                         0.14
                                                0.24
                                                        0.18 0.83165
                                                        0.18 0.83108
              72166
                      0.20
                               0.18
                                        0.14
                                                0.30
              73916
                      0.22
                               0.28
                                        0.20
                                                0.10
                                                        0.20 0.83057
             155896
                      0.16
                               0.18
                                        0.12
                                                0.24
                                                        0.30 0.83054
              65866
                      0.26
                               0.16
                                        0.20
                                                0.20
                                                        0.18 0.83010
              28266
                      0.22
                               0.22
                                        0.14
                                                0.30
                                                        0.12 0.83006
              150556
                      0.28
                               0.14
                                         0.12
                                                0.16
                                                        0.30 0.83004
              74986
                      0.28
                               0.24
                                         0.16
                                                0.12
                                                        0.20 0.83001
  In [ ]: ### COMMENT:
            # Use weights in row 41936: 16, 22, 20, 28, 14.
In [158]: # Refine search.
            lst <- vector("list", length= 5)</pre>
            names(lst) <- c(colnames(train)[-1], "prob01", "prob02")</pre>
            lst[[1]] \leftarrow seq(0.15, 0.17, by=0.01)
            lst[[2]] \leftarrow seq(0.20, 0.24, by=0.01)
            lst[[3]] \leftarrow seq(0.18, 0.21, by=0.01)
            lst[[4]] \leftarrow seq(0.27, 0.31, by=0.01)
            lst[[5]] \leftarrow seq(0.12, 0.19, by=0.01)
            start <- Sys.time()</pre>
            dfc02 <- generate_combs(lst)</pre>
            stop <- Sys.time()</pre>
            # round(stop - start, 2)
            dim(dfc02)
            # 235
            235 5
```

In [159]: # Find the best weights of those in dfc02 (235 rows,

```
# 11 seeds, 5 folds).
           set.seed(1981)
           seed_vector <- sample(1:9999, 11, replace=FALSE)</pre>
           start <- Sys.time()</pre>
           paste("Start time: ", start, sep="")
           dat_result <- gridSearch03(seed_vector, train, dfc02)</pre>
           stop <- Sys.time()</pre>
           round(stop - start, 2)
           # Time difference of 8.52 mins
            'Start time: 2021-05-30 10:22:31'
           Time difference of 8.52 mins
In [160]: best_params <- dat_result[which(dat_result$Acc ==</pre>
                                                max(dat_result$Acc, na.rm=TRUE)),]$row
           length(best_params)
           best_Acc <- dat_result[which(dat_result$Acc ==</pre>
                                                 max(dat_result$Acc, na.rm=TRUE)),]$Acc
In [161]: dfc02[best_params,]
                                    Phenols
                                                  Alcalinity
                                                                   prob01
                                                                                 prob02
                          Hue
                         0.17
                                       0.22
           # 684
                                                         0.19
                                                                     0.28
                                                                                   0.14
           A data.frame: 1 x 5
                  Hue Phenols Alcalinity prob01 prob02
                                  <dbl>
                                                <dbl>
                 <dbl>
                         <dbl>
                                         <dbl>
                  0.17
                          0.22
                                   0.19
                                                 0.14
            684
                                          0.28
In [162]: best Acc
           # 0.8349
           0.83494
           dat01 <- dat result
           dat01 <- dat01[order(dat01$Acc, decreasing=TRUE),]</pre>
           dat01[1:15,]
           A data.frame: 15 x 2
                          Acc
                 <chr>
                        <dbl>
             98
                  684
                      0.83494
            134
                  914 0.83446
            174
                 1211 0.83446
            188
                 1280 0.83440
            152
                  997 0.83345
            141
                  948 0.83342
                 1325 0.83337
            194
            176
                 1221 0.83336
            191
                 1294 0.83336
                 1520 0.83336
            207
```

```
row
                           Acc
                          <dbl>
                  <chr>
                   757 0.83298
             115
                   993 0.83292
             150
             212
                  1565 0 83280
In [164]: | bestrows <- as.character(dat01$row)[1:6]</pre>
            acc <- dat01[1:6, c("Acc")]
            dat02 <- cbind(dfc02[bestrows, ], acc)</pre>
            dat02
            A data.frame: 6 × 6
```

```
Hue Phenols Alcalinity prob01 prob02
                                                       acc
      <dbl>
                <dbl>
                           <dbl>
                                   <dbl>
                                            <dbl>
                                                     <dbl>
 684
       0.17
                 0.22
                            0.19
                                     0.28
                                             0.14 0.83494
 914
       0.16
                 0.24
                            0.18
                                    0.27
                                             0.15 0.83446
1211
       0.16
                 0.23
                            0.18
                                     0.27
                                             0.16 0.83446
                                             0.16 0.83440
1280
       0.16
                 0.21
                            0.19
                                    0.28
 997
       0.15
                 0.22
                            0.20
                                     0.28
                                             0.15 0.83345
 948
       0.17
                 0.20
                            0.21
                                    0.27
                                             0.15 0.83342
```

```
In [181]: # Find the best weights of those in dat02 (6 rows,
          # 1000 seeds, 5 folds).
          ### NOTE: Before running the following, I should
          ### have added rep(0.20, 5) to dat02 to compare
          ### the above weight combinations with no weights.
          set.seed(1931)
          seed_vector <- sample(1:9999, 1000, replace=FALSE)</pre>
          start <- Sys.time()</pre>
          paste("Start time: ", start, sep="")
          dat_result <- gridSearch03(seed_vector, train, dat02[, 1:5])</pre>
          stop <- Sys.time()</pre>
          round(stop - start, 2)
          # Time difference of 20.42 mins
```

'Start time: 2021-05-30 11:19:42'

Time difference of 20.42 mins

```
In [182]: dat_result[order(dat_result$Acc, decreasing=TRUE),]
```

A data.frame: 6 x 2

```
row
                        Acc
               <chr>
                       <dbl>
                1280 0.82947
                 684 0.82931
                 948 0.82923
                 997 0.82907
            3
                1211 0.82896
                 914 0.82874
In [183]: best params <- dat result[which(dat result$Acc ==</pre>
                                                 max(dat_result$Acc, na.rm=TRUE)),]$row
```

```
length(best_params)
           best_Acc <- dat_result[which(dat_result$Acc ==</pre>
                                                max(dat_result$Acc, na.rm=TRUE)),]$Acc
In [184]: dat02[best_params, 1:5]
                                   Phenols
                                                 Alcalinity
                                                                  prob01
                                                                                prob02
           # 1280
                         0.16
                                       0.21
                                                        0.19
                                                                     0.28
                                                                                   0.16
           best Acc
           # 0.8295
           A data.frame: 1 x 5
                   Hue Phenols Alcalinity prob01 prob02
                  <dbl>
                          <dbl>
                                  <dbl>
                                         <dbl>
                                                 <dbl>
            1280
                  0.16
                          0.21
                                    0.19
                                           0.28
                                                  0.16
           0.82947
```

## Get comparative cross-val score for the hybrid model with weights

```
In [192]:
           # Function for obtaining average of confusion matrix
           # accuracy score. This function is called from compute_cvScore_km.
           # We need to take the square root of the weights.
           wghts \leftarrow c(0.16, 0.21, 0.19, 0.28, 0.16)^{\circ}0.5
           names(wghts) <- c(colnames(train)[-1], "prob01", "prob02")</pre>
           get_cvScore_wghts <- function(traindat, valdat) {</pre>
                # Scale traindat for purpose of an svm model.
                svm scaled <- scale(traindat[, -1])</pre>
                svm_centers <- attr(svm_scaled, "scaled:center")</pre>
                svm_scales <- attr(svm_scaled, "scaled:scale")</pre>
                svm_scaled <- as.data.frame(cbind(traindat$Type, svm_scaled),</pre>
                                               row.names=rownames(traindat))
                colnames(svm_scaled) <- colnames(traindat)</pre>
                # This is our current best svm model for the trainset data
                svmod <- svm(I(as.factor(Type)) ~ ., data=svm_scaled, kernel="radial",</pre>
                              gamma= 0.03, cost= 25, scale=FALSE, probability=TRUE)
                preds <- predict(svmod, newdata=svm_scaled, scale=FALSE, probability=TRUE)</pre>
                traindat$prob01 <- as.numeric(attr(preds, "probabilities")[, 2])</pre>
                traindat$prob02 <- as.numeric(attr(preds, "probabilities")[, 3])</pre>
                #################################
                # Scale training set data for the k-means model.
                df <- scale(traindat[, -1], center=TRUE, scale=TRUE)</pre>
               centers <- attr(df, "scaled:center")
scales <- attr(df, "scaled:scale")</pre>
                df <- as.matrix(df)</pre>
                traindat_scaled <- apply(df, MARGIN=2, range01)</pre>
                colnames(traindat_scaled) <- colnames(traindat)[-1]</pre>
                rownames(traindat_scaled) <- rownames(traindat)</pre>
                # Get mins and maxs for scaling of valdat.
                traindat_mins <- as.numeric(apply(df, MARGIN=2, min))</pre>
                traindat_maxs <- as.numeric(apply(df, MARGIN=2, max))</pre>
```

```
###################################
# Apply weights to traindat.
cols <- names(wghts)</pre>
df2 <- t(t(traindat_scaled[, cols]) * as.numeric(wghts[cols]))</pre>
traindat_wghts <- as.data.frame(df2, row.names=rownames(traindat))</pre>
# Prepare valdat.
svmval_scaled <- scale(valdat[, -1], center=svm_centers, scale=svm_scales)</pre>
svmval_scaled <- as.data.frame(svmval_scaled, row.names=rownames(valdat))</pre>
# Compute prob01 and prob02 columns.
preds01_b <- predict(svmod, newdata=svmval_scaled, scale=FALSE, probability=TRUE)</pre>
valdat$prob01 <- as.numeric(attr(preds01_b, "probabilities")[, 2])
valdat$prob02 <- as.numeric(attr(preds01_b, "probabilities")[, 3])</pre>
# Scale valdat.
df02 <- scale(valdat[, -1], center=centers, scale=scales)</pre>
df02 t \leftarrow t(as.matrix(df02))
df02_asList <- split(df02_t, seq(nrow(df02_t)))</pre>
names(df02_asList) <- colnames(valdat)[-1]</pre>
valdat_scaled <- mapply(range02, df02_asList, traindat_mins,</pre>
                         traindat maxs)
# The next step is crucial.
valdat scaled <- as.data.frame(valdat scaled, row.names=rownames(valdat))</pre>
colnames(valdat_scaled) <- colnames(valdat)[-1]</pre>
# Apply weights to valdat_scaled.
df4 <- t(t(valdat_scaled[, cols]) * as.numeric(wghts[cols]))</pre>
valdat wghts <- as.data.frame(df4, row.names=rownames(valdat))</pre>
colnames(valdat_wghts) <- colnames(valdat)[-1]</pre>
# Construct k-means model.
kmod <- kmeans(traindat_wghts, 3, iter.max = 50, nstart=30)</pre>
# See how the clusters are associated with Type level.
dfout <- as.data.frame(cbind(traindat$Type, kmod$cluster),</pre>
                        row.names=rownames(traindat))
colnames(dfout) <- c("Type", "cluster")</pre>
mapping <- get_mapping(dfout)</pre>
###############################
# Apply the k-means model to valdat wghts.
# Each element of the following list is a row of valdat_wghts.
valdat_asList <- split(valdat_wghts[, colnames(kmod$centers)],</pre>
                        seq(nrow(valdat_wghts)))
ctr_list <- vector("list", length= nrow(valdat))</pre>
for(i in 1:nrow(valdat)) {
    ctr_list[[i]] <- kmod$centers</pre>
names(ctr_list) <- rownames(valdat)</pre>
# Get the predictions for the validation set.
cluster_assgns <- mcmapply(getCluster, valdat_asList, ctr_list,</pre>
                            SIMPLIFY=TRUE, USE.NAMES= FALSE, mc.cores=6)
valdat_wghts$cluster <- as.numeric(cluster_assgns)</pre>
```

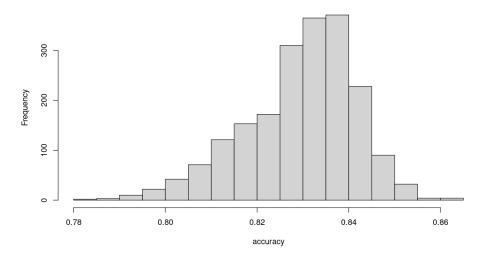
```
valdat_wghts$pred_Type <- NA</pre>
              # Apply mapping to the assigned clusters. Since valdat is
              # quite small (around 35 records if folds = 5), and there
              # are 3 clusters, it is possible that one of the LHS expressions
              # below is NA. I will make the changes if the program fails.
              valdat_wghts[which(valdat_wghts$cluster==1), c("pred_Type")] <- as.numeric(mapping["1"]</pre>
              valdat_wghts[which(valdat_wghts$cluster==2), c("pred_Type")] <- as.numeric(mapping["2"]</pre>
              valdat_wghts[which(valdat_wghts$cluster==3), c("pred_Type")] <- as.numeric(mapping["3"]</pre>
              # Generate confusion matrix for the k-means clusters and
              # the corresponding f-score.
              preds <- as.factor(valdat_wghts$pred_Type)</pre>
              names(preds) <- rownames(valdat)</pre>
              ans <- get_confusion(preds, valdat[, "Type", drop=FALSE])</pre>
              return(ans[[2]])
In [208]: # Use the correct seed, i.e., the seed we have been
          # using for this comparative score.
          set.seed(1931)
          seed_vector <- sample(1:9999, 2000, replace=FALSE)</pre>
          start <- Sys.time()</pre>
          paste("Start time: ", start, sep="")
          dat_result <- compute_cvScore_km(seed_vector, train)</pre>
          stop <- Sys.time()</pre>
          round(stop - start, 2)
          # Time difference of 6.71 mins
           'Start time: 2021-05-30 12:09:57'
          Time difference of 6.54 mins
In [209]: summary(dat result$Acc)
                                      Mean 3rd Qu.
             Min. 1st Qu. Median
                                                       Max.
                    0.821
                                             0.837
            0.780
                             0.831
                                      0.829
                                                      0.860
In [210]: round(mean(dat result$Acc), 4)
          # Accuracy over 2000 seeds is 0.8292. Without weights,
          # this score was 0.8295. svm02 had an average
          # cross-val accuracy score of 0.8278.
          # One thing this shows is that I need far more than 11
          # seeds (giving us 55 folds) in the initial search for
          # weights. But if I were to use many more seeds, the
          # search would take far more time. It would also help to
          # look at some of the 7,800 weight combinations that were
          # not included in the 1000 that were in my sample.
          0.8292
In [211]: round(median(dat_result$Acc), 4)
           round(sd(dat_result$Acc), 6)
          # median: 0.8310
          # sd: 0.012081
          # The median accuracy score for svm02 was 0.8271.
          # Without weights, the hybrid model had a median
          # accuracy score of 0.8314.
          0.831
          0.012081
```

```
In [212]: head(dat_result)
dim(dat_result)
```

A data.frame: 6 x 2

	seed	Acc			
	<int></int>	<dbl></dbl>			
1	6276	0.83238			
2	8252	0.82058			
3	6728	0.83104			
4	9082	0.84316			
5	7952	0.82550			
6	8360	0.84280			
2000 2					

### Distribution of cross-val accuracy scores for hybrid k-means model with weights



### Section 3: Final Comments

We should be able to improve upon the hybrid model that does not use weights, especially given that the standard deviation over the 1000 weight combinations looked at above is 0.00235. Our cross-val mean accuracy score for the hybrid model with no weights is 0.8295. I would expect to see around 1 standard deviation of improvement on this. So we should be able to get a score as high as 0.8318 with appropriate weights. The 0.8318 score would give us a 0.4 percentage point improvement over the svm02 model; while this isn't much, it is better than the 0.17 percentage point improvement that we are currently seeing with the hybrid model, no weights.

Finding good weights is not easy from a computational standpoint. It looks like I need to test each weight combination on far more seeds (the different seeds give us different partitions of the 178 records, i.e., different cross-val folds). It would also be good to look at all of the 8,801 combinations in the above dfc01 dataframe. It is worth trying the method I used in the Addendum of Part 2, a method that looks at the total within-group sum of squares of the different k-means clusterings. In Part 2, this alternative method of finding weights was 7X faster than the method I have been using. (The improvement in speed depends on what we are trying to do with the data and the amount of data we are working with.)

\* \* \* \* \*

# Section 4: Hybrid model with weights found using total withinss

The method of finding weights set out in the Addendum of Part 2 is not guaranteed to work. But it is worth trying out here, in our initial search for weights, when we have the most weight combinations to look through.

This alternative approach to finding weights acquires further validation if we can find a set of weights using the method which improve upon our cross-val mean accuracy score of 0.8295.

This alternative approach is faster because: (a) we no longer have to call getCluster; (b) we no longer have to apply weights to the training set data; (c) we no longer have to call get\_mapping; and (d) we no longer have to apply min-max scaling to the training set. Of these three operations, (a) and (c) are the more expensive ones.

```
In [16]: # Function for computing the tot.withinss for each set of
          # weights in df_params (a dataframe, each row of which is
          # a candidate set of weights). The optimal set of weights
          # will be the set that yields the smallest average (over
          # the folds) for tot.withinss.
          # This function is called from gridSearch07.
          get tot.withinss <- function(traindat, valdat, wghts) {</pre>
               # Scale traindat for purpose of an svm model.
               svm scaled <- scale(traindat[, -1])</pre>
              svm_centers <- attr(svm_scaled, "scaled:center")
svm_scales <- attr(svm_scaled, "scaled:scale")</pre>
               svm scaled <- as.data.frame(cbind(traindat$Type, svm scaled),</pre>
                                               row.names=rownames(traindat))
               colnames(svm scaled) <- colnames(traindat)</pre>
               # This is our current best svm model for the trainset data
               svmod <- svm(I(as.factor(Type)) ~ ., data=svm_scaled, kernel="radial",</pre>
                             gamma= 0.03, cost= 25, scale=FALSE, probability=TRUE)
               preds <- predict(symod, newdata=sym scaled, scale=FALSE, probability=TRUE)</pre>
               traindat$prob01 <- as.numeric(attr(preds, "probabilities")[, 2])
traindat$prob02 <- as.numeric(attr(preds, "probabilities")[, 3])</pre>
               ###############################
               # Scale training set data for the k-means model.
               df <- scale(traindat[, -1], center=TRUE, scale=TRUE)</pre>
              centers <- attr(df, "scaled:center")
scales <- attr(df, "scaled:scale")</pre>
               df <- as.matrix(df)</pre>
               # Get mins and maxs for scaling of valdat.
               traindat_mins <- as.numeric(apply(df, MARGIN=2, min))</pre>
               traindat_maxs <- as.numeric(apply(df, MARGIN=2, max))</pre>
               # Prepare valdat.
               svmval scaled <- scale(valdat[, -1], center=svm centers, scale=svm scales)</pre>
               svmval scaled <- as.data.frame(svmval scaled, row.names=rownames(valdat))</pre>
               # Compute prob01 and prob02 columns.
               preds01_b <- predict(svmod, newdata=svmval_scaled, scale=FALSE, probability=TRUE)</pre>
              valdat$prob01 <- as.numeric(attr(preds01_b, "probabilities")[, 2])
valdat$prob02 <- as.numeric(attr(preds01_b, "probabilities")[, 3])</pre>
               # Scale valdat.
               df02 <- scale(valdat[, -1], center=centers, scale=scales)</pre>
```

```
df02_t \leftarrow t(as.matrix(df02))
df02_asList <- split(df02_t, seq(nrow(df02_t)))</pre>
names(df02_asList) <- colnames(valdat)[-1]</pre>
valdat_scaled <- mapply(range02, df02_asList, traindat_mins,</pre>
                           traindat_maxs)
# The next step is crucial.
valdat_scaled <- as.data.frame(valdat_scaled, row.names=rownames(valdat))</pre>
colnames(valdat_scaled) <- colnames(valdat)[-1]</pre>
# Apply weights to valdat_scaled.
cols <- names(wghts)</pre>
df4 <- t(t(valdat_scaled[, cols]) * as.numeric(wghts[cols]))
valdat_wghts <- as.data.frame(df4, row.names=rownames(valdat))</pre>
colnames(valdat_wghts) <- colnames(valdat)[-1]</pre>
# Construct k-means model on the validation set.
kmod <- kmeans(valdat_wghts, 3, iter.max = 50, nstart=30)</pre>
return(kmod$tot.withinss)
```

```
In [73]: # This grid search searches for the best set of weights to use
          # in our k-means clustering model.
          gridSearch07 <- function(seed_vector, dat, df_params, folds=5) {</pre>
              datout <- rep(NA, 2*nrow(df_params))</pre>
              dim(datout) <- c(nrow(df_params), 2)</pre>
              datout <- as.data.frame(datout)</pre>
              colnames(datout) <- c("row", "tot.withinss")</pre>
              datout$row <- params_rows <- rownames(df_params)</pre>
              # We want the sqrt of the weights.
              df_params <- df_params^0.5</pre>
              ###############################
              # Partition the data into folds.
              segment_size <- round(dim(dat)[1]/folds)</pre>
              diff <- dim(dat)[1] - folds * segment_size</pre>
              last_seg_size <- segment_size + diff</pre>
              segmentsv <- c(rep(segment_size, (folds - 1)), last_seg_size)</pre>
              stopifnot(sum(segmentsv) == dim(dat)[1])
              # Create a dataframe, each row for a distinct seed.
              # Each column of the dataframe is for a distinct set
              # of weights. The entries in the cells are tot.withinss
              # scores.
              seedv len <- length(seed vector)</pre>
              df_scores <- rep(NA, seedv_len*nrow(df_params))</pre>
              dim(df_scores) <- c(seedv_len, nrow(df_params))</pre>
              df scores <- as.data.frame(df scores)</pre>
              colnames(df_scores) <- rownames(df_params)</pre>
              rownames(df_scores) <- as.character(seed_vector)</pre>
              for(h in 1:seedv_len) {
                  # shuffle dat
                  cur_seed <- seed_vector[h]</pre>
                  set.seed(cur seed)
                  smp <- sample(rownames(dat), nrow(dat), replace= FALSE)</pre>
                  dat <- dat[smp,]</pre>
                  # Each element of row_list will be the rows we pick
                  # out for one of the folds. E.g., the first element
                  # of row_list will contain the rows we want for the
```

stop <- Sys.time()
# round(stop - start, 2)</pre>

5

dim(dfc01) # 8,801

8801 5

```
# first fold, the second element of row_list will
                   # contain the rows we want for the second fold, and
                   # so forth.
                   row_list <- vector("list", length=folds)</pre>
                   names(row_list) <- as.character(1:folds)</pre>
                   startpt <- 1
                   for(i in 1:folds) {
                       endpt <- startpt + segmentsv[i] - 1</pre>
                       stopifnot(endpt <= nrow(dat))</pre>
                       row_list[[i]] <- rownames(dat)[startpt:endpt]</pre>
                       startpt <- endpt + 1</pre>
                   }
                   for(i in 1:nrow(df params)) {
                       cur_row <- params_rows[i]</pre>
                       wghts <- as.numeric(df params[i,])</pre>
                       names(wghts) <- colnames(df_params)</pre>
                       train list <- test list <- vector("list", length= folds)
                       for(j in 1:folds) {
                            testdat <- dat[row list[[j]],]</pre>
                            traindat <- dat[which(!(rownames(dat) %in% rownames(testdat))),]</pre>
                            stopifnot((length(rownames(traindat)) + length(rownames(testdat))) == nrow(
                            test_list[[j]] <- testdat</pre>
                            train_list[[j]] <- traindat</pre>
                       # When there are only 5 folds, only 5 cores get used.
                       scores <- mcmapply(get tot.withinss02, train list, test list,</pre>
                                            MoreArgs= list(wghts=wghts),
                                            SIMPLIFY= TRUE, mc.cores=5)
                       # For the current seed, store the average of the tot.withinss
                       # scores, the average taken over the folds.
                       df_scores[as.character(cur_seed), cur_row] <- round(mean(scores), 5)</pre>
                   } # end of for-loop, index i
              } ## end of for-loop, index h
              # Compute the average over the seeds of the tot.withinss scores
              # obtained for each set of parameters in df_params.
              datout$tot.withinss <- round(apply(df scores, MARGIN=2, mean), 5)</pre>
              return(datout)
In [18]: # There are 5 parameter lists to work with. Start by
          # exploring the region around the space where all
          # parameters have an equal weight---in this case, a
          # weight of 0.20.
          lst <- vector("list", length= 5)</pre>
          names(lst) <- c(colnames(train)[-1], "prob01", "prob02")</pre>
          lst[[1]] \leftarrow lst[[2]] \leftarrow lst[[3]] \leftarrow lst[[4]] \leftarrow lst[[5]] \leftarrow seq(0.10, 0.30, by=0.02)
          start <- Sys.time()</pre>
          dfc01 <- generate_combs(lst)</pre>
```

```
In [24]: # Test on a sample of 100.
set.seed(42)
smp <- sample(rownames(dfc01), 100, replace=FALSE)</pre>
```

```
tst_params <- dfc01[smp,]
head(tst_params)</pre>
```

A data.frame: 6 × 5

	Hue	Phenols	Alcalinity	prob01	prob02
	<dbl></dbl>	<dbl></dbl>	<dbl></dbl>	<dbl></dbl>	<dbl></dbl>
47656	0.16	0.28	0.26	0.14	0.16
94566	0.28	0.20	0.10	0.20	0.22
26846	0.20	0.28	0.12	0.28	0.12
10006	0.22	0.24	0.20	0.24	0.10
136376	0.26	0.10	0.20	0.16	0.28
73086	0.12	0.10	0.30	0.30	0.18

```
In [25]: # Find the best weights of those in tst_params.
# Use 120 seeds.

set.seed(1233)
seed_vector <- sample(1:9999, 120, replace=FALSE)

start <- Sys.time()
dat_result <- gridSearch07(seed_vector, train, tst_params)
stop <- Sys.time()
round(stop - start, 2)
# Time difference of 7.82 mins (for 100 rows; 120 seeds each row)</pre>
```

Time difference of 7.82 mins

In [27]: dfc01[best\_params,]

A data.frame: 1 × 5

best\_tot.withinss

 Hue
 Phenois
 Alcalinity
 prob01
 prob02

 <dbl>
 <dbl>
 <dbl>
 <dbl>
 <dbl>

 145126
 0.14
 0.18
 0.1
 0.3
 0.28

0.55

```
In []: ### COMMENT:

# I will search 5000 of the 8801 weight combinations. This
# should take approximately 6.5 hours.
```

```
In [28]: set.seed(42)
          smp <- sample(rownames(dfc01), 5000, replace=FALSE)</pre>
          tst params <- dfc01[smp,]
          remaining <- dfc01[which(!(rownames(dfc01) %in% rownames(tst params))),]
          dim(remaining)
          3801 5
In [29]: save(remaining, file="/home/greg/Documents/stat/github_repos/nudging_kmeans/untested_weight
In [30]: # Find the best weights of those in tst_params.
          # Use 120 seeds.
          set.seed(1233)
          seed_vector <- sample(1:9999, 120, replace=FALSE)</pre>
          start <- Sys.time()</pre>
          paste0("Start time: ", start)
          dat_result <- gridSearch07(seed_vector, train, tst_params)</pre>
          stop <- Sys.time()</pre>
          round(stop - start, 2)
          # Time difference of 7.93 hours (for 5000 rows; 120 seeds each row)
          Time difference of 7.93 hours
In [31]: save(dat result, file="/home/greg/Documents/stat/github repos/nudging kmeans/processed weig
In [34]: best params <- dat result[which(dat result$tot.withinss ==</pre>
                                             min(dat_result$tot.withinss, na.rm=TRUE)),]$row
          length(best_params)
          best_tot.withinss <- round(dat_result[which(dat_result$tot.withinss ==</pre>
                                            min(dat_result$tot.withinss, na.rm=TRUE)),]$tot.withinss, 5
          1
In [35]: dfc01[best_params,]
                               Phenols
                                             Alcalinity
                                                               prob01
                                                                            prob02
          # 159776
                       0.1
                                    0.2
                                                     0.1
                                                                  0.3
                                                                               0.3
          best_tot.withinss
          # 0.5428
          A data.frame: 1 × 5
                  Hue Phenols Alcalinity prob01 prob02
                 <dbl>
                         <dbl>
                                               <dbl>
                                 <dbl>
                                        <dbl>
          159776
                   0.1
                           0.2
                                   0.1
                                          0.3
                                                 0.3
          0.5428
In [36]: dat01 <- dat_result[order(dat_result$tot.withinss, decreasing=FALSE),]</pre>
          dat01[1:15,]
          A data.frame: 15 × 2
                  row tot.withinss
                <chr>
                           <dbl>
          2071 159776
                         0.54280
```

```
row tot.withinss
                   <dbl>
        <chr>
 2384 159756
                  0.54420
 3975 159766
                  0.54421
 1615 145146
                  0.54453
  939 153176
                  0.54456
 2008 157126
                  0.54492
 3441
      159886
                  0.54503
 4654
      139866
                  0.54511
  515 151846
                  0.54563
 2091 153166
                  0.54584
 2748 129196
                  0.54608
 4255 127876
                  0.54618
 1465 115886
                  0.54622
bestrows <- as.character(dat01$row)[1:10]</pre>
withinss.vals <- dat01[1:10, c("tot.withinss")]</pre>
dat01_b <- cbind(dfc01[bestrows, ], withinss.vals)</pre>
dat01_b
```

A data.frame: 10 × 6

In [ ]: ### COMMENTS:

```
Hue Phenols Alcalinity prob01 prob02 withinss.vals
         <dbl>
                  <dbl>
                             <dbl>
                                      <dbl>
                                               <dbl>
                                                              <dbl>
                                                            0.54280
159776
          0.10
                    0.20
                               0.10
                                       0.30
                                                0.30
159756
          0.14
                    0.16
                               0.10
                                       0.30
                                                0.30
                                                            0.54420
159766
          0.12
                    0.18
                               0.10
                                        0.30
                                                0.30
                                                            0.54421
145146
                               0.10
                                        0.30
          0.10
                    0.22
                                                0.28
                                                            0.54453
                                        0.20
153176
          0.10
                    0.30
                               0.10
                                                0.30
                                                            0.54456
157126
          0.12
                    0.22
                               0.10
                                        0.26
                                                0.30
                                                            0.54492
159886
          0.10
                    0.18
                               0.12
                                        0.30
                                                0.30
                                                            0.54503
139866
          0.10
                    0.30
                               0.10
                                        0.22
                                                0.28
                                                            0.54511
151846
          0.12
                    0.30
                               0.10
                                        0.18
                                                0.30
                                                            0.54563
153166
                    0.28
                               0.10
                                        0.20
                                                0.30
                                                            0.54584
         0.12
```

```
# The best weights identified are almost all at the limits
# of the tested parameter space: 0.10 was the smallest
# possible value, and 0.30 was the largest. Thus, we
# might see even smaller total withinss in regions that
# have yet to be tested.

In [41]: # Refine the search a bit.

lst <- vector("list", length= 5)
names(lst) <- c(colnames(train)[-1], "prob01","prob02")

lst[[1]] <- seq(0.08, 0.12, by=0.01)
lst[[2]] <- seq(0.18, 0.22, by=0.01)
lst[[3]] <- seq(0.08, 0.12, by=0.01)
lst[[4]] <- seq(0.28, 0.34, by=0.01)
lst[[5]] <- seq(0.28, 0.34, by=0.02)</pre>
```

```
start <- Sys.time()</pre>
          dfc02 <- generate combs(lst)</pre>
          stop <- Sys.time()</pre>
          # round(stop - start, 2)
          dim(dfc02)
          # 8,801
                       5
          309 5
In [42]: # Add in the no-weights combination.
          dfc02 \leftarrow rbind(dfc02, rep(0.20, 5))
          dim(dfc02)
          310 5
In [45]: # Find the best weights of those in dfc02.
          # Use 120 seeds. Use gridSearch03.
          set.seed(1987)
          seed_vector <- sample(1:9999, 120, replace=FALSE)</pre>
          start <- Sys.time()</pre>
          paste0("Start time: ", start)
          dat_result <- gridSearch03(seed_vector, train, dfc02)</pre>
          stop <- Sys.time()</pre>
          round(stop - start, 2)
          # Time difference of 2.11 hours
          # This time difference tells us that for this 178 record
          # dataset, the alternative method of finding weights is
          # 5.2X faster.
          'Start time: 2021-06-01 09:12:07'
          Time difference of 2.11 hours
In [48]: round(sd(dat_result$Acc), 4)
          # 0.0005
          round((max(dat_result$Acc) - min(dat_result$Acc)), 4)
          # 0.0027
          5e-04
          0.0027
 In [ ]: ### COMMENT:
          # Note the low standard deviation for the 310 weight
          # combinations and the low range. Our "no weights"
          # combination is among the 310.
In [49]: best_params <- dat_result[which(dat_result$Acc ==</pre>
                                            max(dat_result$Acc, na.rm=TRUE)),]$row
          length(best_params)
          best Acc <- dat result[which(dat result$Acc ==</pre>
                                            max(dat result$Acc, na.rm=TRUE)),]$Acc
          2
In [50]: dfc02[best_params,]
                                Phenols
                                             Alcalinity
                       Hue
                                                             prob01
                                                                          prob02
```

A data.frame:  $2 \times 5$ 

```
Hue Phenols Alcalinity prob01 prob02
      <dbl>
               <dbl>
                          <dbl>
                                  <dbl>
                                           <dbl>
 515
       0.12
                 0.20
                           80.0
                                    0.32
                                            0.28
1284
       0.11
                 0.19
                           0.09
                                   0.31
                                            0.30
```

```
In [51]: dat02 <- dat_result[order(dat_result$Acc, decreasing=TRUE),]
dat02[1:10,]</pre>
```

A data.frame: 10 × 2

	row	Acc	
	<chr></chr>	<dbl></dbl>	
50	515	0.83110	
159	1284	0.83110	
46	479	0.83107	
69	635	0.83088	
261	2161	0.83086	
192	1532	0.83082	
205	1773	0.83080	
308	3026	0.83080	
20	315	0.83079	
165	1312	0.83078	

```
In [52]: bestrows <- as.character(dat02$row)[1:3]
acc <- dat02[1:3, c("Acc")]
dat02_b <- cbind(dfc02[bestrows, ], acc)
dat02_b</pre>
```

A data.frame:  $3 \times 6$ 

	Hue	Phenols	Alcalinity	prob01	prob02	acc	
	<dbl></dbl>	<dbl></dbl>	<dbl></dbl>	<dbl></dbl>	<dbl></dbl>	<dbl></dbl>	
515	0.12	0.20	0.08	0.32	0.28	0.83110	
1284	0.11	0.19	0.09	0.31	0.30	0.83110	
479	0.11	0.18	0.12	0.31	0.28	0.83107	

```
In [67]: tail(dfc02)
          A data frame: 6 x 5
                 Hue Phenols Alcalinity prob01 prob02
                <dbl>
                        <dbl>
                                <dbl>
                                       <dbl>
                                              <dbl>
           2926
                 0.08
                         0.18
                                 0.10
                                        0.30
                                               0.34
           3002
                 0.09
                         0.18
                                 0.08
                                        0.31
                                               0.34
           3006
                 0.08
                         0.19
                                 0.08
                                        0.31
                                               0.34
           3026
                 0.08
                         0.18
                                 0.09
                                        0.31
                                               0.34
           3126
                 0.08
                         0.18
                                 80.0
                                        0.32
                                               0.34
            310
                                 0.20
                                        0.20
                                               0.20
                0.20
                         0.20
In [68]: dat02["310", c("Acc")]
          0.83022
 In [ ]: ### COMMENT:
          # Over 120 seeds (600 folds), the current best weights
          # have a score of 0.83110 and the "no-weights" model
          # has a score of 0.83022. This is a difference of
          # 8.8e-04. In other words, the current best weights
          # are not helping much at all to improve the average
          # accuracy score of the k-means hybrid model.
In [53]: # Refine the search.
          dfc03 <- dat02_b[, 1:5]
In [54]: # Find the best weights of those in dfc03.
          # Use 1200 seeds. Use gridSearch03.
          set.seed(1981)
          seed_vector <- sample(1:9999, 1200, replace=FALSE)</pre>
          start <- Sys.time()</pre>
          paste0("Start time: ", start)
          dat_result <- gridSearch03(seed_vector, train, dfc03)</pre>
          stop <- Sys.time()</pre>
          round(stop - start, 2)
          # Time difference of 12.49 mins
          'Start time: 2021-06-01 11:50:04'
          Time difference of 12.49 mins
In [55]: best_params <- dat_result[which(dat_result$Acc ==</pre>
                                             max(dat_result$Acc, na.rm=TRUE)),]$row
          length(best_params)
          best Acc <- dat result[which(dat result$Acc ==</pre>
                                              max(dat_result$Acc, na.rm=TRUE)),]$Acc
In [56]: dat03 <- dat_result[order(dat_result$Acc, decreasing=TRUE),]</pre>
          dat03
```

```
A data.frame: 3 x 2
               row
                      Acc
              <chr>
                     <dbl>
               479 0.83024
               515 0.83002
In [57]: bestrows <- as.character(dat03$row)</pre>
          acc <- dat03[, c("Acc")]
          dat03_b <- cbind(dfc03[bestrows, ], acc)</pre>
          dat03_b
          A data.frame: 3 x 6
                 Hue Phenols Alcalinity prob01 prob02
                                                        acc
                <dbl>
                        <dbl>
                                 <dbl>
                                        <dbl>
                                               <dbl>
                                                       <dbl>
            479
                 0.11
                         0.18
                                  0.12
                                         0.31
                                                0.28 0.83024
                 0.12
                                                0.28 0.83002
            515
                         0.20
                                  0.08
                                         0.32
           1284
                 0.11
                         0.19
                                  0.09
                                         0.31
                                                0.30 0.82997
 In [ ]: | ### COMMENT:
          # As the number of seeds increases, the average
          # accuracy score decreases toward the 0.8295
          # number we are trying to beat. The 0.8295 number
          # is from the hybrid model without weights. It
          # may be that the scaling I am applying, in combination
          # with looking at many thousands of folds of the
          # data (for a data set that is only 178 records),
          # lessens the importance of weights.
```

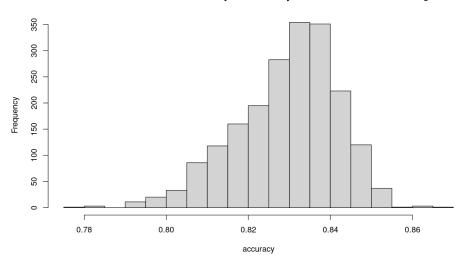
## Get comparative cross-val score using the new weights

```
In [581:
          # Function for obtaining average of confusion matrix
          # accuracy score. This function is called from compute cvScore km.
          # This is the same function as above; only the weights have changed.
          # We need to take the square root of the weights.
          wghts \leftarrow c(0.11, 0.18, 0.12, 0.31, 0.28)^{\circ}0.5
          names(wghts) <- c(colnames(train)[-1], "prob01", "prob02")</pre>
          get_cvScore_wghts <- function(traindat, valdat) {</pre>
              # Scale traindat for purpose of an svm model.
              svm_scaled <- scale(traindat[, -1])</pre>
              svm_centers <- attr(svm_scaled, "scaled:center")
svm_scales <- attr(svm_scaled, "scaled:scale")</pre>
              svm_scaled <- as.data.frame(cbind(traindat$Type, svm_scaled),</pre>
                                              row.names=rownames(traindat))
               colnames(svm_scaled) <- colnames(traindat)</pre>
              # This is our current best svm model for the trainset data
              svmod <- svm(I(as.factor(Type)) ~ ., data=svm_scaled, kernel="radial",</pre>
                             gamma= 0.03, cost= 25, scale=FALSE, probability=TRUE)
              preds <- predict(symod, newdata=sym scaled, scale=FALSE, probability=TRUE)</pre>
               traindat$prob01 <- as.numeric(attr(preds, "probabilities")[, 2])</pre>
               traindat$prob02 <- as.numeric(attr(preds, "probabilities")[, 3])</pre>
              ##############################
               # Scale training set data for the k-means model.
```

```
df <- scale(traindat[, -1], center=TRUE, scale=TRUE)</pre>
centers <- attr(df, "scaled:center")
scales <- attr(df, "scaled:scale")</pre>
df <- as.matrix(df)</pre>
traindat_scaled <- apply(df, MARGIN=2, range01)</pre>
colnames(traindat scaled) <- colnames(traindat)[-1]</pre>
rownames(traindat_scaled) <- rownames(traindat)</pre>
# Get mins and maxs for scaling of valdat.
traindat_mins <- as.numeric(apply(df, MARGIN=2, min))</pre>
traindat_maxs <- as.numeric(apply(df, MARGIN=2, max))</pre>
##################################
# Apply weights to traindat.
cols <- names(wghts)</pre>
df2 <- t(t(traindat_scaled[, cols]) * as.numeric(wghts[cols]))</pre>
traindat_wghts <- as.data.frame(df2, row.names=rownames(traindat))</pre>
# Prepare valdat.
svmval_scaled <- scale(valdat[, -1], center=svm_centers, scale=svm_scales)</pre>
svmval_scaled <- as.data.frame(svmval_scaled, row.names=rownames(valdat))</pre>
# Compute prob01 and prob02 columns.
preds01_b <- predict(svmod, newdata=svmval_scaled, scale=FALSE, probability=TRUE)</pre>
valdat$prob01 <- as.numeric(attr(preds01_b, "probabilities")[, 2])
valdat$prob02 <- as.numeric(attr(preds01_b, "probabilities")[, 3])</pre>
# Scale valdat.
df02 <- scale(valdat[, -1], center=centers, scale=scales)</pre>
df02_t <- t(as.matrix(df02))</pre>
df02_asList <- split(df02_t, seq(nrow(df02_t)))</pre>
names(df02 asList) <- colnames(valdat)[-1]</pre>
valdat_scaled <- mapply(range02, df02_asList, traindat_mins,</pre>
                          traindat_maxs)
# The next step is crucial.
valdat_scaled <- as.data.frame(valdat_scaled, row.names=rownames(valdat))</pre>
colnames(valdat_scaled) <- colnames(valdat)[-1]</pre>
# Apply weights to valdat_scaled.
df4 <- t(t(valdat_scaled[, cols]) * as.numeric(wghts[cols]))</pre>
valdat_wghts <- as.data.frame(df4, row.names=rownames(valdat))</pre>
colnames(valdat_wghts) <- colnames(valdat)[-1]</pre>
# Construct k-means model.
kmod <- kmeans(traindat_wghts, 3, iter.max = 50, nstart=30)</pre>
# See how the clusters are associated with Type level.
dfout <- as.data.frame(cbind(traindat$Type, kmod$cluster),</pre>
                         row.names=rownames(traindat))
colnames(dfout) <- c("Type", "cluster")</pre>
mapping <- get_mapping(dfout)</pre>
####################################
# Apply the k-means model to valdat wghts.
# Each element of the following list is a row of valdat_wghts.
valdat_asList <- split(valdat_wghts[, colnames(kmod$centers)],</pre>
                         seq(nrow(valdat_wghts)))
ctr_list <- vector("list", length= nrow(valdat))</pre>
```

```
for(i in 1:nrow(valdat)) {
                  ctr_list[[i]] <- kmod$centers</pre>
              }
              names(ctr_list) <- rownames(valdat)</pre>
              # Get the predictions for the validation set.
              cluster_assgns <- mcmapply(getCluster, valdat_asList, ctr_list,</pre>
                                          SIMPLIFY=TRUE, USE.NAMES= FALSE, mc.cores=6)
              valdat_wghts$cluster <- as.numeric(cluster_assgns)</pre>
              valdat wghts$pred Type <- NA
              # Apply mapping to the assigned clusters. Since valdat is
              # quite small (around 35 records if folds = 5), and there
              # are 3 clusters, it is possible that one of the LHS expressions
              # below is NA. I will make the changes if the program fails.
              valdat_wghts[which(valdat_wghts$cluster==1), c("pred_Type")] <- as.numeric(mapping["1"]</pre>
              valdat_wghts[which(valdat_wghts$cluster==2), c("pred_Type")] <- as.numeric(mapping["2"]</pre>
              valdat wghts[which(valdat wghts$cluster==3), c("pred Type")] <- as.numeric(mapping["3"]</pre>
              # Generate confusion matrix for the k-means clusters and
              # the corresponding f-score.
              preds <- as.factor(valdat_wghts$pred_Type)</pre>
              names(preds) <- rownames(valdat)</pre>
              ans <- get_confusion(preds, valdat[, "Type", drop=FALSE])</pre>
              return(ans[[2]])
In [60]: # Use the correct seed, i.e., the seed we have been
          # using for this comparative score.
         set.seed(1931)
         seed_vector <- sample(1:9999, 2000, replace=FALSE)</pre>
         start <- Sys.time()</pre>
         paste("Start time: ", start, sep="")
         dat result <- compute cvScore km(seed vector, train)</pre>
         stop <- Sys.time()</pre>
         round(stop - start, 2)
         # Time difference of 7.59 mins
         'Start time: 2021-06-01 12:06:31'
         Time difference of 7.59 mins
In [61]: summary(dat_result$Acc)
         # The interguartile range is 1.6 percentage points.
         # For svm02 this range was 1.7 points.
         # Both distributions have the same maximum value, to
         # 3 significant digits.
            Min. 1st Qu. Median
                                      Mean 3rd Qu.
                                                       Max.
            0.776
                   0.821
                            0.831
                                     0.829
                                             0.837
                                                      0.866
In [62]: round(mean(dat_result$Acc), 4)
         # Accuracy over 2000 seeds is 0.8293. Without weights,
         # this score was 0.8295. svm02 had an average
         # cross-val accuracy score of 0.8278.
         0.8293
In [63]: round(median(dat result$Acc), 4)
          round(sd(dat_result$Acc), 6)
         # median: 0.8311
```

#### Distribution of cross-val accuracy scores for hybrid k-means model with weights



### **Section 4 Comments**

While I have not done an exhaustive search for weights, current results are showing that weights are not helping to improve the accuracy score when this score is measured over a very large number of seeds, or folds. In this case, each of the folds is only 35 or 36 records (20% of the data). We saw the same result, but with a different set of weights, in Section 3---the result being that using weights did not improve upon the original model. Different weights give us different average accuracy scores on validation data, but these differences disappear as we increase the number of sets of validation data to a very large number---in this case, 2000 times 5, or 10,000 folds (sets).

Although I did not foresee it, this asymptotic-like behavior makes sense. I was looking for a stable answer regarding best model, overlooking a different kind of consequence of the fact that a model's performance varies no small amount depending on the data that the model is applied to. If we present each model with enough data, the average performances might not be all that different, especially if---as was the case here--- the type of model is the same in all cases; I was only changing the weights.

Nonetheless, the primary reason for this lack of difference, I suspect, is the scaling I am applying to the data: centering each of the variables, scaling so that each variable has unit variance, and then mapping all values into the range, [0, 1].

The alternative method of finding weights, looking for a minimum in tot.withinss (total within group sum of squares), does seem to work, although I am not yet fully convinced. While the weights I found using this method performed no worse than the weights I found using cross-validation with accuracy scores, we have found that we might be able to say this about any of the sets of weights tested.

# Section 5: Get scores for k-means models, absent min-max scaling

```
In [14]: summary(train[, -1])
```

```
Hue
                              Phenols
                                            Alcalinity
                 :0.480
                          Min. :0.98
          Min.
                                          Min. :10.6
          1st Qu.:0.782
                          1st Qu.:1.74
                                          1st Qu.:17.2
          Median :0.965
                          Median :2.35
                                          Median:19.5
          Mean :0.957
                          Mean :2.30
                                          Mean :19.5
          3rd Qu.:1.120
                          3rd Qu.:2.80
                                          3rd Qu.:21.5
                                 :3.88
                 :1.710
                          Max.
                                          Max.
In [20]: # Before scaling, transform the data a bit so
         # the variable ranges are more alike in terms
         # of range.
         df <- train
         df$Phenols <- (df$Phenols)^0.5</pre>
         df$Alcalinity <- (df$Alcalinity)^0.18</pre>
         summary(df[, -1])
               Hue
                              Phenols
                                            Alcalinity
          Min. :0.480
                          Min. :0.99
                                         Min. :1.53
          1st Qu.:0.782
                          1st Qu.:1.32
                                          1st Qu.:1.67
          Median :0.965
                          Median :1.53
                                          Median :1.71
          Mean :0.957
                          Mean :1.50
                                          Mean :1.70
          3rd Qu.:1.120
                          3rd Qu.:1.67
                                          3rd Qu.:1.74
          Max. :1.710
                          Max. :1.97
                                          Max. :1.84
In [21]: |df_scaled <- scale(df[, -1], center=TRUE, scale=TRUE)</pre>
         df_scaled <- as.data.frame(cbind(df$Type, df_scaled),</pre>
                                     row.names=rownames(df))
         colnames(df scaled) <- colnames(df)</pre>
         summary(df_scaled[, -1])
               Hue
                               Phenols
                                               Alcalinity
          Min. :-2.089
                           Min. :-2.429
                                             Min. :-3.2418
          1st Qu.:-0.765
                           1st Qu.:-0.858
                                             1st Qu.:-0.6406
          Median : 0.033
                           Median : 0.163
                                            Median : 0.0717
          Mean : 0.000
                           Mean : 0.000
                                            Mean : 0.0000
          3rd Qu.: 0.711
                           3rd Qu.: 0.823
                                             3rd Qu.: 0.6370
          Max.
                 : 3.292
                           Max.
                                  : 2.233
                                            Max.
                                                    : 2.6425
In [22]: # Run k-means with number of clusters set to 3.
         set.seed(1233)
         fit_km < - kmeans(df_scaled[, -1], 3, iter.max = 50, nstart = 30)
         print(fit_km$size)
         [1] 73 59 46
In [18]: datout <- as.data.frame(cbind(df_scaled$Type, fit_km$cluster))</pre>
         colnames(datout) <- c("Type", "cluster")</pre>
         rownames(datout) <- rownames(df_scaled)</pre>
         head(datout)
         A data.frame: 6 × 2
              Type cluster
              <dbl>
                    <dbl>
                 2
                       3
          113
          159
                 3
                       3
          21
                 1
                       1
                       2
          131
                 3
                       2
          137
                 3
                       2
          133
                 3
```

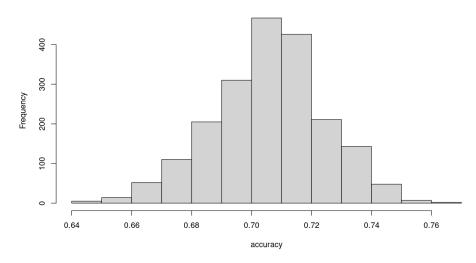
```
In [19]: # We need to map cluster to Type level.
         table(datout$Type, as.factor(datout$cluster))
               1 2 3
            1 49 0 10
            2 24 14 33
            3 0 45 3
In [20]: ans <- as.matrix(table(datout$Type, as.factor(datout$cluster)))</pre>
         print(apply(ans, MARGIN=2, which.max))
         1 2 3
         1 3 2
In [21]: # Apply mapping resulting from fit_km above.
         tmpdat <- datout</pre>
         tmpdat[which(tmpdat$cluster== 1),]$Type <- 1</pre>
         tmpdat[which(tmpdat$cluster== 2),]$Type <- 3</pre>
         tmpdat[which(tmpdat$cluster== 3),]$Type <- 2</pre>
In [23]: # Generate confusion matrix for the k-means clusters.
         # Output accuracy for this confusion matrix.
         preds <- as.factor(tmpdat$Type)</pre>
         names(preds) <- rownames(tmpdat)</pre>
         ans <- get_confusion(preds, df_scaled[, "Type", drop=FALSE])</pre>
         print(ans$matrix)
         print(paste("Accuracy score for the base k-means model: ", as.character(ans[[2]]), sep=""))
         # Section 2 base k-means accuracy score on training set: 0.7135
         # current accuracy score: 0.7135
             1 2 3 class.error
         1 49 10 0
                          0.1695
         2 24 33 14
                          0.5352
         3 0 3 45
                          0.0625
         [1] "Accuracy score for the base k-means model: 0.7135"
```

# Get comparative cross-val score for this new base k-means model

```
# The next step is crucial.
              valdat scaled <- as.data.frame(df02, row.names=rownames(valdat))</pre>
              colnames(valdat_scaled) <- colnames(valdat)[-1]</pre>
              #########################
               # Construct k-means model.
              kmod <- kmeans(df, 3, iter.max = 50, nstart=30)</pre>
              # See how the clusters are associated with Type level.
              dfout <- as.data.frame(cbind(traindat$Type, kmod$cluster))</pre>
              colnames(dfout) <- c("Type", "cluster")</pre>
               rownames(dfout) <- rownames(traindat)</pre>
              mapping <- get_mapping(dfout)</pre>
               #################################
              # Apply the k-means model to valdat_scaled.
              # Each element of the following list is a row of valdat_wghts.
              valdat_asList <- split(valdat_scaled[, colnames(kmod$centers)],</pre>
                                        seq(nrow(valdat scaled)))
              ctr_list <- vector("list", length= nrow(valdat))</pre>
              for(i in 1:nrow(valdat)) {
                   ctr_list[[i]] <- kmod$centers</pre>
              names(ctr list) <- rownames(valdat)</pre>
               # Get the predictions for the validation set.
              cluster_assgns <- mcmapply(getCluster, valdat_asList, ctr_list,</pre>
                                             SIMPLIFY=TRUE, USE.NAMES= FALSE, mc.cores=6)
              valdat scaled$cluster <- as.numeric(cluster assgns)</pre>
              valdat_scaled$pred_Type <- NA</pre>
              # Apply mapping to the assigned clusters. Since valdat is
              # quite small (around 35 records if folds = 5), and there
               # are 3 clusters, it is possible that one of the LHS expressions
              # below is NA. I will make the changes if the program fails.
              valdat_scaled[which(valdat_scaled$cluster==1), c("pred_Type")] <- as.numeric(mapping["1
valdat_scaled[which(valdat_scaled$cluster==2), c("pred_Type")] <- as.numeric(mapping["2</pre>
              valdat_scaled[which(valdat_scaled$cluster==3), c("pred_Type")] <- as.numeric(mapping["3</pre>
              # Generate confusion matrix for the k-means clusters and
              # the corresponding f-score.
              preds <- as.factor(valdat scaled$pred Type)</pre>
              names(preds) <- rownames(valdat)</pre>
              ans <- get_confusion(preds, valdat[, "Type", drop=FALSE])</pre>
              return(ans[[2]])
In [23]: # Again, use the same initial seed that we have
          # been using for this score.
          set.seed(1931)
          seed vector <- sample(1:9999, 2000, replace=FALSE)</pre>
          # Use df instead of train since df has the variable
          # transformations.
          start <- Sys.time()</pre>
          paste("Start time: ", start, sep="")
          ans <- compute_cvScore_km(seed_vector, df)</pre>
          stop <- Sys.time()</pre>
```

```
round(stop - start, 2)
         # Time difference of 5.26 mins
         'Start time: 2021-06-02 10:15:20'
         Time difference of 5.26 mins
In [24]: summary(ans$Acc)
            Min. 1st Qu.
                           Median
                                     Mean 3rd Qu.
                                                      Max.
           0.642
                    0.692
                            0.708
                                     0.706
                                            0.719
                                                     0.765
In [25]: round(mean(ans$Acc), 4)
         # 0.7057
         # In Section 2 above, using the min-max scaling, our
         # score was 0.7095
         0.7057
In [26]: round(median(ans$Acc), 4)
          round(sd(ans$Acc), 6)
         # median: 0.7075
         # sd: 0.018619
         # Section 2 median: 0.7085
         # Section 2 sd: 0.024145
         0.7075
         0.018619
In [27]: options(repr.plot.width= 10, repr.plot.height= 6)
         hist(ans$Acc, breaks=16, xlab="accuracy",
              main="Distribution of cross-val accuracy scores for base k-means model")
```

#### Distribution of cross-val accuracy scores for base k-means model



```
In []: ### COMMENTS:

# The scores are somewhat better when we use the min-max
# scaling on top of the regular scaling. Here I applied
# some transformations to the variables and then applied
# regular scaling. When I did this in Part 2, I then
# found weights and those weights made a real difference
# in the score. So it is worth looking into whether
# weights make a difference in this case, too.
```

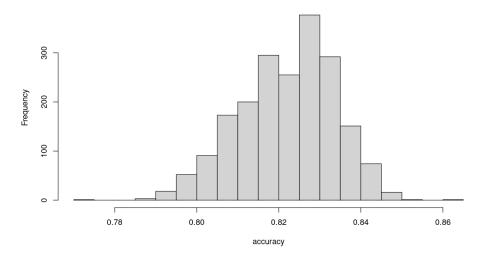
# Get comparative cross-val score for the corresponding hybrid model

```
In [29]: # Function for obtaining average of confusion matrix accuracy
           # score. This function is called from compute_cvScore_km.
           get cvScore hybrid02 <- function(traindat, valdat) {</pre>
               # Scale traindat for purpose of an svm model.
               svm_scaled <- scale(traindat[, -1])
svm_centers <- attr(svm_scaled, "scaled:center")
svm_scales <- attr(svm_scaled, "scaled:scale")</pre>
                svm_scaled <- as.data.frame(cbind(traindat$Type, svm_scaled),</pre>
                                                  row.names=rownames(traindat))
               colnames(svm_scaled) <- colnames(traindat)</pre>
               # This is our current best svm model for the trainset data
               svmod <- svm(I(as.factor(Type)) ~ ., data=svm_scaled, kernel="radial",</pre>
                               gamma= 0.03, cost= 25, scale=FALSE, probability=TRUE)
               preds <- predict(svmod, newdata=svm_scaled, scale=FALSE, probability=TRUE)</pre>
               traindat$prob01 <- as.numeric(attr(preds, "probabilities")[, 2])
traindat$prob02 <- as.numeric(attr(preds, "probabilities")[, 3])</pre>
               # Apply transformations to Phenols and Alcalinity
               traindat$Phenols <- (traindat$Phenols)^0.5</pre>
                traindat$Alcalinity <- (traindat$Alcalinity)^0.18
               ################################
               # Scale training set data for the k-means model.
               traindat_scaled <- scale(traindat[, -1], center=TRUE, scale=TRUE)</pre>
               centers <- attr(traindat_scaled, "scaled:center")
scales <- attr(traindat_scaled, "scaled:scale")</pre>
                rownames(traindat_scaled) <- rownames(traindat)</pre>
                ####################################
               # Prepare valdat for svm modeling.
               svmval_scaled <- scale(valdat[, -1], center=svm_centers, scale=svm_scales)</pre>
                svmval_scaled <- as.data.frame(svmval_scaled, row.names=rownames(valdat))</pre>
               # Compute prob01 and prob02 columns.
               preds01_b <- predict(svmod, newdata=svmval_scaled, scale=FALSE, probability=TRUE)</pre>
               valdat$prob01 <- as.numeric(attr(preds01_b, "probabilities")[, 2])
valdat$prob02 <- as.numeric(attr(preds01_b, "probabilities")[, 3])</pre>
               # Apply transformations to Phenols and Alcalinity
               valdat$Phenols <- (valdat$Phenols)^0.5</pre>
               valdat$Alcalinity <- (valdat$Alcalinity)^0.18</pre>
               # Scale valdat.
               valdat_scaled <- scale(valdat[, -1], center=centers, scale=scales)</pre>
               # The next step is crucial.
               valdat_scaled <- as.data.frame(valdat_scaled, row.names=rownames(valdat))</pre>
               ###################################
               # Construct k-means model.
               kmod <- kmeans(traindat_scaled, 3, iter.max = 50, nstart=30)</pre>
               # See how the clusters are associated with Type level.
               dfout <- as.data.frame(cbind(traindat$Type, kmod$cluster))</pre>
```

```
colnames(dfout) <- c("Type", "cluster")</pre>
              rownames(dfout) <- rownames(traindat)</pre>
              mapping <- get_mapping(dfout)</pre>
              ###############################
              # Apply the k-means model to valdat_scaled.
              # Each element of the following list is a row of valdat_scaled.
              valdat_asList <- split(valdat_scaled[, colnames(kmod$centers)],</pre>
                                      seq(nrow(valdat_scaled)))
              ctr_list <- vector("list", length= nrow(valdat))</pre>
              for(i in 1:nrow(valdat)) {
                  ctr_list[[i]] <- kmod$centers</pre>
              names(ctr list) <- rownames(valdat)</pre>
              # Get the predictions for the validation set.
              cluster_assgns <- mcmapply(getCluster, valdat_asList, ctr_list,</pre>
                                           SIMPLIFY=TRUE, USE.NAMES= FALSE, mc.cores=6)
              valdat_scaled$cluster <- as.numeric(cluster_assgns)</pre>
              valdat_scaled$pred_Type <- NA</pre>
              # Apply mapping to the assigned clusters. Since valdat is
              # quite small (around 35 records if folds = 5), and there
              # are 3 clusters, it is possible that one of the LHS expressions
              # below is NA. I will make the changes if the program fails.
              valdat_scaled[which(valdat_scaled$cluster==1), c("pred_Type")] <- as.numeric(mapping["1</pre>
              valdat_scaled[which(valdat_scaled$cluster==2), c("pred_Type")] <- as.numeric(mapping["2")</pre>
              valdat_scaled[which(valdat_scaled$cluster==3), c("pred_Type")] <- as.numeric(mapping["3</pre>
              # Generate confusion matrix for the k-means clusters and
              # the corresponding f-score.
              preds <- as.factor(valdat_scaled$pred_Type)</pre>
              names(preds) <- rownames(valdat)</pre>
              ans <- get_confusion(preds, valdat[, "Type", drop=FALSE])</pre>
              return(ans[[2]])
In [30]: # Use the same initial seed as we have been using for
          # this score.
         set.seed(1931)
         seed vector <- sample(1:9999, 2000, replace=FALSE)</pre>
         # Here we need to use train instead of df because
         # svm02 is using un-transformed columns.
         start <- Sys.time()</pre>
         paste("Start time: ", start, sep="")
         ans <- compute_cvScore_km(seed_vector, train)</pre>
         stop <- Sys.time()</pre>
          round(stop - start, 2)
         # Time difference of 7.13 mins
          'Start time: 2021-06-02 10:43:32'
         Time difference of 7.13 mins
In [31]: summary(ans$Acc)
            Min. 1st Qu. Median
                                      Mean 3rd Qu.
                                                       Max.
            0.775 0.815
                             0.821
                                      0.822 0.831
                                                       0.865
```

```
In [32]: round(mean(ans$Acc), 4)
         # 0.8219
         # Section 2 score: 0.8295
         # For svm02, this score was 0.8278.
         0.8219
In [33]: round(median(ans$Acc), 4)
         round(sd(ans$Acc), 6)
         # 0.8209
         # 0.011109
         # Section 2 median: 0.8314
         # Section 2 sd: 0.012021
         0.8209
         0.011109
         options(repr.plot.width= 10, repr.plot.height= 6)
         hist(ans$Acc, breaks=14, xlab="accuracy",
              main="Distribution of cross-val accuracy scores for hybrid k-means model")
```

#### Distribution of cross-val accuracy scores for hybrid k-means model



```
### COMMENTS:

# Not only is the min-max scaling taking a bit less time,
# we are also seeing an important (statistically significant)
# difference in the scores. Because I do not think that applying
# weights to this model will get us the boost we need to beat
# the 0.8295 score, I am not going to search for weights. In
# Part 2 with the cow data, when weights were applied to the
# base k-means model, the gain was only 0.007 (i.e., less than
# 1 percentage point). Here in Part 4 the bar is much higher and,
# thus, we are not likely to see a gain even of 0.007 with the
# best weights that we find. (The bar is much higher because we
# have less than half the records to work with and the accuracy
# score we are trying to beat is 11 percentage points higher.)
```

# Get comparative cross-val score for a hybrid model employing pca

The point of employing pca is to reduce the dimensions of train in order to make it easier to find weights. But in what

follows, I see reason to apply min-max scaling in the process, absent any weights. This means, in turn, that weights are unlikely to improve upon the following average cross-val accuracy score.

```
In [35]: # Apply transformations.
         df <- train
         df$Phenols <- (df$Phenols)^0.5</pre>
         df$Alcalinity <- (df$Alcalinity)^0.18</pre>
In [52]: # The R help for prcomp points out that we ought to set
         # scale.=TRUE. If I do not scale, PC1 and PC2 together
         # capture around 99% of the variance. But it is highly
         # unlikely we would see such performance on new data.
         # Scaling should give us a model that performs better on
         # new data.
         pca2 <- prcomp(df[, -1], retx=TRUE, rank.=2, center=TRUE,</pre>
                         scale.=TRUE)
         summary(pca2)
         Importance of first k=2 (out of 3) components:
                                   PC1
         Standard deviation
                                 1.311 0.858
         Proportion of Variance 0.573 0.245
         Cumulative Proportion 0.573 0.818
In [39]: kmpca <- as.data.frame(pca2$x)</pre>
         colnames(kmpca) <- paste0("pc", 1:2)</pre>
         rownames(kmpca) <- rownames(df)</pre>
         summary(kmpca)
                                  pc2
               pc1
          Min. :-0.6176 Min. :-0.43125
          1st Qu.:-0.1952 1st Qu.:-0.10162
                            Median : 0.00229
          Median :-0.0593
          Mean : 0.0000
                             Mean : 0.00000
          3rd Qu.: 0.2143
                             3rd Qu.: 0.09219
                           3rd vu.. ....
Max. : 0.41777
          Max. : 0.6158
 In [ ]: # It makes sense to move the data into the range, [0,1],
         # because that is where the data for prob01 and prob02 lie.
         kmpca scaled <- apply(kmpca, MARGIN=2, range01)</pre>
         summary(kmpca scaled)
In [51]: # See if pca works better with no transformations on
         # the variables.
         pca2_b <- prcomp(train[, -1], retx=TRUE, rank.=2, center=TRUE,</pre>
                           scale.=TRUE)
         summary(pca2_b)
         Importance of first k=2 (out of 3) components:
                                   PC1
                                         PC2
         Standard deviation
                                1.300 0.865
         Proportion of Variance 0.563 0.249
         Cumulative Proportion 0.563 0.813
In [60]: # Function for obtaining average of confusion matrix accuracy
         # score. This function is called from compute_cvScore_km.
         get cvScore hybrid pca <- function(traindat, valdat) {</pre>
             # Scale traindat for purpose of an svm model.
             svm_scaled <- scale(traindat[, -1])</pre>
             svm_centers <- attr(svm_scaled, "scaled:center")</pre>
```

```
svm_scales <- attr(svm_scaled, "scaled:scale")</pre>
svm scaled <- as.data.frame(cbind(traindat$Type, svm scaled),</pre>
                               row.names=rownames(traindat))
colnames(svm_scaled) <- colnames(traindat)</pre>
# This is our current best svm model for the trainset data
svmod <- svm(I(as.factor(Type)) ~ ., data=svm_scaled, kernel="radial",</pre>
              gamma= 0.03, cost= 25, scale=FALSE, probability=TRUE)
preds <- predict(svmod, newdata=svm_scaled, scale=FALSE, probability=TRUE)</pre>
prob01 <- as.numeric(attr(preds, "probabilities")[, 2])
prob02 <- as.numeric(attr(preds, "probabilities")[, 3])</pre>
# Scale the probability columns.
prob01_scaled <- scale(prob01, center=TRUE, scale=TRUE)</pre>
prob01_center <- attr(prob01_scaled, "scaled:center")</pre>
prob01_scale <- attr(prob01_scaled, "scaled:scale")</pre>
prob02_scaled <- scale(prob02, center=TRUE, scale=TRUE)</pre>
prob02 center <- attr(prob02 scaled, "scaled:center")</pre>
prob02 scale <- attr(prob02 scaled, "scaled:scale")</pre>
# Transform columns of traindat.
traindat$Phenols <- (traindat$Phenols)^0.5</pre>
traindat$Alcalinity <- (traindat$Alcalinity)^0.18</pre>
# Apply pca to traindat.
pca <- prcomp(traindat[, -1], retx=TRUE, rank.=2, center=TRUE,</pre>
                scale.=TRUE)
dftmp <- as.data.frame(pca$x, row.names=rownames(traindat))</pre>
# Add in the probability columns.
dftmp$prob01 <- prob01 scaled
dftmp$prob02 <- prob02 scaled
# Apply min-max scaling.
dftmp_scaled <- apply(as.matrix(dftmp), MARGIN=2, range01)</pre>
traindat_scaled <- as.data.frame(dftmp_scaled, row.names=rownames(traindat))</pre>
colnames(traindat_scaled) <- c(paste0("pc", 1:2),"prob01","prob02")</pre>
# Get mins and maxs for scaling of valdat.
traindat_mins <- as.numeric(apply(as.matrix(dftmp), MARGIN=2, min))</pre>
traindat_maxs <- as.numeric(apply(as.matrix(dftmp), MARGIN=2, max))</pre>
################################
# Prepare valdat for svm modeling.
svmval_scaled <- scale(valdat[, -1], center=svm_centers, scale=svm_scales)</pre>
svmval_scaled <- as.data.frame(svmval_scaled, row.names=rownames(valdat))</pre>
# Compute prob01 and prob02 columns.
preds01 b <- predict(symod, newdata=symval scaled, scale=FALSE, probability=TRUE)</pre>
prob01_b <- as.numeric(attr(preds01_b, "probabilities")[, 2])
prob02_b <- as.numeric(attr(preds01_b, "probabilities")[, 3])</pre>
# Scale the probability columns.
prob01b_scaled <- scale(prob01_b, center=prob01_center, scale=prob01_scale)</pre>
prob02b_scaled <- scale(prob02_b, center=prob02_center, scale=prob02_scale)</pre>
# Transform columns of valdat.
valdat$Phenols <- (valdat$Phenols)^0.5</pre>
valdat$Alcalinity <- (valdat$Alcalinity)^0.18</pre>
# Apply pca to valdat.
valpca <- predict(pca, valdat[, -1])</pre>
```

```
# Add in the probability columns.
              df02tmp$prob01 <- prob01b_scaled</pre>
              df02tmp$prob02 <- prob02b_scaled
              # Apply min-max scaling.
              df03_t <- t(as.matrix(df02tmp))</pre>
              df03_asList <- split(df03_t, seq(nrow(df03_t)))</pre>
              names(df03_asList) <- colnames(traindat_scaled)</pre>
              valpca_scaled <- mapply(range02, df03_asList, traindat_mins, traindat_maxs)</pre>
              # The next step is crucial.
              valdat scaled <- as.data.frame(valpca scaled, row.names=rownames(valdat))</pre>
              colnames(valdat_scaled) <- colnames(traindat_scaled)</pre>
              ################################
              # Construct k-means model.
              kmod <- kmeans(traindat_scaled, 3, iter.max = 50, nstart=30)</pre>
              # See how the clusters are associated with Type level.
              dfout <- as.data.frame(cbind(traindat$Type, kmod$cluster))</pre>
              colnames(dfout) <- c("Type", "cluster")</pre>
              rownames(dfout) <- rownames(traindat)</pre>
              mapping <- get_mapping(dfout)</pre>
              #################################
              # Apply the k-means model to valdat_scaled.
              # Each element of the following list is a row of valdat_scaled.
              valdat_asList <- split(valdat_scaled[, colnames(kmod$centers)],</pre>
                                       seq(nrow(valdat_scaled)))
              ctr list <- vector("list", length= nrow(valdat))</pre>
              for(i in 1:nrow(valdat)) {
                  ctr_list[[i]] <- kmod$centers</pre>
              names(ctr_list) <- rownames(valdat)</pre>
              # Get the predictions for the validation set.
              cluster_assgns <- mcmapply(getCluster, valdat_asList, ctr_list,</pre>
                                           SIMPLIFY=TRUE, USE.NAMES= FALSE, mc.cores=6)
              valdat_scaled$cluster <- as.numeric(cluster_assgns)</pre>
              valdat_scaled$pred_Type <- NA</pre>
              # Apply mapping to the assigned clusters. Since valdat is
              \# quite small (around 35 records if folds = 5), and there
              # are 3 clusters, it is possible that one of the LHS expressions
              # below is NA. I will make the changes if the program fails.
              valdat_scaled[which(valdat_scaled$cluster==1), c("pred_Type")] <- as.numeric(mapping["1</pre>
              valdat_scaled[which(valdat_scaled$cluster==2), c("pred_Type")] <- as.numeric(mapping["2</pre>
              valdat_scaled[which(valdat_scaled$cluster==3), c("pred_Type")] <- as.numeric(mapping["3</pre>
              # Generate confusion matrix for the k-means clusters and
              # the corresponding f-score.
              preds <- as.factor(valdat_scaled$pred_Type)</pre>
              names(preds) <- rownames(valdat)</pre>
              ans <- get confusion(preds, valdat[, "Type", drop=FALSE])</pre>
              return(ans[[2]])
In [65]: # Use the same initial seed as we have been using for
         # this score.
         set.seed(1931)
```

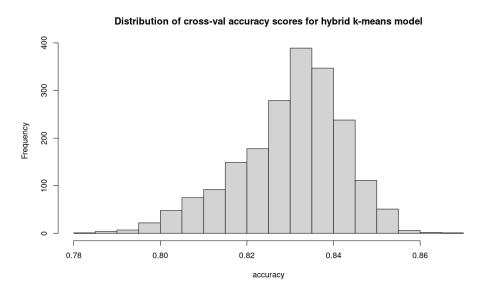
df02tmp <- as.data.frame(valpca, row.names=rownames(valdat))</pre>

```
seed_vector <- sample(1:9999, 2000, replace=FALSE)</pre>
# Here we need to use train instead of df because
# svm02 is using un-transformed columns.
start <- Sys.time()</pre>
paste("Start time: ", start, sep="")
ans <- compute_cvScore_km(seed_vector, train)</pre>
stop <- Sys.time()</pre>
round(stop - start, 2)
# Time difference of 6.65 mins
```

'Start time: 2021-06-02 12:42:29'

```
Time difference of 6.65 mins
In [66]: summary(ans$Acc)
            Min. 1st Qu.
                                    Mean 3rd Qu.
                          Median
                                                     Max.
           0.785
                   0.821
                           0.831
                                    0.830
                                          0.838
                                                    0.865
In [67]: round(mean(ans$Acc), 4)
         # 0.8298
         # Section 2 score: 0.8295
         # For svm02, this score was 0.8278.
         0.8298
In [68]: round(median(ans$Acc), 4)
         round(sd(ans$Acc), 6)
         # 0.8314
         # 0.01239
         # Section 2 median: 0.8314
         # Section 2 sd: 0.012021
         0.8314
         0.01239
```





### See what the variability is in the weights

```
In [70]: # With 4 columns, no weights = 0.25 for each column.
# Keep 0.25 at the center of each sequence.

lst <- vector("list", length= 4)
    names(lst) <- c(paste0("pc", 1:2), "prob01", "prob02")

lst[[1]] <- lst[[2]] <- lst[[3]] <- lst[[4]] <- seq(0.13, 0.37, by=0.02)

start <- Sys.time()
    dfc01 <- generate_combs(lst)
    stop <- Sys.time()
# round(stop - start, 2)

dim(dfc01)
# 1,469 4</pre>
```

```
In [71]: # Function for computing the tot.withinss for each set of
          # weights in df_params (a dataframe, each row of which is
          # a candidate set of weights). The optimal set of weights
          # will be the set that yields the smallest average (over
          # the folds) for tot.withinss.
          # This function is called from gridSearch07.
          get_tot.withinss02 <- function(traindat, valdat, wghts) {</pre>
               # Scale traindat for purpose of an svm model.
              svm scaled <- scale(traindat[, -1])</pre>
              svm_centers <- attr(svm_scaled, "scaled:center")</pre>
               svm scales <- attr(svm scaled, "scaled:scale")</pre>
              svm_scaled <- as.data.frame(cbind(traindat$Type, svm_scaled),</pre>
                                              row.names=rownames(traindat))
              colnames(svm_scaled) <- colnames(traindat)</pre>
              # This is our current best svm model for the trainset data
               svmod <- svm(I(as.factor(Type)) ~ ., data=svm_scaled, kernel="radial",</pre>
                             gamma= 0.03, cost= 25, scale=FALSE, probability=TRUE)
              preds <- predict(svmod, newdata=svm_scaled, scale=FALSE, probability=TRUE)</pre>
              prob01 <- as.numeric(attr(preds, "probabilities")[, 2])</pre>
              prob02 <- as.numeric(attr(preds, "probabilities")[, 3])</pre>
              # Scale the probability columns.
              prob01_scaled <- scale(prob01, center=TRUE, scale=TRUE)</pre>
              prob01_center <- attr(prob01_scaled, "scaled:center")</pre>
              prob01_scale <- attr(prob01_scaled, "scaled:scale")</pre>
              prob02_scaled <- scale(prob02, center=TRUE, scale=TRUE)</pre>
              prob02_center <- attr(prob02_scaled, "scaled:center")
prob02_scale <- attr(prob02_scaled, "scaled:scale")</pre>
               ################################
              # Transform columns of traindat.
              traindat$Phenols <- (traindat$Phenols)^0.5</pre>
              traindat$Alcalinity <- (traindat$Alcalinity)^0.18</pre>
              # Apply pca to traindat.
              pca <- prcomp(traindat[, -1], retx=TRUE, rank.=2, center=TRUE,</pre>
                              scale.=TRUE)
              dftmp <- as.data.frame(pca$x, row.names=rownames(traindat))</pre>
              # Add in the probability columns.
```

```
dftmp$prob01 <- prob01_scaled</pre>
    dftmp$prob02 <- prob02 scaled
    # Apply min-max scaling.
    # dftmp_scaled <- apply(as.matrix(dftmp), MARGIN=2, range01)</pre>
    # traindat scaled <- as.data.frame(dftmp scaled, row.names=rownames(traindat))</pre>
    # colnames(traindat_scaled) <- c(paste0("pc", 1:2),"prob01","prob02")</pre>
    # Get mins and maxs for scaling of valdat.
    traindat_mins <- as.numeric(apply(as.matrix(dftmp), MARGIN=2, min))</pre>
    traindat_maxs <- as.numeric(apply(as.matrix(dftmp), MARGIN=2, max))</pre>
    ###################################
    # Prepare valdat for svm modeling.
    svmval_scaled <- scale(valdat[, -1], center=svm_centers, scale=svm_scales)</pre>
    svmval_scaled <- as.data.frame(svmval_scaled, row.names=rownames(valdat))</pre>
    # Compute prob01 and prob02 columns.
    preds01_b <- predict(svmod, newdata=svmval_scaled, scale=FALSE, probability=TRUE)</pre>
    prob01_b <- as.numeric(attr(preds01_b, "probabilities")[, 2])
prob02_b <- as.numeric(attr(preds01_b, "probabilities")[, 3])</pre>
    # Scale the probability columns.
    prob01b_scaled <- scale(prob01_b, center=prob01_center, scale=prob01_scale)</pre>
    prob02b_scaled <- scale(prob02_b, center=prob02_center, scale=prob02_scale)</pre>
    # Transform columns of valdat.
    valdat$Phenols <- (valdat$Phenols)^0.5</pre>
    valdat$Alcalinity <- (valdat$Alcalinity)^0.18</pre>
    # Apply pca to valdat.
    valpca <- predict(pca, valdat[, -1])</pre>
    df02tmp <- as.data.frame(valpca, row.names=rownames(valdat))</pre>
    # Add in the probability columns.
    df02tmp$prob01 <- prob01b_scaled</pre>
    df02tmp$prob02 <- prob02b_scaled</pre>
    # Apply min-max scaling.
    df03 t <- t(as.matrix(df02tmp))</pre>
    df03_asList <- split(df03_t, seq(nrow(df03_t)))</pre>
    names(df03_asList) <- c(paste0("pc", 1:2),"prob01","prob02")</pre>
    valpca_scaled <- mapply(range02, df03_asList, traindat_mins, traindat_maxs)</pre>
    # The next step is crucial.
    valdat scaled <- as.data.frame(valpca scaled, row.names=rownames(valdat))</pre>
    colnames(valdat scaled) <- c(paste0("pc", 1:2),"prob01","prob02")</pre>
    # Apply weights to valdat_scaled.
    cols <- names(wghts)</pre>
    df4 <- t(t(valdat_scaled[, cols]) * as.numeric(wghts[cols]))</pre>
    # valdat_wghts <- as.data.frame(df4, row.names=rownames(valdat))</pre>
    # colnames(valdat_wghts) <- cols</pre>
    # Construct k-means model on the validation set.
    kmod <- kmeans(df4, 3, iter.max = 50, nstart=30)
    return(kmod$tot.withinss)
}
```

In [77]: # Test on a sample of 100.

```
set.seed(42)
smp <- sample(rownames(dfc01), 100, replace=FALSE)
tst_params <- dfc01[smp,]
head(tst_params)</pre>
```

A data.frame: 6 × 4

	pc1	pc2	prob01	prob02
	<dbl></dbl>	<dbl></dbl>	<dbl></dbl>	<dbl></dbl>
11425	0.33	0.27	0.17	0.23
7201	0.35	0.27	0.19	0.19
22165	0.37	0.15	0.15	0.33
20509	0.27	0.21	0.21	0.31
23329	0.25	0.13	0.29	0.33
22057	0.29	0.25	0.13	0.33

```
In [78]: # Find the best weights of those in tst_params.
# Use 120 seeds.

set.seed(1233)
seed_vector <- sample(1:9999, 120, replace=FALSE)

start <- Sys.time()
paste0("Start time: ", start)
dat_result <- gridSearch07(seed_vector, train, tst_params)
stop <- Sys.time()
round(stop - start, 2)
# Time difference of 7.06 mins (for 100 rows; 120 seeds each row)</pre>
```

'Start time: 2021-06-02 13:53:22'

Time difference of 7.06 mins

In [80]: dfc01[best\_params,]
 best\_tot.withinss

A data.frame: 1 × 4

```
        pc1
        pc2
        prob01
        prob02

        <dbl><dbl><dbl><dbl><dbl><dbl>
        <dbl>

        26377
        0.37
        0.13
        0.13
        0.37
```

0.46

```
In [81]: # Find the best weights of those in dfc01.
# Use 120 seeds. [Approx. time: 1.75 hours]
set.seed(1233)
seed_vector <- sample(1:9999, 120, replace=FALSE)
start <- Sys.time()
paste0("Start time: ", start)</pre>
```

```
dat_result <- gridSearch07(seed_vector, train, dfc01)</pre>
          stop <- Sys.time()</pre>
          round(stop - start, 2)
          datout_01 <- dat_result</pre>
          # Time difference of 1.8 hours.
          'Start time: 2021-06-02 14:03:20'
          Time difference of 1.8 hours
In [82]: best_params <- dat_result[which(dat_result$tot.withinss ==</pre>
                                            min(dat_result$tot.withinss, na.rm=TRUE)),]$row
          length(best_params)
          best_tot.withinss <- round(dat_result[which(dat_result$tot.withinss ==</pre>
                                            min(dat_result$tot.withinss, na.rm=TRUE)),]$tot.withinss, 4
          1
In [83]: dfc01[best_params,]
                                pc2
                        pc1
                                        prob01
                                                     prob02
          # 26377
                       0.37
                               0.13
                                          0.13
                                                       0.37
          best\_tot.withinss
          # 0.4615
```

A data.frame: 1 × 4

 pc1
 pc2
 prob01
 prob02

 <dbl><dbl><dbl><dbl><dbl><dbl>
 <dbl>

 26377
 0.37
 0.13
 0.13
 0.37

0.4615

```
In [84]: datout_01 <- datout_01[order(datout_01$tot.withinss, decreasing=FALSE),]
    datout_01[1:15,]</pre>
```

A data.frame: 15 x 2

	row	tot.withinss	
	<chr></chr>	<dbl></dbl>	
1379	26377	0.46152	
1289	24349	0.46377	
1277	24193	0.46483	
1189	22321	0.46559	
1392	26545	0.46580	
1380	26389	0.46712	
1081	20293	0.46731	
1177	22165	0.46735	
1302	24517	0.46798	
1166	22009	0.46799	
967	18265	0.46822	
1069	20137	0.46933	
1290	24361	0.46950	
1202	22489	0.46966	
849	16237	0.46973	

A data.frame: 6 × 5

```
pc2 prob01 prob02 tot.withinss
         pc1
       <dbl> <dbl>
                       <dbl>
                                <dbl>
                                             <dbl>
26377
        0.37
                0.13
                         0.13
                                 0.37
                                           0.46152
24349
        0.37
                0.13
                         0.15
                                 0.35
                                           0.46377
24193
        0.37
                0.15
                         0.13
                                 0.35
                                           0.46483
22321
        0.37
                0.13
                         0.17
                                 0.33
                                           0.46559
26545
        0.35
                0.13
                         0.15
                                 0.37
                                           0.46580
26389
        0.35
                                 0.37
                                           0.46712
                0.15
```

```
In [16]: # Refine the search, this time using cross-val accuracy scores.

lst <- vector("list", length= 4)
  names(lst) <- c(paste0("pc", 1:2), "prob01", "prob02")

lst[[1]] <- seq(0.36, 0.39, by=0.01)
  lst[[2]] <- seq(0.11, 0.14, by=0.01)
  lst[[3]] <- seq(0.11, 0.14, by=0.01)
  lst[[4]] <- seq(0.36, 0.39, by=0.01)

start <- Sys.time()
  dfc02 <- generate_combs(lst)
  stop <- Sys.time()
  # round(stop - start, 2)</pre>
```

```
dim(dfc02)
# 44 4
```

44 4

```
In [22]: # Function for obtaining average of confusion matrix accuracy
          # score. This function is called from gridSearch03.
          get_cvScore_hybrid_pcaWghts <- function(traindat, valdat, wghts) {</pre>
              # Scale traindat for purpose of an svm model.
              svm scaled <- scale(traindat[, -1])</pre>
              svm_centers <- attr(svm_scaled, "scaled:center")</pre>
              svm_scales <- attr(svm_scaled, "scaled:scale")</pre>
              svm_scaled <- as.data.frame(cbind(traindat$Type, svm_scaled),</pre>
                                              row.names=rownames(traindat))
              colnames(svm_scaled) <- colnames(traindat)</pre>
              # This is our current best svm model for the trainset data
              svmod <- svm(I(as.factor(Type)) ~ ., data=svm_scaled, kernel="radial",</pre>
                             gamma= 0.03, cost= 25, scale=FALSE, probability=TRUE)
              preds <- predict(svmod, newdata=svm_scaled, scale=FALSE, probability=TRUE)</pre>
              prob01 <- as.numeric(attr(preds, "probabilities")[, 2])</pre>
              prob02 <- as.numeric(attr(preds, "probabilities")[, 3])</pre>
              # Scale the probability columns.
              prob01_scaled <- scale(prob01, center=TRUE, scale=TRUE)</pre>
              prob01_center <- attr(prob01_scaled, "scaled:center")</pre>
              prob01_scale <- attr(prob01_scaled, "scaled:scale")</pre>
              prob02_scaled <- scale(prob02, center=TRUE, scale=TRUE)</pre>
              prob02_center <- attr(prob02_scaled, "scaled:center")
prob02_scale <- attr(prob02_scaled, "scaled:scale")</pre>
              #################################
              # Transform columns of traindat.
              traindat$Phenols <- (traindat$Phenols)^0.5</pre>
              traindat$Alcalinity <- (traindat$Alcalinity)^0.18</pre>
              # Apply pca to traindat.
              pca <- prcomp(traindat[, -1], retx=TRUE, rank.=2, center=TRUE,</pre>
                              scale.=TRUE)
              dftmp <- as.data.frame(pca$x, row.names=rownames(traindat))</pre>
              # Add in the probability columns.
              dftmp$prob01 <- prob01_scaled</pre>
              dftmp$prob02 <- prob02 scaled
              # Apply min-max scaling.
              dftmp_scaled <- apply(as.matrix(dftmp), MARGIN=2, range01)</pre>
              traindat_scaled <- as.data.frame(dftmp_scaled, row.names=rownames(traindat))</pre>
              colnames(traindat_scaled) <- c(paste0("pc", 1:2), "prob01", "prob02")</pre>
              # Get mins and maxs for scaling of valdat.
              traindat mins <- as.numeric(apply(as.matrix(dftmp), MARGIN=2, min))</pre>
              traindat maxs <- as.numeric(apply(as.matrix(dftmp), MARGIN=2, max))</pre>
              ###############################
              # Apply weights to traindat. The sqrt should have
              # been taken in the calling function.
              cols <- names(wghts)</pre>
              df2 <- t(t(traindat_scaled[, cols]) * as.numeric(wghts[cols]))</pre>
              traindat wghts <- as.data.frame(df2, row.names=rownames(traindat))</pre>
```

```
# Prepare valdat for svm modeling.
svmval_scaled <- scale(valdat[, -1], center=svm_centers, scale=svm_scales)</pre>
svmval_scaled <- as.data.frame(svmval_scaled, row.names=rownames(valdat))</pre>
# Compute prob01 and prob02 columns.
preds01 b <- predict(symod, newdata=symval scaled, scale=FALSE, probability=TRUE)</pre>
prob01_b <- as.numeric(attr(preds01_b, "probabilities")[, 2])</pre>
prob02_b <- as.numeric(attr(preds01_b, "probabilities")[, 3])</pre>
# Scale the probability columns.
prob01b_scaled <- scale(prob01_b, center=prob01_center, scale=prob01_scale)</pre>
prob02b_scaled <- scale(prob02_b, center=prob02_center, scale=prob02_scale)</pre>
# Transform columns of valdat.
valdat$Phenols <- (valdat$Phenols)^0.5</pre>
valdat$Alcalinity <- (valdat$Alcalinity)^0.18</pre>
# Apply pca to valdat.
valpca <- predict(pca, valdat[, -1])</pre>
df02tmp <- as.data.frame(valpca, row.names=rownames(valdat))</pre>
# Add in the probability columns.
df02tmp$prob01 <- prob01b_scaled</pre>
df02tmp$prob02 <- prob02b_scaled</pre>
# Apply min-max scaling.
df03_t <- t(as.matrix(df02tmp))</pre>
df03_asList <- split(df03_t, seq(nrow(df03_t)))</pre>
names(df03_asList) <- colnames(traindat_scaled)</pre>
valpca_scaled <- mapply(range02, df03_asList, traindat_mins, traindat_maxs)</pre>
# The next step is crucial.
valdat scaled <- as.data.frame(valpca scaled, row.names=rownames(valdat))</pre>
colnames(valdat scaled) <- colnames(traindat scaled)</pre>
# Apply weights to valdat_scaled.
df4 <- t(t(valdat_scaled[, cols]) * as.numeric(wghts[cols]))</pre>
valdat_wghts <- as.data.frame(df4, row.names=rownames(valdat))</pre>
colnames(valdat_wghts) <- colnames(traindat_scaled)</pre>
################################
# Construct k-means model.
kmod <- kmeans(traindat_wghts, 3, iter.max = 50, nstart=30)</pre>
# See how the clusters are associated with Type level.
dfout <- as.data.frame(cbind(traindat$Type, kmod$cluster))</pre>
colnames(dfout) <- c("Type", "cluster")</pre>
rownames(dfout) <- rownames(traindat)</pre>
mapping <- get_mapping(dfout)</pre>
#####################################
# Apply the k-means model to valdat scaled.
# Each element of the following list is a row of valdat_scaled.
valdat_asList <- split(valdat_wghts[, colnames(kmod$centers)],</pre>
                         seq(nrow(valdat_wghts)))
ctr_list <- vector("list", length= nrow(valdat))</pre>
for(i in 1:nrow(valdat)) {
    ctr_list[[i]] <- kmod$centers</pre>
names(ctr_list) <- rownames(valdat)</pre>
```

```
# Get the predictions for the validation set.
               cluster_assgns <- mcmapply(getCluster, valdat_asList, ctr_list,</pre>
                                                SIMPLIFY=TRUE, USE.NAMES= FALSE, mc.cores=6)
               valdat_scaled$cluster <- as.numeric(cluster_assgns)</pre>
               valdat scaled$pred Type <- NA
               # Apply mapping to the assigned clusters. Since valdat is
               # quite small (around 35 records if folds = 5), and there
               # are 3 clusters, it is possible that one of the LHS expressions
               # below is NA. I will make the changes if the program fails.
               valdat_scaled[which(valdat_scaled$cluster==1), c("pred_Type")] <- as.numeric(mapping["1
valdat_scaled[which(valdat_scaled$cluster==2), c("pred_Type")] <- as.numeric(mapping["2
valdat_scaled[which(valdat_scaled$cluster==3), c("pred_Type")] <- as.numeric(mapping["3</pre>
               # Generate confusion matrix for the k-means clusters and
               # the corresponding f-score.
               preds <- as.factor(valdat_scaled$pred_Type)</pre>
               names(preds) <- rownames(valdat)</pre>
               ans <- get_confusion(preds, valdat[, "Type", drop=FALSE])</pre>
                return(ans[[2]])
In [23]: # Find the best weights of those in dfc02.
           # Use 120 seeds.
           set.seed(1233)
           seed vector <- sample(1:9999, 120, replace=FALSE)</pre>
           start <- Sys.time()</pre>
           paste0("Start time: ", start)
           dat_result <- gridSearch03(seed_vector, train, dfc02)</pre>
           stop <- Sys.time()</pre>
           round(stop - start, 2)
           datout_02 <- dat_result</pre>
           # Time difference of 16.12 mins
           'Start time: 2021-06-03 09:24:59'
           Time difference of 16.12 mins
In [24]: best_params <- dat_result[which(dat_result$Acc ==</pre>
                                                 max(dat_result$Acc, na.rm=TRUE)),]$row
           length(best_params)
           best_Acc <- dat_result[which(dat_result$Acc ==</pre>
                                                 max(dat result$Acc, na.rm=TRUE)),]$Acc
In [25]: dfc02[best_params,]
                                        pc2
                                                 prob01
                                                               prob02
                          pc1
                         0.37
           # 58
                                       0.13
                                                   0.14
                                                                  0.36
           best Acc
           A data.frame: 1 × 4
                       pc2 prob01 prob02
                 pc1
               <dbl> <dbl>
                             <dbl>
                                    <dbl>
                              0.14
                                     0.36
                0.37
                      0.13
           0.83172
In [26]: |datout_02 <- datout_02[order(datout_02$Acc, decreasing=TRUE),]</pre>
```

```
datout_02[1:15,]
         A data.frame: 15 x 2
               row
                       Acc
              <chr>
                      <dbl>
           9
                58 0.83172
          32
                169 0.83161
          44
               241 0.83150
          34
                181 0.83141
           7
                52 0.83100
          33
                178 0.83081
          40
               214 0.83077
          22
                   0.83072
          10
                 61 0.83071
                118 0.83070
          41
               217 0.83062
           8
                 55 0.83048
          20
               115 0.83044
          18
                106 0.83034
                 46 0.83033
           6
         bestrows <- (datout 02$row)[1:5]
         dfc03 <- as.data.frame(cbind(dfc02[bestrows,], datout_02[1:5, c("Acc")]),</pre>
                                     row.names=bestrows)
         colnames(dfc03) <- c(colnames(dfc02), "Acc")</pre>
         dfc03
         A data.frame: 5 × 5
                                             Acc
                       pc2 prob01 prob02
                pc1
               <dbl> <dbl>
                            <dbl>
                                    <dbl>
                                           <dbl>
           58
                0.37
                      0.13
                             0.14
                                     0.36 0.83172
          169
                0.36
                      0.13
                             0.13
                                     0.38 0.83161
          241
                0.36
                      0.11
                             0.14
                                     0.39 0.83150
          181
                0.36
                      0.12
                             0.14
                                     0.38 0.83141
           52
                0.39
                      0.11
                             0.14
                                     0.36 0.83100
In [ ]: ### COMMENT:
         # I could continue to search for even better
         # weights. But instead, let's see if we can
         # get a cross-val accuracy score that equals
         # or exceeds 0.8310 with the row 58 weights above.
```

## Get comparative cross-val score for pca hybrid model with weights

```
In [28]: # Function for obtaining average of confusion matrix accuracy
# score. This function is called from get_cvScore_km.

wghts <- c(0.37, 0.13, 0.14, 0.36)^0.5
names(wghts) <- c(paste0("pc", 1:2), "prob01", "prob02")

get_cvScore_pcaHybrid_wghts <- function(traindat, valdat) {</pre>
```

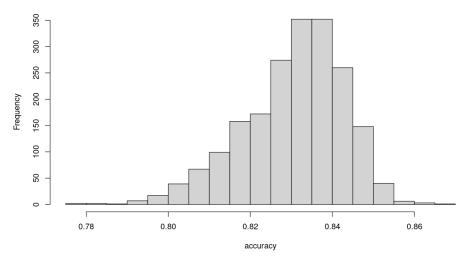
```
# Scale traindat for purpose of an svm model.
svm_scaled <- scale(traindat[, -1])</pre>
svm_centers <- attr(svm_scaled, "scaled:center")
svm_scales <- attr(svm_scaled, "scaled:scale")</pre>
svm_scaled <- as.data.frame(cbind(traindat$Type, svm_scaled),</pre>
                                row.names=rownames(traindat))
colnames(svm_scaled) <- colnames(traindat)</pre>
# This is our current best svm model for the trainset data
svmod <- svm(I(as.factor(Type)) ~ ., data=svm_scaled, kernel="radial",</pre>
               gamma= 0.03, cost= 25, scale=FALSE, probability=TRUE)
preds <- predict(symod, newdata=sym scaled, scale=FALSE, probability=TRUE)</pre>
prob01 <- as.numeric(attr(preds, "probabilities")[, 2])
prob02 <- as.numeric(attr(preds, "probabilities")[, 3])</pre>
# Scale the probability columns.
prob01_scaled <- scale(prob01, center=TRUE, scale=TRUE)</pre>
prob01_center <- attr(prob01_scaled, "scaled:center")</pre>
prob01 scale <- attr(prob01 scaled, "scaled:scale")</pre>
prob02_scaled <- scale(prob02, center=TRUE, scale=TRUE)</pre>
prob02_center <- attr(prob02_scaled, "scaled:center")</pre>
prob02_scale <- attr(prob02_scaled, "scaled:scale")</pre>
##################################
# Transform columns of traindat.
traindat$Phenols <- (traindat$Phenols)^0.5</pre>
traindat$Alcalinity <- (traindat$Alcalinity)^0.18</pre>
# Apply pca to traindat.
pca <- prcomp(traindat[, -1], retx=TRUE, rank.=2, center=TRUE,</pre>
                scale.=TRUE)
dftmp <- as.data.frame(pca$x, row.names=rownames(traindat))</pre>
# Add in the probability columns.
dftmp$prob01 <- prob01_scaled</pre>
dftmp$prob02 <- prob02_scaled</pre>
# Apply min-max scaling.
dftmp scaled <- apply(as.matrix(dftmp), MARGIN=2, range01)</pre>
traindat_scaled <- as.data.frame(dftmp_scaled, row.names=rownames(traindat))</pre>
colnames(traindat_scaled) <- c(paste0("pc", 1:2), "prob01", "prob02")</pre>
# Get mins and maxs for scaling of valdat.
traindat mins <- as.numeric(apply(as.matrix(dftmp), MARGIN=2, min))
traindat maxs <- as.numeric(apply(as.matrix(dftmp), MARGIN=2, max))
###############################
# Apply weights to traindat. The sqrt should have
# been taken in the calling function.
cols <- names(wghts)</pre>
df2 <- t(t(traindat_scaled[, cols]) * as.numeric(wghts[cols]))</pre>
traindat_wghts <- as.data.frame(df2, row.names=rownames(traindat))</pre>
################################
# Prepare valdat for svm modeling.
svmval scaled <- scale(valdat[, -1], center=svm centers, scale=svm scales)</pre>
svmval_scaled <- as.data.frame(svmval_scaled, row.names=rownames(valdat))</pre>
# Compute prob01 and prob02 columns.
preds01_b <- predict(svmod, newdata=svmval_scaled, scale=FALSE, probability=TRUE)</pre>
prob01_b <- as.numeric(attr(preds01_b, "probabilities")[, 2])
prob02_b <- as.numeric(attr(preds01_b, "probabilities")[, 3])</pre>
```

```
# Scale the probability columns.
prob01b_scaled <- scale(prob01_b, center=prob01_center, scale=prob01_scale)</pre>
prob02b_scaled <- scale(prob02_b, center=prob02_center, scale=prob02_scale)</pre>
# Transform columns of valdat.
valdat$Phenols <- (valdat$Phenols)^0.5</pre>
valdat$Alcalinity <- (valdat$Alcalinity)^0.18</pre>
# Apply pca to valdat.
valpca <- predict(pca, valdat[, -1])</pre>
df02tmp <- as.data.frame(valpca, row.names=rownames(valdat))</pre>
# Add in the probability columns.
df02tmp$prob01 <- prob01b_scaled</pre>
df02tmp$prob02 <- prob02b_scaled</pre>
# Apply min-max scaling.
df03_t <- t(as.matrix(df02tmp))</pre>
df03_asList <- split(df03_t, seq(nrow(df03_t)))</pre>
names(df03 asList) <- colnames(traindat scaled)</pre>
valpca scaled <- mapply(range02, df03 asList, traindat mins, traindat maxs)</pre>
# The next step is crucial.
valdat_scaled <- as.data.frame(valpca_scaled, row.names=rownames(valdat))</pre>
colnames(valdat_scaled) <- colnames(traindat_scaled)</pre>
# Apply weights to valdat_scaled.
df4 <- t(t(valdat_scaled[, cols]) * as.numeric(wghts[cols]))
valdat_wghts <- as.data.frame(df4, row.names=rownames(valdat))</pre>
colnames(valdat_wghts) <- colnames(traindat_scaled)</pre>
###############################
# Construct k-means model.
kmod <- kmeans(traindat wghts, 3, iter.max = 50, nstart=30)</pre>
# See how the clusters are associated with Type level.
dfout <- as.data.frame(cbind(traindat$Type, kmod$cluster))</pre>
colnames(dfout) <- c("Type", "cluster")</pre>
rownames(dfout) <- rownames(traindat)</pre>
mapping <- get_mapping(dfout)</pre>
###############################
# Apply the k-means model to valdat_scaled.
# Each element of the following list is a row of valdat_scaled.
valdat_asList <- split(valdat_wghts[, colnames(kmod$centers)],</pre>
                          seq(nrow(valdat wghts)))
ctr_list <- vector("list", length= nrow(valdat))</pre>
for(i in 1:nrow(valdat)) {
    ctr_list[[i]] <- kmod$centers</pre>
names(ctr list) <- rownames(valdat)</pre>
# Get the predictions for the validation set.
cluster_assgns <- mcmapply(getCluster, valdat_asList, ctr_list,</pre>
                              SIMPLIFY=TRUE, USE.NAMES= FALSE, mc.cores=6)
valdat scaled$cluster <- as.numeric(cluster assgns)</pre>
valdat_scaled$pred_Type <- NA</pre>
# Apply mapping to the assigned clusters. Since valdat is
# quite small (around 35 records if folds = 5), and there
# are 3 clusters, it is possible that one of the LHS expressions
```

0.012394

```
# below is NA. I will make the changes if the program fails.
              valdat_scaled[which(valdat_scaled$cluster==1), c("pred_Type")] <- as.numeric(mapping["1</pre>
              valdat_scaled[which(valdat_scaled$cluster==2), c("pred_Type")] <- as.numeric(mapping["2</pre>
              valdat_scaled[which(valdat_scaled$cluster==3), c("pred_Type")] <- as.numeric(mapping["3</pre>
              # Generate confusion matrix for the k-means clusters and
              # the corresponding f-score.
              preds <- as.factor(valdat_scaled$pred_Type)</pre>
              names(preds) <- rownames(valdat)</pre>
              ans <- get_confusion(preds, valdat[, "Type", drop=FALSE])</pre>
              return(ans[[2]])
In [30]: # Again, use the same initial seed that we have
         # been using for this score.
         set.seed(1931)
         seed_vector <- sample(1:9999, 2000, replace=FALSE)</pre>
         start <- Sys.time()</pre>
         paste("Start time: ", start, sep="")
         ans <- compute_cvScore_km(seed_vector, train)</pre>
         stop <- Sys.time()</pre>
         round(stop - start, 2)
         # Time difference of 5.48 mins
         'Start time: 2021-06-03 10:01:28'
         Time difference of 6.43 mins
In [31]: summary(ans$Acc)
             Min. 1st Qu. Median
                                      Mean 3rd Qu.
                                                       Max.
                            0.831
            0.775 0.821
                                     0.830
                                             0.838
                                                       0.865
In [32]: round(mean(ans$Acc), 4)
         # 0.8304
         # svm02's mean accuracy score was 0.8278.
         0.8304
In [33]: round(median(ans$Acc), 4)
          round(sd(ans$Acc), 6)
         # median: 0.8314
         # sd: 0.012394
         0.8314
```

#### Distribution of cross-val accuracy scores for hybrid model with weights



In []:

### Final Comments for Part 4

Excluding the k-means models, the best model we found for the wine dataset was svm02. svm02 has a mean cross-val accuracy score of 0.8278 and a median cross-val accuracy score of 0.8271. We found a hybrid k-means model (without weights) that was able to slightly improve upon this score. This hybrid model has a mean cross-val accuracy score of 0.8295 and a median cross-val accuracy score of 0.8314. (All scores are over 2000 samples.)

Although this hybrid model's improvement upon svm02 is slight, it is statistically significant. If we chose a weaker set of 3 predictors from the wine dataset, the best model accuracy score will decrease. This lowers the bar for the k-means hybrid model; i.e., it may increase the likelihood that the k-means hybrid model will outperform the competing models. In the Part 1 and Part 2 notebooks which worked with the cow data, the bar was lower---around 0.70 for the best non-k-means model. There the hybrid k-means model (with weights) had an average accuracy score of 0.73.

Above, in one of the Comments, I note that in previous work on the wine dataset I chose a different set of 3 predictors for Type. In that work the best non-k-means model had a mean cross-val accuracy score of 0.9056. The best hybrid k-means model I was able to find had a score of 0.9013. In this notebook, the 3 predictors chosen lowered the accuracy score bar to just under 0.83. This was enough that a hybrid k-means model could improve upon the score. In the last section above, I create a different hybrid model that yields a further incremental improvement: over 2000 samples we were able to boost the average score to 0.8304. The median score, however, remained at 0.8314.

The next step would be to choose a third set of 3 predictors, ones that lower the accuracy score of the best models even further, and then see if k-means is able to improve upon the best models, and by how much. Like other models, how well k-means does depends on the predictors chosen, so improvement isn't guaranteed simply because the highest accuracy score of the competing models is lower. But a lower accuracy score does mean more opportunity exists for a model like k-means to give a boost to the best of those lower scores.

In [ ]: