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(54) **METHODS AND COMPOSITIONS FOR PRODUCING SOLVENTS**

(75) Inventors: **Hans P. Blaschek**, Champaign, IL (US); **Steven F. Stoddard**, Cerro Gordo, IL (US); **Zhen Shi**, Champaign, IL (US)

(73) Assignees: **The Board of Trustees of the University of Illinois**, Urbana, IL (US); **Eastman Renewable Materials, LLC**, Kingsport, TN (US)

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(52) **U.S. Cl.**

CPC **C12P 7/065** (2013.01); **C12N 9/001** (2013.01); **C12N 9/006** (2013.01); **C12N 9/1217** (2013.01); **C12P 7/16** (2013.01); **C12P 7/28** (2013.01); **Y02E 50/10** (2013.01); **Y02E 50/17** (2013.01)

(58) **Field of Classification Search**

None

See application file for complete search history.

(56)

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Primary Examiner — Manjunath Rao

Assistant Examiner — William W Moore

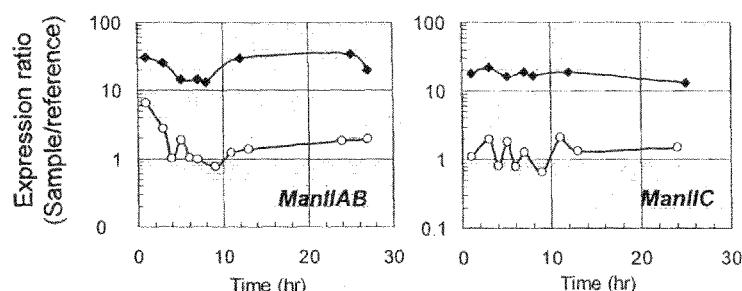
(74) *Attorney, Agent, or Firm* — McDonnell Boehnen Hulbert & Berghoff LLP

(57) **ABSTRACT**

Described herein are methods, compositions and synthetic biology approaches for solvent production, including but not limited to butanol production. Described herein are recombinant bacteria and yeast strains which may be used in production of a solvent, including but not limited to butanol, from lignocellulosic and other plant-based feedstocks. Described herein are methods of producing solvents, including but not limited to butanol, using bacteria and yeast strains. Described herein are methods of producing organisms that display highly efficient butanol production.

16 Claims, 24 Drawing Sheets

Differential expression of sugar transporters in *C. beijerinckii* NCIMB 8052 (♦) and BA101 (○).



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Figure 1. Growth curves (A) and pH profiles (B) for the fermentor cultures of *C. beijerinckii* NCIMB 8052 (♦) and BA101 (○).

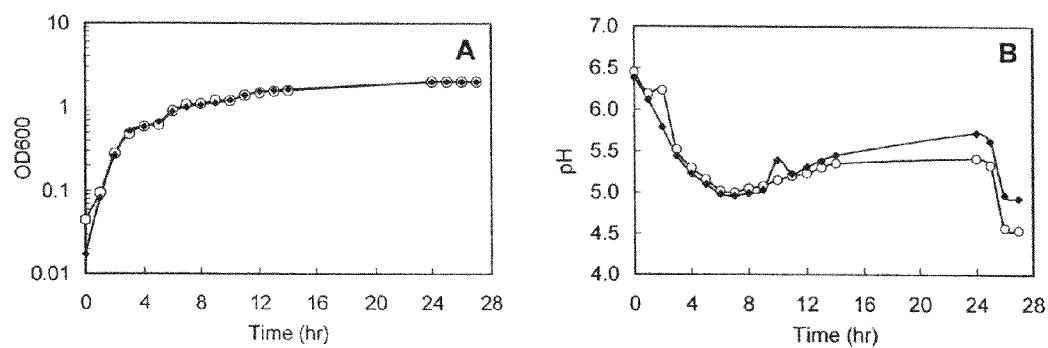


Figure 2. Product formation in the fermentor cultures of *C. beijerinckii* NCIMB 8052 (♦) and BA101 (○).

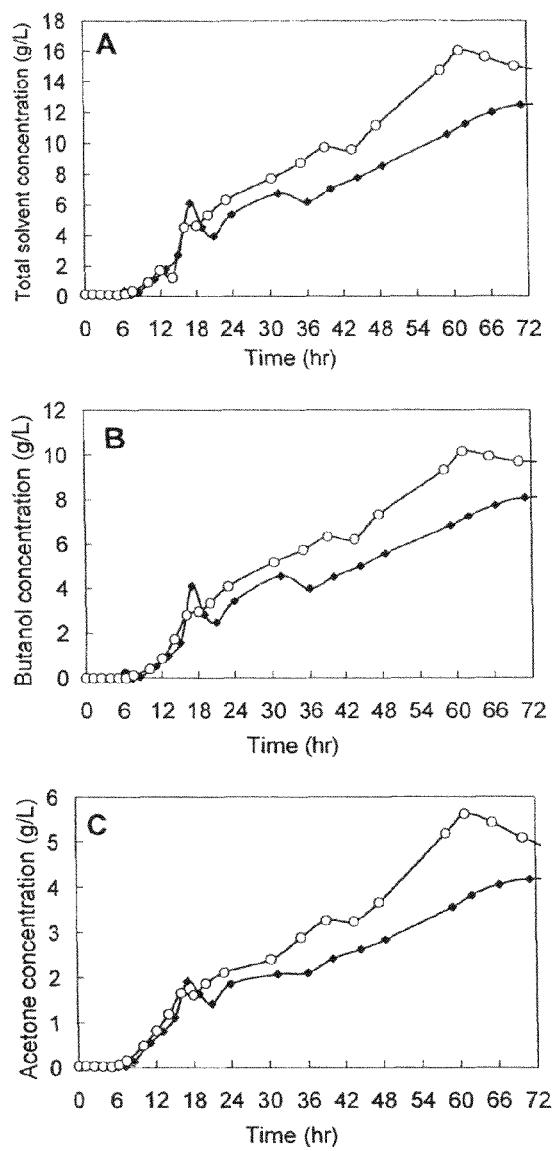


Figure 3. Gene expression profiles analyzed by DNA microarray for *C. beijerinckii* NCIMB 8052 (A) and BA101 (B) over the time course of fermentation.

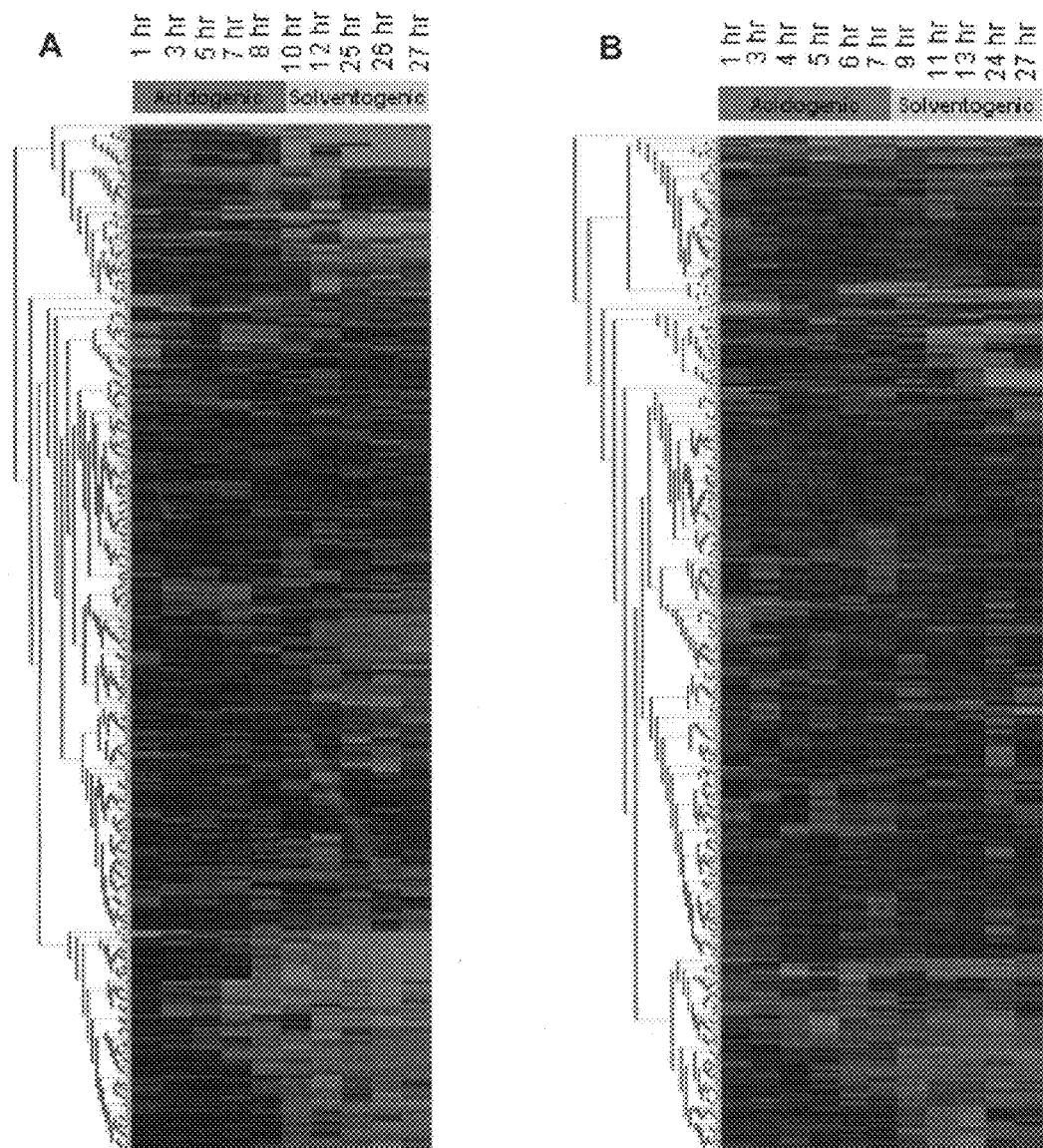


Figure 4. Differential expression of solventogenic genes in *C. beijerinckii* NCIMB 8052 (♦) and BA101 (o).

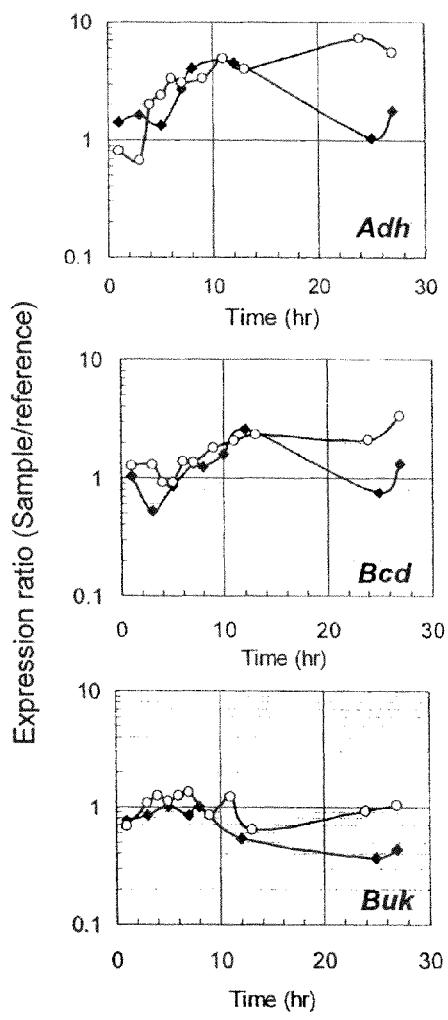


Figure 5. Differential expression of sugar transporters in *C. beijerinckii* NCIMB 8052 (♦) and BA101 (o).

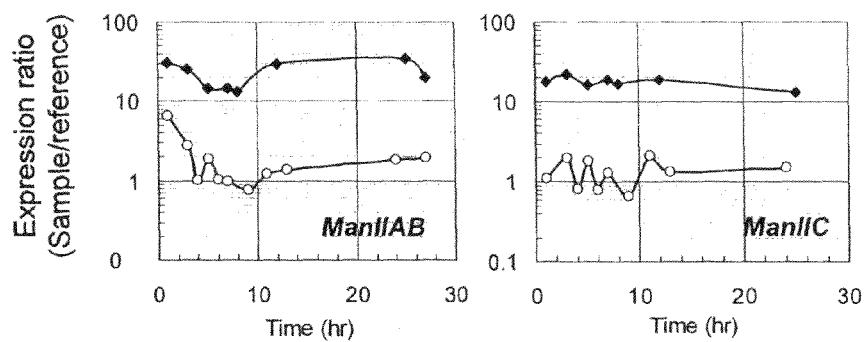


Figure 6. Differential expression of sporulation genes in *C. beijerinckii* NCIMB 8052 (♦) and BA101 (o).

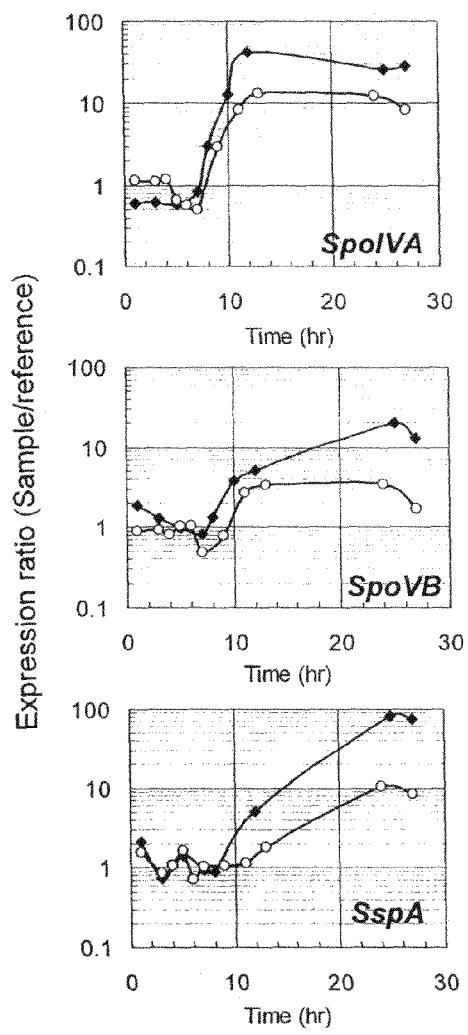


Figure 7. Differential expression of chemotaxis genes in *C. beijerinckii* NCIMB 8052 (♦) and BA101 (○).

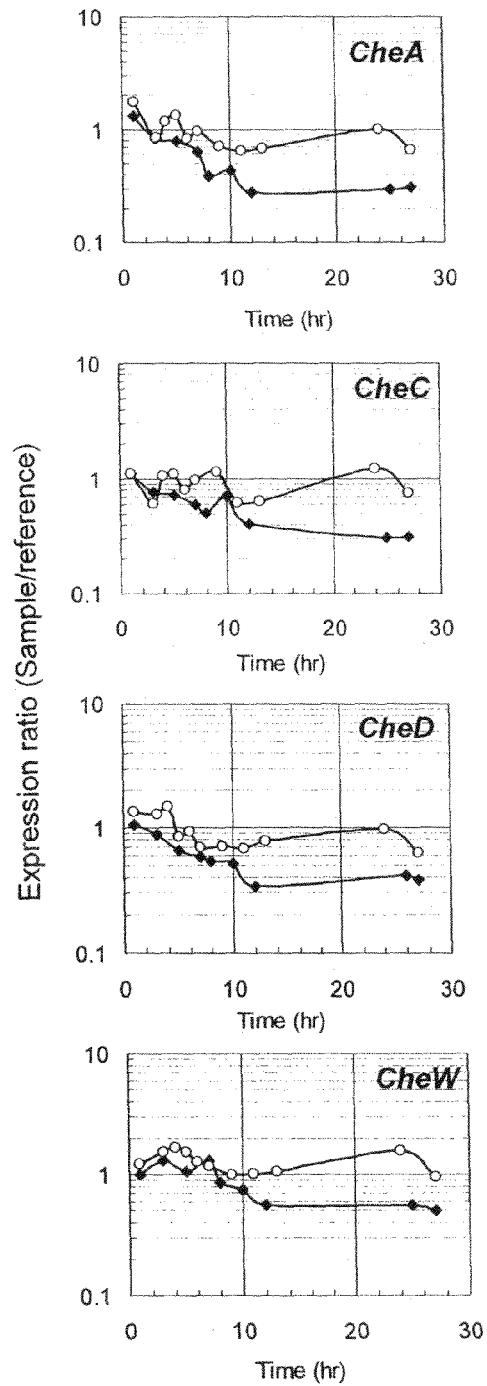


Figure 8. Solventogenic genes with comparable expression kinetics in *C. beijerinckii* NCIMB 8052 (♦) and BA101 (o).

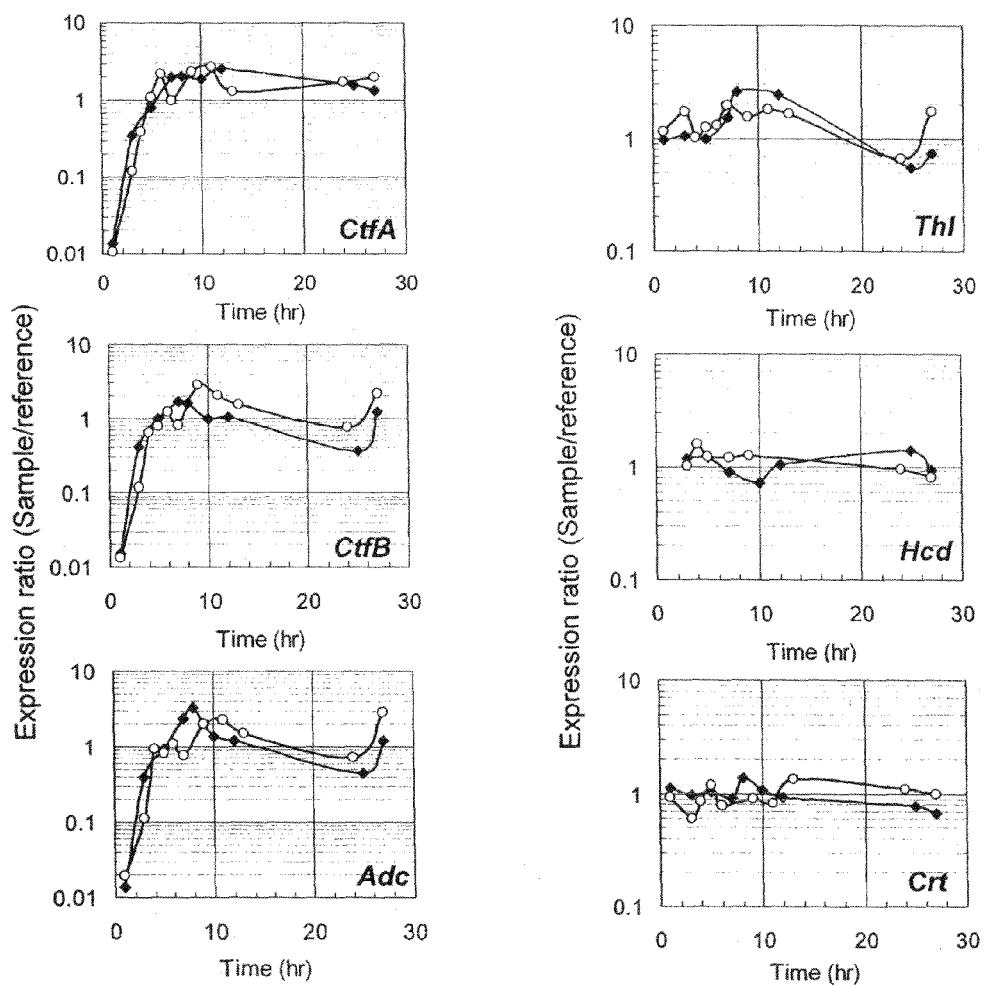


Figure 9. Reactions in the clostridial solventogenic pathway.

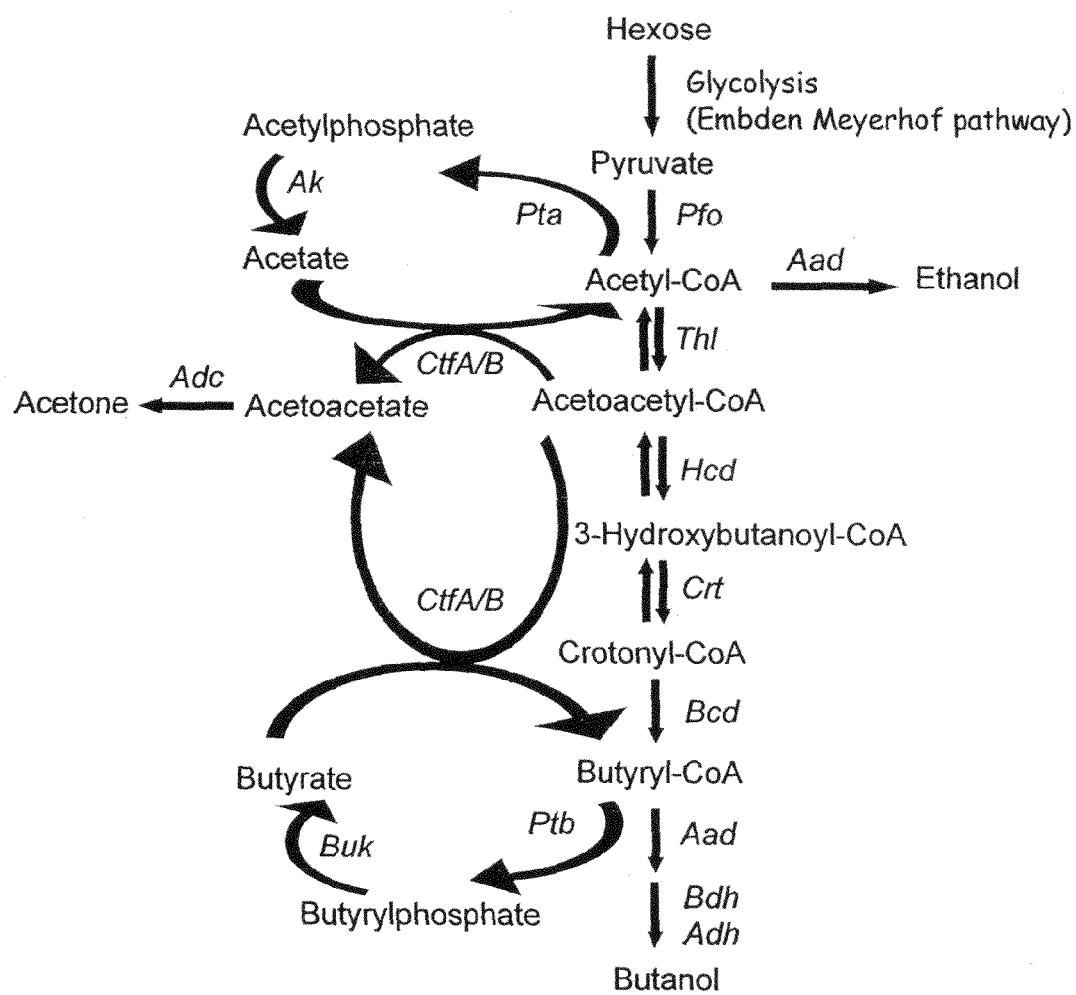


Figure 10. Adh (Alcohol dehydrogenase) *C. beijerinckii* NCIMB 8052 cDNA sequence

Figure 11. Bed (Butyryl-CoA dehydrogenase) *C. beijerinckii* NCIMB 8052 cDNA sequence

atgaattttaagtttaactagagaacaacaatttagtacaacaaatggtagagaattcgca
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ggagcaggcggagatgttcttcataatataattgtctgtgagaattatcaaaaagttgt
ggtactacaggagttattttcagcgcatacatcattatgtgcatcagtaattaatgaa
aatggaaactaacaacaaagagcaaaatatttacctgatcttgcagcggtaaaaagatc
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gctgtattagaaggggaccattatgtattaaatggttcaaaaatcttcataacaaatggt
ggagttgctgaaactttcataatattgttatgacagataagagtcaaggaacaaaagga
atttctgcattcatagtagaaaagtttcccaggattctcaatagggaaaatttagaaaac
aagatggaatcagagcatcttcaactactgagtttagttatggaaaactgcatacgtacca
aaagaaaaaccttagcaaagaaggtaaaggattttgtatagcaatgaaaactcttgat
ggaggaagaatttgtatagctgctcaagctttaggtattgcagaaggagctttgaagaa
gctgttaactatataatgaaaagaaaacaattttgtaaaccattatcagcattccaagga
ttacaatggtatatactgtaatggatgttaaatccaaagctgctaaataacttagtatac
ctagctgcaacaaagaagcaagctggtgagccttactcagtggatgctgcaagagctaaa
ttatttgctgcagatgttcaatggaaagttacaactaaggcagttcaaatcttggtgga
tatggttacactaaagaatacccagtagaaagaatgatgagagatgctaaaatatgcgaa
atctacgaaggaacttcagaagttcaaaaagatggttatcgcaggaagcatttaagatag

Figure 12. Buk (Butyrate kinase) *C. beijerinckii* NCIMB 8052 cDNA sequence

atgtcatataagctattaataatcaatccagggttcaacatcaacaaagattgggtttac
gaaggagaaaaggaactattgaagaaactttagacacacaatgaagaataaagaga
tatgatacaatatgtcaatttgatatttagaaaagaagtatattaatgttcttaaa
gaaaagaatttgatataaagactctaagtctattgttgtagaggtggaatgcttaga
ccagttgaaggtggaacatatgcagtaatgatgcaatggttgaagattaaaagttgga
gttcaaggacctcatgctctaaccattggcgaataattgccaagtcaattggagatgaa
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gcggtagctaaagatatggaaaagaaagtggacaaggatatgaaaacctaattttgt
gttgcatacatatgggtggaggcgttcagtttgtgctcacaatcatggaaaagttgtcgat
gtaaataatgcatttagatggagatggccattctcaccagaaagagctggatcagttcca
attggtgatttagttaaatgtgttttagtgaaaatataatgtgaagcagaagtatatggc
aaggctgtaggaaaaggtggattgttggttatctaaacacaaatgatgtaaaaggtgtt
attgataagatggaagaaggagataaagaatgtgaatcaatatacaaagcatttttat
caaatttcaaaagcaatcgagaaaatgtcagttgttattagaaggtaaagttgatcaaatt
attttaccggaggaattgcataactcacaacacttggccagaccttaaagcaaaagtt
aatgtgatagccccagttacagttatcctggagaagatgaattacttgctctagctcaa
ggtgctataagagtacttgatggagaacaagctaaaggttactag

Figure 13. CheA (Chemotaxis protein) *C. beijerinckii* NCIMB 8052 cDNA sequence

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aacgaatcattattagatttagagcaaaatccagatgatacagataaggtaatgaaata
tttagagttgcccacacaattaaggggatggcagcgactatggattactgatttagca
gaacttaccataagatgaaagatgtatttagctgaaatttagagaaggaaaactaaaagta
actcaggatgttagtaacagtattttgattgccttgatacattagaaaagatggttgat
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gagtttgattttgagttgaaatttgatttagtaactaaaatactgttagatgaaatatta
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gcttaaggcagagcaaaaaggaataaacacagaaggattgtcagatagtgatattaa
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caagcttacttgcataaaatgttaggagagggaaacattagcaatatacattaggattt
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tatagagaaaatgtatattccatttagtaagatgaaactttagatataaggctgat
aatactgataagaaattttgtataatgttaatgttaggtgataaaaactatgatgat
gtcgattcattacttggacaacaagaatgttataaagccactcgaaaaacattaa
aatttagatcagtatattggtgcactatacttggttaatggttagtaacgctgatact
gatgttagggcattattataa

Figure 14. CheC (Chemotaxis protein) *C. beijerinckii* NCIMB 8052 cDNA sequence

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gtggagtattctaatttaagcacttgcaatttagatgcacttaagaagttctaacata  
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gtagcttggAACAGTTGTAAGAGTGTGGCGATATTCAAGGAAATATGCTTTAGTTC  
GAACAGGAAACAGCTGAAAATGTAATAAAAAAATTAGTTGGAAGCAAGCAGTCTCCAGAA  
AGTGAATGGAAAGTTCTGTATTATGCGAGATAGCAAATATAATCAGCATCATATG  
AATGCTATTGCACAGTTAACTAATCTTGTATGGCACCATCAGTGCCTGCAACTTCATT  
GATATGTTAGTGCAATACTCACAACATACATTATTGAGTCCAATCAATATGATGAATAT  
AATCTAGATATTGAAACTGTATTCTTAGACAGTGATAACAGAAGAGAAATAGGAGGACAT  
TTTATTATATCCCTATGCCGGGTTCGTTAGAGAAAATTGAAATCTATAGGAATAAT  
TAA
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Figure 15. CheD (Chemotaxis protein) *C. beijerinckii* NCIMB 8052 cDNA sequence

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gggtgtaaaaagagaaatctcgtagctaaatagctggaggagcttcgatgtttaatttt
tctgataagagcataataagtgatattggaaaaaggaatagtgtatgcagtgaagaaaaagg
ttaaaaagaagagtcaattcctataattgctgaagagataggtggaaataaaggtagaact
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gaattgttag
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Figure 16. ManIIAB (Mannose-specific PTS system component IIAB) *C. beijerinckii* NCIMB 8052 cDNA sequence

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ttaaaaaaaaataaaataaa
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Figure 17. ManIIC (Mannose-specific PTS system component IIC) *C. beijerinckii* NCIMB 8052 cDNA sequence

```
atgactttaaatatagttcaaattatattagtcatttaatagcatttttagctggtgta  
gaaggtagttagatgaattccaattcaccaaccgataattgcttgtacattaattggc  
ttgggttacaggtatattactaccatgcttaatcttaggtggtaactcttcaaatacgcc  
tttagttggcaaatattggtgctgttagcaccgtatgcagcattagccgttgtgca  
tctgcaattatatttagttcttgaggtaaggtaaggcaggagttgtttagcaatcgct  
attgtctgttcctttagcagttgcaggattattataacaattattgtcgtagacttgct  
acagcggttcgtacatttatggatgtctgctgctaaagaaggaaatcttagagctattgat  
atgtggcaatcgctgtcttgcataaggtaacgtattgcgattccagcagcacta  
gtattagcaatcggtgcaggtcctatttagttcattacttgctgtatgcctacttggta  
actgtgttttagcaattggtggtggatgttttagctgtgtttagcaatggtaatc  
aacatgatggctacaaaagaagtatggccattttcgcaattgtttgtttagcaact  
gtttcacaaattacacttatcgacttggtaataggtagcttttagcacttcttac  
tttagcacttagcaaacaaggtaggtggctcaggtaatggtgaaattcaaatactggtagtcc  
tttagggatctaatacgatagataactaa
```

Figure 18. SpoIVa (Stage IV sporulation protein A) *C. beijerinckii* NCIMB 8052 cDNA sequence

```
atgggagggaaaaatgtggacaatttaacatatacaaagatatagccatagaacc  
ggggacatatatgttaggagtgcgttaggaccaggtaggacagggaaatctacatttattaaa  
aagttcatggatcttatgtaatcacctaaaattgataacttataaaaaggaaagagca  
aaagatgagttaccacagagtggctcaggaaaaatattcacacaacagaacctaattt  
gtacctaattgaagcagttagaaataatttagggatgaaataaaatttaaagtaaggatg  
gttactgcgttaggatattgtaaaaggagcatttaggatatttagatgttaggaaact  
aaaatggttcatacaccatggtatgattacgaaataccctttgaggatgcagcagaaatt  
gggacaagaaaagtataaaagaccattcaacaataggatttagtaattacaactgatgga  
agtataactgaaatagatagaatgactatgttagaaccagaagaaagagaatataatgaa  
ttacagtcaactaaataaccatttattgttagtttaataactaataagcctaatttcaa  
gaaactaaggcattgaaaaaagaatttagaaagataatataatgttaactgttcaagttatg  
gatgtttataatatggaagaacaagatattgaggattttaaacacgtacttaaggaa  
ttccctgtaaaagaaataatattgtatgcctaatggtagaaaaatggaatgtaac  
cattggctaaaaaagattttcaacataattatgaaacatgactgtcaagatgtctctaag  
gtaagagacataaaatactgcttaatgatggatttggaaatgaaagactttatggaaaggca  
accataaatgaaagtgtttggtagtgaaacagcaaaaatctatgaaacctaaggat  
ggaattttctataagggtcttagtgagatgtatgtaaatggtagttcagaaagtgc  
ttgctcagcataataatgacctaagtcatgcaaaatgtgaatgtataaggtaaagat  
gcattggaagatgtaaagagagagcgctatggattggtagcaccacaactttcgaaatg  
aagtttggggcctgtaaaatgttaaacaaggtatataatggagttaaatggca  
agtgcctcaagcttacatcttataaaagtgtgatattaaactgagataagccctataatg  
ggatcagaaaaggaatcggaagaacttagttaaaggccttctagaacaatttggaaacagat  
ccagcccttggcaaaagtaatgttggaaaatctttagaagtgttagttaaagaa  
ggattacaaaacaagcttatataaaatgccagaagatgttcaagttaaagattcaaaagact  
ttgcagaaaatttataatgaaaggtaatggtagttaatctgtataatactttaa
```

Figure 19. SpoVB (Stage V sporulation protein B) *C. beijerinckii* NCIMB 8052 cDNA sequence

atgaaaaaaacaatccctaataagagggagcataattcttggagttgcaggaatattaaca
agattttaggtctattcttagatggccactaataatgttaattggagatgagggatt
ggatattatcaaatgtcatacccgctatatatgttttatagctatggcttcggagtt
ccagttgcaatttctaagatgatcatcagaaaagaatgcaacaaatgatatatatggaagc
tttgaagtaatgaaagaatctgcatttaatgacaataataggacaggcacaacttta
gcattattcttttgc当地accatagtattatgttgaagtggatccaaaagcgtat
tattcattaatcggaatatcattgcaccaattgtaatatcattgtactatattttaga
ggattcttcaagggttacaaaatgactccgtcagctatcacagataatagagcag
attggtagagtcatattcggtgtggacttgctgtatatttactacctagaggcatagag
tactcagcagtggcagcattggagcaactgcgggagctgttcttggaggagcttat
ttatactcgaattataaaagagtaaagaaacgttatgccattaaaaaaaataaaagcaat
ccagaaatattaaatactatattaaagattgcaatccaatatcattagttactacagta
tcaagtattatgaatttaatagattccatatttagtaccacaaaaactattagatgcggga
ttcacaaaatgtacaatcaactgtattatatgcgc当地acaggtaaagcgtctgtatt
gtaaatattccattaactcttctatggctattgtacatctctatcattcatagct
gagaatttcatacttaagaaacagaaagagctaaaagtagatgc当地atgtaaaa
atggcatcagaattgttccgtgcactttgggttatttttagctgaaccagta
atgaagtttatattccaggcagggttgaaggaatagagatattaaatatttattcatta
acaattccattataataattactcaaacaacaacagcaataactacaaggaacaggcat
tatataaaaactgttattaaaccttgcattgggtttagttaacattgttattaaatgtt
gtttagttccatgc当地atgtttaacatataatggctgttggc当地atgtttagct
tatttaacagtaagtatttaatataatgtgatgatgaaatttacactaagagtaagactt
aatttatacggaaatattaataaaaccttgc当地atccaggattatgttatttgc
ttaataagttataatattttatataagaataacaatttagtaatggatattttgc当地
tctatattttgggtatgatgatgatattataatgataattgttattcaaggatatttaat
gttgaagaaataagagatgatattaaagaaagtaa

Figure 20. SspA (Small acid-soluble spore protein) *C. beijerinckii* NCIMB 8052 cDNA sequence

```
ttgatgtatatgtcatcaaataacagtggaaagaaatagaacattagtagtaccagaagcaaag  
gcaggattaaacagataaaaactgagggtgcctcagaagtggattaagtgattatgaa  
aacatcgataaaggaagccttcattcaagacaaaaatggatatgttggcggttatatggta  
aaacatatgattcaagattacgaacaaggcttaagtaa
```

Figure 21A. Cbei_0322 DNA sequence

atgaatttaagttaactagagaacaacaatttagtacaacaaatggtagagaattcgcagtaa
atgaagttaagccaatagctgcccaaattcgacgaaacagaaaatccctatggaaaacgttga
aaaaatggctaagcttaaaatgatgggatcccattttaaagaatttggagcaggcgga
gatgttcttcataataattgtgttgcagaattatcaaaagttgtgtactacaggagtt
ttcttcagcqcatatcattatgtcatcagtaattaatgaaactaacgaacaaag
agaaaaatattcacatgtttcagcggtaaaaagatcggtcttcggattaaacagaacca
ggtgtgttgcagatgtcgcaggacaacaaactgtgttgcattatgttat
taaatggtcaaaaatctcataacaaatggtagttgttgcattaaactttcataatattgttat
gacagataagagtcaaggaacaaaaggaatttgcattcatagtagtagaaaagtttattccagga
ttctcaataggaaaatttagaaaacaagatggaatcagagcatctcaactactgagtt
tggaaaactgcatactaccaaaagaaaacctacttagcaaaagaaggtaaggatttgatgc
aatggaaaactttgtatggaggaagaattggatagctgtcaagcttttaggtattgcagaagga
gttttgaagaagctgttaactatgaaagaaaacaatttggtaaaccattatcagcat
tccaaggattacaatggtatatactgttcaagatggatgttaaaatccaagctgtctaaataacttagt
atacctagctgcacaaagaagcaagctggtagcgttactcagtgatgtcaagagctaaa
ttatttgctgcagatgttgcattgttgcattgttgcattttggatatgc
gttacactaaagaataaccaggatgatgttgcattttggatatgc
aggaacttcagaagttcaaaagatggtatcgcaggaagcattttaaagatag
(SEQ ID NO: 12)

Figure 21B. Cbei_0322 protein sequence

mnfqltreqqlvqqmvrefavnevkpiaeideserfpmenvekmaklkmmgipfskefggagg
dvlsyiisveelskvcgtqvilsahtslicasvinengtnqrakylpdlcsgkkigafgltep
qagtdaagqqtavlegdhyvlngskifitnggvaetfiifamtdksqgtkgisafiveksfpg
fsigklenkmqirasssttelvmencivpkennlskegkfgiamktldggrigiaaqalgiaeag
afeeavnymkerkqfgkplsafqglqwyiaeemdvdkiqaakylvylaatkkqagepysvdaarak
lfaadvamevtkavqifgggygtkeypvermmrdakiceiyegtsevqkmviagsilr
(SEQ ID NO: 13)

Figure 22A. DNA sequence of Cbei_1722 adh gene

atggcacgtttactttaccaagagacatttatcatggagaaggagcacttgaggcacttaaaa
ctttaaaaggtaagaaagcttcttagtagttggcggatcaatgaaaagatttggatttct
taaacaagttaagattttaaaagaagcaggaatgaaatgaaattttgaaggtgtgaa
ccagatccatcaagtggaaacagtaatgaaaggcgagaagctatgagaaacttgcgttgcatt
ggatagttgcaatgggtggaggatcaccaattgtatgcgtcaaaggctatgtggatattctacga
ataccaggattttactttgaacaacgcgttgcattttggattaccagaccttagacaaaaaa
gctaatgttgcgttgcatttcataacaaggcggtacagctacagaatgttgcatttcagtt
tcacaaatttattcagaaaaattaaatccttagctgatttaaacataactccagatatacg
aatagttgatccagcacttgctcaaactatgccaaaaactttaacagctcatactggatggat
gcattaactcacgtatagaagcatacactgcatacttcataatttctcagatccattag
caattaaagctgttagaaatggttcaagaaaatttaatcaaattgttgcatttttttttttttttt
tagaaatctaatt
atagttcactcaatggctcataagggttgtgtgttattccatattcctcatggatgtgcaatgg
ctatatttttaccatatgtatt
tgcgagagccttaaaaatt
attaatggatt
aaagattttaaagctaatctttcattttatcgctcataacgcgtttagatgtcatgcacaggatc
aaatccttagagaaaatagatgttgcataatgaaaaatttttttttttttttttttttttttttt
aaagtttaatt
(SEQ ID NO: 14)

Figure 22B. Predicted amino Acid sequence of Cbei_1722 adh protein

MARFTLPRDLYHGEAGALEVLKTLKGKAFVVVGGGSMKRGFLKQVEDYLTEAGMEVELFEGVE
PDPSVETVMKGAEAMRNFEPEWIVAMGGGSPIDAACAMWIFYEPDFTFEQAVVPFGLPELRQK
AKFVAIPSTSGTATEVTAFSVITNYTEKIKYPLADFNITPDIAIVDPVLAQTMPKTLTAHTGMD
ALTHAIEAYTASLRSNFSDPLAIKALQMVKENLIKSFEGDKEARNLMHEAQCLAGMAFSNALLG
IVHSMAHKVGAVFHIPHGCANAIFLPYVIQYNRTKCEDRYADIARALKLEGNTDSELTDLSLGM
INKMNSDLNIPHSMKEYGVTEEDFKANLSFIAHNAVLDACTGSNPREIDDATMEKLFFECTYYGT
KVEL
(SEQ ID NO: 15)

Figure 23A. Cbei_3111 DNA sequence

atgcatatgtcatcaaataatagtggaaagaaatagaacatttagtaccagaagcaaaacaaggat
taaacagattaaaactgaggttgcttcagaagttggattaagcaatttatgaaagcatggataa
aggaaacaccttcttcaagacaaaatggatatgttggcgatatatggtaaaacatatgatcgaa
gattatgaacaaggctttaagtaa
(SEQ ID NO: 16)

Figure 23B. Cbei_3111 protein sequence

mhmssnnsgrnrtlvpeakqqlnrlkteinasevglsnsyesmdkgngnlssrqngyvggymvkhmiedyeqqglk
(SEQ ID NO: 17)

Figure 24A. Cbei_3250 DNA seq

atgtcatcaaataatagtggaaagaaacagaacattagtagcagaagcaaaacaaggattaaaca
gattaaaaactgaggttgcttcagaagttaggattacatgattacaaaatcaagataaaggaaa
tttatcttcaagacaaaatggatacggttgcggatacatggtaagcacatgattgaaagctac
gaacaaggtttaaagtaa
(SEQ ID NO: 18)

Figure 24B. Cbei_3250 Protein seq

MSSNNNSGRNRTLVPEAKQGLNRLKTEVASEVGLHDYENQDKGNLSSRQNGYVGGYMVKHMIESY
EQGLK
(SEQ ID NO: 19)

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METHODS AND COMPOSITIONS FOR PRODUCING SOLVENTS**CROSS-REFERENCE TO RELATED APPLICATIONS**

This application claims the benefit of priority to U.S. Provisional Application Ser. No. 60/930,775, filed May 17, 2007 the contents of which are incorporated by reference herein in their entirety.

INCORPORATION-BY-REFERENCE OF MATERIAL SUBMITTED ELECTRONICALLY

Incorporated by reference in its entirety herein is a computer-readable nucleotide/amino acid sequence listing submitted concurrently herewith and identified as follows: 27,804 bytes ASCII (Text) file named "703562Replacement-SequenceListing.TXT," created on Jul. 22, 2010.

STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH OR DEVELOPMENT

This invention was made with United States Government support under USDA grant number 2001-35504-10668. The Government may retain certain rights in the invention.

FIELD OF INVENTION

The compositions and methods described herein pertain to the generation of solvents, including but not limited to the generation of butanol. Specifically, the invention relates to genetic modification of solventogenic microorganisms to enhance production of solvents. More specifically, the invention relates to genetic modification of solventogenic clostridia to enhance efficiency of production of butanol.

BACKGROUND OF THE INVENTION

With the inevitable depletion of petroleum reserves, fast-growing global populations, and widespread industrialization, there has been an increasing worldwide interest in renewable energies. There is a growing consensus that producing liquid biofuels such as ethanol from renewable and inexpensive lignocellulosic-based plant materials (biomass) has a great potential to meet a large portion of this nation's energy demand in the transportation sector. Moreover, producing biofuels from biomass will simultaneously address three important societal concerns: security of supply (biofuels can be produced locally in sustainable systems), lower greenhouse gas (biofuels recycle carbon dioxide), and support of agriculture. The U.S. Department of Energy (DOE) has set a goal to replace 30% of the liquid transportation fuel with biofuels by 2030.

Similar to ethanol, butanol has many favorable attributes as a fuel molecule. However, it is an underexploited biofuel. Butanol can be produced as a co-product with ethanol and acetone from carbohydrates through fermentation by several solventogenic Clostridia. Compared to the currently popular fuel additive, ethanol, butanol has several advantages. It contains around 22% oxygen which when used as a fuel will result in more complete combustion and low exhaust smoke. In addition, it has a higher energy content (BTU/volume) than ethanol, is more miscible with gasoline and diesel, and has a lower vapor pressure and solubility characteristics which would allow for it to be shipped by pipeline, unlike ethanol.

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Solventogenic clostridia are well-known as natural producers of organic solvents via fermentation process. *C. acetobutylicum* and *C. beijerinckii* are among the prominent solvent-producing strains capable of producing acetone and butanol as the main fermentation products (Jones, D. T., and D. R. Woods. 1986. Acetone-butanol fermentation revisited. *Microbiol. Mol. Biol. Rev.* 50: 484-524.) Efforts have been made to improve the Clostridia-based butanol fermentation processes by developing new strains and downstream technologies. For example, as described in U.S. Pat. No. 6,358,717, which is incorporated herein by reference in its entirety, Blaschek and others used chemical mutagenesis to develop a mutant strain of *Clostridium beijerinckii*, BA101 with higher butanol concentration. To circumvent butanol inhibition, Blaschek and others also developed various downstream processes including gas stripping, pervaporation, and liquid-liquid extraction. See, e.g., Ezeji, T. C., Qureshi, N. & Blaschek, H. P. Butanol fermentation research: Upstream and downstream manipulations. *Chem Rec* 4, 305-314 (2004); US Pat. Pub. No. 2005/0089979; Qureshi et al., Butanol production using *Clostridium beijerinckii* BA101 hyper-butanol producing mutant strain and recovery by pervaporation, *Appl Biochem Biotech* 84-6, 225-235 (2000); Formanek et al., Enhanced butanol production by *Clostridium beijerinckii* BA101 grown in semidefined P2 medium containing 6 percent maltodextrin or glucose. *Applied and Env. Microbiol.* 63(6):2306-2310 (1997); and Ezeji et al., Acetone butanol ethanol (ABE) production from concentrated substrate: reduction in substrate inhibition by fed-batch technique and product inhibition by gas stripping, *Appl Microbiol Biot* 63, 653-658 (2004), each of which is incorporated herein by reference in its entirety.

The butanol biosynthesis pathway of the solvent producing Clostridia has been studied, and some of the enzymes involved therein have been purified and characterized. See, e.g., Boynton et al., Cloning, sequencing, and expression of clustered genes encoding beta-hydroxybutyryl-coenzyme A (CoA) dehydrogenase, crotonase, and butyryl-CoA dehydrogenase from *Clostridium acetobutylicum* ATCC 824, *Journal of Bacteriology* 178, 3015-3024 (1996); Petersen & Bennett, Cloning of the *Clostridium acetobutylicum* ATCC 824 Acetyl Coenzyme-a Acetyltransferase (Thiolase-Ec 2.3.1.9) Gene, *Applied and Environmental Microbiology* 57, 2735-2741 (1991); Petersen et al., Molecular-Cloning of an Alcohol (Butanol) Dehydrogenase Gene-Cluster from *Clostridium acetobutylicum* ATCC-824, *Journal of Bacteriology* 173, 1831-1834 (1991); and Durre et al., Solventogenic Enzymes of *Clostridium acetobutylicum*—Catalytic Properties, Genetic Organization, and Transcriptional Regulation, *Fems Microbiol Rev* 17, 251-262 (1995), each of which is incorporated herein by reference in its entirety.

Butanol fermentation has traditionally been constrained by self-limitation of the reaction due to the toxic effect of the product on the microorganism involved in the process. There is a need for producing solventogenic microorganisms such as clostridia that achieve increased efficiency in the production of bio-butanol.

BRIEF SUMMARY OF THE INVENTION

Described herein are methods, systems and synthetic biology approaches for solvent production, including but not limited to butanol production. Described herein are recombinant bacteria and yeast strains which may be used in production of butanol from lignocellulosic and other plant-based

feedstocks. Described herein are methods of producing solvents, including but not limited to butanol, using recombinant bacteria and yeast strains.

Described herein are genetically-modified solventogenic organism strains comprising altered expression or structure of a gene relative to the original organism strain, wherein such genetic modifications result in increased efficiency of solvent production. Described herein are genetically-modified solventogenic clostridia strains comprising altered expression or structure of a gene relative to the clostridia strain prior to its genetic modification, wherein such genetic modifications result in increased efficiency of butanol production. In some modifications the clostridia species is *Clostridium beijerinckii* which is an anaerobic bacterium known for the fermentative production of acetone and butanol. In some embodiments, the genetic modifications are introduced by genetic recombination. In some embodiments, the genetic modifications are introduced by nucleic acid transformation.

Described herein are methods for producing genetically-modified solventogenic organism strains wherein such genetic modifications result in increased efficiency of solvent production. Described herein are methods for identifying genetic signatures associated with increased efficiency of butanol production wherein the genetic signatures include, but are not limited to, increased or decreased expression of genes related to butanol production pathway and variants thereof, and modified or altered sequences of genes involved in or related to the butanol production pathway. Genes and sequence variants thereof that have been identified in relation to increased efficiency of solvent production are used to transform bacteria (e.g., clostridia) or other microorganisms and increased or decreased expression of these genes are correlated with more efficient butanol production by these recombinant solventogenic organisms.

Increased efficiency of solvent production can be determined in any number of ways including but not limited to: concentration (weight/volume) of solvent in fermentation medium, yield (weight/weight) of solvent per amount of substrate, and rate of solvent formation (weight/volume/time).

Described herein are recombinant solventogenic organism strains comprising increased expression of a gene selected from the group consisting of Adh, Bcd, and Buk and variants thereof, relative to the organism strain prior to its transformation.

Described herein are recombinant solventogenic organisms comprising increased expression of a gene selected from the group consisting of CheA, CheC, and CheD and variants thereof relative to the organism strain prior to its transformation.

Described herein are recombinant solventogenic organisms comprising decreased expression of a gene selected from the group consisting of ManIIAB and ManIIC and variants thereof relative to the organism strain prior to its transformation.

Described herein are recombinant solventogenic organisms comprising decreased expression of a gene selected from the group consisting of SpoIVA, SpoVB, and SspA and variants thereof relative to the organism strain prior to its transformation.

In some variations, the recombinant solventogenic organisms described herein comprise a heterologous nucleic acid sequence. In some variations, the recombinant solventogenic organisms described herein comprise an introduced heterologous nucleic acid. In some variations, expression of the heterologous nucleic acid sequence is controlled by an inducible

promoter. In some variations, expression of the heterologous nucleic acid sequence is controlled by a constitutive promoter.

In some variations, the recombinant solventogenic organisms described herein comprise an mRNA resulting from transcription of the heterologous nucleic acid sequence, wherein the mRNA accumulates to a higher or lower level relative to the organism strain prior to transformation.

In some variations, the recombinant solventogenic organisms described herein comprise a protein resulting from the heterologous nucleic acid, and the protein accumulates to a higher or lower level relative to the organism strain prior to its transformation.

In some variations, the recombinant solventogenic organisms described herein comprise a protein with an altered activation state which is correlated with increased production of a solvent, relative to the organism strain prior to its transformation.

In some variations, the recombinant solventogenic organisms described herein are yeast.

In some variations, the recombinant solventogenic organisms described herein are bacteria. In some variations, the recombinant solventogenic organisms described herein are *Escherichia*. In some variations, the recombinant solventogenic organisms described herein are *Escherichia coli*. In some variations, the recombinant solventogenic organisms described herein are *Clostridium*. In some variations, the recombinant solventogenic organisms described herein are *Clostridium beijerinckii*. In some variations, the recombinant solventogenic organisms described herein are *Clostridium acetobutylicum*.

In some variations, the recombinant solventogenic organisms described herein are cellulolytic.

In some variations, the recombinant solventogenic organisms described herein are non-cellulolytic.

In some variations, the recombinant solventogenic organisms described herein comprise an siRNA, DNAzyme, or antisense nucleic acid.

In some variations, the recombinant solventogenic organisms described herein comprise a heterologous nucleic acid from a *Clostridium*. In some variations, the recombinant solventogenic organisms described herein comprise a heterologous nucleic acid from a solventogenic *Clostridium*. In some variations, the recombinant solventogenic organisms described herein comprise a heterologous nucleic acid from a *Clostridium beijerinckii*. In some variations, the recombinant solventogenic organisms described herein comprise a heterologous nucleic acid from *Clostridium beijerinckii* 8052. In some variations, the recombinant solventogenic organisms described herein comprise a heterologous nucleic acid from *Clostridium beijerinckii* BA101.

In some variations, the recombinant solventogenic organisms described herein produce butanol. In some variations, the recombinant solventogenic organisms described herein produce ethanol. In some variations, the recombinant solventogenic organisms described herein produce acetone.

Described herein are methods of producing a solvent comprising culturing the recombinant solventogenic organisms described herein.

Described herein are methods for producing butanol, comprising culturing the recombinant solventogenic organisms described herein.

Described herein are methods for producing ethanol, comprising culturing the recombinant solventogenic organisms described herein.

Described herein are methods of identifying a gene related to production of a solvent comprising culturing cells in a

medium comprising a material which can be acted on to produce the solvent, comprising measuring the level of the solvent, and correlating an accumulation of a specific mRNA population via microarray with production of the solvent.

Described herein are methods of identifying the solventogenic potential of an organism comprising culturing cells in a medium comprising a material which can be acted on to produce the solvent, and correlating an accumulation of an mRNA population selected from the group consisting of Adh, Bcd, Buk, CheA, CheC, CheD, ManIIAB, ManIIC, SpoIVA, SpoVB, and SspA mRNA. In some variations the organism is yeast. In some variations the organism is bacteria. In some variations the organism is an *Escherichia coli*. In some variations the organism is a *Clostridium*. In some variations the organism is a *Clostridium beijerinckii*. In some variations the organism is a *Clostridium acetobutylicum*. In some variations the organism is cellulolytic. In some variations the organism is non-cellulolytic. In some variations the organism is recombinant.

These and other embodiments, features and advantages will become more apparent to those skilled in the art when taken with reference to the following more detailed description of the invention in conjunction with the accompanying drawings.

BRIEF DESCRIPTION OF THE DRAWINGS

The patent or application file contains at least one drawing executed in color. Copies of this patent or patent application publication with color drawing(s) will be provided by the Office upon request and payment of the necessary fee.

The accompanying drawings, which are incorporated herein and constitute part of this specification, illustrate presently preferred embodiments of the invention, and, together with the general description given above and the detailed description given below, serve to explain features of the invention.

FIG. 1A and FIG. 1B depict growth curves (A) and pH profiles (B), respectively, for the fermentor cultures of *C. beijerinckii* NCIMB 8052 (♦) and *C. beijerinckii* BA101 (○). This figure is described in Example 2.

FIG. 2A, FIG. 2B and FIG. 2C depict formation of total solvents (A), butanol (B), and acetone (C), respectively, in the fermentor cultures of *C. beijerinckii* NCIMB 8052 (♦) and *C. beijerinckii* BA101 (○). Time courses are shown for the production of solvents in *C. beijerinckii* BA101 in comparison with *C. beijerinckii* NCIMB 8052. This figure is described in Example 2.

FIG. 3A and FIG. 3B depict mRNA accumulation profiles analyzed by DNA microarray for *C. beijerinckii* NCIMB 8052 and *C. beijerinckii* BA101, respectively, over the time course of fermentation. This figure is in color, and is described in Example 4.

FIG. 4 quantitatively depicts differential mRNA accumulation of solventogenic genes in *C. beijerinckii* NCIMB 8052 (♦) and *C. beijerinckii* BA101 (○). Increased expression in *C. beijerinckii* BA101 during the solventogenic stage is shown for alcohol dehydrogenase (Adh), butyryl-CoA dehydrogenase (Bcd) and butyrate kinase (Buk). This figure is described in Example 4.

FIG. 5 depicts differential mRNA accumulation of sugar transporters in *C. beijerinckii* NCIMB 8052 (♦) and *C. beijerinckii* BA101 (○). Components of mannose-family phosphoenolpyruvate (PEP)-dependent phosphotransferase system IIA, IIB (ManIIAB) and IIC (ManIIC) were significantly down-regulated in *C. beijerinckii* BA101. This figure is described in Example 4.

FIG. 6 depicts differential mRNA accumulation of sporulation genes in *C. beijerinckii* NCIMB 8052 (♦) and *C. beijerinckii* BA101 (○). Induction of late stage sporulation factors was much weaker in *C. beijerinckii* BA101 than in the wild-type strain. Lowered activation in *C. beijerinckii* BA101 through the solventogenic phase is shown for coat morphosis sporulation protein (SpoIVA), Stage V sporulation protein B (SpoVB) and small acid-soluble spore protein (SspA). This figure is described in Example 4.

FIG. 7 depicts differential mRNA accumulation of chemotaxis genes in *C. beijerinckii* NCIMB 8052 (♦) and *C. beijerinckii* BA101 (○). Higher expression levels of CheA, CheC, CheD and CheW in a chemotaxis gene cluster are shown for *C. beijerinckii* BA101 during the solventogenic stage.

FIG. 8 depicts solventogenic mRNAs with comparable accumulation kinetics in *C. beijerinckii* NCIMB 8052 (♦) and *C. beijerinckii* BA101 (○). Expression of aceto-acetyl CoA:acetate-butyrate CoA transferase subunit α/β (CtfA/B) and acetoacetate decarboxylase (Adc) were highly activated

at the onset of solventogenic phase in *C. beijerinckii* BA101 and *C. beijerinckii* NCIMB 8052. Changes in expression levels were much smaller for thiolase (Thl), 3-hydroxybutyryl-CoA dehydrogenase (Hcd) and crotonase (Crt) in *C. beijerinckii* BA101 and *C. beijerinckii* NCIMB 8052. This figure is described in Example 4.

FIG. 9 depicts reactions in the clostridial solventogenic pathway. Genes involved in catalyzing the conversion of intermediate metabolites are indicated.

FIG. 10 shows the Adh (Alcohol dehydrogenase) gene Cbei_2181 of *C. beijerinckii* NCIMB 8052 DNA sequence (SEQ ID NO: 1).

FIG. 11 shows the Bcd (Butyryl-CoA dehydrogenase) gene Cbei_2035 of *C. beijerinckii* NCIMB 8052 DNA sequence (SEQ ID NO: 2).

FIG. 12 shows the Buk (Butyrate kinase) *C. beijerinckii* NCIMB 8052 DNA sequence (SEQ ID NO: 3).

FIG. 13 shows the CheA (Chemotaxis protein) *C. beijerinckii* NCIMB 8052 DNA sequence (SEQ ID NO: 4).

FIG. 14 shows the CheC (Chemotaxis protein) *C. beijerinckii* NCIMB 8052 DNA sequence (SEQ ID NO: 5).

FIG. 15 shows the CheD (Chemotaxis protein) *C. beijerinckii* NCIMB 8052 DNA sequence (SEQ ID NO: 6).

FIG. 16 shows the ManIIAB (Mannose-specific PTS system component IIAB) *C. beijerinckii* NCIMB 8052 DNA sequence (SEQ ID NO: 7).

FIG. 17 shows the ManIIC (Mannose-specific PTS system component IIC) *C. beijerinckii* NCIMB 8052 DNA sequence (SEQ ID NO: 8).

FIG. 18 shows the SpoIVA (Stage 1V sporulation protein A) *C. beijerinckii* NCIMB 8052 DNA sequence (SEQ ID NO: 9).

FIG. 19 shows the SpoVB (Stage V sporulation protein B) *C. beijerinckii* NCIMB 8052 DNA sequence (SEQ ID NO: 10).

FIG. 20 shows the SspA (Small acid-soluble spore protein) *C. beijerinckii* NCIMB 8052 DNA sequence (SEQ ID NO: 11).

The DNA sequence (SEQ ID NO: 12) of the Cbei_0322 gene homologous to Bcd (Butyryl-CoA dehydrogenase) gene Cbei_2035 of *C. beijerinckii* NCIMB 8052 is shown in FIG. 21A and the protein sequence of Cbei_0322 (SEQ ID NO: 13) shown in FIG. 21B.

The DNA sequence (SEQ ID NO: 14) of the Cbei_1722 gene homologous to the Adh (Alcohol dehydrogenase) gene Cbei_2181 of *C. beijerinckii* NCIMB 8052 is shown in FIG. 22A and predicted amino acid sequence (SEQ ID NO: 15) of Cbei_1722 is shown in FIG. 22B.

The DNA sequence of Cbei_3111 (SEQ ID NO: 16) homologous to SspA (Small acid-soluble spore protein) gene Cbei_3080 of *C. beijerinckii* NCIMB 8052 is shown in FIG. 23A and the protein sequence of Cbei_3111 (SEQ ID NO: 17) shown in FIG. 23B.

The DNA sequence of Cbei_3250 (SEQ ID NO: 18) homologous to SspA (Small acid-soluble spore protein) gene Cbei_3080 of *C. beijerinckii* NCIMB 8052 is shown in FIG. 24A and the protein sequence of Cbei_3250 (SEQ ID NO: 19) shown in FIG. 24B.

DETAILED DESCRIPTION OF THE INVENTION

The detailed description illustrates by way of example, not by way of limitation, the principles of the invention. It is to be understood that this invention is not limited to the particular methodology, protocols, cell lines, constructs, and reagents described herein and as such may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention, which will be limited only by the appended claims. Hence, the invention is not limited to the preferred embodiments described exemplarily herein. Moreover, this description will clearly enable one skilled in the art to make and use the invention, and describes several embodiments, adaptations, variations, alternatives and uses of the invention, including what is presently believed by applicant to be the best mode of carrying out the invention.

As used herein and in the appended claims, the singular forms "a," "an," and "the" include plural reference unless the context clearly indicates otherwise. For example, reference to "alcohol dehydrogenase" is a reference to one or more such proteins and includes variants and equivalents thereof known to those skilled in the art.

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood to one of ordinary skill in the art to which this invention belongs. Although any methods, devices, and materials similar or equivalent to those described herein can be used in the practice or testing of the invention, the preferred methods, devices and materials are now described. The publications discussed herein are provided solely for their disclosure prior to the filing date of the present application. Nothing herein is to be construed as an admission that the inventors are not entitled to antedate such disclosure by virtue of prior invention or for any other reason.

This invention utilizes routine techniques in the field of recombinant genetics. Basic texts disclosing the general methods of use in this invention include Sambrook et al., Molecular Cloning, A Laboratory Manual (3rd ed. 2001); Kriegler, Gene Transfer and Expression: A Laboratory Manual (1990); and Current Protocols in Molecular Biology (Ausubel et al., eds., 1994). General texts which describe molecular biological techniques include Berger and Kimmel, Guide to Molecular Cloning Techniques, Methods in Enzymology volume 152 Academic Press, Inc., San Diego, Calif. (Berger); Sambrook et al., Molecular Cloning—A Laboratory Manual (2nd Ed.), Vol. 1-3, Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y., 1989 ("Sambrook") and Current Protocols in Molecular Biology, F. M. Ausubel et al., eds., Current Protocols, a joint venture between Greene Publishing Associates, Inc. and John Wiley & Sons, Inc., (supplemented through 1999) ("Ausubel").

Described herein are 1) organisms for use in the methods and compositions described herein; 2) methods of identifying organisms for use in the methods and compositions described

herein, 3) methods of modifying organisms, 4) methods of preparing substrates, 5) methods of processing cellulose to sugars, 6) methods of generating solvents from sugars, and 7) methods of optimizing organisms for use in industrial applications.

Described herein are methods for identifying genetic signatures (increased or decreased expression of gene(s) or, variant gene sequences) associated with a mutated clostridia (*C. beijerinckii* BA101) that exhibits butanol production with increased efficiency relative to the wild type clostridia (*C. beijerinckii* NCIMB 8052)). Methods for modifying clostridia or other organisms to acquire such genetic signatures wherein acquisition of the genetic signatures results in increased efficiency of ethanol production are described herein.

Organisms for Use in the Methods and Compositions Described Herein

In the broadest sense, any prokaryotic or eukaryotic organism capable of adaptation for use in the methods and compositions described herein may be used in the methods and compositions described herein.

In some variations, bacteria, fungi, yeast or other organisms which are initially solventogenic are used in the methods and compositions described herein. As used herein, a solventogenic organism is an organism that is at least partially capable of producing a solvent such as butanol, ethanol, acetone, or isopropanol. Non-limiting examples of solventogenic microorganisms include *Clostridium* species, such as *C. beijerinckii*, *C. beijerinckii* 8052, *C. beijerinckii* BA101, *C. acetobutylicum*, *C. pasteurianum*, *C. butyricum*, *C. sporogenes*, *C. felsenium*, *C. saccharobyticum*, *C. saccharoperbutylacetonicum*, *C. tetanomorphum*, *C. aurantibutyricum*, *C. cavaeris*, *C. puniceum* and *C. thermosaccharolyticum* (Durre, P., Formation of solvents in Clostridia in 'Handbook on Clostridia', P. Durre (ed.), CRC Press-Taylor & Francis Group, Boca Raton, Fla., USA, 2005), as well as *C. algidixylanolyticum* (D M Broda, et al., *Clostridium algidixylanolyticum* sp. nov., a psychrotolerant, xylan-degrading, spore-forming bacterium. Int J Syst Evol Microbiol 50: 623-631, 2000), *C. thermopapryolyticum* (B S Mendez, et. al., *Clostridium thermopapryolyticum* sp. nov., a cellulolytic thermophile. Int J Syst Bacteriol 41 (2):281-283, 1991) and *C. carboxydivorans* (J S Liou, et. al., *Clostridium carboxydivorans* sp. nov., a solvent-producing *clostridium* isolated from an agricultural settling lagoon, and reclassification of the acetogen *Clostridium scatologenes* strain SL1 as *Clostridium drakei* sp. nov. Int J Syst Evol Microbiol 55: 2085-2091, 2005), and some non-*Clostridium* species such as *Anaerobacter polyendosporus* (V I Duda, et. al., A new anaerobic bacterium, forming up to five endospores per cell—*Anaerobacter polyendosporus* gen. et sp. nov. Arch Microbiol 148(2):121-127, 1987; A V Siunov, et. al., Phylogenetic status of *Anaerobacter polyendosporus*, an anaerobic, polysporogenous bacterium. Int J Syst Bacteriol 49: 1119-1124, 1999), *Butyrabacterium methylotrophicum* (J G Zeikus, et. al., Isolation and characterization of a new, methylotrophic, acidogenic anaerobe, the Marburg strain. Curr Microbiol 3(6):381-386, 1980; G-J Shen, et. al., Biochemical basis for carbon monoxide tolerance and butanol production by *Butyrabacterium methylotrophicum*. Appl Microbiol Biotechnol 51: 827-832, 1999), *Thermoanaerobacterium thermosaccharolyticum* and *Thermoanaerobacterium* strain Mel9 (M D Collins, et. al., The phylogeny of the genus *Clostridium*: proposal of five new genera and eleven new species combinations. Int J Syst Bacteriol 44: 812-826, 1994; P G Stroot, et. al., Description of a new butanol-producing thermophile *Thermoanaerobacterium* strain Mel9. In Abstracts of the 99th Meeting of the American Society for

Microbiology, 1999), and *Thermohydrogenium kirishiense* (EV Zacharova, et. al., *Thermohydrogenium kirishiense* gen. nov. and sp. nov., a new anaerobic thermophilic bacterium. Arch Microbiol 160: 492-497, 1993).

Anaerobic spore-forming bacteria belonging to the genus *Clostridium* have been useful in industrial applications including enzyme and solvent production. Among saccharolytic butyric acid-producing clostridia, there are a number of species capable of producing significant amounts of neutral solvents during the later stages of a batch fermentation under appropriate conditions. The strain used most extensively for the production of acetone and butanol are generally classified as *C. acetobutylicum*. A number of different species of butanol-producing clostridia are recognized based on differences in the type and ratio of the solvents produced. *C. beijerinckii* (*C. butylicum*) produces solvents in approximately the same ratio as *C. acetobutylicum*, however isopropanol is produced in place of acetone. *C. aurantibutyricum* produces both acetone and isopropanol in addition to butanol. *C. tetanomorpha* produces almost equimolar amounts of butanol and ethanol but no other solvents. (Jones and Woods (1986) *supra*).

Advantages of using *C. beijerinckii* over *C. acetobutylicum* include broader substrate range and better pH range, ability to produce butanol during log-phase growth, stability with respect to strain degeneration, and ability to use a variety of substrates to produce butanol. Moreover, the solventogenic genes on *C. beijerinckii* are located on the chromosome, whereas the genes are located on a plasmid in *C. acetobutylicum*. Thus *C. beijerinckii* is more genetically stable.

In some variations, bacteria, fungi, yeast or other organisms which are not initially solventogenic are used in the methods and compositions described herein.

Non-limiting examples of the organisms described herein include *Clostridium* sp. In some variations the *Clostridium* is *C. phytofermentans*, *C. thermohydrosulfuricum*, *C. absonum*, *C. absonum*, *C. acidisolii*, *C. akagii*, *C. algidixylanolyticum*, *C. bowmanii*, *C. cellulolyticum*, *C. cylindrosporum*, *C. diolis*, *C. estertheticum*, *C. estertheticum*, *C. estertheticum*, *C. frigidicarnis*, *C. frigidicarnis*, *C. frigor*, *C. glycolicum*, *C. papyrosolvens*, *C. perfringens*, *C. pseudotetanicum*, *C. psychrophilum*, *C. rubrum*, *C. sardinense*, *C. sardinense*, *C. thermocellum*, *C. celerecrescens*, *C. lentocellum*, *C. polysaccharolyticum*, *C. populeti*, *C. thermohydrosulfuricum*, *C. thermocellum*, *C. cellulovorans*, or *C. josui*.

In some variations, the organisms described herein include *Escherichia* sp., including *E. coli*, *Saccharomyces* sp., including *S. cerevisiae*, and various Cyanobacteria.

In some variations, the organisms described herein include *Aspergillus* sp., *Bacillus* sp., *Brevibacterium* sp., *Clostridium* sp., *Corynebacterium* sp., *Gluconobacter* sp., *Pseudomonas* sp., *Rhodococcus* sp., *Streptomyces* sp., *Xanthomonas* sp., *Candida* sp., and *Zymomonas* sp.

In some variations the organisms described herein include *Acidithiobacillus* sp., *Acinetobacter* sp., *Allochromatium* sp., *Azotobacter* sp., *Bacillus* sp., *Bdellovibrio* sp., *Cellulomonas* sp., *Desulfovibrio* sp., *Geobacillus* sp., *Gluconobacter* sp., *Kocuria* sp., *Lactobacillus* sp., *Leuconostoc* sp., *Myxococcus* sp., *Pediococcus* sp., *Propionibacterium* sp., *Pseudomonas* sp., *Raoultella* sp., *Rhizobium* sp., *Rhodospirillum* sp., *Sporosarcina* sp., *Streptomyces* sp., *Thermus* sp., *Thiobacillus* sp., *Variovorax* sp., *Vibrio* sp., *Wautersia* sp., and *Zymomonas* sp.

In some variations the organisms described herein include *Selenomonas* sp., *Methanobrevibacter* sp., *Ruminococcus* sp., *Fibrobacter* sp., *Prevotella* sp., *Treponema* sp., *Azospirillum* sp., *Cellulomonas* sp., and *Trichoderma* sp.

In some variations the organisms described herein include *Acremonium* sp., *Alternaria* sp., *Aureobasidium* sp., *Botrytis* sp., *Chaetomium* sp., *Dipodascus* sp., *Endomyces* sp., *Eremascus* sp., *Geotrichum* sp., *Hemicola* sp., *Neurospora* sp., *Penicillium* sp., *Pichia* sp., *Schizosaccharomyces* sp., *Sordaria* sp., and *Sordaria* sp.

In some variations the organisms described herein are cellulolytic. In some variations the organisms described herein are non-cellulolytic.

10 Methods of Identifying Organisms

Described herein are methods of identifying organisms for use in the methods and compositions described herein. Unless the context clearly indicates otherwise, any organism described herein may be identified by the methods described herein.

In some variations, organisms are screened for their ability to produce a particular product or products from one or more starting materials. In some variations, a culture medium or organisms in a culture medium are screened for the presence, absence, or level of a particular product. In some variations, a culture medium or organisms in a culture medium are screened for the presence, absence or level of a particular solvent, including but not limited to butanol, ethanol, or acetone. By way of nonlimiting example, screening for products or solvents may be via HPLC, mass spectrometry, GC, immunoassay, activity assay, or other methods known by those of skill in the art.

In some variations, an organism is screened for the presence, absence, or amount of a particular gene or gene product.

In some variations, DNA is screened for the presence, or absence, or copy number of a particular gene. By way of nonlimiting example, screening of DNA may be via Southern blot hybridization, PCR, microarray, or other methods known by those of skill in the art. In some variations genomic or non-genomic DNA is screened via microarray for the presence or absence of a particular gene.

In some variations, an organism's mRNA is screened for the presence, absence, or amount of a particular mRNA species. By way of nonlimiting example, screening of mRNA may be via Northern blot hybridization, PCR, microarray, or other methods known by those of skill in the art. In some variations, an organism's mRNA is screened via microarray for the presence, absence, or amount of a particular mRNA. In some variations, an organism's mRNA is screened via the method described in Example 4 for the presence, absence or amount of a particular mRNA species.

In some variations, an organism's mRNA is screened for the presence of a particular mRNA species.

In some variations, an organism's mRNA is screened for an amount of a particular mRNA species. In some variations, a recombinant organism's mRNA is screened for an amount of a particular mRNA species, relative to the organism strain prior to its transformation.

In some variations, a recombinant organism is screened for a decreased level of a particular mRNA species, relative to the organism strain prior to its transformation. In some variations, a recombinant organism is screened for an amount of decrease in level of a particular mRNA species, relative to the organism strain prior to its transformation, wherein the decreased mRNA species is used by a pathway that limits the ability of the recombinant organism to produce a preferred solvent. In some variations the amount of decrease of the mRNA species is 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 90%, 95%, 100%, relative to the organism strain prior to its transformation.

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In some variations, an organism's mRNA is screened for an increased level of a particular mRNA species. In some variations, a recombinant organism's mRNA is screened for an increased level of a particular mRNA species, relative to the organism strain prior to its transformation. In some variations, a recombinant organism's mRNA is screened for an increased level of a particular mRNA species relative to the organism strain prior to its transformation, wherein the level of the particular mRNA species is increased at least 1.5-fold, 2-fold, 4-fold, 10-fold, 25-fold, 50-fold, 100-fold relative to the organism strain prior to its transformation. In some variations, a recombinant organism's mRNA is screened for an increased level of a particular mRNA species relative to the organism strain prior to its transformation, wherein the level of the particular mRNA species is increased at least 2-fold. In some variations, a recombinant organism's mRNA is screened for an increased level of a particular mRNA species relative to the organism strain prior to its transformation, wherein the level of the particular mRNA species is increased at least 5-fold. In some variations, a recombinant organism's mRNA is screened for an increased level of a particular mRNA species relative to the organism strain prior to its transformation, wherein the level of the particular mRNA species is increased at least 10-fold. In some variations, a recombinant organism's mRNA is screened for an increased level of a particular mRNA species relative to the organism strain prior to its transformation, wherein the level of the particular mRNA species is increased at least 15-fold. In some variations, a recombinant organism's mRNA is screened for an increased level of a particular mRNA species relative to the organism strain prior to its transformation, wherein the level of the particular mRNA species is increased at least 20-fold.

In some variations, an organism's proteins are screened for the presence, absence, or amount of a particular protein, or activation state of a particular protein. By way of nonlimiting example, screening of proteins may be via Western blot hybridization, immunoassay, activity assay, microarray, various fluorescence and flow cytometry methods including fluorescence-activated cell sorting, or other methods known by those of skill in the art.

In some variations, an organism's proteins are screened for an amount of a particular protein species. In some variations, a recombinant organism's proteins are screened for an amount of a particular protein species, relative to the organism strain prior to its transformation.

In some variations, a recombinant organism is screened for a decreased level of a particular protein species, relative to the organism strain prior to its transformation. In some variations, a recombinant organism is screened for a decrease in amount of a particular protein species, relative to the organism strain prior to its transformation, wherein the decreased protein species is used by a pathway that limits the ability of the recombinant organism to produce a preferred solvent. In some variations the amount of decrease of the protein species is 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 90%, 95%, 100%, relative to the organism strain prior to its transformation.

In some variations, an organism's proteins are screened for an increased level of a particular protein species. In some variations, a recombinant organism strain's proteins are screened for an increased level of a particular protein species, relative to the organism strain prior to its transformation. In some variations, a recombinant organism's proteins are screened for an increased level of a particular protein species relative to the organism strain prior to its transformation, wherein the level of the particular protein species is increased

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about 1.5-fold, 2-fold, 5-fold, 10-fold, 20-fold, 30-fold, 40-fold, 50-fold, 60-fold, 80-fold, or 100-fold relative to the organism strain prior to its transformation. In some variations, a recombinant organism's proteins are screened for an increased level of a particular protein species relative to the organism strain prior to its transformation, wherein the level of the particular protein species is increased at least 2-fold. In some variations, a recombinant organism's proteins are screened for an increased level of a particular protein species relative to the organism strain prior to its transformation, wherein the level of the particular protein species is increased at least 5-fold. In some variations, a recombinant organism's proteins are screened for an increased level of a particular protein species relative to the organism strain prior to its transformation, wherein the level of the particular protein species is increased at least 10-fold. In some variations, a recombinant organism's proteins are screened for an increased level of a particular protein species relative to the organism strain prior to its transformation, wherein the level of the particular protein species is increased at least 20-fold. In some variations, a recombinant organism's proteins are screened for an increased level of a particular protein species relative to the organism strain prior to its transformation, wherein the level of the particular protein species is increased up to about 80-fold.

In some variations, an organism's proteins are screened for a level of a particular activated protein species. In some variations a protein is activated by phosphorylation, dephosphorylation, cleavage, refolding, or association with another molecule, including but not limited to another protein.

In some variations, an organism's proteins are screened for a level of a particular activated protein species. In some variations, a recombinant organism's proteins are screened for a level of a particular activated protein species, relative to the organism strain prior to its transformation.

In some variations, a recombinant organism is screened for a decreased level of a particular activated protein species, relative to the organism strain prior to its transformation. In some variations, a recombinant organism is screened for a decrease in level of a particular activated protein species, relative to the organism strain prior to its transformation, wherein the decreased activated protein species is used by a pathway that limits the ability of the recombinant organism to produce a preferred solvent. In some variations the amount of decrease of the activated protein species is 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 90%, 95%, 100%, relative to the organism strain prior to its transformation.

In some variations, a recombinant organism's proteins are screened for an increased level of a particular activated protein species, relative to the organism strain prior to its transformation. In some variations, a recombinant organism's proteins are screened for an increased level of a particular activated protein species relative to the organism strain prior to its transformation, wherein the level of the particular activated protein species is increased at least 1.5-fold, 2-fold, 5-fold, 10-fold, 20-fold, 30-fold, 40-fold, 50-fold, 60-fold, 80-fold, or 100-fold relative to the organism strain prior to its transformation. In some variations, a recombinant organism's proteins are screened for an increased level of a particular activated protein species relative to the organism strain prior to its transformation, wherein the level of the particular activated protein species is increased at least 1.5-fold. In some variations, a recombinant organism's proteins are screened for an increased level of a particular activated protein species relative to the organism strain prior to its transformation, wherein the level of the particular activated protein species is increased

tein species is increased at least 5-fold. In some variations, a recombinant organism's proteins are screened for an increased level of a particular activated protein species relative to the organism strain prior to its transformation, wherein the level of the particular activated protein species is increased at least 15-fold. In some variations, a recombinant organism's proteins are screened for an increased level of a particular activated protein species relative to the organism strain prior to its transformation, wherein the level of the particular activated protein species is increased at least 20-fold. In some variations, a recombinant organism's proteins are screened for an increased level of a particular activated protein species relative to the organism strain prior to its transformation, wherein the level of the particular activated protein species is increased up to about 80-fold.

In some variations, an organism is screened for a level of a particular solvent. In some variations, a recombinant organism is screened for a level of a particular solvent, relative to the organism strain prior to its transformation.

In some variations, a recombinant organism is screened for a decreased level of a particular solvent, relative to the organism strain prior to its transformation. In some variations, a recombinant organism is screened for a decrease in level of a particular solvent, relative to the organism strain prior to its transformation, wherein the decreased solvent is generated by a pathway that limits the ability of the recombinant organism to produce a preferred solvent. In some variations, the solvent which has been decreased is ethanol. In some variations, the solvent which has been decreased is acetone. In some variations, the solvent which has been decreased is butanol. In some variations the amount of decrease is 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 90%, 95%, 100%, relative to the organism strain prior to its transformation.

Increased efficiency of solvent production can be determined in any number of ways including but not limited to: concentration (weight/volume) of solvent in fermentation medium, yield (weight/weight) of solvent per amount of substrate, and rate of solvent formation (weight/volume/time).

In one aspect of the invention, a recombinant organism strain is screened for an increased level of a particular solvent, relative to the organism strain prior to its transformation.

In some variations, recombinant solventogenic organism strains are screened for producing an increased amount of a particular solvent relative to the organism strain prior to its transformation, wherein the amount of the particular solvent is increased at least 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2.0, 2.2, 2.4, 2.6, 2.8, 3.0, 3.2, 3.4, 3.6, 3.8, 4.0, 4.5, 5.0, 5.5, 6.0, 6.5, 7.0, 7.5, 8.0, 8.5, 9.0, 9.5, 10, 15, 20, 40, 60, 80, or 100-fold over that in the organism strain prior to its transformation.

Where the concentration of the solvent in the organism strain prior to its transformation is 10 g/L, the recombinant solventogenic organism strains are screened for having concentrations of the solvent of about 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, or 100 g/L.

In some variations, recombinant solventogenic organism strains are screened for producing an increased yield of a particular solvent per amount of the substrate, relative to the organism strain prior to its transformation. Where the yield of solvent in the organism strain prior to its transformation is about 20 g/100 g of substrate, recombinant solventogenic organism strains of the present invention are screened for producing yields of: 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 44, 48, 50, 52, 56, 60, 64, 68, 72, 76, or 80 g solvent per 100 g substrate.

In some variations, recombinant organism strains are screened for displaying an increased rate of formation of a particular solvent, relative to the organism strain prior to its transformation. Where the rate of formation of solvent in the organism strain prior to its transformation is about 0.2 g/L/hour of substrate recombinant solventogenic organism strains are screened for producing rates of solvent formation of: 0.24, 0.26, 0.28, 0.3, 0.32, 0.34, 0.36, 0.38, 0.4, 0.44, 0.48, 0.52, 0.56, 0.6, 0.64, 0.68, 0.72, 0.76, 0.8, 0.9, 1, 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2, 3, 4, 8, or 12 g/L/hr.

In some variations, a recombinant organism is screened for an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased at least 0.05%, 0.1%, 0.15%, 0.2%, 0.25%, 0.3%, 0.35%, 0.4%, 0.45%, 0.5%, 0.55%, 0.6%, 0.65%, 0.7%, 0.75%, 0.8%, 0.9%, 0.95%, 1%, 1.1%, 1.2%, 1.3%, 1.4%, 1.5%, 1.6%, 1.7%, 1.8%, 1.9%, 2%, 2.1%, 2.2%, 2.3%, 2.4%, 2.5%, 2.75%, 3%, 3.25%, 3.5%, 3.75%, 4%, 4.25%, 4.5%, or 5%. In some variations, a recombinant organism is screened for an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased at least 5%, 0.10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 90%, 95%, 100%, 125%, 150%, 175%, 200%, 225%, 250%, 275%, 300%, 325%, 350%, 375%, 400%, 425%, 450%, 475%, 500%, 600%, 700%, 800%, 900%, or 1000%. In some variations, a recombinant organism is screened for an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased at least 25%. In some variations, a recombinant organism is screened for an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased at least 50%. In some variations, a recombinant organism is screened for an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased at least 75%. In some variations, a recombinant organism is screened for an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased at least 100%. In some variations, a recombinant organism is screened for an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased at least 200%. In some variations, a recombinant organism is screened for an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased between 0.05-500%. In some variations, a recombinant organism is screened for an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased between 0.05-300%. In some variations, a recombinant organism is screened for an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased between 0.5-500%. In some variations, a recombinant organism is screened for an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased between 5-500%. In some variations, a recombinant organism is screened for an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased between 100-500%. In some variations, a recombinant organism is screened for an increased level of a particular solvent

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relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased between 10-100%. In some variations, a recombinant organism is screened for an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased between 500-1000%. In some variations, the solvent is butanol.

In some variations the solventogenic potential of an organism is evaluated by screening for the presence, absence, or amount of a particular DNA sequence, mRNA sequence, protein, reaction product or solvent. By way of nonlimiting example, the presence, absence, or amount of a particular DNA sequence, mRNA sequence, protein, reaction product or solvent related to reactions or reaction pathways used in the generation of solvents may be evaluated. By way of nonlimiting example, the presence, absence, or amount of a particular DNA sequence, mRNA sequence, protein, reaction product or solvent related to reactions or reaction pathways used in the tolerance to solvents may be evaluated. In some variations, the presence, absence, or amount of a particular DNA sequence, mRNA sequence, protein, reaction product or solvent related to sugar transporters relevant to the production of solvents is evaluated. In some variations, the presence, absence, or amount of a particular DNA sequence, mRNA sequence, protein, reaction product or solvent related to sporulation activities may be evaluated. In some variations, the presence, absence, or amount of a particular DNA sequence, mRNA sequence, protein, reaction product or solvent related to chemotaxis may be evaluated.

In some variations the solventogenic potential of an organism is evaluated by screening for the presence, absence, or amount of a combination of particular DNA sequences, mRNA sequences, proteins, products or solvents.

In some variations, the solventogenic potential of an organism is evaluated by transiently or stably transforming the organism with one or more genes related to production of a solvent, and screening for a particular product or solvent. In some variations, the solventogenic potential of an organism is evaluated by transiently or stably transforming the organism with one or more of the genes described herein, including but not limited to the genes described in the methods of processing cellulose to sugars, methods of generating solvents from sugars, and methods of optimizing organisms for use in industrial applications sections.

Methods of Modifying Organisms

In some variations, the organisms for use in the compositions and methods described herein are modified in order to improve their ability to produce a solvent, including but not limited to butanol, ethanol, or acetone. In some variations, the organisms for use in the compositions and methods described herein are genetically-modified in order to improve their ability to produce a solvent. In some variations, genetic material is introduced into the organisms for use in the compositions and methods described herein in order to improve their ability to produce a solvent.

Described herein are recombinant solventogenic organisms. In some variations the recombinant solventogenic organisms described herein have increased or decreased expression of a gene product relative to the organism strain prior to its transformation. An "organism strain prior to its transformation," as used herein refers to the starting organism strain that was transformed, which transformation yielded the recombinant organism.

For the purposes of this invention, the term "transformation" is used broadly encompass all methods for introducing a particular nucleic acid sequence into an organism. Thus, the

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term "transformation" indicates the genetic alteration of a cell resulting from the uptake and expression of foreign genetic material (DNA). Methods for uptake of foreign DNA include transduction, a process in which bacterial DNA is moved from one bacterium to another by a bacteriophage and bacterial conjugation wherein a living bacterial cell transfers genetic material through cell-to-cell contact.

The term "transformation" also indicates the genetic alteration of a cell resulting from the uptake and expression of a specific genetic sequence (altered or heterologous nucleic acid sequence) without uptake of a foreign genetic material. The latter would include, but is not limited to, sequence alterations induced by site-directed mutagenesis or genetic recombination.

Information about site-directed mutagenesis is found in the following publications and references cited within: Ling et al., *Approaches to DNA mutagenesis: an overview*, Anal Biochem. 254(2): 157-178 (1997); Dale et al., *Oligonucleotide-directed random mutagenesis using the phosphorothioate method*, Methods Mol. Biol. 57: 369-374 (1996); Smith, *In vitro mutagenesis*, Ann. Rev. Genet. 19: 423-462 (1985); Botstein & Shortle, *Strategies and applications of in vitro mutagenesis*, Science 229: 1193-1201 (1985); Kunkel et al., *Rapid and efficient site-specific mutagenesis without phenotypic selection*, Methods in Enzymol. 154, 367-382 (1987); Zoller & Smith, *Oligonucleotide-directed mutagenesis: a simple method using two oligonucleotide primers and a single-stranded DNA template*, Methods in Enzymol. 154: 329-350 (1987).

A solventogenic organism, as used herein, is an organism capable of producing one or more solvents, including but not limited to butanol, ethanol, isopropanol or acetone.

A "recombinant organism," as used herein, is a non-naturally occurring organism with an introduced nucleic acid sequence. The introduced nucleic acid sequence may be integrated into the organism's chromosome, or separate from the organism's chromosome. As nonlimiting examples, the introduced nucleic acid may be a plasmid, a vector, a virus, a viral particle, a bacteriophage, an artificial chromosome, a mini-chromosome, or a linear strand of single stranded or double stranded nucleic acid. A nucleic acid sequence may also be introduced by site directed mutagenesis or genetic recombination.

In some variations the introduced nucleic acid is a heterologous nucleic acid. A "heterologous nucleic acid," as used herein, refers to a sequence of nucleic acids derived from an organism strain different from the organism strain into which the nucleic acid is introduced.

There are many known methods of transiently or stably introducing nucleic acid into organisms. There are well-established strategies for nucleic acid transformation of bacteria in the literature, including those described in Mercenier and Chassy, *Strategies for the development of bacterial transformation systems*, Biochimie 70, 503-517 (1988), Trevors et al., *Electrotransformation of Bacteria by Plasmid DNA*, in Guide to Electroporation and Electrofusion, Ed. Chang, Chassy, Saunders and Sowers, Academic Press (1992), and Dower et al., *Protocols for the Transformation of Bacteria by Electroporation*, Ed. Chang, Chassy, Saunders and Sowers, Academic Press (1992), each of which is incorporated herein by reference in its entirety for all purposes.

There are well-established transformation systems for *Clostridium* sp. in the literature, including Blaschek and White, *Genetic systems development in the clostridia*, FEMS Microbiology Reviews 17, 349-356 (1995); Chen et al., *Factors involved in the transformation of previously non-transformable Clostridium perfringens type B*, FEMS Microbiol

Lett. 140(2-3):185-91 (1996); Phillips-Jones, *Introduction of Recombinant DNA into Clostridium spp.*, in Electroporation Protocols for Microorganisms, Ed. Jac Nickoloff, Humana Press (1995); Young et al., *Genetic Methods in Clostridia*, in Methods in Microbiology, Vol. 29, Ed. Margaret Smith and R. Elizabeth Sockett, Academic Press (1999); and Rood, *Genetic Analysis in Clostridium perfringens*, in *The Clostridia: Molecular Biology and Pathogenesis*, Ed. Rood, McClane, Songer and Titball, Academic Press (1997), each of which is incorporated herein by reference in its entirety for all purposes.

Nucleic acid molecules may be introduced into the yeast cells by standard yeast transformation methods such as Lithium acetate/single-stranded carrier DNA/polyethylene glycol method; Frozen Yeast Protocol using frozen yeast cells that are competent for transformation after thawing; Gene Gun Transformation using gold or tungsten nanoparticles coated with DNA that can be shot into cells; and Protoplast Transformation. See e.g., Sambrook et al., *Molecular Cloning: A Laboratory Manual*, 3d Ed., Cold Spring Harbor Press, Plainsview, N.Y. (2000). The transforming DNA may or may not be integrated into the genome of the yeast cell. Upon the co-transformation of a linearized vector and a nucleic acid molecule into a yeast cell, the nucleic acid molecule is inserted into the insertion site via gap repair, an endogenous homologous recombination system in *S. cerevisiae*.

By way of nonlimiting example, to construct a solvent-producing clostridia, including but not limited to a *C. beijerinckii*, *C. beijerinckii* NCIMB 8052, or *C. beijerinckii* BA101 strain, one or more genes related to solvent production may be expressed or overexpressed. Such genes may be isolated from another organism, including but not limited to different clostridia or butanol-producing clostridia. Those of skill in the art are familiar with the tools for genetic manipulation of clostridia, including but not limited to appropriate source DNA, promoters, enhancers, terminators, integration vectors, autonomously replicating vectors, transformation systems, enhanced- or site-specific recombination systems, transposons, mobile intron systems and culture media.

In some variations, an organism described herein is transformed with one or more genes expressible in the organism.

In some variations, an organism described herein is transformed with a gene from a cellulolytic organism. In some variations, an organism described herein is transformed with a gene from a non-cellulolytic organism.

In some variations, an organism described herein is transformed with a gene from a *Clostridium* strain. In some variations, an organism described herein is transformed with a gene from *Clostridium beijerinckii*.

In some variations, an organism described herein is transformed with one or more genes which have been altered so as to be better expressed in the organism. In some variations, an organism described herein is transformed with one or more genes which have been codon optimized for use in the organism. In some variations, an organism described herein is transformed with one or more genes which have been altered via site-directed mutagenesis to improve production of a particular solvent in the organism.

In some variations, an organism described herein is modified by random mutagenesis to improve production of a particular solvent in the organism.

In some variations, an organism described herein is transformed with one or more genes under the control of an inducible promoter. In some variations, an organism described herein is transformed with one or more genes under the control of a constitutive promoter.

In some variations, one or more of genes of interest is amplified via PCR from a solventogenic organism such as a *clostridium* or, more specifically, *C. beijerinckii* or *C. beijerinckii* BA101. In some variations a promoter active in clostridia is used. In some variations a terminator active in clostridia is used. In some variations an integration vector which allows insertion of genes into clostridia is used. In some variations a self-replicating or suicide vector which allows expression of heterologous genes in clostridia is used. 10 (Flavia Ramirez; M S Thesis; University of Illinois—Urbana Champaign). In some variations, potential transformants bearing the target gene will be identified via one or more selectable or detectable markers. In some variations, potential transformants are analyzed by Southern blot hybridization, 15 PCR, and/or activity assay. The engineered *Clostridia* strain may further be evaluated for solvent production, including but not limited to butanol, ethanol or acetone production.

In some variations, a yeast strain is used in a process to produce one or more solvents. Described herein are yeast 20 strains wherein metabolic engineering and/or functional genomics have been utilized to optimize the yeast strain's solventogenic potential. Compared to a native butanol-producing host, such as Clostridia, the yeast *Saccharomyces cerevisiae* has several advantages. For example, *S. cerevisiae* 25 is robust, displays a different tolerance to concentrations of product and inhibitors present in lignocellulosic hydrolysates, and is viable at a somewhat different pH range. In addition, yeast has a short doubling time, its genetics and physiology is well-studied, and many genetic engineering tools are available. 30

There are well-established strategies for transformation of yeast in the literature, including those described in Becker and Guarente, *Protocol for High-Efficiency Yeast Transformation*, in Guide to Electroporation and Electrofusion, Ed. 35 Chang, Chassy, Saunders and Sowers, Academic Press (1992), which is incorporated herein by reference in its entirety for all purposes.

By way of nonlimiting example, to construct a solvent-producing yeast, including but not limited to a *S. cerevisiae* 40 strain, one or more genes related to solvent production may be expressed or overexpressed. Such genes may be isolated from another organism, including but not limited to the native butanol producer clostridia. Those of skill in the art are familiar with the tools for genetic manipulation of yeast, including 45 but not limited to appropriate source DNA, promoters, enhancers, terminators, integration vectors, transformation systems, and culture media.

The present invention relates to methods of obtaining the disclosed nucleic acid molecules and proteins and of using 50 the disclosed nucleic acid molecules, proteins, fragments of proteins for gene identification and analysis, preparation of constructs, transformation of cells.

The term "an isolated nucleic acid" refers to a nucleic acid 55 that is no longer accompanied by some of materials with which it is associated in its natural state or to a nucleic acid the structure of which is not identical to that of any of naturally occurring nucleic acid. Examples of an isolated nucleic acid include: DNA which has the sequence of part of a naturally occurring genomic DNA molecules, but are not flanked by two coding sequences that flank that part of the molecule in the genome of the organism in which it naturally occurs; a nucleic acid incorporated into a vector or into the genomic DNA of a prokaryote or eukaryote in a manner such that the resulting molecule is not identical to any naturally occurring 60 vector or genomic DNA; a separate molecule such as a DNA, a genomic fragment, a fragment produced by polymerase chain reaction (PCR), or a restriction fragment; recombinant 65

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DNAs; and synthetic DNAs. An isolated nucleic acid may also be comprised of one or more segments of DNA, genomic DNA or synthetic DNA.

In some variations, one or more of genes of interest is amplified via PCR from a solventogenic organism such as a *clostridium* or, more specifically, *C. beijerinckii* or *C. beijerinckii* BA101. In some variations a promoter active in yeast, such as PyK or PGK, is used. In some variations a terminator active in yeast, such as CYC1 terminator, is used. In some variations a yeast delta integration vector which allows sequential insertion of multiple cloned genes into the yeast dispersed chromosomal sites is used. In some variations, potential transformants bearing the target gene will be identified via one or more selectable or detectable markers. In some variations, potential transformants are analyzed by Southern blot hybridization, PCR, and/or activity assay. The engineered yeast or *S. cerevisiae* strain may further be evaluated for solvent production, including but not limited to butanol, ethanol or acetone production. In some variations the engineered yeast or *S. cerevisiae* strain is evaluated for butanol production.

In some variations, an organism described herein is optimized to decrease production of one or more gene products which compete with or are otherwise detrimental to the production of solvents. In some variations an organism described herein is transformed with a nucleic acid to decrease or impair expression of one or more gene products which compete with or are otherwise detrimental to the production of solvents.

In some variations, siRNA, DNAzymes, antisense, promoter inactivation, repressors, or other methods known by those of skill in the art are used to decrease production of one or more gene products which compete with or are otherwise detrimental to the production of solvents.

In some variations, a recombinant organism described herein has an altered level of a particular solvent. In some variations, a recombinant organism has an altered level of a particular solvent, relative to the organism strain prior to its transformation.

In some variations, a recombinant organism comprises a decreased level of a particular solvent, relative to the organism strain prior to its transformation. In some variations, a recombinant organism comprises a decrease in level of a particular solvent, relative to the organism strain prior to its transformation, wherein the decrease in level of the particular solvent is used by a pathway that limits the ability of the recombinant organism to produce a preferred solvent. In some variations, the solvent which has been decreased is ethanol. In some variations, the solvent which has been decreased is acetone. In some variations, the solvent which has been decreased is butanol. In some variations the amount of decrease is 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 90%, 95%, 100%, relative to the organism strain prior to its transformation.

In some variations, an organism comprises an altered level of a particular mRNA species. In some variations, a recombinant organism comprises an altered level of a particular mRNA species, relative to the organism strain prior to its transformation.

In some variations, a recombinant organism comprises a decreased level of a particular mRNA species, relative to the organism strain prior to its transformation. In some variations, a recombinant organism comprises a decrease in level of a particular mRNA species, relative to the organism strain prior to its transformation, wherein the decreased mRNA species is used by a pathway that limits the ability of the recombinant organism to produce a preferred solvent. In

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some variations the amount of decrease of the mRNA species is 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 90%, 95%, 100%, relative to the organism strain prior to its transformation.

5 In some variations, an organism comprises an altered amount of a particular protein species. In some variations, a recombinant organism comprises an altered amount of a particular protein species, relative to the organism strain prior to its transformation.

10 In some variations, a recombinant organism comprises a decreased level of a particular protein species, relative to the organism strain prior to its transformation. In some variations, a recombinant organism comprises a decrease in level of a particular protein species, relative to the organism strain prior to its transformation, wherein the decreased protein species is used by a pathway that limits the ability of the recombinant organism to produce a preferred solvent. In some variations the amount of decrease of the protein species is 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 90%, 95%, 100%, relative to the organism strain prior to its transformation.

15 In some variations, an organism comprises an altered level of a particular activated protein species. In some variations, a recombinant organism comprises an altered level of a particular activated protein species, relative to the organism strain prior to its transformation.

20 In some variations, a recombinant organism comprises a decreased level of a particular activated protein species, relative to the organism strain prior to its transformation. In some variations, a recombinant organism comprises a decrease in level of a particular activated protein species, relative to the organism strain prior to its transformation, wherein the decreased activated protein species is used by a pathway that limits the ability of the recombinant organism to produce a preferred solvent. In some variations the amount of decrease of the activated protein species is 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 90%, 95%, 100%, relative to the organism strain prior to its transformation.

25 In one aspect of the invention, a recombinant organism species produces a particular solvent with increased efficiency. Increased efficiency of solvent production can be determined in any number of ways including but not limited to: concentration (weight/volume) of solvent in fermentation medium, yield (weight/weight) of solvent per amount of substrate, and rate of solvent formation (weight/volume/time).

30 In one aspect of the invention, a recombinant organism strain is screened for an increased level of a particular solvent, relative to the organism strain prior to its transformation. In some variations, a recombinant organism according to the present invention shows an increased amount of a particular solvent relative to the organism strain prior to its transformation, wherein the amount of the particular solvent is increased

35 at least 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2.0, 2.2, 2.4, 2.6, 2.8, 3.0, 3.2, 3.4, 3.6, 3.8, 4.0, 4.5, 5.0, 5.5, 6.0, 6.5, 7.0, 7.5, 8.0, 8.5, 9.0, 9.5, 10, 15, 20, 40, 60, 80, or 100-fold over that in the organism strain prior to its transformation. Where the concentration of the solvent in the organism strain prior to its transformation is 10 g/L, the concentration of the solvent in the recombinant solventogenic organism strain of the present invention is 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 45, 50, 55, 60, 65, 70, 75, 80,

40 85, 90, 95, or 100 g/L. In some embodiments of the invention, the solvent concentration in a culture of the recombinant solventogenic organism strain is about 10, 20, 30, 40, 50, or 60 g/L.

In some variations, a recombinant organism strain according to the present invention produces an increased yield of a particular solvent per amount of the substrate, relative to the organism strain prior to its transformation. Where the yield of solvent in the organism strain prior to its transformation is about 20 g/100 g of substrate, a recombinant solventogenic organism strain of the present invention produces yields of: 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 44, 48, 50, 52, 56, 60, 64, 68, 72, 76, or 80 g solvent per 100 g substrate. In some embodiments of the invention, the yield from a culture of the recombinant solventogenic organism strain is about 24, 30, 40, 50, or 60 g/100 g of substrate.

In some variations, a recombinant organism strain according to the present invention displays an increased rate of formation of a particular solvent, relative to the organism strain prior to its transformation. Where the rate of formation of solvent in the organism strain prior to its transformation is about 0.2 g/L/hour of substrate a recombinant solventogenic organism strain of the present invention produces rates of solvent formation of: 0.24, 0.26, 0.28, 0.3, 0.32, 0.34, 0.36, 0.38, 0.4, 0.44, 0.48, 0.52, 0.56, 0.6, 0.64, 0.68, 0.72, 0.76, 0.8, 0.9, 1, 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9, 2, 3, 4, 8, or 12 g/L/hr. In some embodiments of the invention, the rate of solvent formation from a culture of the recombinant solventogenic organism strain is about 0.24, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9, 1, 1.2, 1.4, 1.6, 1.8, 2.0, 2.2, 2.4, 2.6, 2.8, or 3.0 g/L/hour.

In some variations, a recombinant organism comprises an increased level of a particular solvent, relative to the organism strain prior to its transformation. In some variations, a recombinant organism comprises an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased at least 0.05%, 0.1%, 0.15%, 0.2%, 0.25%, 0.3%, 0.35%, 0.4%, 0.45%, 0.5%, 0.55%, 0.6%, 0.65%, 0.7%, 0.75%, 0.8%, 0.9%, 0.95%, 1%, 1.1%, 1.2%, 1.3%, 1.4%, 1.5%, 1.6%, 1.7%, 1.8%, 1.9%, 2%, 2.1%, 2.2%, 2.3%, 2.4%, 2.5%, 2.75%, 3%, 3.25%, 3.5%, 3.75%, 4%, 4.25%, 4.5%, or 5%. In some variations, a recombinant organism comprises an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased at least 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 90%, 95%, 100%, 125%, 150%, 175%, 200%, 225%, 250%, 275%, 300%, 325%, 350%, 375%, 400%, 425%, 450%, 475%, 500%, 600%, 700%, 800%, 900%, or 1000%. In some variations, a recombinant organism comprises an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased at least 25%. In some variations, a recombinant organism comprises an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased at least 50%. In some variations, a recombinant organism comprises an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased at least 75%. In some variations, a recombinant organism comprises an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased at least 100%. In some variations, a recombinant organism comprises an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased at least 200%. In some variations, a recombinant organism comprises an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased at least 200%.

transformation, wherein the level of the particular solvent is increased between 0.05-500%. In some variations, a recombinant organism comprises an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased between 0.05-300%. In some variations, a recombinant organism comprises an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased between 0.5-500%. In some variations, a recombinant organism comprises an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased between 5-500%. In some variations, a recombinant organism comprises an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased between 100-500%. In some variations, a recombinant organism comprises an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased between 10-100%. In some variations, a recombinant organism comprises an increased level of a particular solvent relative to the organism strain prior to its transformation, wherein the level of the particular solvent is increased between 500-1000%. In some variations, the solvent is butanol. In some variations, the solvent is ethanol. In some variations, the solvent is acetone.

In some variations, an organism comprises an increased level of a particular mRNA species. In some variations, a recombinant organism comprises an increased level of a particular mRNA species, relative to the organism strain prior to its transformation. In some variations, a recombinant organism comprises an increased level of a particular mRNA species relative to the organism strain prior to its transformation, wherein the level of the particular mRNA species is increased at least 1.5-fold, 2-fold, 4-fold, 10-fold, 25-fold, 50-fold, or 100-fold relative to the organism strain prior to its transformation. In some variations, a recombinant organism comprises an increased level of a particular mRNA species relative to the organism strain prior to its transformation, wherein the level of the particular mRNA species is increased at least 2-fold. In some variations, a recombinant organism comprises an increased level of a particular mRNA species relative to the organism strain prior to its transformation, wherein the level of the particular mRNA species is increased at least 5-fold. In some variations, a recombinant organism comprises an increased level of a particular mRNA species relative to the organism strain prior to its transformation, wherein the level of the particular mRNA species is increased at least 10-fold. In some variations, a recombinant organism comprises an increased level of a particular mRNA species relative to the organism strain prior to its transformation, wherein the level of the particular mRNA species is increased at least 15-fold. In some variations, a recombinant organism comprises an increased level of a particular mRNA species relative to the organism strain prior to its transformation, wherein the level of the particular mRNA species is increased at least 20-fold. In some variations, a recombinant organism comprises an increased level of a particular mRNA species relative to the organism strain prior to its transformation, wherein the level of the particular mRNA species is increased at least 40-fold. In some variations, a recombinant organism comprises an increased level of a particular mRNA species relative to the organism strain prior to its transformation, wherein the level of the particular mRNA species is increased at least 60-fold.

In some variations, an organism comprises an increased level of a particular protein species. In some variations, a

recombinant organism comprises an increased level of a particular protein species, relative to the organism strain prior to its transformation. In some variations, a recombinant organism comprises an increased level of a particular protein species relative to the organism strain prior to its transformation, wherein the level of the particular protein species is increased at least 1.5-fold, 2-fold, 5-fold, 10-fold, 20-fold, 30-fold, 40-fold, 50-fold, 60-fold, 80-fold, or 100-fold relative to the organism strain prior to its transformation. In some variations, a recombinant organism comprises an increased level of a particular protein species relative to the organism strain prior to its transformation, wherein the level of the particular protein species is increased at least 1.5-fold. In some variations, a recombinant organism comprises an increased level of a particular protein species relative to the organism strain prior to its transformation, wherein the level of the particular protein species is increased at least 5-fold. In some variations, a recombinant organism comprises an increased level of a particular protein species relative to the organism strain prior to its transformation, wherein the level of the particular protein species is increased at least 10-fold. In some variations, a recombinant organism comprises an increased level of a particular protein species relative to the organism strain prior to its transformation, wherein the level of the particular protein species is increased at least 20-fold. In some variations, a recombinant organism comprises an increased level of a particular protein species relative to the organism strain prior to its transformation, wherein the level of the particular protein species is increased at least 40-fold. In some variations, a recombinant organism comprises an increased level of a particular protein species relative to the organism strain prior to its transformation, wherein the level of the particular protein species is increased at least 60-fold. In some variations, a recombinant organism comprises an increased level of a particular protein species relative to the organism strain prior to its transformation, wherein the level of the particular protein species is increased at least 80-fold. In some variations, a recombinant organism comprises an increased level of a particular protein species relative to the organism strain prior to its transformation, wherein the level of the particular protein species is increased at least 100-fold.

In some variations, the amount of a particular protein species in the organism strain prior to its transformation is 0.10 percent of the total protein in a cell. The amount of the particular protein species in the recombinant solventogenic organism strain is about 0.2, 0.5, 1.0, 2.0, 4.0, 6.0, 8.0, 10.0 percent of the total protein in the cell.

In some variations, an organism comprises an increased level of a particular activated protein species. In some variations, a recombinant organism comprises an increased level of a particular activated protein species, relative to the organism strain prior to its transformation. In some variations, a recombinant organism comprises an increased level of a particular activated protein species relative to the organism strain prior to its transformation, wherein the level of the particular activated protein species is increased at least 1.5-fold, 2-fold, 5-fold, 10-fold, 20-fold, 30-fold, 40-fold, 50-fold, 60-fold, 80-fold, or 100-fold relative to the organism strain prior to its transformation. In some variations, a recombinant organism comprises an increased level of a particular activated protein species relative to the organism strain prior to its transformation, wherein the level of the particular activated protein species is increased at least 1.5-fold. In some variations, a recombinant organism comprises an increased level of a particular activated protein species relative to the organism strain prior to its transformation, wherein the level of the particular activated protein species is increased at least 5-fold. In some

variations, a recombinant organism comprises an increased level of a particular activated protein species relative to the organism strain prior to its transformation, wherein the level of the particular activated protein species is increased at least 10-fold. In some variations, a recombinant organism comprises an increased level of a particular activated protein species relative to the organism strain prior to its transformation, wherein the level of the particular activated protein species is increased at least 20-fold. In some variations, a recombinant organism comprises an increased level of a particular activated protein species relative to the organism strain prior to its transformation, wherein the level of the particular activated protein species is increased at least 40-fold. In some variations, a recombinant organism comprises an increased level of a particular activated protein species relative to the organism strain prior to its transformation, wherein the level of the particular activated protein species is increased at least 60-fold. In some variations, a recombinant organism comprises an increased level of a particular activated protein species relative to the organism strain prior to its transformation, wherein the level of the particular activated protein species is increased at least 80-fold. In some variations, a recombinant organism comprises an increased level of a particular activated protein species relative to the organism strain prior to its transformation, wherein the level of the particular activated protein species is increased at least 100-fold.

In some variations, the amount of a particular activated protein species in the organism strain prior to its transformation is 0.10 percent of the total protein in a cell. The amount of the particular activated protein species in the recombinant solventogenic organism strain is about 0.2, 0.5, 1.0, 2.0, 4.0, 6.0, 8.0, 10.0 percent of the total protein in the cell.

Modifying Clostridia for Increasing Efficiency of Butanol Production.

U.S. Pat. No. 6,358,717 discloses a method of producing high levels of butanol using a fermentation process that employs a mutant strain of *Clostridium beijerinckii*. *Clostridium beijerinckii* BA101 (ATCC No. PTA-1550) is a hyper-butanol producing strain formed by mutagenesis of the wild type *Clostridium beijerinckii* NCIMB 8052. (Annous, B. A., and H. P. Blaschek. 1991. Isolation and characterization of *Clostridium acetobutylicum* mutants with enhanced amylolytic activity. *Appl. Environ. Microbiol.* 57: 2544-2548; Formanek, J., R. Mackie, and H. P. Blaschek. 1997. Enhanced butanol production by *Clostridium beijerinckii* BA101 grown in semidefined P2 medium containing 6 percent maltodextrin or glucose. *Appl. Environ. Microbiol.* 63: 2306-2310.)

In one aspect of the invention, gene expression profiles of *C. beijerinckii* BA101 and the wild type *C. beijerinckii* NCIMB 8052 are compared. Profiles of expression of solventogenic genes are compared between the hyper-butanol producing *C. beijerinckii* BA101 and the wild type *C. beijerinckii* NCIMB 8052. Typically, gene expression profiles are compared using standard microarray techniques.

Microarrays comprising nucleic acid probes comprising the sequence of one or more genes of *C. beijerinckii* BA101 or the wild type *C. beijerinckii* NCIMB 8052 are arrayed on a surface of the microarray. The genome of the wild type *Clostridium beijerinckii* 8052 is about 6.0 Mbp and the sequence is available at www.jgi.doe.gov (GenBank accession number CP000721); thus probes corresponding to genes of the wild type *C. beijerinckii* NCIMB 8052 are readily obtained. Methods for fabricating and using microarrays is found in U.S. Pat. No. 5,807,522, which is herein incorporated by reference. Instructions for constructing microarray hardware (e.g., arrayers and scanners) using commercially available parts can be found at <http://cmgm.stanford.edu/pbr/>

own/and in Cheung et al., 1999, Nat. Genet. Supplement 21: 15-19, which are herein incorporated by reference. Additional discussions of microarray technology and protocols for preparing samples and performing microarray experiments are found in M. Schena (ed.), DNA Microarrays: A Practical Approach, Oxford University Press, Oxford, UK, 1999. Descriptions of how to use an arrayer and the associated software are found at <http://cmgm.stanford.edu/pbrown/mguide/a-rayerHTML/ArrayerDocs.html>, which is herein incorporated by reference.

In a typical microarray experiment, a microarray is hybridized with differentially labeled RNA, DNA, or DNA populations derived from two different samples. Most commonly RNA is isolated from cells or tissues of interest and is reverse transcribed to yield DNA. Labeling is usually performed during reverse transcription by incorporating a labeled nucleotide in the reaction mixture. Although various labels can be used, most commonly the nucleotide is conjugated with the fluorescent dyes Cy3 or Cy5. For example, Cy5-dUTP and Cy3-dUTP can be used. DNA derived from one sample (representing, for example, a particular cell type or growth condition) is labeled with one fluorophore while DNA derived from a second sample (representing, for example, a different or mutant cell type, or growth condition) is labeled with the second fluorophore. Similar amounts of labeled material from the two samples are cohybridized to the microarray. In the case of a microarray experiment in which the samples are labeled with Cy5 (which fluoresces red) and Cy3 (which fluoresces green), the primary data (obtained by scanning the microarray using a detector capable of quantitatively detecting fluorescence intensity) are ratios of fluorescence intensity (red/green, R/G). These ratios represent the relative concentrations of DNA molecules that hybridized to the DNA probes represented on the microarray and thus reflect the relative expression levels of the mRNA corresponding to each DNA probe/gene represented on the microarray.

Differential expression of genes, especially solventogenic genes, are compared between *C. beijerinckii* BA101 and the wild type *C. beijerinckii* NCIMB 8052. In some embodiments, expression profiles are correlated with solvent production and butanol production phases, respectively of *C. beijerinckii* BA101, *C. beijerinckii* NCIMB 8052 or both. Sets of genes that are differentially expressed between the wild type and hyper-butanol mutant are identified. Genes in the hyper-butanol mutant *C. beijerinckii* BA101 show increased or decreased expression relative to genes of the wild type *C. beijerinckii* NCIMB 8052. In one aspect these genes are involved in one or more solvent production-related pathways such as solventogenesis, chemotaxis, motility, sporulation and sugar transport.

In one aspect of the invention, one or more of these genes are identified and their expression profiles corresponding to a hyper-butanol producing state is replicated in a *Clostridium*, preferably in a *Clostridium beijerinckii*. This can be accomplished in a number of ways including, but not limited to, transforming a microorganism such as clostridia with the gene under the control of a constitutive or inducible promoter. The promoter is designed to replicate the increased or decreased gene expression (relative to wild type) observed in the hyper-butanol producing mutant. In one aspect the organism transformed with a wild type gene from *Clostridium beijerinckii* NCIMB 8052, whose genetic (DNA) sequence is publicly available.

In one aspect of the invention, the sequences of *Clostridium beijerinckii* NCIMB 8052 and hyper-butanol producing *Clostridium beijerinckii* BA101 are compared. *Clostridium beijerinckii* BA101 is publicly available (ATCC

No. PTA-1550) and may be sequenced using methods known to those of skill in the art. In some variations, a recombinant organism is transformed with one or more genes from *Clostridium beijerinckii* BA101 that has a sequence different from the corresponding gene in *Clostridium beijerinckii* NCIMB 8052. Where the expression of the gene is altered in BA101 relative to the wild-type, a suitable promoter is operably linked to the gene sequence prior to transformation. The promoter is able to be used to replicate the gene expression profile in BA101 in the recombinant organism.

In one aspect of the invention, the genes related to the solvent productions pathways identified by this analysis include homologous genes with at least 70, 75, 80, 83, 85, 90, 95, 97, 99 or 100% homology with the known sequence of a gene in wild type *C. beijerinckii* NCIMB 8052.

In another embodiment of the invention, homologous polynucleotides are identified by the ability to hybridize under moderate to high stringency conditions to a polynucleotide sequence provided herein, or a fragment thereof, or a complementary sequence thereof. Hybridization techniques are well known in the art of molecular biology. High stringency conditions are known in the art. See, for example, Maniatis et al., Molecular Cloning: A Laboratory Manual, 2d Edition, 1989, and Short Protocols in Molecular Biology, ed. Ausubel, et al. Stringent conditions are sequence-dependent and will be different in different circumstances. Longer sequences hybridize specifically at higher temperatures. An extensive guide to the hybridization of nucleic acids is found in Tijssen, Techniques in Biochemistry and Molecular Biology—Hybridization with Nucleic Acid Probes, “Overview of principles of hybridization and the strategy of nucleic acid assays” (1993). Generally, stringent conditions are selected to be about 5-10° C. lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength pH. The T_m is the temperature (under defined ionic strength, pH and nucleic acid concentration) at which 50% of the probes complementary to the target hybridize to the target sequence at equilibrium (as the target sequences are present in excess, at T_m , 50% of the probes are occupied at equilibrium). Stringent conditions will be those in which the salt concentration is less than about 1.0 M sodium ion, typically about 0.01 to 1.0 M sodium ion concentration (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30° C. for short probes (e.g. 10 to 50 nucleotides) and at least about 60° C. for longer probes (e.g. greater than 50 nucleotides). In another embodiment, less stringent hybridization conditions are used. For example, moderate or low stringency conditions may be used, as are known in the art. (See Maniatis and Ausubel, supra, and Tijssen, supra). For purposes of illustration, suitable moderately stringent conditions for testing the hybridization of a polynucleotide of this invention with other polynucleotides include prewashing in a solution of 5×SSC (“saline sodium citrate”; 9 mM NaCl, 0.9 mM sodium citrate), 0.5% SDS, 1.0 mM EDTA (pH 8.0); hybridizing at 50-60° C., 5×SSC, overnight; followed by washing twice at 65° C. for 20 minutes with each of 2×, 0.5× and 0.2×SSC containing 0.1% SDS. One skilled in the art will understand that the stringency of hybridization can be readily manipulated, such as by altering the salt content of the hybridization solution and/or the temperature at which the hybridization is performed. For example, in another embodiment, suitable highly stringent hybridization conditions include those described above, with the exception that the temperature of hybridization is increased, e.g., to 60-65° C., or 65-70° C. Stringent conditions may also be achieved with the addition of destabilizing agents such as formamide.

Identification of homologous genes can also be performed by optimal alignment of sequences for comparison to analyze sequence identity (homology) known in the art. Homology in this context means sequence similarity or identity, with identity being preferred. This homology is determined using standard techniques known in the art, including, but not limited to, the local homology algorithm of Smith & Waterman, *Adv. Appl. Math.* 2: 482 (1981), by the homology alignment algorithm of Needleman & Wunsch, *J. Mol. Biol.* 48: 443 (1970), by the search for similarity method of Pearson & Lipman, *PNAS USA* 85: 2444 (1988), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Drive, Madison, Wis.), the Best Fit sequence program described by Devereux et al., *Nucl. Acid Res.* 12: 387-395 (1984), preferably using the default settings, or by inspection. One example of a useful algorithm is PILEUP. PILEUP creates a multiple sequence alignment from a group of related sequences using progressive, pairwise alignments. It can also plot a tree showing the clustering relationships used to create the alignment. PILEUP uses a simplification of the progressive alignment method of Feng & Doolittle, *J. Mol. Evol.* 35: 351-360 (1987); the method is similar to that described by Higgins & Sharp CABIOS 5: 151-153 (1989). Useful PILEUP parameters include a default gap weight of 3.00, a default gap length weight of 0.10, and weighted end gaps. Another example of a useful algorithm is the BLAST (Basic Local Alignment Search Tool) algorithm, described in Altschul et al., *J. Mol. Biol.* 215, 403-410, (1990) and Karlin et al., *PNAS USA* 90: 5873-5787 (1993). A particularly useful BLAST program is the WU-BLAST-2 program which was obtained from Altschul et al., *Methods in Enzymology*, 266: 460-480 (1996); <http://blast.wustl.edu/>. WU-BLAST-2 uses several search parameters, most of which are set to the default values. The adjustable parameters are set with the following values: overlap span=1, overlap fraction=0.125, word threshold (T)=11. The HSP S and HSP S2 parameters are dynamic values and are established by the program itself depending upon the composition of the particular sequence and composition of the particular database against which the sequence of interest is being searched; however, the values may be adjusted to increase sensitivity. A percent amino acid sequence identity value is determined by the number of matching identical residues divided by the total number of residues of the "longer" sequence in the aligned region. The "longer" sequence is the one having the most actual residues in the aligned region (gaps introduced by WU-Blast-2 to maximize the alignment score are ignored). Thus, "percent (%) nucleic acid sequence identity" is defined as the percentage of nucleotide residues in a candidate sequence that are identical with the nucleotide residues of a particular nucleic acid. A preferred method utilizes the BLASTN module of WU-BLAST-2 set to the default parameters, with overlap span and overlap fraction set to 1 and 0.125, respectively.

The nucleic acids of the present invention that are identified by altered expression or nucleotide sequence in the hyper-butanol producing clostridia can be used to isolate nucleic acids encoding homologous proteins from other strains of the same or other species and microorganisms, such as Clostridia, *Escherichia*, *Saccharomyces*, etc. Isolation of homologous genes using sequence-dependent protocols is well known in the art. Examples of sequence-dependent protocols include, but are not limited to, methods of nucleic acid hybridization, and methods of DNA and RNA amplification as exemplified by various uses of nucleic acid amplification technologies (e.g., polymerase chain reaction, ligase chain

reaction). For example, genes encoding homologous proteins, either as DNA's or genomic DNA's, could be isolated directly by using all or a portion of the nucleic acids of the present invention as DNA hybridization probes to screen DNA or genomic libraries from any desired organism employing methodology well known to those skilled in the art. Methods for forming such libraries are well known in the art (Sambrook et al., *Molecular Cloning: A Laboratory Manual*; Cold Spring Harbor Laboratory Press, Cold Spring Harbor, 1989).

Nucleic acids of interest may also be synthesized, either completely or in part, especially where it is desirable to provide host-preferred sequences, by well-known techniques. See, e.g., Carruthers et al. (*Cold Spring Harbor Symp. Quant. Biol.* 47: 411-418, 1982) and Adams et al. (*J. Am. Chem. Soc.* 105: 661, 1983). Thus, all or a portion of the nucleic acids of the present invention may be synthesized using codons preferred by a selected host.

Genes and homologs and variants thereof that are identified as having a role in hyper-production of butanol can be used for transforming host species or organisms for the high efficiency production of butanol. In one aspect, the nucleic acids used for transformation comprise the sequence of the gene as well as an operably linked constitutive or inducible promoter that can be used to regulate expression of the gene. Specific examples of methods for modifying clostridia for increasing efficiency of butanol production are provided infra.

Methods of Preparing Substrates

In addition to conventional starch (maize, wheat, millet, rye, etc.) or sugar (molasses) substrates saccharolytic clostridia are able to utilize many different carbohydrates. (See Jones and Woods, 1986, *supra*.) Solvent production starting materials such as biomass, plant-based, cellulosic, lignocellulosic or hemicellulosic materials may be directly entered into the solvent production process. However, often such materials are pretreated to convert lignocellulosic biomass into a form which is more accessible to cellulolytic and fermentation processes. Pretreatment typically includes one or more of increasing the surface area to volume ratio by, for example comminution; steam treatment, acid hydrolysis, or enzymatic treatment. Those of skill in the art are familiar with these and other pretreatment methods.

Methods of Processing Cellulose to Sugars

Cellulosic and hemicellulosic materials may be converted to downstream products such as fermentable sugars by various methods. In some variations, biomass, lignocellulosic, or cellulosic materials are converted to downstream products such as fermentable sugars via a method which does not require living bacteria, yeast, or other organisms.

In some variations, biomass, lignocellulosic, or cellulosic materials are converted to downstream products such as fermentable sugars via a method which utilizes living bacteria, yeast, or other organisms.

In some variations, any organism capable of processing biomass, lignocellulosic, or cellulosic materials to one or more useful downstream products, including but not limited to fermentable sugars, is used in the methods described herein. In some variations, any organism capable of processing cellulose to one or more useful downstream products, including but not limited to fermentable sugars, is used in the methods described herein.

In some variations, a cellulolytic yeast, bacteria or other organism, including but not limited to *Clostridia*, *Saccharomyces*, or *Escherichia* strains, are naturally or through genetic manipulation made capable of processing biomass, lignocel-

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lulosic, or cellulosic materials to one or more useful downstream products, including but not limited to fermentable sugars.

In some variations, a solventogenic organism is transformed with one or more genes or regulatory sequences controlling expression of a gene relating to the conversion of biomass, lignocellulosic, or cellulosic materials to one or more useful downstream products, including but not limited to fermentable sugars.

In some variations, a solventogenic organism is transformed with one or more heterologous genes or heterologous regulatory sequences controlling a gene relating to the conversion of biomass, lignocellulosic, or cellulosic materials to one or more useful downstream products, including but not limited to fermentable sugars.

In some variations, a solventogenic organism is transformed with one or more genes relating to activation or inactivation of a gene product involved in the conversion of biomass, lignocellulosic, or cellulosic materials to one or more useful downstream products, including but not limited to fermentable sugars.

In some variations, a solventogenic organism is transformed with one or more cellulolytic genes. In some variations, a solventogenic organism is transformed with one or more genes involved in generating a functional cellulosome complex. In some variations, a solventogenic organism is transformed with all of the genes involved in a cellulosome complex.

In some variations, a solventogenic organism is transformed with one or more secretable cellulolytic genes. In some variations, a non-solventogenic organism is transformed with one or more secretable cellulolytic genes. In some variations, a solventogenic organism is transformed with one or more secretable cellulolytic genes. In some variations, a solventogenic organism is transformed with all of the secretable cellulolytic genes necessary to convert biomass, lignocellulosic, or cellulosic materials to one or more useful downstream products, including but not limited to fermentable sugars.

In some variations, a solventogenic organism is transformed with one or more cellulolytic genes. In some variations, a solventogenic organism is transformed with one or more genes involved in generating a functional cellulosome complex. In some variations, a solventogenic organism is transformed with all of the genes involved in a cellulosome complex.

In some variations, a solventogenic organism is transformed with one or more genes encoding one or more enzymes that cut at random at internal amorphous sites in a cellulose polysaccharide chain. In some variations, a solventogenic organism is transformed with one or more genes encoding one or more endoglucanases or 1,4-beta-D-glucan-4-glucanohydrolases.

In some variations, a solventogenic organism is transformed with one or more genes encoding one or more enzymes that process reducing or nonreducing ends of cellulose polysaccharide chains to hexoses such as glucose, or cellobiose. In some variations, a solventogenic organism is transformed with one or more genes encoding one or more exoglucanases. In some variations, a solventogenic organism is transformed with one or more genes encoding one or more 1,4-beta-D-glucan glucanohydrolases, cellobextrinases, 1,4-beta-D-glucan cellobiohydrolases, or cellobiohydrolases.

In some variations, a solventogenic organism is transformed with one or more genes encoding one or more beta-glucosidases or beta-glucoside glucohydrolases.

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In some variations, a solventogenic organism is transformed with one or more genes encoding one or more scaffold-type proteins.

In some variations, a solventogenic organism is transformed with one or more genes or regulatory sequences which decrease or impair the activity of one or more pathways which decrease or impair the solventogenic potential of a solventogenic organism. In some variations, a solventogenic organism is transformed with one or more heterologous genes or heterologous regulatory sequences which decrease or impair the activity of one or more pathways which decrease or impair the solventogenic potential of a solventogenic organism. In some variations, a solventogenic organism is transformed with one or more genes or regulatory sequences which decrease or impair the activity of one or more pathways which decrease or impair the solventogenic potential of a *Clostridium* strain, including but not limited to *C. beijerinckii* or *C. beijerinckii* BA101.

Methods of Generating Solvents from Sugars

20 Cellulosic materials are typically converted into a mixture of hexose sugars, such as glucose and mannose, and pentose sugars, such as xylose and arabinose. These sugars may then be acted upon to generate one or more solvents.

In some variations, the organisms described herein are optimized to ferment one or more hexose or pentose sugars to solvents, for example butanol, ethanol, or acetone. In some variations, the organisms described herein are optimized to ferment all major hexose or pentose sugars to solvents. In some variations, the organisms described herein are optimized to ferment one or more of glucose, mannose, xylose or arabinose. In some variations, the organisms described herein are optimized to ferment glucose. In some variations, the organisms described herein are optimized to ferment mannose. In some variations, the organisms described herein are optimized to ferment xylose. In some variations, the organisms described herein are optimized to ferment arabinose.

In some variations the organism that converts one or more hexose or pentose sugars to a solvent, including but not limited to butanol, is also capable of converting cellulosic material to hexose or pentose sugars, with or without pretreatment.

In some variations the organism that converts one or more hexose or pentose sugars to a solvent, including but not limited to butanol, is not capable of converting cellulosic material to hexose or pentose sugars, with or without pretreatment.

45 In some variations the process utilizing an organism that converts one or more hexose or pentose sugars to a solvent, including but not limited to butanol, includes simultaneous or sequential use of a second organism or strain that is capable of converting cellulosic material to hexose or pentose sugars, with or without pretreatment.

In some variations, the organisms described herein are optimized to ferment one or more hexose or pentose sugars by increasing or facilitating the organism's use of favored pathways. In some variations, the organisms described herein are optimized to ferment one or more of glucose, mannose, xylose or arabinose by increasing or facilitating the organism's use of favored pathways.

55 In some variations, the organisms described herein are optimized to ferment one or more hexose or pentose sugars by decreasing or impairing use of pathways which decrease or impair production of a solvent of interest. In some variations, the organisms described herein are optimized to ferment one or more of glucose, mannose, xylose or arabinose by decreasing or impairing use of pathways which decrease or impair production of a solvent of interest.

In some variations, an organism described herein is transformed with one or more genes involved in the metabolic

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pathway of a particular hexose or pentose sugar. In some variations, an organism described herein is transformed with all genes involved in the metabolic pathway of a particular hexose or pentose sugar. In some variations, an organism described herein is transformed with one or more genes involved in the metabolic pathway of glucose. In some variations, an organism described herein is transformed with all genes involved in the metabolic pathway of glucose. In some variations, an organism described herein is transformed with one or more genes involved in the metabolic pathway of mannose. In some variations, an organism described herein is transformed with all genes involved in the metabolic pathway of mannose. In some variations, an organism described herein is transformed with one or more genes involved in the metabolic pathway of xylose. In some variations, an organism described herein is transformed with all genes involved in the metabolic pathway of xylose. In some variations, an organism described herein is transformed with one or more genes involved in the metabolic pathway of arabinose. In some variations, an organism described herein is transformed with all genes involved in the metabolic pathway of arabinose.

In some variations, an organism described herein is transformed with one or more genes involved in the metabolic pathway of a particular hexose or pentose sugar from a bacteria. In some variations, an organism described herein is transformed with one or more genes involved in the metabolic pathway of a particular hexose or pentose sugar from a *Neurospora* strain, including but not limited to *N. crassa*.

In some variations, the titer, yield and productivity of solvent production is increased by optimizing the various metabolic pathways involved in the biosynthesis of one or more solvents of interest, including but not limited to butanol, ethanol, and acetone. In some variations, the titer, yield and productivity of butanol production is increased by optimizing the various metabolic pathways involved in the biosynthesis of butanol. In some variations, metabolic flux analysis is used to identify the rate-limiting steps in solvent synthesis in an organism described herein, including but not limited to a *Clostridium* or *S. cerevisiae* strain.

By way of nonlimiting example, for a linear pathway, the level of final product is related to the overall flux through the pathway. An optimized solvent biosynthetic pathway should have increased overall flux through the pathway without significant accumulation of pathway intermediates. Various analytical instruments may be used to determine the concentrations of key metabolites in the metabolic pathways involved in the biosynthesis of the solvent at fermentation conditions and identify the rate-limiting enzymes. Non-limiting examples of analytical instruments include GC-MS, HPLC-MS, HPLC (stand alone), Piezorray robotic printer (non-contact microarray printing onto membranes, plates, and slides), UV/visible/fluorescence microplate reader, and chemiluminometer microplate reader. To trace the metabolites, C-14 based isotopic labeling methods in combination with either LC-MS or NMR may be used.

Once the rate-limiting enzymes are identified in an organism described herein, overexpression of the one or more genes limiting the overall flux may be used to determine its effect on the concentrations of pathway intermediates and the final solvent product. If the product concentration is increased, then the overexpressed gene or genes are indeed positively correlated with solvent production. Non-limiting examples of strategies to balance gene expression include manipulation of promoter strength, ribosomal binding site (RBS) strength, gene location in an operon, and mRNA stability.

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The effect of various sporulation, motility, and sugar transport genes may be similarly evaluated. For example, increasing or decreasing the expression of one or more genes relating to sporulation, motility, and sugar transport may be used to determine their effect on the concentrations of pathway intermediates and the final solvent product. If the product concentration is increased, then the gene or genes with increased or decreased expression are correlated with solvent production. Non-limiting examples of strategies to balance gene expression include manipulation of promoter strength, ribosomal binding site (RBS) strength, gene location in an operon, and mRNA stability.

Solventogenic Genes

Acid concentration and reducing state are also known to influence the production of solvents and hence, impact the expression of solvent-related genes in *Clostridium*. Genes involved in solvent production and butanol production are identified in FIG. 9.

As demonstrated in FIG. 4, alcohol dehydrogenase (Adh), butyryl-CoA dehydrogenase (Bcd) and butyrate kinase (Buk) are expressed at altered (higher or lower) levels during the solventogenic stage in BA101 compared with the wild-type *C. beijerinckii* strain.

In some variations, an organism described herein is optimized to increase production of an enzyme in the solventogenic pathway. In some variations, an organism described herein is transformed with a gene encoding an enzyme in the solventogenic pathway. In some variations an organism described herein is transformed with a gene encoding an enzyme in the solventogenic pathway to overexpress the enzyme.

In some variations, an organism described herein is optimized to increase production of all of the enzymes described herein in the butanol solventogenic pathway. In some variations, an organism described herein is transformed with all of the enzymes described herein in the butanol solventogenic pathway. In some variations an organism described herein is transformed with a gene encoding all of the enzymes described herein in the butanol solventogenic pathway to overexpress the enzymes.

Alcohol dehydrogenase (Adh) encodes an important terminal enzyme required for alcohol production. Thus, increased Adh expression may directly contribute to elevated butanol synthesis in BA101. In some variations, an organism described herein is optimized to increase production of Adh. In some variations, an organism described herein is transformed with an Adh gene. In some variations an organism described herein is transformed with an Adh gene to overexpress Adh. In some variations, an organism described herein is transformed with an Adh gene from a microbial organism to overexpress Adh. In some variations, an organism described herein is transformed with an Adh gene from a *Clostridium* sp. to overexpress Adh. In some variations, an organism described herein is transformed with an Adh gene from *Clostridium beijerinckii* to overexpress Adh. In some variations, an organism described herein is transformed with a nucleic acid which results in an increase in expression of the Adh gene whose DNA sequence is shown in FIG. 10. In some variations, an organism described herein is transformed with an Adh gene whose DNA sequence is shown in FIG. 10 to overexpress Adh.

In some variations, an organism described herein is transformed with an Adh gene whose DNA sequence is at least 60-100% identical to the DNA sequence shown in FIG. 10 or complement thereof. In some variations, an organism described herein is transformed with an Adh gene whose DNA sequence is at least 80-100% identical to the DNA

sequence shown in FIG. 10 or complement thereof. In some variations, an organism described herein is transformed with an Adh gene whose DNA sequence is at least 90-100% identical to the DNA sequence shown in FIG. 10 or complement thereof. In some variations, an organism described herein is transformed with an Adh gene whose DNA sequence is at least 60, at least 65, at least 70, at least 75, at least 80, at least 85, at least 90, at least 95, or 100% identical to the DNA sequence shown in FIG. 10 or complement thereof. In some variations, an organism described herein is transformed with an Adh gene whose DNA sequence is at least 80% identical to the DNA sequence shown in FIG. 10 or complement thereof. In some variations, an organism described herein is transformed with an Adh gene whose DNA sequence is at least 85% identical to the DNA sequence shown in FIG. 10 or complement thereof. In some variations, an organism described herein is transformed with an Adh gene whose DNA sequence is at least 90% identical to the DNA sequence shown in FIG. 10 (SEQ ID NO: 1) or complement thereof. The *Clostridium beijerinckii* NCIMB 8052 published genome identifies this adh gene as Cbei_2181.

The *C. beijerinckii* NCIMB 8052 published genome identifies the adh gene shown in FIG. 10 as Cbei_2181 (SEQ ID NO: 1). NCBI BLAST search against the *C. beijerinckii* NCIMB 8052 genome revealed another *C. beijerinckii* NCIMB 8052 gene that is a close homolog of Cbei_2181 at both the DNA sequence and the protein sequence levels. The DNA sequence (SEQ ID NO: 14) is shown in FIG. 22A and predicted amino acid sequence (SEQ ID NO: 15) of Cbei_1722 is shown in FIG. 22B.

At the DNA level the Cbei_1722 adh gene shows 90% identity to Cbei_2181 with 1% gaps in the alignment. At the protein level the Cbei_1722 adh protein shows 93% amino acid identity to Cbei_2181, with 97% similarity and zero gaps. The DNA and protein alignments both show an “Expect value” of zero, suggesting the two enzymes are either functionally equivalent, or nearly so. The Cbei_1722 adh gene is annotated at an “iron-containing alcohol dehydrogenase”. Multiple isozymes of the class of adh enzymes are known to exist in solvent-forming *Clostridium* species and are known to be induced or de-repressed near the onset of solvent formation (Walter K A, Bennett G N, Papoutsakis E T; Molecular characterization of two *Clostridium acetobutylicum* ATCC 824 butanol dehydrogenase isozyme genes; J. Bacteriol. 1992 November; 174(22):7149-58). It is postulated that Cbei_1722 could be used in the same manner as the Cbei_2181 adh gene.

Butyryl-CoA dehydrogenase (Bcd) catalyzes the formation of butyryl-CoA, an immediate precursor for butanol. Higher Bcd expression in BA101 may lead to increased butyryl-CoA production, which in turn may improve the formation of butanol. In some variations, an organism described herein is optimized to increase production of Bcd. In some variations, an organism described herein is transformed with a Bcd gene. Some variations an organism described herein is transformed with a Bcd gene to overexpress Bcd. In some variations, an organism described herein is transformed with a Bcd gene from a microbial organism to overexpress Bcd. In some variations, an organism described herein is transformed with a Bcd gene from a *Clostridium* sp. to overexpress Bcd. In some variations, an organism described herein is transformed with a Bcd gene from *Clostridium beijerinckii* to overexpress Bcd. In some variations, an organism described herein is transformed with a nucleic acid which results in an increase in expression of the Bcd gene whose DNA sequence is shown in FIG. 11. In some variations, an organism described herein is transformed with a Bcd gene whose DNA sequence is shown

in FIG. 11 (SEQ ID NO: 2) to overexpress Bcd. The *Clostridium beijerinckii* NCIMB 8052 published genome identifies this bcd gene as Cbei_2035.

In some variations, an organism described herein is transformed with a Bcd gene whose DNA sequence is at least 60-100% identical to the DNA sequence shown in FIG. 11 or complement thereof. In some variations, an organism described herein is transformed with a Bcd gene whose DNA sequence is at least 80-100% identical to the DNA sequence shown in FIG. 11 or complement thereof. In some variations, an organism described herein is transformed with a Bcd gene whose DNA sequence is at least 90-100% identical to the DNA sequence shown in FIG. 11 or complement thereof. In some variations, an organism described herein is transformed with a Bcd gene whose DNA sequence is at least 60, at least 65, at least 70, at least 75, at least 80, at least 85, at least 90, at least 95, or 100% identical to the DNA sequence shown in FIG. 11 or complement thereof. In some variations, an organism described herein is transformed with a Bcd gene whose DNA sequence is at least 80% identical to the DNA sequence shown in FIG. 11 or complement thereof. In some variations, an organism described herein is transformed with a Bcd gene whose DNA sequence is at least 85% identical to the DNA sequence shown in FIG. 11 or complement thereof. In some variations, an organism described herein is transformed with a Bcd gene whose DNA sequence is at least 90% identical to the DNA sequence shown in FIG. 11 or complement thereof. In some variations, an organism described herein is transformed with a Bcd gene whose DNA sequence is at least 95% identical to the DNA sequence shown in FIG. 11 or complement thereof. In some variations, an organism described herein is transformed with a Bcd gene whose DNA sequence is at least 100% identical to the DNA sequence shown in FIG. 11 or complement thereof.

The *C. beijerinckii* NCIMB 8052 published genome identifies the bcd gene shown in FIG. 11 as Cbei_2035 (SEQ ID NO: 2). Other genes identified in the *C. beijerinckii* NCIMB 8052 published genome that are close homologs of the bcd gene Cbei_2035 (SEQ ID NO: 2) include Cbei_0322. The DNA sequence of Cbei_0322 (SEQ ID NO: 12) shown in FIG. 21A and the protein sequence of Cbei_0322 (SEQ ID NO: 13) shown in FIG. 21B.

Cbei_0322 shows 98% identity to Cbei_2035 at the DNA level and 98% identity at the protein sequence level, with no gaps. The close homology suggests that gene Cbei_0322 could show Bcd activity. Cbei_0322 is annotated as a “acyl-CoA dehydrogenase domain protein” which is consistent with its being a bcd gene. Cbei_2035 (SEQ ID NO: 2) is also annotated as “acyl-CoA dehydrogenase domain protein” in the GenBank record. While it is possible that the native role of Cbei_0322 protein may be in a pathway other than solvent production, such as for instance the metabolism of other fatty acids, its close homology to Cbei_2035 suggests that even if that were true, it could be used as a functional Bcd gene under the control of an appropriate promoter.

Butyrate kinase (Buk) is a key enzyme in butyrate synthesis. Increased Buk activity in BA101 may allow the generation of higher amounts of butyrate, which can then be converted into butyryl-CoA and further into butanol. In some variations, an organism described herein is optimized to increase production of Buk. In some variations, an organism described herein is transformed with a Buk gene. In some variations an organism described herein is transformed with a Buk gene to overexpress Buk. In some variations, an organism described herein is transformed with a Buk gene from a microbial organism to overexpress Buk. In some variations, an organism described herein is transformed with a Buk gene from a *Clostridium* sp. to overexpress Buk. In some variations, an organism described herein is transformed with a Buk gene from *Clostridium beijerinckii* to overexpress Buk. In some variations, an organism described herein is transformed with a nucleic acid which results in an increase in expression of the Buk gene whose DNA sequence is shown in FIG. 12. In

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some variations, an organism described herein is transformed with a Buk gene whose DNA sequence is shown in FIG. 12 or overexpress Buk.

In some variations, an organism described herein is transformed with a Buk gene whose DNA sequence is at least 60-100% identical to the DNA sequence shown in FIG. 12 or complement thereof. In some variations, an organism described herein is transformed with a Buk gene whose DNA sequence is at least 80-100% identical to the DNA sequence shown in FIG. 12 or complement thereof. In some variations, an organism described herein is transformed with a Buk gene whose DNA sequence is at least 90-100% identical to the DNA sequence shown in FIG. 12 or complement thereof. In some variations, an organism described herein is transformed with a Buk gene whose DNA sequence is at least 60, at least 65, at least 70, at least 75, at least 80, at least 85, at least 90, at least 95, or 100% identical to the DNA sequence shown in FIG. 12 or complement thereof. In some variations, an organism described herein is transformed with a Buk gene whose DNA sequence is at least 80% identical to the DNA sequence shown in FIG. 12 or complement thereof. In some variations, an organism described herein is transformed with a Buk gene whose DNA sequence is at least 85% identical to the DNA sequence shown in FIG. 12 or complement thereof. In some variations, an organism described herein is transformed with a Buk gene whose DNA sequence is at least 90% identical to the DNA sequence shown in FIG. 12 or complement thereof.

In some variations, an organism described herein is optimized to increase expression of any one or more of Adh, Bcd, or Buk. In some variations, an organism described herein is transformed with genes encoding any one or more of Adh, Bcd, or Buk. In some variations an organism described herein is transformed with genes encoding each of Adh, Bcd, and Buk to overexpress Adh, Bcd, and Buk. In some variations an organism described herein is transformed with a nucleic acid which increases expression of any one or more of Adh, Bcd, or Buk.

Genes of Solvent Production Pathway

As demonstrated in FIG. 8, expression of aceto-acetyl CoA:acetate-butyrat CoA transferase subunit α/β (CtfA/B) and acetoacetate decarboxylase (Adc) was highly activated at the onset of solventogenic phase in BA101 and the wild-type strain. Changes in expression levels were much smaller for thiolase (Thl), 3-hydroxybutyryl-CoA dehydrogenase (Hcd) and crotonase (Crt) in BA101 and the wild-type strain.

Despite the somewhat comparable expression kinetics of CtfA/B, Adc, Thl, Hcd and Crt in the BA101 strain relative to the wild type parent, altering (increasing or decreasing) the expression of these genes may prove useful in increasing solvent production in the organisms described herein.

In some variations, an organism described herein is optimized to increase production of one or more solvents by changing the expression of any one or more of CtfA/B, Adc, Thl, Hcd and Crt. In some variations, an organism described herein is optimized to increase production of one or more solvents by increasing the expression of one or more of CtfA/B, Adc, Thl, Hcd and Crt. In some variations, an organism described herein is optimized to increase production of one or more solvents by decreasing the expression of one or more of CtfA/B, Adc, Thl, Hcd and Crt. In some variations, an organism described herein is transformed with genes encoding any one or more of CtfA/B, Adc, Thl, Hcd and Crt.

In some variations, an organism described herein is optimized to decrease production of one or more gene products which compete with or are otherwise detrimental to the production of solvents. In some variations an organism described herein is transformed with a nucleic acid to decrease or impair

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expression of one or more gene products which compete with or are otherwise detrimental to the production of solvents.

In some variations an organism described herein is transformed with a nucleic acid to decrease or impair expression of Adc. In some variations an organism described herein is transformed with a nucleic acid to increase expression of Adc. In some variations, an organism described herein is transformed with an Adc gene from a microbial organism to over-express Adc. In some variations, an organism described herein is transformed with an Adc gene from a *Clostridium* sp. to overexpress Adc. In some variations, an organism described herein is transformed with an Adc gene from *Clostridium beijerinckii* to overexpress Adc.

In some variations, an organism described herein is transformed with an Adc gene whose DNA sequence is at least 60-100% identical to that of the *Clostridium beijerinckii* NCIMB 8052 gene. In some variations, an organism described herein is transformed with an Adc gene whose DNA sequence is at least 60-100% identical to that of the *Clostridium beijerinckii* BA101 gene. In some variations, an organism described herein is transformed with an Adc gene whose DNA sequence is at least 60, at least 65, at least 70, at least 75, at least 80, at least 85, at least 90, at least 95, or 100% identical to that of the *Clostridium beijerinckii* NCIMB 80 gene. In some variations, an organism described herein is transformed with an Adc gene whose DNA sequence is at least 60, at least 65, at least 70, at least 75, at least 80, at least 85, at least 90, at least 95, or 100% identical to that of the *Clostridium beijerinckii* BA101 gene.

In some variations an organism described herein is transformed with a nucleic acid to decrease or impair expression of CtfA/B. In some variations an organism described herein is transformed with a nucleic acid to increase expression of CtfA/B. In some variations, an organism described herein is transformed with a CtfA/B gene from a microbial organism to overexpress CtfA/B. In some variations, an organism described herein is transformed with a CtfA/B gene from a *Clostridium* sp. to overexpress CtfA/B. In some variations, an organism described herein is transformed with a CtfA/B gene from *Clostridium beijerinckii* to overexpress CtfA/B.

In some variations, an organism described herein is transformed with a CtfA/B gene whose DNA sequence is at least 60-100% identical to that of the *Clostridium beijerinckii* NCIMB 8052 gene. In some variations, an organism described herein is transformed with a CtfA/B gene whose DNA sequence is at least 60-100% identical to that of the *Clostridium beijerinckii* BA101 gene. In some variations, an organism described herein is transformed with a CtfA/B gene whose DNA sequence is at least 60, at least 65, at least 70, at least 75, at least 80, at least 85, at least 90, at least 95, or 100% identical to that of the *Clostridium beijerinckii* NCIMB 80 gene. In some variations, an organism described herein is transformed with a CtfA/B gene whose DNA sequence is at least 60, at least 65, at least 70, at least 75, at least 80, at least 85, at least 90, at least 95, or 100% identical to that of the *Clostridium beijerinckii* BA101 gene.

In some variations an organism described herein is transformed with a nucleic acid to decrease or impair expression of a gene product leading to production of a solvent other than butanol. In some variations an organism described herein is transformed with a nucleic acid to decrease or impair expression of a gene product leading to production of a solvent other than ethanol. In some variations an organism described herein is transformed with a nucleic acid to decrease or impair expression of a gene product leading to production of a solvent other than acetone.

Sugar Transport Genes

As demonstrated in FIG. 5, sugar transporters in the phosphoenolpyruvate-dependent phosphoryltransferase system (PTS) are down-regulated in BA101 relative to the wild-type strain. BA101 shows significantly lower expression of mannose-type PTS components ManIIAB and ManIIC, which mediate broad spectrum sugar uptake across the cell membrane.

In some variations, an organism described herein is optimized to decrease production of a gene product relating to one or more specific sugar transporters. In some variations, an organism described herein is transformed with a gene that decreases or knocks out the expression or activity of a gene product relating to one or more specific sugar transporters.

In some variations, an organism described herein is optimized to decrease production of all of the gene products described herein relating to one or more specific sugar transporters. In some variations, an organism described herein is transformed with a gene that decreases or knocks out the expression or activity of all of the gene products described herein relating to one or more specific sugar transporters.

In some variations, an organism described herein is optimized to decrease production of ManIIAB. In some variations, an organism described herein is transformed with a nucleic acid to decrease expression of a ManIIAB gene. In some variations an organism described herein is transformed with a nucleic acid to decrease expression of a ManIIAB gene via antisense, siRNA, or DNAzyme technology. In some variations, an organism described herein is transformed with a gene from a microbial organism to decrease expression of ManIIAB. In some variations, an organism described herein is transformed with a gene from a *Clostridium* sp. to decrease expression of ManIIAB. In some variations, an organism described herein is transformed with a gene from *Clostridium beijerinckii* to decrease expression of ManIIAB. In some variations, an organism described herein is transformed with a nucleic acid which results in a decrease in expression of the ManIIAB gene whose DNA sequence is shown in FIG. 16.

In some variations, an organism described herein is optimized to decrease production of ManIIC. In some variations, an organism described herein is transformed with a ManIIC gene. In some variations an organism described herein is transformed with a ManIIC gene to overexpress ManIIC.

In some variations, an organism described herein is optimized to decrease production of ManIIC. In some variations, an organism described herein is transformed with a nucleic acid to decrease expression of a ManIIC gene. In some variations an organism described herein is transformed with a nucleic acid to decrease expression of a ManIIC gene via antisense, siRNA, or DNAzyme technology. In some variations, an organism described herein is transformed with a gene from a microbial organism to decrease expression of ManIIC. In some variations, an organism described herein is transformed with a gene from a *Clostridium* sp. to decrease expression of ManIIC. In some variations, an organism described herein is transformed with a gene from *Clostridium beijerinckii* to decrease expression of ManIIC. In some variations, an organism described herein is transformed with a nucleic acid which results in a decrease in expression of the ManIIC gene whose DNA sequence is shown in FIG. 17.

Sporulation Genes

Sporulation genes are activated as cells reach stationary phase and enter solventogenic stage. Sporulation is generally believed to be necessary for solvent formation. As demonstrated in FIG. 6, among a cascade of sporulation events, BA101 is found defective in late stage sporulation. In contrast to large fold induction in the wild-type, activation is much

weaker in BA101 for genes encoding sporulation proteins necessary for the completion of spore formation and spore stability. These proteins include spore coat assembly protein SpoIV, spore cortex synthesis protein SpoVB and spore DNA packaging protein SspA. Deficiency in sporulation possibly prolongs the clostridial form and thereby allows extended solventogenesis in BA101, which may give rise to enhanced butanol formation.

In some variations, an organism described herein is optimized to decrease production of a gene product relating to sporulation. In some variations, an organism described herein is transformed with a gene that decreases or knocks out the expression or activity of a gene product relating to sporulation.

In some variations, an organism described herein is optimized to decrease production of all of the gene products described herein relating to sporulation. In some variations, an organism described herein is transformed with a gene that decreases or knocks out the expression or activity of all of the gene products described herein relating to sporulation.

In some variations, an organism described herein is optimized to decrease production of SpoIVA. In some variations, an organism described herein is transformed with a gene that decreases or knocks out the expression or activity of SpoIVA. In some variations, an organism described herein is transformed with a nucleic acid to decrease expression of a SpoIVA gene. In some variations an organism described herein is transformed with a nucleic acid to decrease expression of a ManIIAB gene via antisense, siRNA, or DNAzyme technology. In some variations, an organism described herein is transformed with a nucleic acid sequence from a microbial organism to decrease expression of SpoIVA. In some variations, an organism described herein is transformed with a gene from a *Clostridium* sp. to decrease expression of SpoIVA. In some variations, an organism described herein is transformed with a gene from *Clostridium beijerinckii* to decrease expression of SpoIVA. In some variations, an organism described herein is transformed with a nucleic acid which results in a decrease in expression of the SpoIVA gene whose DNA sequence is shown in FIG. 18.

In some variations, an organism described herein is optimized to decrease production of SpoVB. In some variations, an organism described herein is transformed with a gene that decreases or knocks out the expression or activity of SpoVB. In some variations, an organism described herein is transformed with a nucleic acid to decrease expression of a SpoVB gene. In some variations an organism described herein is transformed with a nucleic acid to decrease expression of a ManIIAB gene via antisense, siRNA, or DNAzyme technology. In some variations, an organism described herein is transformed with a gene from a *Clostridium* sp. to decrease expression of SpoVB. In some variations, an organism described herein is transformed with a gene from *Clostridium beijerinckii* to decrease expression of SpoVB. In some variations, an organism described herein is transformed with a nucleic acid which results in a decrease in expression of the SpoVB gene whose DNA sequence is shown in FIG. 19.

In some variations, an organism described herein is optimized to decrease production of SspA. In some variations, an organism described herein is transformed with a gene that decreases or knocks out the expression or activity of SspA. In some variations, an organism described herein is transformed with a nucleic acid to decrease expression of an SspA gene. In some variations an organism described herein is transformed with a nucleic acid to decrease expression of an SspA gene via antisense, siRNA, or DNAzyme technology. In some variations an organism described herein is transformed with an

antisense nucleic acid to decrease expression of an SspA gene. In some variations, an organism described herein is transformed with a nucleic acid sequence from a microbial organism to decrease expression of SspA. In some variations, an organism described herein is transformed with a gene from a *Clostridium* sp. to decrease expression of SspA. In some variations, an organism described herein is transformed with a gene from *Clostridium beijerinckii* to decrease expression of SspA. In some variations, an organism described herein is transformed with a nucleic acid which results in a decrease in expression of the SspA gene whose DNA sequence is shown in FIG. 20 (SEQ ID NO: 11). The *Clostridium beijerinckii* NCIMB 8052 published genome identifies this SspA gene as Cbei_3080.

In some variations, an organism described herein is transformed with a SspA gene whose DNA sequence is at least 60-100% identical to the DNA sequence shown in FIG. 20 or complement thereof. In some variations, an organism described herein is transformed with a SspA gene whose DNA sequence is at least 80-100% identical to the DNA sequence shown in FIG. 20 or complement thereof. In some variations, an organism described herein is transformed with a SspA gene whose DNA sequence is at least 90-100% identical to the DNA sequence shown in FIG. 20 or complement thereof. In some variations, an organism described herein is transformed with a SspA gene whose DNA sequence is at least 60, at least 65, at least 70, at least 75, at least 80, at least 85, at least 90, at least 95, or 100% identical to the DNA sequence shown in FIG. 20 or complement thereof. In some variations, an organism described herein is transformed with a SspA gene whose DNA sequence is at least 80% identical to the DNA sequence shown in FIG. 20 or complement thereof. In some variations, an organism described herein is transformed with a SspA gene whose DNA sequence is at least 85% identical to the DNA sequence shown in FIG. 20 or complement thereof. In some variations, an organism described herein is transformed with a SspA gene whose DNA sequence is at least 90% identical to the DNA sequence shown in FIG. 20 or complement thereof.

The *C. beijerinckii* NCIMB 8052 published genome identifies the SspA gene shown in FIG. 20 as Cbei_3080 (SEQ ID NO:11). It is annotated in GenBank as a “small acid-soluble spore protein, alpha/beta type.”

Other genes identified in the *C. beijerinckii* NCIMB 8052 published genome that are close homologs of the sspA gene Cbei_3080 (SEQ ID NO:11) include Cbei_3111 and Cbei_3250. They belong to a family of highly conserved spore proteins that are present in this organism and are annotated with the same function—“small acid-soluble spore protein alpha/beta type”—as is Cbei_3080 (SEQ ID NO:11) shown in FIG. 20. At the protein sequence level Cbei_3111 is 98% similar and 91% identical to Cbei_3080. Cbei_3250 is 94% similar and 91% identical.

The utility of Cbei_3111 and Cbei_3250 would be the same as that taught for Cbei_3080 in the patent, which is to reduce or eliminate their expression through a variety of methods.

The DNA sequence of Cbei_3111 (SEQ ID NO: 16) shown in FIG. 23A and the protein sequence of Cbei_3111 (SEQ ID NO: 17) shown in FIG. 23B.

The DNA sequence of Cbei_3250 (SEQ ID NO: 18) shown in FIG. 24A and the protein sequence of Cbei_3250 (SEQ ID NO: 19) shown in FIG. 24B.

Chemotaxis Genes

As demonstrated in FIG. 7, BA101 has higher expression of chemotaxis and motility genes than the wild-type strain. Genes in a chemotaxis operon CheA, CheC, CheD and CheW

become repressed in the wild-type during the solventogenic phase, while their expression levels remain stable in BA101. As highly solventogenic clostridia are generally associated with high motility, BA101 appears to remain in a motile form which may be favorable to solvent production.

In some variations, an organism described herein is optimized to increase production of one or more chemotaxis or motility genes. In some variations, an organism described herein is transformed with a gene encoding one or more chemotaxis or motility genes. In some variations an organism described herein is transformed with a gene encoding one or more chemotaxis or motility genes to overexpress one or more of the chemotaxis or motility genes.

In some variations, an organism described herein is optimized to increase production of all of the chemotaxis or motility genes described herein. In some variations, an organism described herein is transformed with genes encoding all of the chemotaxis or motility genes described herein. In some variations an organism described herein is transformed with genes encoding all of the chemotaxis or motility genes described herein to overexpress all of the chemotaxis or motility genes described herein.

In some variations, an organism described herein is optimized to increase production of CheA. In some variations, an organism described herein is transformed with a CheA gene. In some variations an organism described herein is transformed with a CheA gene to overexpress CheA. In some variations, an organism described herein is optimized to increase production of CheA in the solventogenic phase. In some variations, an organism described herein is transformed with a CheA gene. In some variations an organism described herein is transformed with a CheA gene to overexpress CheA in the solventogenic phase. In some variations, an organism described herein is transformed with a CheA gene from a microbial organism to overexpress CheA. In some variations, an organism described herein is transformed with a CheA gene from a *Clostridium* sp. to overexpress CheA. In some variations, an organism described herein is transformed with a CheA gene from *Clostridium beijerinckii* to overexpress CheA. In some variations, an organism described herein is transformed with a nucleic acid which results in an increase in expression of the CheA gene whose DNA sequence is shown in FIG. 13. In some variations, an organism described herein is transformed with a CheA gene whose DNA sequence is shown in FIG. 13 to overexpress CheA.

In some variations, an organism described herein is transformed with a CheA gene whose DNA sequence is at least 60-100% identical to the DNA sequence shown in FIG. 13 or complement thereof. In some variations, an organism described herein is transformed with a CheA gene whose DNA sequence is at least 80-100% identical to the DNA sequence shown in FIG. 13 or complement thereof. In some variations, an organism described herein is transformed with a CheA gene whose DNA sequence is at least 90-100% identical to the DNA sequence shown in FIG. 13 or complement thereof. In some variations, an organism described herein is transformed with a CheA gene whose DNA sequence is at least 60, at least 65, at least 70, at least 75, at least 80, at least 85, at least 90, at least 95, or 100% identical to the DNA sequence shown in FIG. 13 or complement thereof. In some variations, an organism described herein is transformed with a CheA gene whose DNA sequence is at least 80% identical to the DNA sequence shown in FIG. 13. In some variations, an organism described herein is transformed with a CheA gene whose DNA sequence is at least 85% identical to the DNA sequence shown in FIG. 13 or complement thereof. In some variations, an organism described herein is transformed with

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a CheA gene whose DNA sequence is at least 90% identical to the DNA sequence shown in FIG. 13 or complement thereof.

In some variations, an organism described herein is optimized to increase production of CheC. In some variations, an organism described herein is transformed with a CheC gene. In some variations an organism described herein is transformed with a CheC gene to overexpress CheC. In some variations, an organism described herein is optimized to increase production of CheC in the solventogenic phase. In some variations, an organism described herein is transformed with a CheC gene. In some variations an organism described herein is transformed with a CheC gene to overexpress CheC in the solventogenic phase. In some variations, an organism described herein is transformed with a CheC gene from a microbial organism to overexpress CheC. In some variations, an organism described herein is transformed with a CheC gene from a *Clostridium* sp. to overexpress CheC. In some variations, an organism described herein is transformed with a CheC gene from *Clostridium beijerinckii* to overexpress CheC. In some variations, an organism described herein is transformed with a nucleic acid which results in an increase in expression of the CheC gene whose DNA sequence is shown in FIG. 14. In some variations, an organism described herein is transformed with a CheC gene whose DNA sequence is shown in FIG. 14 to overexpress CheC.

In some variations, an organism described herein is transformed with a CheC gene whose DNA sequence is at least 60-100% identical to the DNA sequence shown in FIG. 14 or complement thereof. In some variations, an organism described herein is transformed with a CheC gene whose DNA sequence is at least 80-100% identical to the DNA sequence shown in FIG. 14 or complement thereof. In some variations, an organism described herein is transformed with a CheC gene whose DNA sequence is at least 90-100% identical to the DNA sequence shown in FIG. 14 or complement thereof. In some variations, an organism described herein is transformed with a CheC gene whose DNA sequence is at least 60, at least 65, at least 70, at least 75, at least 80, at least 85, at least 90, at least 95, or 100% identical to the DNA sequence shown in FIG. 14 or complement thereof. In some variations, an organism described herein is transformed with a CheC gene whose DNA sequence is at least 80% identical to the DNA sequence shown in FIG. 14 or complement thereof. In some variations, an organism described herein is transformed with a CheC gene whose DNA sequence is at least 85% identical to the DNA sequence shown in FIG. 14 or complement thereof. In some variations, an organism described herein is transformed with a CheC gene whose DNA sequence is at least 90% identical to the DNA sequence shown in FIG. 14 or complement thereof.

In some variations, an organism described herein is optimized to increase production of CheD. In some variations, an organism described herein is transformed with a CheD gene. In some variations an organism described herein is transformed with a CheD gene to overexpress CheD. In some variations, an organism described herein is optimized to increase production of CheD in the solventogenic phase. In some variations, an organism described herein is transformed with a CheD gene. In some variations an organism described herein is transformed with a CheD gene to overexpress CheD in the solventogenic phase.

In some variations, an organism described herein is optimized to increase production of CheW. In some variations, an organism described herein is transformed with a CheW gene. In some variations an organism described herein is transformed with a CheW gene to overexpress CheW. In some

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variations, an organism described herein is optimized to increase production of CheW in the solventogenic phase. In some variations, an organism described herein is transformed with a CheW gene. In some variations an organism described herein is transformed with a CheW gene to overexpress CheW in the solventogenic phase. In some variations, an organism described herein is transformed with a CheW gene from a microbial organism to overexpress CheW. In some variations, an organism described herein is transformed with a CheW gene from a *Clostridium* sp. to overexpress CheW. In some variations, an organism described herein is transformed with a CheW gene from *Clostridium beijerinckii* to overexpress CheW. In some variations, an organism described herein is transformed with a nucleic acid which results in an increase in expression of the CheW gene whose DNA sequence is shown in FIG. 15. In some variations, an organism described herein is transformed with a CheW gene whose DNA sequence is shown in FIG. 15 to overexpress CheW.

In some variations, an organism described herein is transformed with a CheW gene whose DNA sequence is at least 60-100% identical to the DNA sequence shown in FIG. 15 or complement thereof. In some variations, an organism described herein is transformed with a CheW gene whose DNA sequence is at least 80-100% identical to the DNA sequence shown in FIG. 15 or complement thereof. In some variations, an organism described herein is transformed with a CheW gene whose DNA sequence is at least 90-100% identical to the DNA sequence shown in FIG. 15 or complement thereof. In some variations, an organism described herein is transformed with a CheW gene whose DNA sequence is at least 60, at least 65, at least 70, at least 75, at least 80, at least 85, at least 90, at least 95, or 100% identical to the DNA sequence shown in FIG. 15 or complement thereof. In some variations, an organism described herein is transformed with a CheW gene whose DNA sequence is at least 80% identical to the DNA sequence shown in FIG. 15 or complement thereof. In some variations, an organism described herein is transformed with a CheW gene whose DNA sequence is at least 85% identical to the DNA sequence shown in FIG. 15 or complement thereof. In some variations, an organism described herein is transformed with a CheW gene whose DNA sequence is at least 90% identical to the DNA sequence shown in FIG. 15 or complement thereof.

To develop an organism that can tolerate various inhibitors and products in the solvent production process, analysis of the mechanism of tolerance may be investigated. In some variations DNA microarray analysis is used to study the global or selected expression profiles of an organism described herein when exposed to various inhibitors or products in order to identify the organism's genetic responses. In addition, microarray analysis may be used to examine specific enzymes (glycolytic and non-glycolytic) that may be inhibited by these degradation compounds. Enzymes of particular interest include alcohol dehydrogenase, phosphofructokinase, glucokinase, galactokinase, aldehyde dehydrogenase, pyruvate dehydrogenase complex, butyryl-CoA dehydrogenase, butyrate kinase, etc.

In some variations, an organism described herein is optimized for alcohol dehydrogenase tolerance to inhibitors and products in the solvent production process. In some variations, an organism described herein is optimized for butyryl-CoA dehydrogenase tolerance to inhibitors and products in the solvent production process. In some variations, an organism described herein is optimized for butyrate kinase tolerance to inhibitors and products in the solvent production process.

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In some variations, an organism described herein is optimized for phosphofructokinase tolerance to inhibitors and products in the solvent production process. In some variations, an organism described herein is optimized for glucokinase tolerance to inhibitors and products in the solvent production process. In some variations, an organism described herein is optimized for galactokinase tolerance to inhibitors and products in the solvent production process. In some variations, an organism described herein is optimized for aldehyde dehydrogenase tolerance to inhibitors and products in the solvent production process. In some variations, an organism described herein is optimized for pyruvate dehydrogenase complex tolerance to inhibitors and products in the solvent production process.

Methods of Optimizing Organisms for Use in Industrial Applications

In some variations, an organism described herein is optimized so as to be more tolerant of industrial conditions. In some variations, an organism described herein is subjected to a selection process under the industrial condition of interest, and the most adapted cells are identified. In some variations, an organism described herein is subjected to mutagenesis, subsequently subjected to a selection process under the industrial condition of interest, and the most adapted cells are identified. In some variations an organism described herein is transformed with one or more genes or regulatory sequences giving increased tolerance or resistance to an industrial condition of interest, and the most adapted cells are identified.

In some variations, an organism described herein is optimized to increase tolerance or resistance to one or more aspects or by-products of pretreatment. In some variations, an organism described herein is optimized to increase tolerance or resistance to one or more of salt, acetate, furfural, hydroxymethylfurfural, acetic acid, ferulic acid, glucuronic acid, rhoumaric acid, and phenolic compounds.

In some variations, an organism described herein is optimized to increase tolerance or resistance to rhoumaric acid. In some variations, an organism described herein is optimized to increase tolerance or resistance to ferulic acid.

In some variations, an organism described herein is optimized to increase tolerance or resistance to salt.

In some variations, an organism described herein is optimized to increase tolerance or resistance to one or more intermediates or products generated in the solventogenic process.

In some variations, an organism described herein is optimized to increase tolerance or resistance to one or more specific solvent recovery methods, including but not limited to gas stripping and adsorption or selective membranes.

In some variations, an organism described herein is optimized to increase tolerance or resistance to one or more temperatures utilized in the solventogenic process.

In some variations, an organism described herein is optimized to increase tolerance or resistance to one or more salts encountered in the solventogenic process.

In some variations, an organism described herein is optimized to increase tolerance or resistance to one or more pH conditions utilized in the solventogenic process.

In some variations, an organism described herein is optimized to increase tolerance or resistance to one or more continuous processing conditions utilized in the solventogenic process.

In some variations, an organism described herein is optimized to increase tolerance or resistance to one or more solvents generated in the solventogenic process.

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In some variations, an organism described herein is optimized to increase tolerance or resistance to one or more feedstock materials in the solventogenic process.

EXAMPLES

Clostridial fermentation cultures were grown for both *C. beijerinckii* NCIMB 8052 and the hyper-butanol-producing mutant BA101 ATCC No. PTA-1550. Samples were collected at various time points over the course of fermentation. Total RNA was isolated from each time point sample. Dye-labeled DNA was generated by reverse transcription from total RNA and used as a sample probe in microarray hybridization. An RNA pool was constructed by mixing samples obtained from different stages of cell growth. Dye-labeled DNA probe derived from this RNA pool was used as a reference probe in microarray hybridization.

The DNA microarray included ~500 predicted protein-coding genes based on the draft sequence of *C. beijerinckii* NCIMB 8052 provided by the Joint Genome Institute, available at www.jgi.doe.gov and at GenBank as accession number CP000721. The array represented 10 functional classes covering ~10% of the genome.

Example 1

Bacterial Strains and Fermentation Protocols

30 Bacterial strains and growth conditions. *C. beijerinckii* NCIMB 8052 is the wild-type strain. BA101 is the hyper butanol-producing mutant strain. Stocks of the wild-type and BA101 spores were stored in sterile nanopure H₂O at 4° C.

35 Fermentation protocols. 1 ml *C. beijerinckii* spore suspensions were heat shocked at 80° C. for 10 min, and inoculated into 100 ml tryptone-glucose-yeast extract (TGY) media containing 3% tryptone, 2% glucose, 1% yeast extract and 0.1% L-cysteine-HCl. The TGY culture was grown at 35° C. for 12 hrs in an anaerobic chamber (Coy Laboratory Products) maintained under a gas mixture of 85% N₂, 10% CO₂ and 5% H₂. The culture was diluted 10⁶-10⁷ fold into 0.45% liquefied TGY-agar and the mixture was allowed to solidify in plates in the anaerobic chamber. Plates were incubated at 35° C. for 2-3 days. Individual colonies developed on the plates were inoculated into 30 ml cooked meat medium (CMM, Oxoid #CM0081) plus added 1% glucose. The CMM culture was grown at 35° C. for 9 hrs in the anaerobic chamber. Subsequently, 10 ml CMM culture was inoculated into fresh 100 ml TGY media and grown at 35° C. for 3 hrs in the anaerobic chamber. An aliquot of 20 ml TGY pre-culture was inoculated into 1.7 liter P2 media containing P2 solutions supplemented with 6% glucose and 0.1% yeast extract in a fermentation reactor (New Brunswick Scientific). The P2 culture was grown at 35° C. under nitrogen flow. Fermentation samples were taken at various time points for analysis.

Example 2

Fermentation Sample Analysis

60 Aliquots of 1 ml fermentation culture grown in P2 media were collected at various time points for both *C. beijerinckii* NCIMB 8052 and BA101.

65 Cell growth was monitored by measuring the absorbance at 600 nm with a spectrophotometer (Beckman Coulter). Results are depicted in FIG. 1A. The growth curve for the two strains was very similar under these conditions.

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Changes in pH were monitored by sampling the liquid culture using a pH meter. Results are depicted in FIG. 1B. The pH of the liquid culture was similar under these conditions, though the *C. beijerinckii* NCIMB 8052 liquid culture had a higher pH at the later timepoints.

Culture supernatants were analyzed for solvent and acid contents using gas chromatography (Agilent Technologies). Results are shown in FIG. 2A, FIG. 2B, and FIG. 2C. Total solvents were similar in the two strains until about 20 hours, after which point the level of solvents was consistently higher in the BA101 strain.

Example 3

RNA Sampling and Isolation

Aliquots of 10 ml fermentation culture in P2 media were obtained at various time points for both *C. beijerinckii* NCIMB 8052 and BA101. Cells were pelleted by centrifuging at 4000 g for 10 min. Total RNA was extracted from the cell pellets using a RNeasy mini kit (Qiagen) according to the manufacturer's protocol. RNA quality was determined with nanochip on an Agilent 2100 Bioanalyzer (Agilent Technologies). RNA concentration was quantified by measuring A260 using a UV/vis spectrophotometer (Bioteck Instruments). Purified samples were stored in aliquots at -80° C.

To make a reference for comparing gene expression in the time course samples, a RNA pool was prepared and used to

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generate an oppositely labeled probe in microarray hybridization. To do so, a group of 500 ml static flask cultures were grown in P2 media for *C. beijerinckii* NCIMB 8052. The cultures were harvested at different stages of cell growth over the course of fermentation and total RNA was extracted from each cell pellet. An RNA pool was generated by mixing equal quantities of purified RNA from each growth phase, and this mixture was used to create a reference probe for microarray hybridization.

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Example 4

Microarray Construction

15 DNA microarray was constructed by spotting long oligonucleotide probes onto a glass slide (UIUC Functional Genomics Keck Center). A 70-mer probe was selected for a single predicted open reading frame (ORF) in the sequenced *C. beijerinckii* genome (Illumina). Each probe was printed in 20 duplicate on the array slide. Each array includes 485 predicted ORFs representing 10 functional classes and approximately 1/10 of the genome based on the draft sequence assembly of *C. beijerinckii* NCIMB 8052 (Joint Genome Institute). The *C. beijerinckii* NCIMB 8052 genes included in the microarray analysis are shown in Table 1, below.

25 Each gene is associated with a unique gene ID according to the JGI annotation available at the time when the list was compiled for microarray construction.

TABLE 1

Gene name	Gene ID
Transcriptional regulator AbrB	1
Probable glucose kinase	11
SpoOA protein (CheY-like receiver domain and HTH-type DNA binding domain)	54
SpoIVB	55
Exonuclease VII small subunit	62
Exonuclease VII large subunit	63
Critical stage III sporulation protein AH	67
Stage III sporulation protein AG, SpoIIIAG	68
Stage III sporulation protein AF, SpoIIIAF, putative	69
Stage III sporulation protein AE, SpoIIIAE	70
Stage III sporulation protein AD SpoIIIAAD	71
Stage III sporulation protein AC, SpoIIAC	72
Stage III sporulation protein AB, SpoIIIB	73
Stage III sporulation protein AA, SpoIIIAA	74
CDP-diglyceride synthetase	90
Pseudouridine synthase	102
Riboflavin kinase/FAD synthase	103
Ribosomal Protein S15	104
Periplasmic serine protease, YMFB <i>B. subtilis</i> ortholog	109
Sporulation protein SpoIIIE, DNA segregation ATPase	110
Predicted Fe—S oxidoreductase	111
Catabolic acetolactate synthase	139
Aspartyl/asparaginyl-tRNA synthetase	168
Ribose 5-phosphate isomerase A	175
Putative alternative nitrogenase molybdenum-iron protein, NifD- or NifE-like	193
Putative alternative FeMo-cofactor synthesis protein, NifB-like	196
Putative alternative nitrogenase iron protein, NifH-like	201
Putative alternative nitrogenase molybdenum-iron protein, NifD- or NifE-like	204
Stage V sporulation protein	217
Asparagine synthase, N-terminal domain	218
ABC-type multi-drug/protein/lipid transport system, membrane ATPase component	225
NH ₃ -dependent NAD synthase fused to amidohydrolase domain	228
DSBH domain-containing protein	229
RecG helicase	235
Phosphopantetheine adenyltransferase	237
Phosphotransacetylase	241
Acetate kinase	242
Acy carrier protein ACP	246
ADP-glucose pyrophosphorylase	253
ADP-glucose pyrophosphorylase	254
Glycogen phosphorylase	256
Glycogen synthase, GlgA	257

TABLE 1-continued

Gene name	Gene ID
L-lactate dehydrogenase	290
Acyl-coA dehydrogenase: butyryl-CoA dehydrogenase	292
Formate acetyltransferase	293
Pyruvate-formate lyase	295
6-Phosphofructokinase	306
RecA recombinase, ATPase	310
Stage V sporulation protein S, SpoVS	312
Beta-galactosidase	324
Beta-galactosidase	328
DNA-dependent RNA polymerase sigma subunit	345
Specialized DNA-dependent RNA polymerase sigma subunit	346
Response regulator (CheY-like receiver domain and HTH-type DNA-binding domain)	350
Permease component of ATP-dependent phosphate uptake system	354
Fe—S oxidoreductase, related to NitB/MoaA family with PDZ N-terminal domain	358
Glycerol 3-phosphate dehydrogenase	360
Coat morphogenesis sporulation protein SpoIVA	361
Uncharacterized stress-induced protein, YicC family	364
RNA polymerase-associated protein RpoZ, omega subunit, YLOH <i>B. subtilis</i> ortholog	367
Flavoprotein involved in pantothenate metabolism, YLOI <i>B. subtilis</i> ortholog	368
Primosomal protein N', superfamily II helicase	369
Ribulose-phosphate 3-epimerase	378
Ribosomal protein L28	380
Ribosomal protein L2	410
Adenylate kinase	428
DNA-dependent RNA polymerase alpha subunit	436
ABC-type transporter, ATPase component, cobalt transporters subfamily	439
Probable spore cortex lytic enzyme	455
Phosphotransbutyrylase, Ptb	459
Butyrate kinase, Burk	460
Flagellar motor switch protein, FliG	471
Ethanolamine utilization protein, EutE	499
Ribose 5-phosphate isomerase A	537
Alpha-L-arabinofuranosidase	544
Putative pyruvate kinase	555
Critical small acid-soluble spore protein, alpha/beta type	559
ATPases with chaperone activity ClpC, two ATP-binding domains	587
RNA methyltransferase TrmII family, group 3	597
Acyl-coA dehydrogenase: butyryl-CoA dehydrogenase	617
Critical probable spore coat protein	650
Putative spore coat protein	651
Spore coat protein S	652
Mannosyl transferase	653
Probable spore coat protein	654
Stage II sporulation protein	662
DNA gyrase (topoisomerase II) subunit A	671
DNA gyrase (topoisomerase II) subunit B	672
RecF, ABC family ATPase	674
DNA polymerase III beta subunit	676
DNA replication initiator protein, ATPase	677
Stage III sporulation protein J, SpoIII J	681
SpoIII J-associated protein	682
Stage O sporulation protein J, SpoOJ	686
SpoOJ activation inhibitor	687
Stage O sporulation protein J, SpoOJ	688
Single strand DNA-binding protein Ssb	697
Uncharacterized conserved protein, CotF <i>B. subtilis</i> ortholog	718
VWA domain-containing CoxE-like protein family	731
Membrane permease, predicted cation efflux pumps	741
Predicted Co/Zn/Cd cation transporter	759
Regulatory protein TenI	775
Uncharacterized protein containing two CBS domains	779
Transcriptional regulator, LysR family	793
Phosphoglycerate mutase family protein	826
3-Oxoacyl-(acyl carrier protein) reductase	834
Alcohol dehydrogenase	873
Possible phosphoglycerate mutase	875
Uncharacterized oxidoreductase, Fe-dependent alcohol dehydrogenase family	902
Fructose-bisphosphate aldolase	930
Probable tagatose-6-phosphate kinase, AgaZ	972
Probable tagatose-6-phosphate kinase	973
Small acid-soluble spore protein beta	1029
Small acid-soluble spore protein	1030
Fructose-1,6-bisphosphatase, YYDE <i>B. subtilis</i> ortholog	1033
Cytosine deaminase	1074
Cyclopropane fatty acid synthase	1079
ABC-type probable sulfate transporter, periplasmic binding protein	1097
Bifunctional enzyme phosphoribosyl-formyl-glycinamide (FGAM) synthase	1223
3-Oxoacyl-[acyl-carrier-protein] synthase III	1238

TABLE 1-continued

Gene name	Gene ID
Dioxygenase	1239
Malonyl CoA-acyl carrier protein transacylase	1240
3-Oxoacyl-[acyl-carrier-protein] reductase	1241
3-Oxoacyl-[acyl-carrier-protein] synthase II	1242
Acetyl-CoA carboxylase	1243
FabZ	1244
Acetyl-CoA carboxylase: biotin carboxylase	1245
Acetyl-CoA carboxylase subunit beta	1246
Acetyl-CoA carboxylase carboxyl transferase subunit alpha	1247
Predicted endonuclease involved in recombination	1274
Ferric uptake regulation protein	1276
DNA-dependent RNA polymerase sigma subunit	1283
Cell division GTPase FtsZ	1286
Recombination protein RecR	1313
DNA-directed DNA polymerase III chain, DnaX	1315
Pyruvate carboxylase	1324
Xylan 1,4-beta-xylosidase	1336
Sigma factor SigK processing regulatory protein, BofA <i>B. subtilis</i> ortholog	1359
Phosphoenolpyruvate synthase	1376
Pyruvate water dikinase	1379
Spore coat protein CotJC	1382
Histidine kinase	1385
Long-chain fatty acid-CoA ligase	1407
4-Hydroxybutyryl-CoA dehydratase	1411
Arsenate reductase, ArsC, tyrosine-phosphatase family enzyme	1422
Spore coat peptide assembly protein CotJB	1434
Transketolase	1450
Bifunctional D-arabino 3-hexulose-6-phosphate formaldehyde lyase/phosphohexuloisomerase	1453
Beta-glucosidase	1475
ABC transporter, ATP-binding component	1477
Xylose isomerase	1504
xylulose kinase	1505
Transaldolase, putative	1507
3-Oxoacyl-[acyl-carrier-protein] reductase	1519
Activator of 2-hydroxyglutaryl-CoA dehydratase	1526
NADH-dependent butanol dehydrogenase BDH II	1542
MDR-type permease	1577
Response regulator (CheY-like receiver domain and DNA-binding HTH domain)	1599
Regulator of stationary/sporulation gene expression AbrB-like gene	1615
Phosphoglycerate mutase	1662
Critical small acid-soluble spore protein, alpha/beta type	1685
Small acid-soluble spore protein SspA	1699
SleC	1704
Stage V sporulation protein T, transcriptional regulator AbrB homolog	1745
Ribose 5-phosphate isomerase B	1773
Thiolase, acetyl-CoA acetyltransferase	1777
Stage III sporulation protein D, spore protease Gpr-related protein	1788
Hypothetical protein	1790
Spore protease Gpr-related protein, YYAC <i>B. subtilis</i> ortholog	1792
Predicted iron-binding protein, hemerythrin	1829
Critical small acid-soluble spore protein	1840
Pyruvate kinase	1851
Alcohol dehydrogenase, zinc-dependent	1873
Transketolase, N-terminal section	1874
Transketolase, C-terminal section	1875
Ribulose-phosphate 3-epimerase	1876
Ribose 5-phosphate isomerase B	1877
ABC-type transport system, ATPase component	1887
Long-chain fatty acid-CoA ligase	1903
Malonyl CoA-acyl carrier protein transacylase	1906
Small acid-soluble spore protein beta	1927
Histidinol-phosphate aminotransferase	1941
1-Phosphofructokinase	1972
Pyruvate ferredoxin oxidoreductase	1982
Predicted oxidoreductase, GSP39 <i>B. subtilis</i> ortholog	1988
Uncharacterized protein, YPUG <i>B. subtilis</i> ortholog	2004
Putative 4-cys ferredoxin	2009
SpoU	2018
Predicted S-adenosylmethionine-dependent methyltransferase	2022
Stage V sporulation protein D, SpoVD, FtsI/pbp family	2024
Stage V sporulation protein D, SpoVD, FtsI/pbp family	2025
Stage V sporulation protein E, SpoVE	2029
Chemotaxis motility protein B, MotB	2038
Chemotaxis motility protein A, MotA	2039
Butyryl-CoA dehydrogenase	2135
Homocitrate synthase subunit alpha, NifV	2156
Putative NirJ1 protein	2161

TABLE 1-continued

Gene name	Gene ID
Putative [2Fe—2S] ferredoxin, FdxA	2162
FeMo-cofactor synthesis protein, NifN	2163
FeMo cofactor synthesis protein, NifE	2164
Nitrogenase molybdenum-iron protein beta subunit, NifK	2165
Nitrogenase molybdenum-iron protein alpha subunit, NifD	2166
GlnB-like protein-1	2168
Nitrogenase iron protein, NifH	2169
Sporulation factor SpoIIM	2206
3-Oxoacyl-(acyl carrier protein) reductase	2207
Aldehyde dehydrogenase; alcohol dehydrogenase	2247
FAD/FMN-containing dehydrogenase	2254
Pyruvate formate-lyase	2257
Pyruvate formate-lyase activating enzyme	2258
8-Oxoguanine-DNA glycosylases	2268
Co-chaperonin GroES, Hsp10 family	2270
Chaperonin GroEL, Hsp60 family	2271
Glucose-6-phosphate isomerase	2283
3-Oxoacyl-[acyl-carrier protein] reductase	2303
Streptogramin B lactonase	2386
Hypothetical cytosolic protein	2399
Acetyl-CoA acetyltransferase, thiolase	2402
MDR-type permease, probably tetracycline-resistance protein	2412
Malic enzyme	2425
Predicted aldo/keto reductase, YTBE/YVGN <i>B. subtilis</i> ortholog	2496
Phosphoenolpyruvate synthase	2500
Glucose kinase	2501
Membrane-associated methyl-accepting chemotaxis protein with HAMP domain	2547
Chemotaxis protein CheW	2548
Chemotaxis protein methyltransferase, CheR	2553
Chemotaxis protein CheA	2555
Flagellar motor protein MotB	2556
Flagellar motor component MotA	2557
Beta-glucosidase	2559
Pyruvate kinase	2577
Enolase	2578
2,3-Biphosphoglycerate-independent phosphoglycerate mutase gene	2579
Transketolase, C-terminal section	2596
Transketolase, N-terminal section	2597
tRNA-processing ribonuclease	2605
Protein containing Zn-finger domain	2624
SOS regulatory protein LexA	2626
DNA mismatch repair enzyme, MutL	2630
Mismatch repair protein MutS, ATPase	2634
Ketopantoate hydroxymethyltransferase	2674
Alpha-galactosidases/6-phospho-beta-glucosidase, family 4 glycosyl hydrolase	2726
Stage II sporulation protein	2738
Stage V sporulation protein B	2745
Stage V sporulation protein T, SpoVT	2746
Stage V sporulation protein	2754
HD-GYP hydrolase domain-containing protein	2760
Sporule maturation protein	2782
Pyruvate carboxylase PYKA	2785
Pyruvate formate lyase-activating enzyme	2795
HD-GYP hydrolase domain-containing protein	2801
Short-chain dehydrogenase: 3-oxoacyl-[acyl-carrier protein] reductase	2805
Transcriptional regulator TetR/AcrR family	2813
Phosphatidylserine decarboxylase	2814
Mannose/fructose-specific phosphotransferase system component IIC	2839
Mannose-specific phosphotransferase system component IIAB	2840
Pyruvate formate-lyase	2846
Pyruvate formate-lyase activating enzyme	2850
Acyl-acyl carrier protein thioesterase	2861
Putative acyl-CoA ligase	2868
Aldehyde dehydrogenase, NAD-dependent dehydrogenase family	2878
Zinc-containing alcohol dehydrogenase, long-chain	2891
Putative transcription activator, Stc-like	2892
Cation transport P-type ATPase	2906
Septum site-determining protein, MinD	2941
Stage V sporulation protein E	2943
Putative stage IV sporulation protein FB	2945
Biotin carboxylase: acetyl-CoA carboxylase, putative	2948
Protein of unknown function LDUF464 superfamily	2955
Putative kinase	2970
Ribulose-phosphate 3-epimerase	2973
Alcohol dehydrogenase, zinc-dependent	2988
Transketolase, N-terminal section	2989
Transketolase, C-terminal section	2990
Ribulose-phosphate 3-epimerase	2991

TABLE 1-continued

Gene name	Gene ID
Ribose 5-phosphate isomerase B	2992
Ribulose-phosphate 3-epimerase family protein	2995
Similar to ribulose-5-phosphate 3-epimerase	2996
Stage V sporulation protein R, SpoVR	3012
6-Phosphofructokinase	3028
VanW-like protein family	3037
Glyceraldehyde 3-phosphate dehydrogenase	3041
Phosphoglycerate kinase	3042
Triosephosphate isomerase	3043
2,3-Bisphosphoglycerate-independent phosphoglycerate mutase	3044
Phosphopyruvate hydratase	3046
ABC-type sulfate transporter, ATPase component	3054
Putative alternative nitrogenase molybdenum-iron protein, NifD- or NifE-like	3056
ABC-type probable sulfate transporter, permease protein	3059
Pyruvate formate lyase-activating enzyme	3069
Pyruvate formate lyase-activating enzyme	3070
Ferredoxin	3075
Critical peptidase S16, ATP-dependent protease	3077
HD-GYP hydrolase domain-containing protein	3100
Muconate cycloisomerase-related protein, YKGB <i>B. subtilis</i> ortholog	3102
Glutamyl-tRNA reductase	3107
Hydroxymethylbilane syntase (porphobilinogen deaminase)	3109
Uroporphyrinogen III syntase	3110
Delta-aminolevulinic acid dehydratase (porphobilinogen synthase)	3111
Glutamate-1-semialdehyde aminotransferase	3112
Possible cysteine desulphurase from NiS family	3135
FKBP-type peptidyl-prolyl cis-transisomerase (trigger factor)	3149
Critical ClpX, ATPase regulatory subunit	3151
ATP-dependent Lon protease	3153
Spore cortex protein	3209
Sporulation protein B	3210
Membrane-associated sensory transduction histidine kinase (with HAMP domain)	3255
Response regulator (CheY-like receiver domain and HTH DNA-binding domain)	3256
Hydrogenase expression/formation protein HypE	3277
Fructose-bisphosphate aldolase class I	3310
Beta-xylosidase	3318
Small acid-soluble spore protein SspA	3349
Small acid-soluble spore protein, alpha/beta type	3380
Stage O sporulation protein J, putative	3416
Putative transcription activator Stc	3418
Alcohol dehydrogenase	3419
Putative electron-transfer protein HydG	3420
Alcohol dehydrogenase, iron-containing	3432
Critical small acid-soluble spore protein, alpha/beta type	3461
Probable enoyl-CoA hydratase	3466
Probable enoyl-CoA hydratase	3467
Alcohol dehydrogenase, zinc-containing	3477
Possible stage V sporulation protein, SpoVT	3499
Acyl-CoA dehydrogenase, short-chain specific: butyryl-CoA dehydrogenase	3508
Transaldolase	3637
Acetyl-CoA carboxylase (biotin carboxylase subunit)	3649
Acetyl-CoA carboxylase biotin carboxyl carrier protein	3650
L-lactate dehydrogenase	3682
Phosphoglycerate mutase	3691
Uncharacterized conserved protein YHAD family	3755
L-lactate dehydrogenase	3774
L-serine dehydratase, iron-sulfur-dependent, beta subunit	3775
Beta-glucosidase	3801
Transcriptional regulator of NagC/XylR (ROK) family, sugar kinase	3813
Fructose bisphosphatase	3818
Propionate-CoA transferase	3820
Crotonase	3821
Fructose-1,6-bisphosphate aldolase	3828
Phosphoglucomutase	3831
Accessory regulator protein B	3855
Histidine kinase-like ATPase	3856
Accessory regulatory protein A	3857
Flagellar biosynthesis related protein	3885
Spore coat protein, putative	3889
Critical spore coat protein, CotF-related	3890
Spore coat protein, putative	3891
Critical spore coat protein, CotF-related	3892
(R)-2-hydroxyglutaryl-CoA dehydratase activator-related protein	3926
Glucose kinase	3978
3-Hydroxybutyryl-CoA dehydrogenase	3988
ABC transporter, ATP-binding protein	3993
Aid CoA-acylating aldehyde dehydrogenase	3999
Butyrate-acetoacetate CoA-transferase subunit A	4000

TABLE 1-continued

Gene name	Gene ID
Butyrate-acetoacetate CoA-transferase subunit B	4001
Acetoacetate decarboxylase	4002
ABC-type transport system, ATPase component	4022
Phosphoenolpyruvate synthase/pyruvate phosphate dikinase	4025
Pyruvate water dikinase	4028
Zinc-binding dehydrogenase: alcohol dehydrogenase	4030
Histidine kinase-like ATPase	4032
Response regulator (CheY-like receiver domain and HTH DNA-binding domain)	4033
Short-chain dehydrogenase: 3-oxoacyl-[acyl-carrier protein] reductase	4069
Nitroreductase family protein	4070
Phosphoglycerate mutase	4085
Chemotaxis protein CheW	4116
Alpha-glucosidase	4142
Thioredoxin reductase	4148
Malic enzyme	4150
Anaerobic sulfite reductase subunit B	4154
Anti-anti SigF	4182
Anti-sigma factor F, Stage II sporulation protein AB	4183
Sporulation-specific sigma factor F	4184
Critical SpoVA protein	4185
IMP dehydrogenase/GMP reductase: Stage V sporulation protein AD	4186
Stage V sporulation protein AE, SpoVAE	4187
Spore protease Gpr	4192
Stage II sporulation protein P, SpoIP	4193
Transcriptional regulator of heat shock genes, Hrc A	4198
Molecular chaperone DnaK, Hsp70 family	4200
Molecular chaperones DnaJ, Hsp40 family	4201
Ferrodoxin-nitrite reductase	4202
Stage IV sporulation protein	4211
Spore coat protein S	4220
Predicted dehydrogenase of short-chain alcohol dehydrogenase family	4238
TPR repeats-containing protein	4350
Alpha-galactosidase	4383
Alpha-galactosidase	4384
Thiamine biosynthesis enzyme ThiH	4386
Spore photoproduct lyase SpIB	4463
Melibiase (alpha-galactosidase)	4465
Cysteine synthase/cystathione-beta synthase, CysK	4468
DNA gyrase subunit B	4500
DNA gyrase subunit A	4501
SsDNA exonuclease RecJ	4503
Pyruvate: ferredoxin oxidoreductase	4506
Chemotaxis protein CheW	4513
Chemotaxis protein CheD	4514
Chemotaxis protein CheB, containing CheY-like receiver domain and HTH DNA-binding domain	4515
Chemotaxis protein methyltransferase CheR	4516
Chemotaxis histidine kinase CheA, containing CheW-like adaptor domain	4517
Chemotaxis protein CheC	4518
Chemotaxis signal transduction protein CheW	4520
Flagellar switch protein FliM	4521
Flagellar switch protein FliY, containing CheC-like domain	4522
Flagellar hook-associated protein FlgK	4526
Flagellar hook-associated protein 3	4527
Carbon storage regulator	4529
Flagellar protein FliS	4532
Flagellar cap protein FliD, putative	4533
Possible hook-associated protein, flagellin family	4535
Spore coat polysaccharide biosynthesis protein	4543
FlaG	4544
Chemotaxis motility protein A, MotA	4551
Chemotaxis motility protein B, MotB	4552
Flagellar basal body rod protein FlgB	4553
Flagellar basal body rod protein FlgC	4554
Flagellar assembly protein FliH, putative	4558
Flagellar-type ATPase	4559
Flagellar export protein FliJ	4560
Flagellar hook assembly protein FlgD, putative	4562
Flagellar hook protein flgE	4564
Flagellar protein FlbD	4565
Flagellar basal body-associated protein FliL	4566
Flagellar biosynthesis protein FlpP	4568
Flagellar biosynthesis protein FliQ	4569
Flagellar biosynthesis protein FliA	4571
Flagellar GTP-binding protein FlhF	4572
Sigma factor of SigD/WhiG family	4575
Flagellar basal body rod protein	4578
General secretion pathway protein, pilin family	4608

TABLE 1-continued

Gene name	Gene ID
Ferredoxin	4635
Sulfate adenylate transferase, CysD subfamily	4636
GTPase, sulfate adenylate transferase subunit	4637
HD-GYP domain-containing-protein	4638
Chemotaxis protein CheW	4639
Chemotaxis protein methyltransferase CheR	4642
Chemotaxis protein/ glutamate methylesterase	4643
CheY-like receiver domains, putative	4649
ABC transporter, ATP-binding protein	4656
Hsp 90	4663
Uncharacterized conserved protein	4670
ATP-dependent Clp proteinase	4671
Deoxyribose-phosphate aldolase	4679
HD-GYP hydrolase domain-containing protein	4683
Beta-xylosidase, family 43 glycosyl hydrolase	4696
Hsp 18	4699
Glycerol dehydrogenase	4730
L-lactate dehydrogenase	4749
Pyruvate formate-lyase	4760
Glycerol dehydratase activator	4761
Critical IMP dehydrogenase/GMP reductase	4775
Alcohol dehydrogenase/ acetaldehyde dehydrogenase	4776
2-Oxoacid: ferredoxin oxidoreductase, alpha subunit	4779
3-Oxoacyl-[acyl-carrier-protein] synthase III	4789
Activator of 2-hydroxyglutaryl-CoA dehydratase	4794
Predicted permease	4797
Chemotaxis protein CheY homolog	4801
Chemotaxis protein cheA	4802
Chemotaxis protein Chew	4803
Transcriptional regulator, Lrp family	4811
Critical endopeptidase Clp	4819
3-Oxoacyl-[acyl-carrier protein] synthase	4831
Lactate dehydrogenase	4866
Small acid-soluble spore protein SspC2	4927
L-lactate dehydrogenase	4951
Phosphoglycerate mutase	4961
Alpha-xylosidase	4968
Aldehyde dehydrogenase (NAD ⁺)	4974
Critical bacterial regulatory protein MarR	4976
Topoisomerase I	4983
Acetyl-CoA: acetoacetyl-CoA transferase alpha subunit	4992
Pyruvate kinase, barrel domain	5003
Critical heat shock protein DnaJ, N-terminal domain	5005
Butyryl-CoA dehydrogenase, putative	5011
Oligopeptide transport permease protein	5044

Microarray DNA Probe Labeling and Hybridization.

Two-color microarray hybridization was performed using the aminoallyl labeling procedure adapted from a TIGR protocol (UIUC Functional Genomics Keck Center). Briefly, 3 µg of purified total RNA were primed with random hexamers (Pharmacia) and used as templates for DNA synthesis using aminoallyl dNTPs (Ambion) and Superscript III reverse transcriptase (Invitrogen) in each labeling reaction. The aminoallyl-labeled DNAs were coupled to Cy3 or Cy5 dye esters (Molecular Probes), and oppositely dye-labeled probes were hybridized on an array simultaneously. To compare gene expression in the time course of fermentation, one of the dye-labeled probes was generated from samples collected at individual time points, whereas the other dye-labeled control probe was derived from the RNA pool as described above.

Microarray hybridization was performed using one array for each sample collected in the fermentation time course. Briefly, the slides were rehydrated, UV cross-linked, and pre-hybridized in 5×SSC, 0.1% (w/v) SDS and 1% (w/v) BSA at 42° C. for 45 min. The slides were then hybridized with a mixture of oppositely labeled DNA probes in hybrid-

ization buffer (Ambion) at 42° C. for 16-48 hrs. After hybridization, the slides were washed with 1×SSC and 0.2% (w/v) SDS at 42° C. for 5 min, followed by a second wash in 0.1×SSC and 0.2% (w/v) SDS at room temperature for 5 min, and a last wash in 0.1×SSC for 5 min. The slides were dried and immediately scanned on an Axon 4000B scanner (UIUC Functional Genomics Keck Center). Features in each array were extracted using GenePix Pro 6.0.

Results are depicted in FIG. 3A and FIG. 3B for *C. beijerinckii* NCIMB 8052 and BA101, respectively. Expression level is indicated by intensity of the color bar (green to red) based on log₂ transformation of the normalized expression ratio determined for each gene at individual time point. Temporal expression patterns are visualized with hierarchical clustering for the transition of fermentation cultures from acidogenesis to solventogenesis

Microarray Data Analysis.

Data generated from microarray experiments were processed and visualized using the TM4 suite (TIGR). Briefly, the expression ratio (Cy5/Cy3) for a gene in each sample was determined based on quantification of the fluorescence inten-

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sity for each spot on the array using GenePix Pro 6.0. The expression ratios obtained from all the genes on each array were normalized using Midas (TIGR). LOWESS intensity-based normalization was applied in most cases. Normalized expression ratios for a gene obtained at the analyzed time points were used to construct the temporal profiles of gene expression over the course of fermentation for *C. beijerinckii* NCIMB 8052 and BA101, respectively. Global expression patterns were analyzed by average linkage hierarchical clustering with Euclidean distance matrices and visualized colorimetrically using TMEV (TIGR).

Results for mRNA accumulation levels of various enzymes in the Clostridial solventogenic pathway were quantitatively depicted in FIG. 4. Differential mRNA accumulation of solventogenic genes was compared in *C. beijerinckii* NCIMB 8052 (♦) versus BA101 (○). Increased expression in BA101 during the solventogenic stage was observed for alcohol dehydrogenase (Adh), butyryl-CoA dehydrogenase (Bcd) and butyrate kinase (Buk).

Results for mRNA accumulation levels of various sugar transporters were quantitatively depicted in FIG. 5. Differential mRNA accumulation of sugar transporters was compared in *C. beijerinckii* NCIMB 8052 (♦) and BA101 (○). Components of mannose-family phosphoenolpyruvate (PEP)-dependent phosphotransferase system IIA, IIB (ManIAB) and IIC (ManIIC) were significantly down-regulated in BA101.

Results for mRNA accumulation levels of various sporulation genes were quantitatively depicted in FIG. 6. Differen-

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tial expression of sporulation genes was compared in *C. beijerinckii* NCIMB 8052 (♦) and BA101 (○). Induction of late stage sporulation factors was much weaker in BA101 than in the wild-type strain. Lowered activation in BA101 through the solventogenic phase was observed for coat morphosis sporulation protein (SpoIVa), Stage V sporulation protein B (SpoVB) and small acid-soluble spore protein (SspA).

Results for mRNA accumulation levels of various chemotaxis genes were quantitatively depicted in FIG. 7. Differential expression of chemotaxis genes was compared in *C. beijerinckii* NCIMB 8052 (♦) and BA101 (○). Higher expression levels of CheA, CheC, CheD and CheW in a chemotaxis gene cluster were observed for BA101 during the solventogenic stage.

Results for mRNA accumulation levels of various solventogenic genes were quantitatively depicted in FIG. 8. Solventogenic genes with comparable expression kinetics were compared in *C. beijerinckii* NCIMB 8052 (♦) and BA101 (○). Expression of aceto-acetyl CoA:acetate-butyrat CoA transferase subunit α/β (CtfA/B) and acetoacetate decarboxylase (Adc) was highly activated at the onset of solventogenic phase in BA101 and the wild-type strain. Changes in expression levels were much smaller for thiolase (Thl), 3-hydroxybutyryl-CoA dehydrogenase (Hcd) and crotonase (Crt) in BA101 and the wild-type strain.

Tables 2A and 2B show subsets of genes that were found to be differentially expressed between *C. beijerinckii* NCIMB 8052 and BA101.

TABLE 2 (A)

Genes with increased expression in BA101 compared with the wild-type strain.		
Functional class	Gene name	Gene product activity
Solventogenesis	Alcohol dehydrogenase	Catalyzing the reduction of aldehyde to alcohol
	Butyryl-CoA dehydrogenase	Catalyzing the reduction of crotonyl-CoA to butyryl-CoA
	Butyrate kinase	Catalyzing the generation of butyrate from butyrylphosphate with concurrent ATP synthesis
Chemotaxis	CheA	Chemotaxis sensory transducer, histidine kinase
	CheC	Chemotaxis protein
	CheD	Chemotaxis methylation system protein
	CheW	Chemotaxis protein, histidine kinase

TABLE 2 (B)

Genes with reduced expression in BA101 relative to the wild-type strain.		
Functional class	Gene name	Gene product activity
Sporulation	Coat morphosis sporulation protein SpoIVa	Spore coat assembly
Sugar transporters	Stage V sporulation protein B SpoVB	Spore cortex biosynthesis
	Small acid-soluble spore protein SspA	Packaging and protection of spore DNA
Sugar transporters	Mannose-specific phosphoenolpyruvate-dependent phosphotransferase system component IIAB	Mediating phosphoryl relay for the modification of incoming sugar
	Mannose/fructose-specific phosphoenolpyruvate-dependent phosphotransferase system component IIC	Mediating sugar transport across the membrane through permease

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Example 5

General Methods Used in the Examples

PCR primers are designed using the PrimerSelect features of the DNASTAR suite of molecular biology programs from DNASTar, Inc. (Madison, Wis.). Techniques of primer design are known in the art (PCR Primer Design, 2007, Anton Yuryev editor, Humana Press).

PCR products are amplified using Takara Ex TaqTM DNA Polymerase from Takara Bio USA (Madison, Wis.), and a Gene Amp[®] PCR system 9700 thermocycler from Applied Biosystems (Foster City, Calif.). Other DNA polymerase products for PCR provide suitable alternatives. Cycling parameters can vary according to the specific primers and DNA sequences being amplified. In general the methods and parameters are known in the art. (PCR Protocols, 2nd edition, 2003, John M. S. Bartlett and David Stirling editors, Humana Press; PCR: The Basics, 2nd edition, 2006, M. J. McPherson and S. G. Moller, Taylor & Francis publisher).

For colony PCR, fresh colonies are picked from Petri plates and suspended in a 50-100 µL of ultrapure water or 10 mM Tris, pH 7.5. 1-5 µL of the cell suspension is substituted for the purified DNA in a normal PCR reaction mixture. The initial PCR heat cycle of the process may be extended in some cases, for example 10 min at 94° C., to aid in cell lysis.

The isolation and purification of plasmid DNA, chromosomal DNA, DNA fragments from preparative agarose gels and PCR products is accomplished using commercial kits that are available from various suppliers. Examples of two such suppliers are Qiagen Inc. (Valencia, Calif.) and MO BIO Laboratories (Carlsbad, Calif.). Examples of Qiagen kits for some applications are "QIAprep" for plasmid DNA, "QIAquick[®]" for purifying DNA fragments from agarose gels, and "QIAquick[®]" or "MinElute[®]" for purifying PCR products. Chromosomal DNA preparations (genomic DNA) are prepared using the "UltraClean Soil DNA Isolation" kit from MO BIO Laboratories.

For introduction of DNA into *Clostridium* hosts by electroporation (transformation), a culture of the *Clostridium* strain is grown to an OD₆₀₀ of 0.8, then washed for two cycles with 15% polyethylene glycol (PEG). Electroporation is done in the presence of 10 µg of plasmid DNA using a cuvette with a 2 mm path in a Bio-Rad Gene PulserTM exponential decay generator set (BioRad, Richmond, Calif.) for 2.0 kV (10 kV/cm), 200 ohms and 4.5 ms. Electroporation parameters may vary from strain to strain. Those skilled in the art will be capable of adjusting parameters as needed (Molecular Cloning: A laboratory manual, 3rd edition, 2001, Joseph Sambrook and David W. Russell, Cold Spring Harbor Laboratory Press; Handbook on Clostridia, 2005, Peter Durre editor, Taylor & Francis publisher).

General cloning methods such as use of restriction endonucleases, DNA ligase and other nucleic acid modification techniques, separative techniques such as agarose or polyacrylamide gel electrophoresis, and the like are known in the art and comprehensive guides are available (Methods for General and Molecular Microbiology, 3rd edition, 2007, C. A. Reddy editor in chief, ASM Press; Molecular Genetics of Bacteria, 2nd edition, 2003, Larry Snyder and Wendy Champness, ASM Press; Molecular Cloning: A laboratory manual, 3rd edition, 2001, Joseph Sambrook and David W. Russell, Cold Spring Harbor Laboratory Press).

Example 6

Construction of Strains of Solventogenic Clostridia wherein spoIVa Gene Expression is Deficient

A mutant derivative of *Clostridium beijerinckii* strain NCIMB 8052 is constructed wherein the function of the

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spoIVa gene encoded by SEQ ID NO: 9 (locus_tag Cbei_1136 of GenBank CP000721) is destroyed by insertion of a plasmid bearing a cloned fragment of the spoIVa gene DNA into the chromosome, so as to disrupt the coding sequence of the gene. Insertion of the plasmid into the chromosome takes place by single-cross-over homologous recombination between the chromosomal spoIVa gene and the cloned spoIVa fragment.

A spore suspension of *Clostridium beijerinckii* strain NCIMB 8052 is heat shocked for 10 minutes at 80° C., placed on ice briefly, moved into a Coy[®] anaerobic chamber (Coy Laboratory Products, Grass Lake, Mich.) containing an atmosphere of 85% N₂, 10% CO₂ and 5% H₂, and then used to inoculate 10 mL of TGY medium in an 18 mm diameter test tube. The culture is grown at 35° C. to an OD₆₀₀ of about 0.6 to 0.8. A 1.0 mL portion of this culture is used to inoculate another 10 mL of TGY, which is grown to about 0.6 OD₆₀₀, or to a density that yields good chromosomal DNA preparations. The culture is then harvested and processed to prepare purified chromosomal DNA using the "UltraCleanTM Soil DNA Isolation" kit and protocols from MO BIO Laboratories.

PCR primers incorporating terminal XmaI restriction endonuclease sites are designed using the PrimerSelectTM software package of DNASTAR Inc. (Madison, Wis.) so as to amplify an internal fragment of the spoIVa gene of preferably 250-600 bp in length, ideally in the central part of the coding region of the gene; for example, the 3' one-third of the gene preferably is avoided to prevent partially functioning spoIVa gene product in the resulting mutants.

The chosen internal fragment of the spoIVa gene is amplified by the PCR reaction using the purified chromosomal DNA preparation and the chosen PCR primers. The amplified spoIVa internal fragment with the terminal XmaI sites is purified from the finished PCR mixture using Qiagen MinElute[®] spin columns or a similar product. Alternatively the fragment could be separated using a preparative agarose gel and purified from a gel slice using Qiagen Qiaquick[®] kits. The purified spoIVa fragment is restriction digested with XmaI to generate cohesive ends, and reisolated from an agarose gel.

Plasmid pAK102 (A Y Kim and H P Blaschek, 1993, J. Bacteriol. 175: 3838-43) was constructed by ligation of HindIII-linearized plasmid pUC19 and a 2.3-kb HindIII erythromycin resistance gene fragment from plasmid pVA677 (F L Macrina et. Al, 1980, J. Bacteriol. 143: 1425-1435). pAK102 encodes resistance to ampicillin and erythromycin, and replicates autonomously in *E. coli* but not in *Clostridium* species; thus in *Clostridium*, pAK102 is a "suicide vector." Plasmids of equivalent function could be prepared from common *E. coli* vectors and common sources of the erythromycin resistance gene functional in *Clostridium* (Methods for General and Molecular Microbiology, 3rd edition, 2007, C. A. Reddy editor in chief, ASM Press; Clostridia, 1989, Nigel P. Minton and David J. Clarke editors, Plenum Press). Plasmid pAK102 DNA is purified from a transformed *E. coli* DH5alpha host that is routinely grown under 50 µg/mL of ampicillin selection, using a Qiagen Qiaprep[®] kit. The pAK102 DNA is linearized by digestion with XmaI and the purified internal fragment of the spoIVa gene is cloned into the vector using DNA ligase.

The ligation mixture is electroporated into *E. coli* DH5alpha and transformants are recovered by growth on LB agar petri plates as colonies that are resistant to 50-100 µg/mL of ampicillin. The transformants are screened to determine the size of the *Clostridium* fragment inserted into the plasmid. To do this, colony PCR is performed using the same primers that were used above, and PCR reaction products are sepa-

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rated by electrophoresis on 1% to 1.5% agarose gels. Transformants that show only the expected fragment size, and not multiples of that size, are selected for the next step and are labeled "pAK102/spoIV".

Plasmid pAK102/spoIV DNA is purified from the chosen *E. coli* transformant, using a Qiagen Qiaprep® kit. The plasmid DNA is used to transform strain *C. beijerinckii* NCIMB 8052 by electroporation. Transformants are initially allowed to recover by growth in TGY medium without antibiotic selection for 3 hours at 35°C., then spread on TGY-1.5% agar plate medium containing 25 µg/mL of erythromycin. Alternatively, erythromycin concentrations as low as 10 µg/mL might be considered for the initial selective plates. Following their initial recovery, erythromycin resistant strains are propagated in the presence of 10-40 µg/L of erythromycin. Because the pAK102 vector is incapable of independent replication in *Clostridium* species, transformants are expected to retain antibiotic resistance by virtue of having integrated the pAK102/spoIV construct into the chromosome, at a site bounded by the endpoints of the cloned spoIV fragment. The proper insertion of the plasmid, and its position within the spoIV gene is verified by DNA sequencing of spoIV gene target region.

The resulting strains, which are mutants of *C. beijerinckii* NCIMB 8052 having disrupted or impaired spoIV function, are tested in fermentations for solvent formation in P2 medium as in Example 1, except that 10-25 µg/L of erythromycin is added to the fermentation medium for every 24 hours of elapsed culture time. The preservation and routine propagation of the spoIV mutant strains in the lab, as well as other strains that may be defective in the formation of normal spores, may require the making and use of frozen cultures of vegetative cells in medium containing 15% glycerol, or 0.1% DMSO, or other cryoprotectives. Such methods are known to those who are skilled in the art (Methods for General and Molecular Microbiology, 3rd edition, 2007, C.A. Reddy (editor in chief), ASM Press) and could be used if necessary to prevent the emergence of degenerated strains by excessive serial propagation over time.

In the general manner of this example, derivatives of NCIMB 8052, or BA101 or other solventogenic *Clostridium* species and strains, are constructed having mutations in other genes that are targeted for various degrees of disrupted function; for instance mutants bearing defective spoVB, sspA, manIIB or manIIC genes or their close homologs, or where expression of the normal gene is driven by reduced-strength promoters. In the case of *Clostridium* species having active restriction-modification systems, such as for example *C. acetobutylicum* ATCC 824 and other strains, steps to overcome the transformation barrier imposed by the restriction systems are added to the above protocol. Typically these involve prior methylation of the transforming DNA by various in vitro DNA methylation reactions, or by propagation of the DNA/vector in hosts that methylate the DNA but do not restrict it. Procedures for such modification are common in the research literature of solventogenic clostridia (Handbook on *Clostridia*, 2005, Peter Durre (editor), Taylor & Francis publisher).

Example 7

Construction of Solventogenic Clostridia Engineered for Constitutive Expression of the adh Gene at High Levels from a Heterologous Promoter

A derivative of *Clostridium beijerinckii* strain NCIMB 8052 or BA101 is constructed whereby the NCIMB 8052 adh

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gene (SEQ ID NO: 1, Cbei_2181 of GenBank CP000721) is constitutively expressed at increased levels by a combination of transcription from the promoter of the ferredoxin gene of *Clostridium pasteurianum* ATCC 6013, and by gene amplification on a replicative multicopy plasmid.

Plasmid pMTL500E is a multicopy *E. coli/Clostridium* shuttle vector that encodes erythromycin resistance and which is stably maintained in *Clostridium* strains including *C. beijerinckii* 8052 (AM López-Contreras, et. al., 2001, 10 *Clostridium beijerinckii* cells expressing *Neocallimastix patriciarum* glycoside hydrolases show enhanced lichenan utilization and solvent production, Appl Environ Microbiol. 67: 5127-33; A Y Kim, et. al., Heterologous expression of endo-beta-1,4-D-glucanase from *Clostridium cellulovorans* 15 in *Clostridium acetobutylicum* ATCC 824 following transformation of the engB gene, 1994, Appl Environ Microbiol. 60: 337-40; Handbook on Clostridia, 2005, Peter Durre editor, Taylor & Francis publisher).

The promoter and ribosome binding site (RBS) from the 20 ferredoxin gene (fd) from *Clostridium pasteurianum* ATCC 6013 (GenBank accession number M11214) has been shown to be capable of driving the constitutive expression of heterologous genes to very high levels in multiple *Clostridium* species, including *C. beijerinckii* strain NCIMB 8052; (M C 25 Graves and J C Rabinowitz, 1986, In vivo and in vitro transcription of the *Clostridium pasteurianum* ferredoxin gene. Evidence for "extended" promoter elements in gram-positive organisms, J Biol. Chem. 1986 261: 11409-15; Minton N P, et. al., 1995, Chemotherapeutic tumour targeting using 30 clostridial spores, FEMS Microbiol Rev. 17: 357-64; U.S. Pat. No. 6,652,849 (2003)).

To begin, plasmid pMTL500E DNA is linearized with restriction endonuclease XmaI. Alternatively, another restriction site within the multiple cloning site (MCS) of the vector 35 could also be used, provided XmaI in the remainder of the example is also replaced by that restriction enzyme.

A DNA fragment carrying the fd promoter and RBS sequences is prepared by oligonucleotide synthesis using the published DNA sequence for the fd promoter and RBS binding region (GenBank accession number M11214), starting at the 5' end from the first base of the source sequence (-168 relative to the fd gene start codon) but incorporating an XmaI site upstream of that, and replacing the sequence "TTCATG" with "CATATG" (an NdeI site) where "ATG" is the ferredoxin 40 gene start codon, and terminating at the 3' end with any string of non-homologous bases. Alternatively an fd promoter/RBS 45 fragment featuring the same subterminal restriction sites could be prepared by PCR amplification from *Clostridium pasteurianum* ATCC 6013 chromosomal DNA template. The complete adh gene from *C. beijerinckii* strain NCIMB 8052 50 chromosomal DNA template is amplified by PCR using a forward primer that includes a subterminal NdeI site, wherein the "ATG" of the NdeI site is also the ATG start codon for the adh gene, and where the reverse primer includes a subterminal 55 XmaI site. It should be noted that in this example, and in Example 8 and other examples incorporating this promoter replacement tactic, that there are alternative restriction recognition sites incorporating ATC sequences that could be chosen for the promoter-RBS-gene fusion, for example 60 restriction endonucleases Nb.BsrDI or BsrDI.

The synthesized fd promoter/RBS fragment and the PCR-ed adh gene fragment are purified, then digested with NdeI and ligated together, creating a "fd promoter/RBS/adh gene" 65 fragment having subterminal XmaI sites. This is digested with XmaI and ligated into the linearized pMTL500E plasmid. The reaction products are used to transform *E. coli* DH5alpha. Ampicillin resistant colonies are selected and the

transformant colonies are screened by DNA sequencing to confirm the presence of the correct "fd promoter-RBS-adh gene" insert. The new plasmid is purified from the *E. coli* transformant and is used to electroporate *C. beijerinckii* strain NCIMB 8052 or BA101. Erythromycin resistant transformant colonies are recovered as in Example 6.

Alternatively, the plasmid pMTL500F, which already has the fd promoter sequence positioned upstream of an MCS (page 141, Chapter 6, in *The Clostridia and Biotechnology*, 1993, D. R. Woods editor, Butterworth-Heinemann), could be adapted as the cloning vector for the adh gene provided that the details of the method preserve a functioning RBS for expression of the cloned adh gene.

The resulting strains express adh constitutively due to the use of the heterologous fd promoter, and due to gene amplification on the multicopy vector. The expression of adh in the new strains is confirmed to be constitutive, and is quantitated by enzyme assay. The new strains are tested in fermentations for solvent formation as in Example 1, including the addition of erythromycin to the fermentation medium for every 24 hours of elapsed culture time.

Other promoters for constitutive gene expression are known in the art and would be suitable for use in this example; for instance, the ptb (phosphobutyl transferase) gene promoter from *C. acetobutylicum* has been used to drive constitutive expression of the LacI in several *Clostridium* species—sufficient to suppress the fd promoter when under control of the LacZ operator (J T Heap, et. al., 2007, *The ClosTron: a universal gene knock-out system for the genus Clostridium*, *J Microbiol Methods* 70: 452-64). Consequently, if tuning of the level of expression of the adh gene or other cloned genes is required to achieve the best result, other promoters can be tried as a means of achieving that end.

If further tuning of the expression level of the cloned adh is found to be required, the method of cloning the gene is repeated with minor modifications to the DNA sequence of the RBS site, so as to alter the efficiency of ribosome binding and the level of functional gene product in the cell. (See page 167, *The Clostridia and Biotechnology*, 1993, D. R. Woods editor, Butterworth-Heinemann).

The following shows the DNA sequence in the RBS region of the native fd and adh genes, where the upper-case letters are the start codons of the genes and the Shine-Dalgarno sequences of the RBS region are underlined. Tuning of the expression level of the cloned genes is accomplished by altering either the sequence in the underlined regions, and the spacing between those regions and the ATG start codon.

adh ttttaggaggaa atat_{TTT} ATG (SEQ ID NO: 20)

fd tttaaggaggtgtattttcATG (SEQ ID NO: 21)

fd-adh tttaaggaggtgtatttcatATG SEQ ID NO: 22)
(new)

In the general manner of this example, derivatives of *C. beijerinckii* strain NCIMB 8052, or BA101 or other solventogenic *Clostridium* species and strains, are constructed having an increased level of expression, or constitutive expression of other genes and their homologs, for instance the bcd, buk, cheA, cheC and cheD genes. In the case of *Clostridium* species having active restriction-modification systems, such as for example *C. acetobutylicum* ATCC 824 and other strains, steps to overcome the transformation barrier imposed by the restriction systems are added to the above protocol as in example 6.

Construction of Solventogenic Clostridia Engineered
for Constitutive Expression of the adh Gene in
Single Copy Number from a Heterologous Promoter

The methods of Example 6 and Example 7 can be combined and modified to achieve constitutive expression of the adh gene, at a level that is lower than expression from a multicopy plasmid. This is achieved by integrating the fd promoter-RBS-adh gene construct into the chromosome of the *Clostridium* host. The expression level of the adh gene may be higher than the untransformed parent strain, or it may be lower than the untransformed parent strain, depending upon the native level of expression of the adh gene in the untransformed strain, and upon modifications to the fd promoter and RBS sequences of the engineered strain.

DNA of plasmid pAK102 DNA is prepared and linearized by digestion with XmaI as in Example 6.

A DNA fragment carrying the fd promoter and RBS sequences, engineered at the ATG start codon to contain an NdeI site, is constructed as in Example 7.

A fragment of the adh gene from *C. beijerinckii* strain NCIMB 8052 (SEQ ID NO: 1, Cbei_2181 of GenBank CP000721), consisting of the 5' one-third to one-half of the gene, is generated by PCR amplification from chromosomal DNA, incorporating the 5' NdeI site and 3' XmaI site as in Example 7.

The fd-RBS fragment is ligated to the adh fragment at their NdeI sites, and then the fd-RBS-adh fragment is inserted by ligation into the XmaI site of plasmid pAK102. The new plasmid construct is recovered and verified, and then electroporated into *Clostridium beijerinckii* NCIMB 8052 or BA101 hosts and selected by erythromycin resistance as in Example 6. The resulting erythromycin resistant transformants are single-cross-over products between the cloned adh 5' fragment on the plasmid, and the adh gene on the chromosome. The structure of the expected construct, in order from 5' to 3' of the top strand of the genome sequence, would be as shown below.

5'-partial adh gene—pAK102 vector—fd promoter—RBS—complete adh gene—3'

The erythromycin resistant transformants are checked by DNA sequencing to verify the expected structure.

The isolated new strains are maintained under erythromycin selection to prevent reversion by homologous crossing-out of the plasmid. The strains are assayed for constitutive expression of adh enzyme, and for levels of solvent and acid formation in batch fermentation experiments. Due to its presence in single copy number, the level of expression of adh would be expected to be less than the strains of Example 7. As in example 7, further tuning of cellular levels of the Adh enzyme could be accomplished by varying the constitutive promoter that is used (for example, ptb) or by changing the sequence of the RBS region of the construct.

In the general manner of this example, derivatives of *Clostridium beijerinckii* NCIMB 8052, or BA101 or other solventogenic *Clostridium* species and strains, are constructed having various levels of constitutive expression of other genes and their homologs, for instance the bcd, buk, cheA, cheC and cheD genes. In the case of *Clostridium* species having active restriction-modification systems, such as for example *C. acetobutylicum* ATCC 824 and other strains, steps to overcome the transformation barrier imposed by the restriction systems are added to the above protocol as in example 6.

Construction of Solventogenic Clostridia Engineered
for Reduced Expression of the *sspA* Gene Relative
to the Untransformed Strain

Constitutive expression from the heterologous fd promoter, driving the expression of a single copy of a gene as taught in Example 8, can be modified to adjust the level of expression of the engineered gene. Such modification also could be in the direction of lowered expression relative to the untransformed host. This is advantageous in the instance where reduced gene expression is beneficial to solvent formation, but where complete elimination of gene expression produces undesirable effects.

By introducing changes to the DNA sequence of the fd promoter, the level of transcription of the gene is reduced leading to a reduction in mRNA levels for the gene in the cell and lower levels of functional gene product. By altering the DNA sequence corresponding to the RBS and the spacing between the RBS and the ATG start codon of the gene, the level of translation of the mRNA can be reduced, also leading to accumulation of less functional gene product in the cell. A combination of the mRNA reduction and translation reduction could lead any degree of reduction of gene expression without producing a full “knockout” affect.

DNA of plasmid pAK102 DNA is prepared and linearized by digestion with XmaI as in Example 6.

The 5' one-half of the *sspA* gene from strain *Clostridium beijerinckii* NCIMB 8052 (SEQ ID NO: 11, Cbei_3080 of GenBank CP000721), is PCR-amplified from chromosomal DNA template, using a primer design that incorporates a 5' NdeI site and a 3' XmaI site as in Example 7. Being that *sspA* is a short gene (210 bases), if suitable primers cannot be found, then a Clal restriction site that exists near the middle of the gene is used to cleave the PCR amplification product and the 5' half of the *sspA* gene is purified from an agarose gel.

A DNA fragment carrying the fd promoter and RBS sequences, engineered at the ATG start codon to contain an NdeI site, is synthesized as in Example 7, including the creation of the 5' XmaI and 3' NdeI sites, except that instead of a single DNA sequence, a collection of oligonucleotide species is produced having various nucleotide base changes in the fd promoter and RBS sites.

The fd promoter of *C. pasteurianum* ATCC 6013 (GenBank M11214) has been characterized. It displays “minus-10” and “minus-35” sequences that are not unlike those described for normal promoters of other gram-positive bacteria (M C Graves and J C Rabinowitz, 1986, J Biol. Chem. 1986 261: 11409-15; page 287, The Clostridia and Biotechnology, 1993, D. R. Woods editor, Butterworth-Heinemann). In particular, base changes introduced in the regions of minus-75 to minus-67, and minus-57 to minus-46 relative to the ATG start codon of the fd gene could impact promoter strength. Changes made to the RBS site at bases minus-17 to minus-11 alter the efficiency of translation of mRNA to protein. These bases are underlined in the DNA sequence below, which shows the fd promoter and RBS region of the oligonucleotide to be synthesized (the “atg” start codon is shown in lower-case). By introducing one or several different changes in the underlined regions in the sequence of each fd-RBS DNA oligo that is synthesized, a mixture of oligonucleotides bearing different mutations in the region is produced.

(SEQ ID NO: 23)
5 ' _TTTAAAAAGTTAAAAACATGATAACAATAAGTTATGGTAAACTTATG
ATTAAAATTTAAGGAGGTGTATTCATatg_3 '

5 The mixture of synthesized fd-RBS fragments bearing the different mutations is ligated to the *sspA* fragment at their NdeI sites, and then the fd-RBS-*sspA* fragment is ligated into the XmaI site of plasmid pAK102. In the case of using Clal to generate the *sspA* fragment, a blunt end ligation is done to close the plasmid. The new plasmid construct is recovered and verified, and then electroporated into *Clostridium beijerinckii* NCIMB 8052 or BA101 hosts and selected by erythromycin resistance as in Example 6. The resulting erythromycin resistant transformants are single-cross-over products between the cloned *sspA* 5' fragment on the plasmid, and the *sspA* gene on the chromosome. The isolated new strains are maintained under erythromycin selection to prevent reversion by homologous crossing-out of the plasmid. As alluded to in Example 6, maintenance of the culture using techniques other than spore propagation, such as frozen glycerol stocks of vegetative cells, might be necessary for some isolates.

20 The total collection of erythromycin-resistant isolates would comprise a collection of strains showing varying levels of expression of the *sspA* gene. The isolates are screened in fermentations for their ability to produce more solvent or produce solvent more efficiently, or faster, as in Example 6. Candidates that show improved solvent forming properties or other desirable phenotypes are further characterized to determine the location of the inserted DNA in the chromosome, and the extent of expression of the *sspA* gene at both the transcriptional level (abundance of mRNA) and translational level (abundance of SspA protein), and to characterize the sporulation and morphological properties of the new strains.

25 In the general manner of this example, derivatives of *Clostridium beijerinckii* NCIMB 8052, or BA101 or other solventogenic *Clostridium* species and strains, are constructed having a reduced level of expression of genes that are targeted for various degrees of reduction; for instance 30 mutants showing reduced expression of *spoIVa*, *spoVB*, *manIIAB* or *manIIC* genes or their homologs. In the case of *Clostridium* species having active restriction-modification systems, such as for example *C. acetobutylicum* ATCC 824 35 and other strains, steps to overcome the transformation barrier imposed by the restriction systems are added to the above protocol as in example 6.

Example 10

Construction of Solventogenic Clostridia Engineered
for Inducible Expression of the *bcd* Gene

Plasmid pMTL5401F is a *Clostridium/E. coli* shuttle vector designed for inducible expression of cloned genes (JT 55 Heap, et. al., 2007, The ClosTron: a universal gene knock-out system for the genus *Clostridium*, J Microbiol Methods.: 452-64). For the purpose of this example its essential elements are the ferredoxin gene fd promoter fused to the operator of the lacZ operon (the promoter/operator combination is 60 called “fac”), the lacI repressor gene under the control of the *C. acetobutylicum* ptb gene promoter, plasmid replication functions for *E. coli* and *Clostridium* hosts, and ampicillin and erythromycin resistance genes for selection in *E. coli* and *Clostridium* hosts. In this system the LacZ gene product 65 represses transcription initiation at the fd promoter due to the close proximity of the lac operator to the fd promoter. In the presence of the lactose analog IPTG (isopropyl-beta-D-

thiogalactopyranoside), the LacI repressor fails to bind its operator and the fd promoter can then function. In this system, genes cloned downstream of the plasmid fac promoter/operator are repressed until IPTG is added to the system, at which time the promoter is induced and the gene is expressed.

Plasmid pMTL5401F can be used for the inducible expression of the bcd gene (SEQ ID NO:2, Cbei_2035 from GenBank CP000721). To do this a DNA fragment bearing the full-length bcd gene and including about 25 bases upstream of the bcd gene (to include the gene's RBS site, but no more), and having terminating restriction sites to control the length of DNA upstream and downstream of the gene, is prepared by PCR amplification from *C. beijerinckii* strain NCIMB 8052 or strain BA101 chromosomal DNA template. This fragment is then inserted into the linearized pMTL5401F vector so as to bring the bcd gene and its RBS under the control of the fac promoter. Plasmid clones having the proper structure are then recovered and confirmed as in Example 7, and are labeled plasmid "pfac-bcd."

C. beijerinckii strain NCIMB 8052 or BA101 is then transformed with pfac-bcd by electroporation and erythromycin resistant colonies are selected as in the other examples, and are maintained under erythromycin selection. Transformants are verified by DNA sequencing and then tested for levels of bcd enzyme expression, and for solvent production in batch fermentations at various timepoints before and after induction of bcd gene expression by addition of IPTG to the culture. IPTG could be tried in the concentration range of 0.5 mM to 2 mM, but higher concentrations could be tried if required.

As an alternative to the LacI/lacZ operator system, other inducible promoter/operator systems for use in *Clostridium* species have also been described and shown to function, for instance the adaptation of the xylose-inducible system from *Staphylococcus xylosus* for use in *C. acetobutylicum* (L. Girbal, et. al., 2003, Development of a sensitive gene expression reporter system and an inducible promoter-repressor system for *Clostridium acetobutylicum*, Appl Environ Microbiol.: 4985-8). To use this system the xylA promoter-operator sequence is PCR amplified from chromosomal DNA of *S. xylosus* strains DSM 20267. This could be cloned into an appropriate vector, such as pMTL500E, or low-copy number derivatives of the same replicon such as pMTL502E (page 45, Handbook on Clostridia, 2005, Peter Durre editor, Taylor & Francis publisher) and used for xylose-inducible expression of cloned genes for solvent production.

Alternatively, gene expression microarrays could be used to search entire *Clostridium* genomes for promoters matching certain desired expression characteristics, including constitutive promoters, promoters of various strength for low-level, intermediate-level or high-level expression of genes, promoters responding to specific external factors such as chemical compounds that are added or that are present in fermentation substrates, or promoters that follow certain desirable temporal patterns of transcription initiation in the specific fermentation process that is being developed. To accomplish this, high-density microarrays representing entire genomes at high resolution would be prepared; for example arrays supplied by Roche NimbleGen, Inc. could be used. Messenger RNA to be amplified for final interrogation of the arrays would be isolated from cultures of *Clostridium beijerinckii* NCIMB 8052 or BA101, or other *Clostridium* strains, under multiple different conditions, the exact conditions depending on the promoter-control objectives of the work. A time-course of the culture could be used to discover promoters that show a temporal pattern of activity. Promoters that respond to specific added inducers, for example xylose or arabinose, or furfural or HMF, etc., could be discovered by comparing

samples prepared before and after addition of those substances. Constitutive promoters would be those that show relatively little change in activity in time-course experiments or in response to other challenges. The specific promoters of interest would be discovered by the pattern of expression of the genes downstream of the promoters; in other words, one would analyze the microarray data to find specific genes which expression reflects the desired patterns, then clone regions upstream of those genes, or operons in the case of apparent co-transcription of contiguous genes, to discover the exact promoter that displays the wanted characteristics.

In the general manner of this example, derivatives of *C. beijerinckii* strain NCIMB 8052, or BA101 or other solventogenic *Clostridium* species and strains, are constructed having inducible expression of other genes and their homologs, for instance the adh, buk, cheA, cheC and cheD genes. In the case of *Clostridium* species having active restriction-modification systems, such as for example *C. acetobutylicum* ATCC 824 and other strains, steps to overcome the transformation barrier imposed by the restriction systems are added to the above protocol as in example 6.

Example 11

Construction of *E. coli* Strains Engineered to Express the adh Gene from a Solventogenic *Clostridium*

Strains of *E. coli* are constructed that constitutively express the adh gene of *C. beijerinckii* strain NCIMB 8052 from a strong constitutive promoter. The strains are constructed by insertion of an *E. coli* promoter-*Clostridium* adh gene construct into the lacZ gene in the chromosome of *E. coli*, from a linear DNA fragment, by double crossover recombination into lacZ.

The Ptac and Ptrc promoters are constitutive synthetic promoters that are often used for engineered expression of genes in *E. coli* (Herman A. De Boer, et al., The tac promoter: a functional hybrid derived from the trp and lac promoters. Proc. Natl. Acad. Sci. 1983).

The adh gene (SEQ ID NO:1, Cbei_2181 of GenBank CP000721) is amplified by PCR using chromosomal DNA from *C. beijerinckii* NCIMB 8052 as template. The forward PCR primer is designed so as to incorporate the Ptac promoter or the Ptrc promoter sequence and the lacZ ribosome binding site, properly positioned in relation to the ATG start codon of the *Clostridium* adh gene to support expression of adh in *E. coli*. The reverse primer is designed with a terminal HindIII restriction site to facilitating its subsequent ligation to a tetracycline resistance gene fragment (Tet^R) from pBR322 or a related vector.

A DNA fragment bearing the HindIII site and Tet^R gene and its native promoter is prepared by PCR amplification from a suitable vector, such as pBR322 for example.

The Ptac-adh and Tet^R fragments are ligated together at their HindIII sites, yielding a linear DNA fragment carrying the Tet^R gene and the adh gene, with the genes oriented for divergent transcription. The Tet^R-Ptac-adh or Tet^R-Ptrc-adh fragment is ligated into a pGEM-T vector (Promega Corporation), disrupting the lacZ sequence of that vector.

Using a Ptac promoter construct as an example, the pGEM-T lacZ::Ptac-adh-Tet^R plasmid is then linearized and electroporated into an *E. coli* recB recC sbcB host, which supports transformation and recombination with linear DNA molecules (Winans, S. C., Elledge, S. J., Krueger, J. H. & Walker, G. C., 1985, *J. Bacteriol.* 161: 1219-1221). A double crossover recombination event between the linearized plas-

mid and the lacZ gene of the *E. coli* chromosome results in insertional inactivation of the host lacZ gene, and a Tet^R lacZ phenotype for transformants. Stable transformants are selected with tetracycline, and tested for the lacZ phenotype, and for insertion of the expected structure into the chromosome by DNA sequencing.

In the general manner of this example, other solvent pathway genes from various *Clostridium* species and strains, for instance the bcd, buk, cheA, cheC and cheD genes, could be cloned and expressed in *E. coli* hosts.

All publications and patent applications cited in this specification are herein incorporated by reference as if each individual publication or patent application were specifically and individually indicated to be incorporated by reference.

Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be readily apparent to those of ordinary skill in the art in light of the teachings of this invention that certain changes and modifications may be made thereto without departing from the spirit or scope of the appended claims.

SEQUENCE LISTING

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attgctgttc cttagcagt	tgccaggat	ttatataacaa ttatgttgc	360
acagcggtcg tacattttat	ggatgctgt	gctaaagaag gaaatcttag	420
atgtggcaaa tcgctgttat	ttgtctacaa	ggtatacgtt ttgcgattcc	480
gtattagcaa tcgggtcagg	tccttattgt	tcattacttg ctgctatgcc	540
actgggtgtt tagcaatttg	tgggtggatg	gtttagtgc ttggttatgc	600
aacatgtatgg ctacaaaaga	agtagtgccaa	ttcttcgcaaa ttggtttgtt	660
gtttcacaaa ttacacttat	cggacttggt	gcaataggtt tagctttac	720
ttagcactta gcaaaacaagg	tggctcaggat	aatgggtggaa attcaaatac	780
ttagggatc taatagatag	atactaa		807

<210> SEQ ID NO 9

<211> LENGTH: 1494

<212> TYPE: DNA

<213> ORGANISM: Clostridium beijerinckii

<400> SEQUENCE: 9

atgggaggaga aaaaatgttca	caattttaac	atatacaaaat atatagccga	tagaacccaa	60
ggggacatat atgttaggat	cgttaggatca	gttaggacag ggaaatctac	atttattaaa	120
aagttcatgg atctttaggt	aatacctaaa	attgataata cttataaaaa	ggaaagagca	180
aaagatgagt taccacagag	tggctcaggaa	aaaaatattt acacaacaga	acctaaattt	240
gtacctaattt aagcgttgc	aataaatttta	ggggatgaaa taaaattttaa	agtaaggatg	300
gttgactgcg taggatataat	tgtaaaagga	gcatttaggat	atcttagatgg tgaggaaact	360
aaaatgggttc atacaccatg	gtatgattac	gaaatacctt ttgaggatgc	agcagaaattt	420
gggacaagaa aagttataaa	agaccattca	acaataggat	tagtattac aactgtatgg	480
agtataactg gaatagatag	aatgactat	gtagaaccag	aagaaagagt aatataatgaa	540
ttacagtcac taaataaacc	atttatttgc	gttcttaata	ctaataagcc taattctcaa	600

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gaaaactaagg cattaaaaaa agaatttagaa gataaatata atgttaactgt tcaagttatg	660
gatgtttata atatggaga acaagatatt gaggatttat ttaaacacgt acttaaggaa	720
ttccctgtaa aagaataaaa tattgatatg cctgaatggt tagaaaaatt ggaatgtAAC	780
cattggctaa aaaaagattt tttcaacata attatgaaca tgagtcaaga tgtctctaag	840
gtaagagaca taaaatactg cttaaatgtat tttgaaaatg aagactttat gggaaaggca	900
accataaaatg aagttgattt gggttagtgg aacagaaaaa tatctatgaa acctaaggat	960
ggaattttct ataaggttct tagtgagata tgtaatttag aagttggttc agaaaagtgc	1020
ttgctcagca taataaatga cctaaatgtat gcaaaatgtg aatatgataa ggttaaagat	1080
gcatttggaaag atgttaagaga gagcggctat ggattggttg caccacaact ttccggaaatg	1140
aagtttgagg agcctgaaat agttaaaca ggtataaaat atggagttaa attaaaggca	1200
agtgcctcaa gcttacatct tataaagtgt gatattaaaa ctgagataag ccctataatg	1260
ggatcagaaa aggaatcgga agaacttagta aaaggccttc tagaacaatt tgaaaacagat	1320
ccagccottc tttggcaaag taatatgttt ggaaaatott tagaagtgtt agttaaagaa	1380
ggattacaaa acaagctata taaaatgcca gaagatgttc aagttaagat tc当地agact	1440
ttgcagaaaa ttattatga aggttaatggt ggattaatct gtataatact ttAA	1494

<210> SEQ ID NO 10

<211> LENGTH: 1536

<212> TYPE: DNA

<213> ORGANISM: Clostridium beijerinckii

<400> SEQUENCE: 10

atggaaaaac aatccctaat aagaggggc ATAATTCTTG gagttgcagg aatattaaca	60
agatTTTtag gtctattttt tagatggcca ctaataatgt taattggaga tgagggatt	120
ggatattatc aatgtcata cccgtatATG atgtttttt tagctatggc ttctggagtt	180
ccagttgcaa tttctaaAGAT gatacggaa aagaatgcaa caaatgatATG atggcAGC	240
tttgaagtaa tgaaAGAATC tgctatTTTA atgacaataa tagggacagg cacaacttta	300
gcattattct ttttgcAAA accaatAGTA ttatTTTG AGTGGGATCC AAAAGCGTAT	360
tattcattaa tcggatATC atttgcACCA attgttaatC CATTGTAAC tatatttGAG	420
ggatttttc aagggttACA aaatATGACT CGTCAGCTA tatCACAGAT aatAGAGCAG	480
attggtagAG tcataATTGCG TGTGGACTT GCTGTATTT tactacCTAG aggcatAGAG	540
tactcAGcAG GTGGTGCAGC atttggAGCA ACTGCGGGAG CTGTTCTGG aggAGCTTAT	600
ttataACTCGA attataAAAG agtAAAGAAA CGTTATGCCA ttAAAAAAAT AAAAGCAAT	660
ccagAAAT taaatACtAt attAAAGATT GCAATACCAA tatCATTAGG tactACAGTA	720
tcaAGTATTAA tgaatttAAAGT agattCCATT ttGATACCA aaaaACTATT agatGCGGGA	780
ttcacAAATG tacaatCAAC tGTTATTATG GCGCAATTAA CAGGTAAAGC GTCTGTAA	840
gtAAATATTc cattaACTCT ttCTATGGCT atttGTAACt CTCTAAATTCC tatCATAGCT	900
gagaATTCA tacttaAGAA acAGAAAGAG CTAaaaaAGTA aatAGATGC ATCTATGAAA	960
atGGCATCAG taattGCTAT tCCGTGCACT ttGGGTtAT tCTTTTAGC tGAACCAGTA	1020
atGAAGTTA tattCCAGG CAGGTTGAA ggaatAGAGA tattAAATAA ttatCATTAA	1080
acaattcCTT ttataataat tactcaaaca acaacAGCAA tactacaagg aacAGGGCAT	1140
tatataAAAC ctGTTATTAA CCTCTTGATT gggTGTtGA ttAAAATTGT attAACGTGG	1200

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gttagtagttc	ctatgcaaat	gtttaacata	tatggctgt	ttttggcgag	ctttggagct	1260
tatTTAACAG	taagtatttt	aaatatAGTg	atgatGAAAT	ttacactaAG	agtaAGACTT	1320
aattttatacG	aaatatttaat	aaaacCTTGC	tatgcATCCA	gtattatGAT	gttaATTGTA	1380
ttaataaGTT	ataatatttt	atataAGGAAT	acaattAGTA	atggaatATC	ttgcttgaca	1440
tctatatTTT	tgggtatGAT	agtatatatTT	ataatGATAA	ttgtattCAA	ggtatttaAT	1500
qttqaacAAA	taaqacataq	attaaaaqa	aqtAA			1536

<210> SEQ ID NO 11
<211> LENGTH: 219
<212> TYPE: DNA
<213> ORGANISM: Clostridium beijerinckii

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<400> SEQUENCE: 11

ttgatgtata tgtcatcaa taacagtgg agaaatagaa cattagtacc agaagcaaag 60
gcaggattaa acagattaa aactgaggtt gcttcagaag ttggattaag tgattatgaa 120
aacatcgata aaggaagcct ttcttcaaga caaatggat atgttggcgg ttatatgta 180
aaacatatgtt ttcacgatca cgaacaaggctttaactaa 219
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<210> SEQ ID NO 12
<211> LENGTH: 1140
<212> TYPE: DNA
<213> ORGANISM: Clostridium beijerinckii

<210> SEQ ID NO 13
<211> LENGTH: 388
<212> TYPE: PRT
<213> ORGANISM: Clostridium beijerinckii

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<400> SEQUENCE: 13

Met Ala Arg Phe Thr Leu Pro Arg Asp Leu Tyr His Gly Glu Gly Ala
 1 5 10 15

Leu Glu Val Leu Lys Thr Leu Lys Gly Lys Lys Ala Phe Val Val Val
 20 25 30

Gly Gly Gly Ser Met Lys Arg Phe Gly Phe Leu Lys Gln Val Glu Asp
 35 40 45

Tyr Leu Thr Glu Ala Gly Met Glu Val Glu Leu Phe Glu Gly Val Glu
 50 55 60

Pro Asp Pro Ser Val Glu Thr Val Met Lys Gly Ala Glu Ala Met Arg
 65 70 75 80

Asn Phe Glu Pro Asp Trp Ile Val Ala Met Gly Gly Ser Pro Ile
 85 90 95

Asp Ala Ala Lys Ala Met Trp Ile Phe Tyr Glu Tyr Pro Asp Phe Thr
 100 105 110

Phe Glu Gln Ala Val Val Pro Phe Gly Leu Pro Glu Leu Arg Gln Lys
 115 120 125

Ala Lys Phe Val Ala Ile Pro Ser Thr Ser Gly Thr Ala Thr Glu Val
 130 135 140

Thr Ala Phe Ser Val Ile Thr Asn Tyr Thr Glu Lys Ile Lys Tyr Pro
 145 150 155 160

Leu Ala Asp Phe Asn Ile Thr Pro Asp Ile Ala Ile Val Asp Pro Val
 165 170 175

Leu Ala Gln Thr Met Pro Lys Thr Leu Thr Ala His Thr Gly Met Asp
 180 185 190

Ala Leu Thr His Ala Ile Glu Ala Tyr Thr Ala Ser Leu Arg Ser Asn
 195 200 205

Phe Ser Asp Pro Leu Ala Ile Lys Ala Leu Gln Met Val Gln Glu Asn
 210 215 220

Leu Ile Lys Ser Phe Glu Gly Asp Lys Glu Ala Arg Asn Leu Met His
 225 230 235 240

Glu Ala Gln Cys Leu Ala Gly Met Ala Phe Ser Asn Ala Leu Leu Gly
 245 250 255

Ile Val His Ser Met Ala His Lys Val Gly Ala Val Phe His Ile Pro
 260 265 270

His Gly Cys Ala Asn Ala Ile Phe Leu Pro Tyr Val Ile Gln Tyr Asn
 275 280 285

Arg Thr Lys Cys Glu Asp Arg Tyr Ala Asp Ile Ala Arg Ala Leu Lys
 290 295 300

Leu Glu Gly Asn Thr Asp Ser Glu Leu Thr Asp Ser Leu Ile Gly Met
 305 310 315 320

Ile Asn Lys Met Asn Ser Asp Leu Asn Ile Pro His Ser Met Lys Glu
 325 330 335

Tyr Gly Val Thr Glu Glu Asp Phe Lys Ala Asn Leu Ser Phe Ile Ala
 340 345 350

His Asn Ala Val Leu Asp Ala Cys Thr Gly Ser Asn Pro Arg Glu Ile
 355 360 365

Asp Asp Ala Thr Met Glu Lys Leu Phe Glu Cys Thr Tyr Tyr Gly Thr
 370 375 380

Lys Val Glu Leu
 385

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<210> SEQ ID NO 14
<211> LENGTH: 1167
<212> TYPE: DNA
<213> ORGANISM: Clostridium beijerinckii

<400> SEQUENCE: 14

atggcacgtt ttactttacc tagggacttg tatcatggag aaggggcact tgaagtactt      60
aaaactttaa aaggcaaaaa agcttttcta gttgttggtg gaggatcaat gaaaagattt      120
ggttttctta aacaagttga agattattta acagaagcag gaatggaagt agaactattt      180
gaaggagtag aaccagatcc atcagtagaa acagtaatga agggtgccga agcaatgaga      240
aacttcgagc ctgattggat agttgtatg ggtggaggat caccataaga tgctgcaaaa      300
gtctatgtgaa tattctatga atacccagat ttacttttgc aacaagctgt tgttccattt      360
ggttttaccag agcttagaca aaaagctaaa ttgttagcta ttccatcaac aagtggtaca      420
gtacagaaag ttacagctt ttcaagtata acaaattaca cagaaaagat taaatatcct      480
tttagctgatt ttaacataac tccagatata gcaatagttg atccagttt agctcaaact      540
atgccaaaaa cattaacagc tcatactgga atggatgcac taactcatgc tatagaagca      600
tatactgcat cgcttagatc aaatttttct gatcctttag caattaaggc attgcaaatg      660
gtacaagaaa attaatcaa gtctttgaa ggagacaagg aagctagaaa tctaattgcat      720
gaagctcaat gtttagcagg aatggcattt tctaattgcat tacttggat agttcattca      780
atggctcaca aggttggtgc tgtattccat attcctcatg gatgtgcgaa tgctatattc      840
ttaccatatg taattcaata taatagaaca aaatgcgaag atagatatgc tgatattgct      900
agagcattaa aattagaagg aaacacagat tcagaattaa ctgattcatt aattggatg      960
attaataaaa tgaatagtga tttaaatatt cctcattca tgaaagaata tggagttact      1020
gaagaagatt tttaaagcaaa tctttcattt attgctcata atgcagtgtt agatgcattgt      1080
acaggatcaa atcctagaga aatagatgtatgc gcgacaatgg aaaaactatt tgaatgcaca      1140
tattatggga caaagggttga actataag      1167

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<210> SEQ ID NO 15
<211> LENGTH: 388
<212> TYPE: PRT
<213> ORGANISM: *Clostridium beijerinckii*

<400> SEQUENCE: 15

Leu Glu Val Leu Lys Thr Leu Lys Gly Lys Lys Ala Phe Val Val Val
20 25 30

Gly Gly Gly Ser Met Lys Arg Phe Gly Phe Leu Lys Gln Val Glu Asp
35 40 45

50	55	60													
Pro	Asp	Pro	Ser	Val	Glu	Thr	Val	Met	Lys	Gly	Ala	Glu	Ala	Met	Arg

Asn Phe Glu Pro Asp Trp Ile Val Ala Met Gly Gly Gly Ser Pro Ile

Asp Ala Ala Lys Ala Met Trp Ile Phe Tyr Glu Tyr Pro Asp Phe Thr
 100 105 110

Phe	Glu	Gln	Ala	Val	Val	Pro	Phe	Gly	Leu	Pro	Glu	Leu	Arg	Gln	Lys
115							120						125		

-continued

Ala Lys Phe Val Ala Ile Pro Ser Thr Ser Gly Thr Ala Thr Glu Val
 130 135 140
 Thr Ala Phe Ser Val Ile Thr Asn Tyr Thr Glu Lys Ile Lys Tyr Pro
 145 150 155 160
 Leu Ala Asp Phe Asn Ile Thr Pro Asp Ile Ala Ile Val Asp Pro Val
 165 170 175
 Leu Ala Gln Thr Met Pro Lys Thr Leu Thr Ala His Thr Gly Met Asp
 180 185 190
 Ala Leu Thr His Ala Ile Glu Ala Tyr Thr Ala Ser Leu Arg Ser Asn
 195 200 205
 Phe Ser Asp Pro Leu Ala Ile Lys Ala Leu Gln Met Val Gln Glu Asn
 210 215 220
 Leu Ile Lys Ser Phe Glu Gly Asp Lys Glu Ala Arg Asn Leu Met His
 225 230 235 240
 Glu Ala Gln Cys Leu Ala Gly Met Ala Phe Ser Asn Ala Leu Leu Gly
 245 250 255
 Ile Val His Ser Met Ala His Lys Val Gly Ala Val Phe His Ile Pro
 260 265 270
 His Gly Cys Ala Asn Ala Ile Phe Leu Pro Tyr Val Ile Gln Tyr Asn
 275 280 285
 Arg Thr Lys Cys Glu Asp Arg Tyr Ala Asp Ile Ala Arg Ala Leu Lys
 290 295 300
 Leu Glu Gly Asn Thr Asp Ser Glu Leu Thr Asp Ser Leu Ile Gly Met
 305 310 315 320
 Ile Asn Lys Met Asn Ser Asp Leu Asn Ile Pro His Ser Met Lys Glu
 325 330 335
 Tyr Gly Val Thr Glu Glu Asp Phe Lys Ala Asn Leu Ser Phe Ile Ala
 340 345 350
 His Asn Ala Val Leu Asp Ala Cys Thr Gly Ser Asn Pro Arg Glu Ile
 355 360 365
 Asp Asp Ala Thr Met Glu Lys Leu Phe Glu Cys Thr Tyr Tyr Gly Thr
 370 375 380
 Lys Val Glu Leu
 385

<210> SEQ ID NO 16
 <211> LENGTH: 216
 <212> TYPE: DNA
 <213> ORGANISM: Clostridium beijerinckii
 <400> SEQUENCE: 16

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atgcatatgt catcaaataa tagtggaga aatagaacat tagtaccaga agcaaaacaa      60
ggattaaaca gattaaaaac tgagggttgc tcagaagttg gattaagcaa ttatgaaagc      120
atggataaaag gaaaccttgc ttcaagacaa aatggatatg ttggcgata tatggtaaaa      180
catatgatcg aagattatga acaaggtctt aagtaa                            216
  
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<210> SEQ ID NO 17
 <211> LENGTH: 71
 <212> TYPE: PRT
 <213> ORGANISM: Clostridium beijerinckii
 <400> SEQUENCE: 17

Met His Met Ser Ser Asn Asn Ser Gly Arg Asn Arg Thr Leu Val Pro			
1	5	10	15

Glu Ala Lys Gln Gly Leu Asn Arg Leu Lys Thr Glu Val Ala Ser Glu

-continued

20 25 30

Val Gly Leu Ser Asn Tyr Glu Ser Met Asp Lys Gly Asn Leu Ser Ser
 35 40 45

Arg Gln Asn Gly Tyr Val Gly Gly Tyr Met Val Lys His Met Ile Glu
 50 55 60

Asp Tyr Glu Gln Gly Leu Lys
 65 70

<210> SEQ ID NO 18

<211> LENGTH: 210

<212> TYPE: DNA

<213> ORGANISM: Clostridium beijerinckii

<400> SEQUENCE: 18

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aacagattaa aaactgaggt tgcttcagaa gtaggattac atgattacga aaatcaagat	120
aaaggaaatt tatcttcaag acaaaatgg a tacgttggcg gatacatggt taagcacatg	180
attgaaagct acgaacaagg tttaaagtaa	210

<210> SEQ ID NO 19

<211> LENGTH: 69

<212> TYPE: PRT

<213> ORGANISM: Clostridium beijerinckii

<400> SEQUENCE: 19

Met Ser Ser Asn Asn Ser Gly Arg Asn Arg Thr Leu Val Pro Glu Ala	
1 5 10 15	

Lys Gln Gly Leu Asn Arg Leu Lys Thr Glu Val Ala Ser Glu Val Gly	
20 25 30	

Leu His Asp Tyr Glu Asn Gln Asp Lys Gly Asn Leu Ser Ser Arg Gln	
35 40 45	

Asn Gly Tyr Val Gly Gly Tyr Met Val Lys His Met Ile Glu Ser Tyr	
50 55 60	

Glu Gln Gly Leu Lys
 65

<210> SEQ ID NO 20

<211> LENGTH: 21

<212> TYPE: DNA

<213> ORGANISM: Clostridium beijerinckii

<400> SEQUENCE: 20

ttttagagg aaatatttat g 21

<210> SEQ ID NO 21

<211> LENGTH: 23

<212> TYPE: DNA

<213> ORGANISM: Clostridium beijerinckii

<400> SEQUENCE: 21

tttaaggagg tgtattttc atg 23

<210> SEQ ID NO 22

<211> LENGTH: 23

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic construct

<400> SEQUENCE: 22

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tttaaggagg tgtatttcat atg	23
<210> SEQ ID NO 23	
<211> LENGTH: 78	
<212> TYPE: DNA	
<213> ORGANISM: Clostridium pasteurianum	
<400> SEQUENCE: 23	
tttaaaaagt taaaaacat gatacaataa gttatggtaa acttatgatt aaaattttaa	60
ggaggtgtat ttcatatg	78

What is claimed is:

1. A method of producing butanol comprising contacting a carbohydrate-comprising substrate with recombinant *Clostridia* or *Escherichia coli* bacteria transformed with a nucleic acid that results in a decreased level or activity of ManIIAB polypeptide or ManIIC polypeptide compared to the bacteria prior to transformation with a substrate, such that the substrate is metabolized to butanol.
2. The method claim 1, wherein the bacterium prior to transformation is selected from the group consisting of *Clostridium beijerinckii*, *Clostridium acetobutylicum*, *Clostridium saccharobutylicum*, and *Clostridium saccharo-erbutylacetonicum*.
3. The method of claim 2, wherein the bacterium prior to transformation is *Clostridium beijerinckii*.
4. The method of claim 3, wherein the bacterium prior to transformation is *Clostridium beijerinckii* NCIMB 8052 or a variant thereof.
5. The method of claim 4, wherein the variant is *Clostridium beijerinckii* BA101.
6. The method of claim 4, wherein the bacterium prior to transformation is *Clostridium beijerinckii* NCIMB 8052.
7. The method of claim 1, wherein the ManIIAB polypeptide is encoded by SEQ ID NO:7.
8. The method of claim 1, wherein the ManIIC polypeptide is encoded by SEQ ID NO:8.
9. The method of claim 1, wherein the transforming nucleic acid encodes an antisense RNA.
10. The method of claim 1, wherein the transforming nucleic acid knocks out or reduces the expression or activity

of the ManIIAB polypeptide or ManIIC polypeptide of the bacterium when compared to the bacterium prior to its transformation.

11. The method of claim 1, wherein the level of the ManIIAB polypeptide or ManIIC polypeptide is decreased as compared to the bacterium prior to transformation by manipulation of one or more of a promoter strength, a ribosomal binding site strength, and a regulatory region.

12. The method of claim 1, wherein the recombinant bacterium further comprises an increased level of one or more of an alcohol dehydrogenase and butyryl-CoA dehydrogenase compared to the bacterium prior to transformation.

13. The method of claim 12, wherein the recombinant bacterium is transformed with one or more additional nucleic acids that results in the increased expression of an alcohol dehydrogenase, a butyryl-CoA dehydrogenase, or both an alcohol dehydrogenase and a butyryl-CoA dehydrogenase compared to the bacterium prior to transformation.

14. The method of claim 13, wherein the one or more additional nucleic acids comprises a nucleic acid sequence with 80% or more identity to SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:12, or SEQ ID NO:14.

15. The method of claim 14, wherein the one or more additional nucleic acids comprises SEQ ID NO:1 or SEQ ID NO:14.

16. The method of claim 14, wherein the one or more additional nucleic acids comprises SEQ ID NO:2 or SEQ ID NO:12.

* * * * *

UNITED STATES PATENT AND TRADEMARK OFFICE
CERTIFICATE OF CORRECTION

PATENT NO. : 9,080,187 B2
APPLICATION NO. : 12/154027
DATED : July 14, 2015
INVENTOR(S) : Blaschek et al.

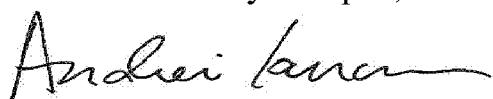
Page 1 of 1

It is certified that error appears in the above-identified patent and that said Letters Patent is hereby corrected as shown below:

In the Specification

Column 1 In Statement Regarding Federally Sponsored Research or Development, delete "This invention was made with United States Government support under USDA grant number 2001-35504-10668. The government may retain certain rights in the invention.", insert -- This invention was made with government support under grant number 2001-35504-10668 awarded by the United States Department of Agriculture. The government has certain rights in the invention. --

Signed and Sealed this
Seventeenth Day of April, 2018



Andrei Iancu
Director of the United States Patent and Trademark Office