

# Algorithms for the analysis of complex genomes

Michael Schatz

Oct 18, 2013  
CSHL In House



# Introductions



Srividya "Sri"  
Ramakrishnan

DOE Systems Biology  
Knowledgebase

Worlds fastest  
genomics pipelines



Tyler Garvin

WSBS

Interactive CNV  
and QC of single  
cell sequencing



Greg Vulture

CSHL URP / NYU

Mathematics of  
genomic architecture  
and heterozygosity



Aspyn Palatnick  
CSH HS  
iGenomics

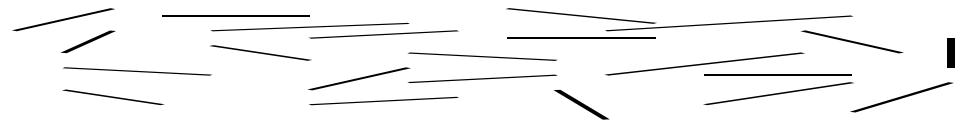


# Outline

- I. Read length & assembly complexity
2. Single molecule assembly of rice
3. De novo indel mutations in autism

# Assembling a Genome

## 1. Shear & Sequence DNA



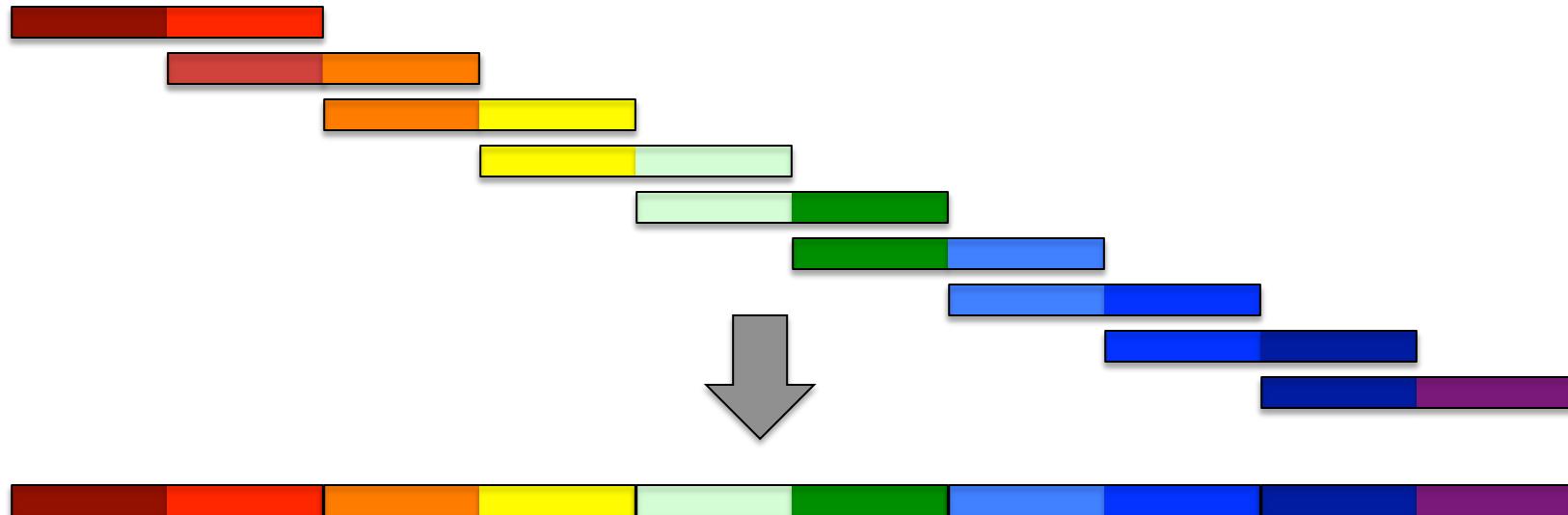
## 2. Construct assembly graph from overlapping reads

...AGCCTAGGGATGCGCGACACGT

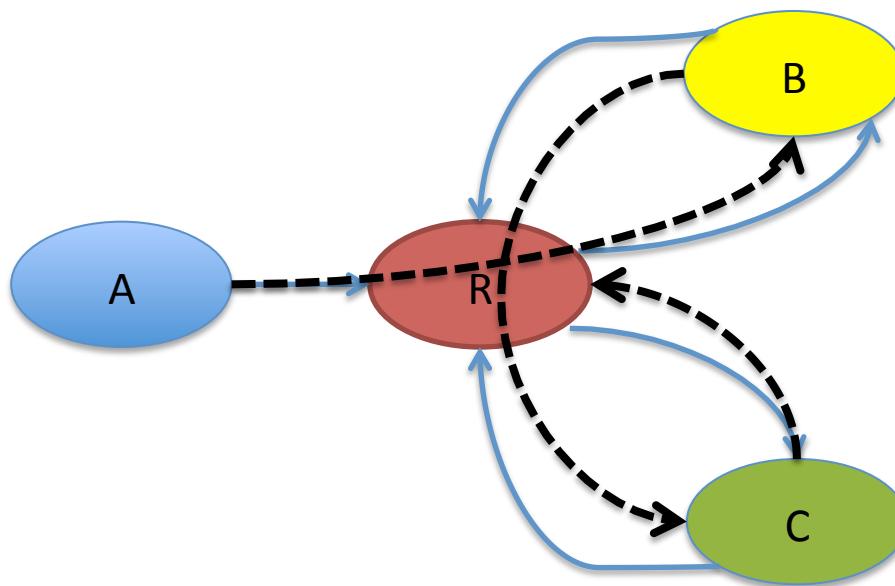
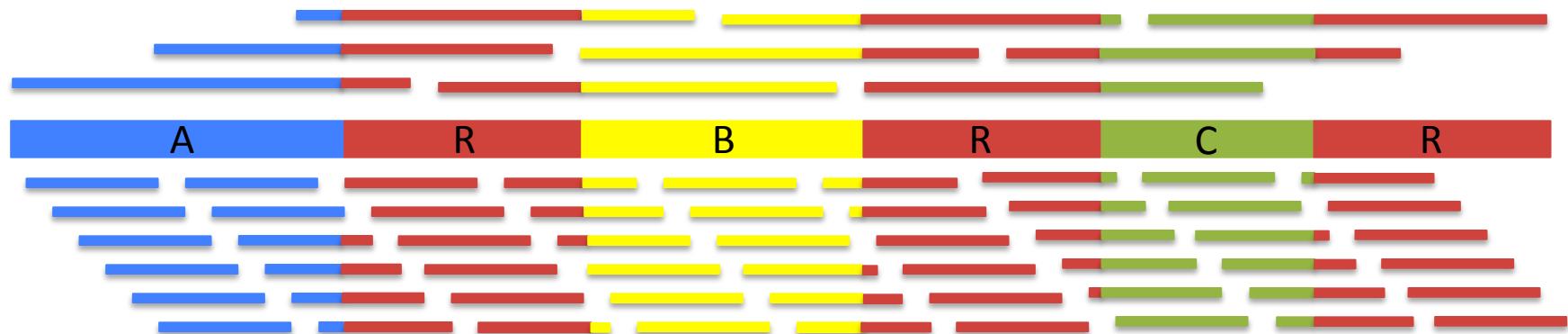
GGATGCGCGACACGT CGCATATCCGGTTGGT CAACCTCGGACGGAC

CAACCTCGGACGGAC CTCAGCGAA...

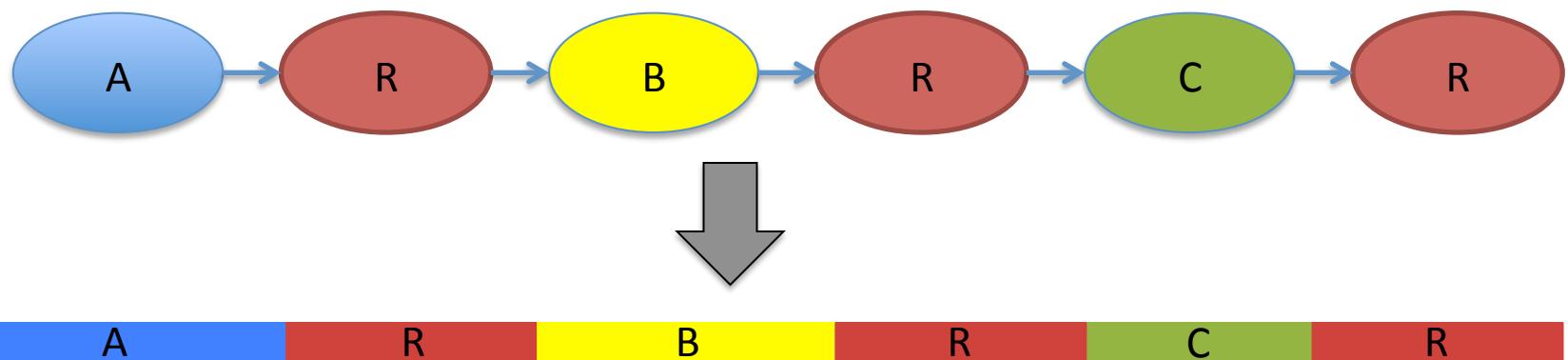
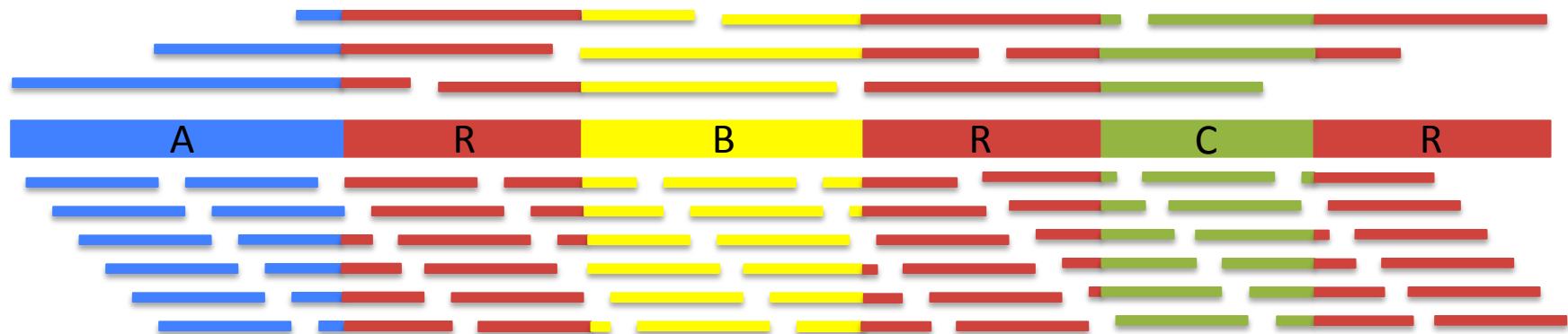
## 3. Simplify assembly graph



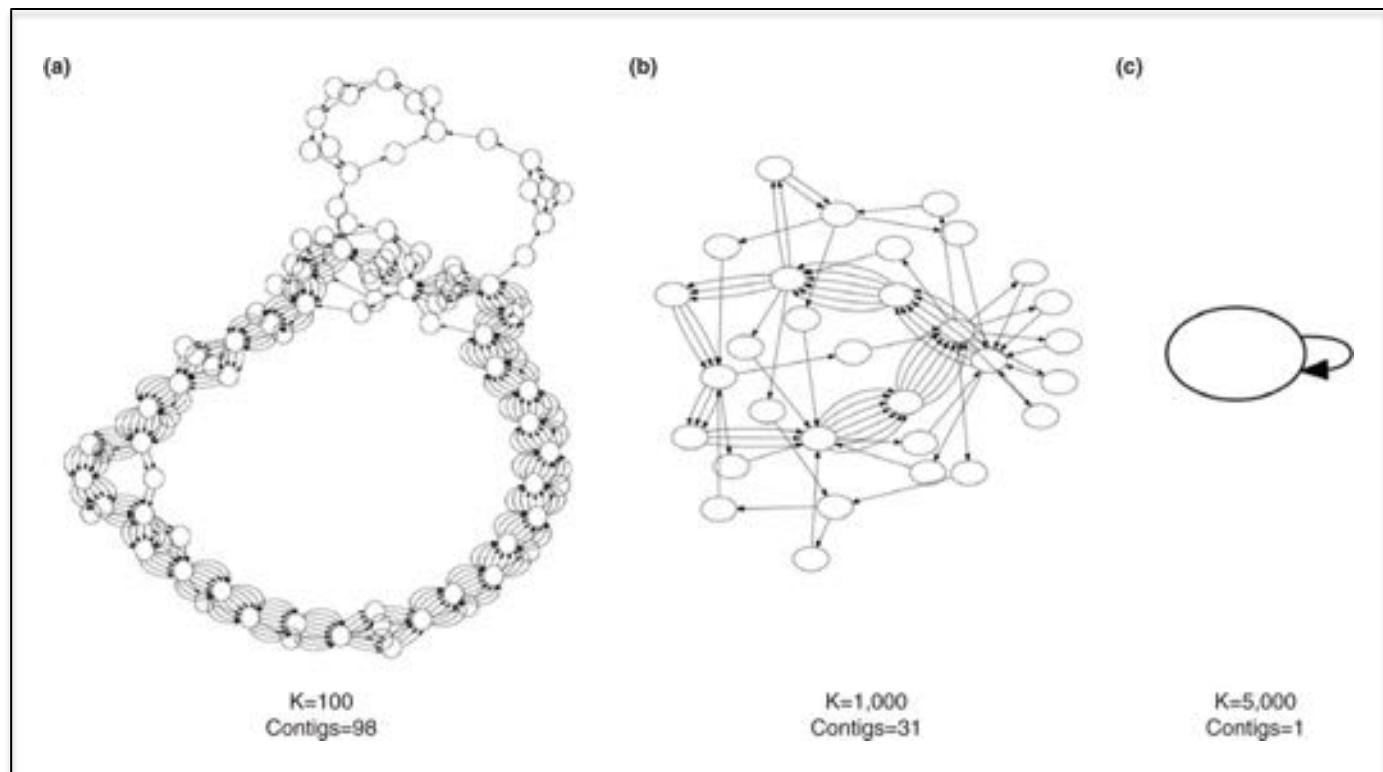
# Assembly Complexity



# Assembly Complexity



# Reducing Complexity



Longer reads span more repeats, simplifying the assembly problem

- Idealized assembly of *B. anthracis* reduces to a single contig with 5kb reads
- Exact improvement depends on the specific genome

## The advantages of SMRT sequencing

Roberts, RJ, Carneiro, MO, Schatz, MC (2013) *Genome Biology*. 14:405

# N50 size

Def: 50% of the genome is in contigs as large as the N50 value

Example: 1 Mbp genome



N50 size = 30 kbp

$$(300k + 100k + 45k + 45k + 30k = 520k \geq 500\text{ kbp})$$

Note:

A “good” N50 size is a moving target relative to other recent publications. 10-20 kbp contig N50 is currently a typical value for most “simple” genomes.

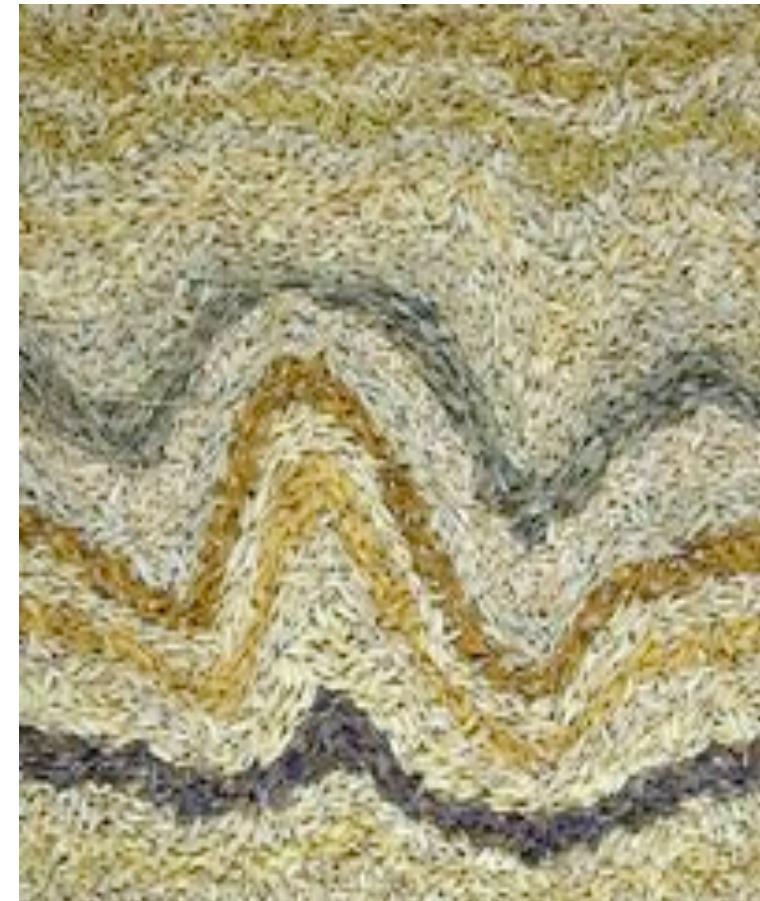
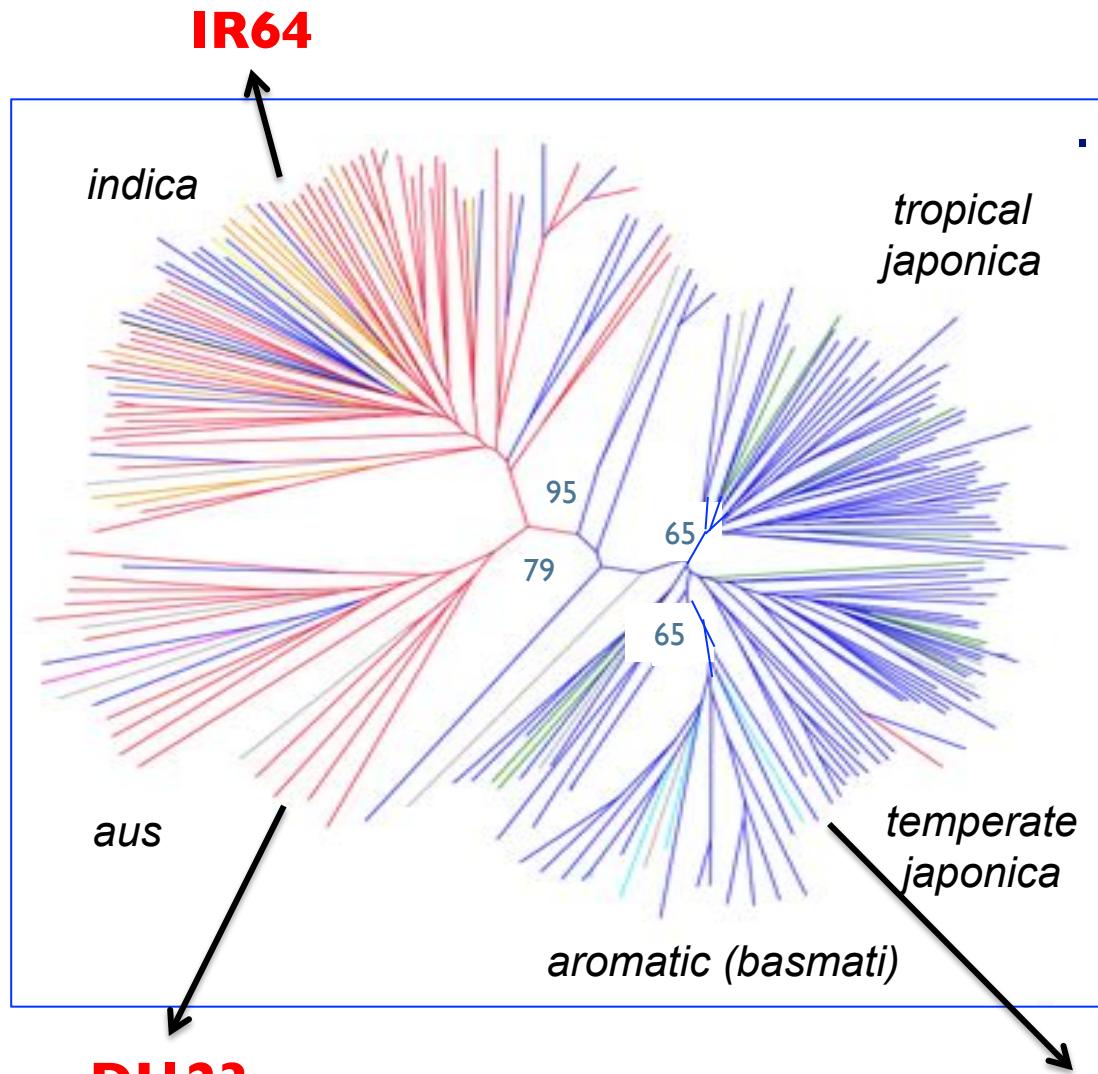


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# Population structure of *Oryza sativa*

3 varieties selected for *de novo* sequencing

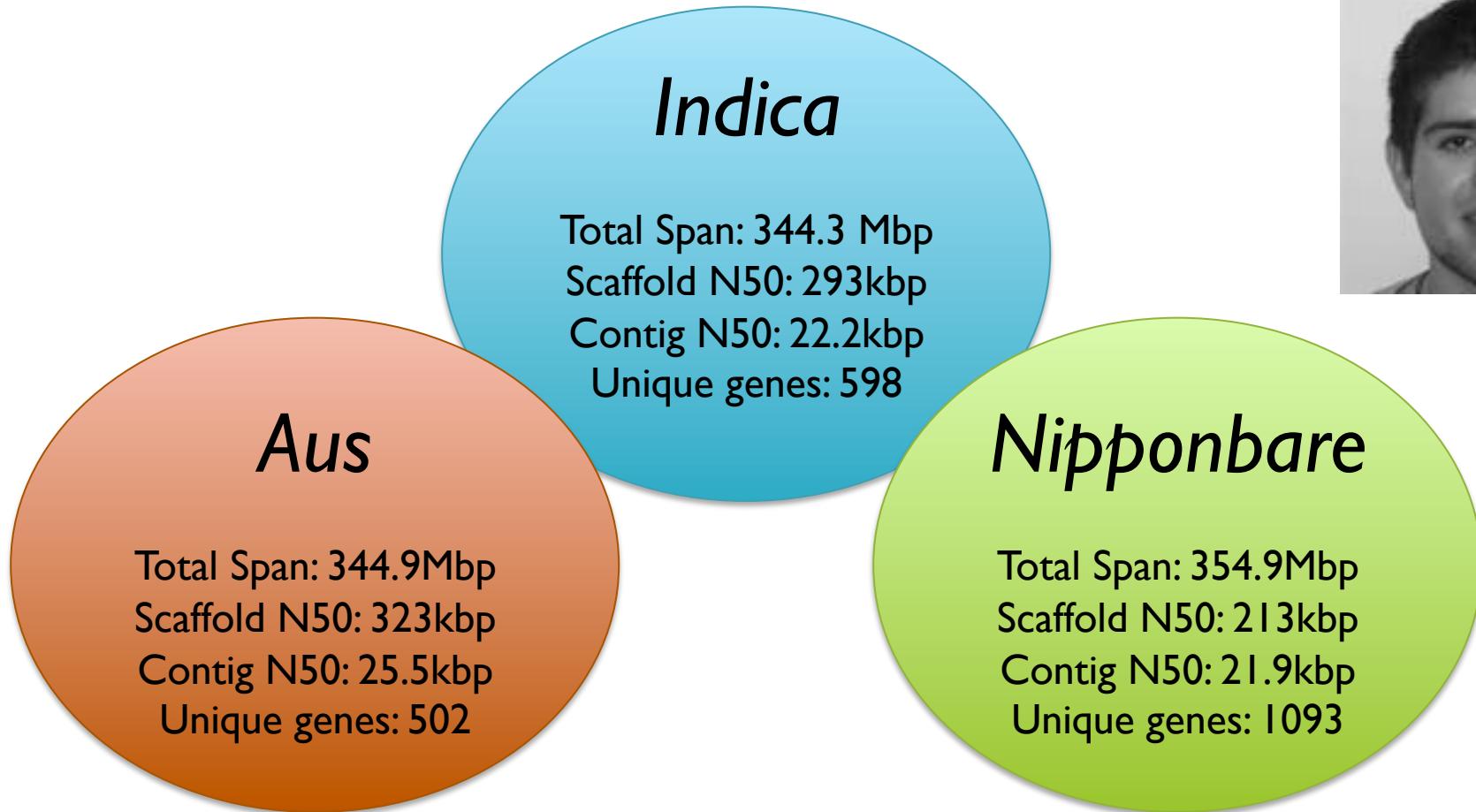


**IR64**

Garris et al. (2005)  
Genetics 169: 1631–1638

**Nipponbare**

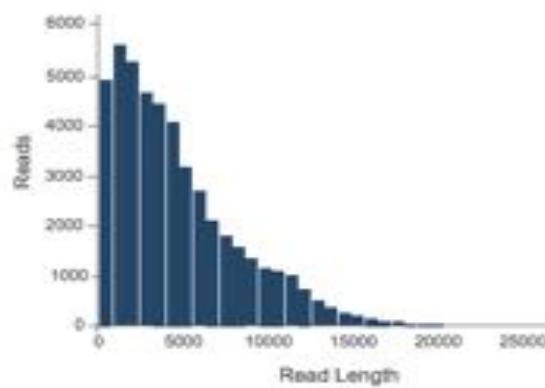
# Assembly and Annotation



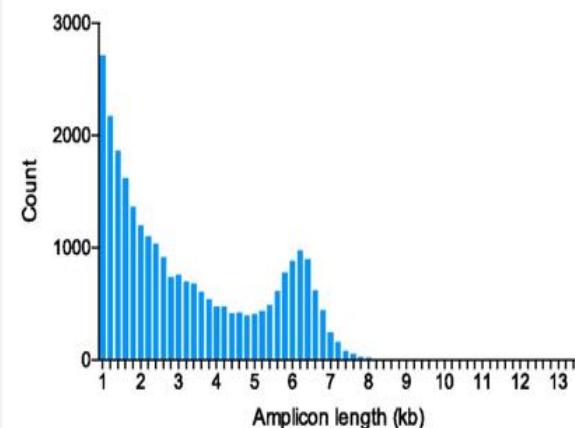
**New whole genome de novo assemblies of three divergent strains of rice documents novel gene space of Aus and Indica subpopulations**  
Schatz, MC, McCombie, WR, Ware, DW, McCouch, S, et al (2013) *In preparation*

# Single Molecule Sequencing Technology

PacBio RS II



Moleculo



Oxford Nanopore



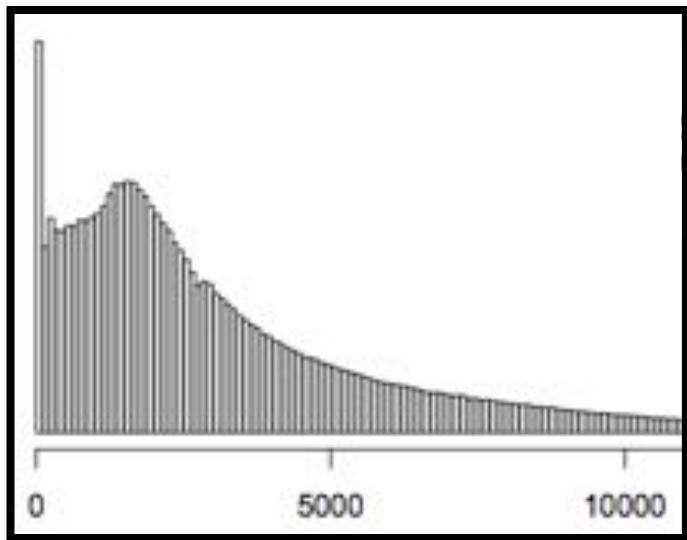
Clive G. Brown @Clive\_G\_Brown

I've reluctantly rejoined twitter purely so that I can make one tweet - when the appropriate time arises ...

Expand

9 Oct

# SMRT Sequencing Data



Match	83.7%
Insertions	11.5%
Deletions	3.4%
Mismatch	1.4%

TTGTAAGCAGTTGAAACTATGTGT GGATTAGATAAAGAACATGAAAG  
TTGTAAGCAGTTGAAACTATGTGT - GATTAG-ATAAAGAACATGGAAAG  
  
ATTATAAAA-CAGTTGATCCATT-AGAAGA-AAACGCAAAAGGC GGCTAGG  
| | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |  
A-TATAAAATCAGTTGATCCATTAGAA-AGAACGC-AAAGGC-GCTAGG  
  
CACCTTGAATGT AATCGCACTTGAAAGAACAGATT TATTCCGGGCCCG  
| | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |  
C-ACCTTG-ATGT-AT--CACTTGAAAGAACAGATT TATTCCGGGCCCG  
  
TAACGAATCAAGATTCTGAAACACAT-AT ACAACCTCCAAAA-CACAA  
| | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |  
T-ACGAATC-AGATTCTGAAACACATGAT---ACCTCCAAAA GCACAA  
  
-AGGAGGGAAAGGGGGGAATATCT-ATAAAAGATTACAAATT AGA-TGA  
| | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |  
GAGGAGG-AA---GAATATCTGAT-AAAGATTACAAATT-GAGTGA  
  
ACT-AATTACACAAATA-AATAACACTTTA-ACAGAATTGAT-GGAA-GTT  
| | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |  
ACTAAATTACACAA-ATAATAACACTTTAGACAAATTGATGGAAAGGTT  
  
TCGGAGAGATCCAACAAATGGC-ATCGCCTTGA-GTTAC-AATCAA  
| | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |  
TC-GAGAGATCC-AAACAAT-GGC GATCG-CTTGACGTTACAAATCAA  
  
ATCCAGTGGAAAATATAAT TTATGCAATCCAGGAACCTATTACAATTAG  
| | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |  
ATCCAGT-GAAAATATA--TTATGC-ATCCA-GAACCTATTACAATTAG

## Sample of 100k reads aligned with BLASR requiring >100bp alignment

# PacBio Error Correction: HGAP



- With 50-100x of Pacbio coverage, virtually all of the errors can be eliminated
  - Works well for Microbial genomes: single contig per chromosome routinely achieved
  - Difficult to scale up for use with eukaryotic genomes

**Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data**  
Chin, CS et al. (2013) *Nature Methods*. 10: 563-569

# Hybrid Sequencing



**Illumina**  
*Sequencing by Synthesis*

High throughput (60Gbp/day)  
High accuracy (~99%)  
Short reads (~100bp)



**Pacific Biosciences**  
*SMRT Sequencing*

Lower throughput (1 Gbp/day)  
Lower accuracy (~85%)  
Long reads (5kbp+)

# Hybrid Error Correction: PacBioToCA

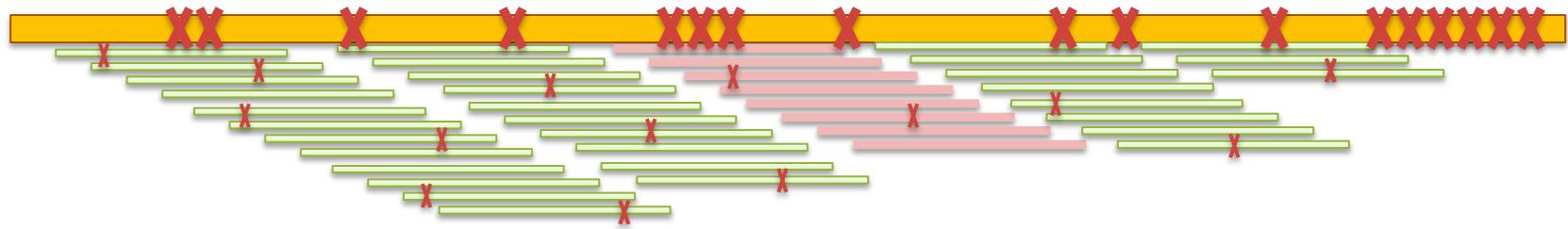
<http://wgs-assembler.sf.net>

## I. Correction Pipeline

1. Map short reads to long reads
2. Trim long reads at coverage gaps
3. Compute consensus for each long read



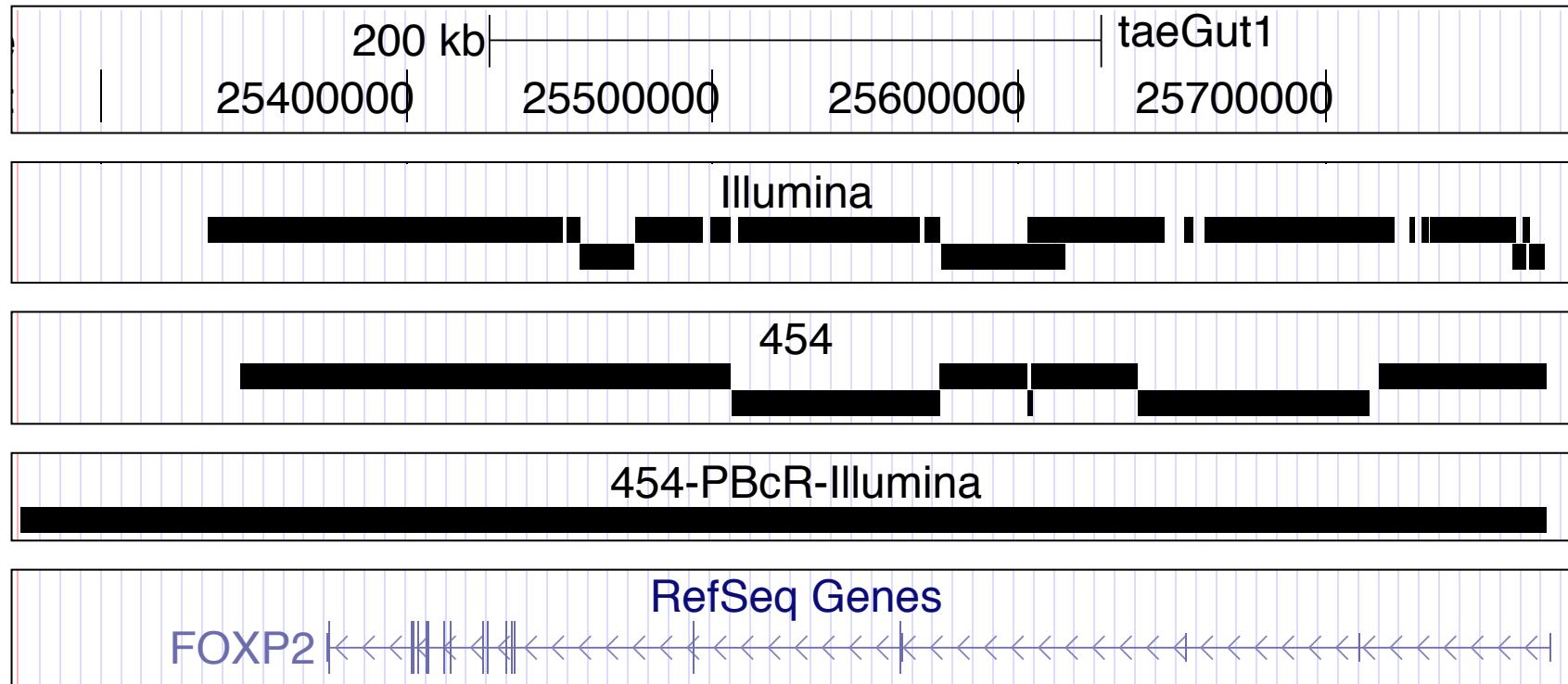
## 2. Error corrected reads can be easily assembled, aligned



**Hybrid error correction and de novo assembly of single-molecule sequencing reads.**  
Koren, S, Schatz, MC, et al. (2012) *Nature Biotechnology*. doi:10.1038/nbt.2280

# Improved Gene Reconstruction

FOXP2 assembled in a single contig in the PacBio parrot assembly

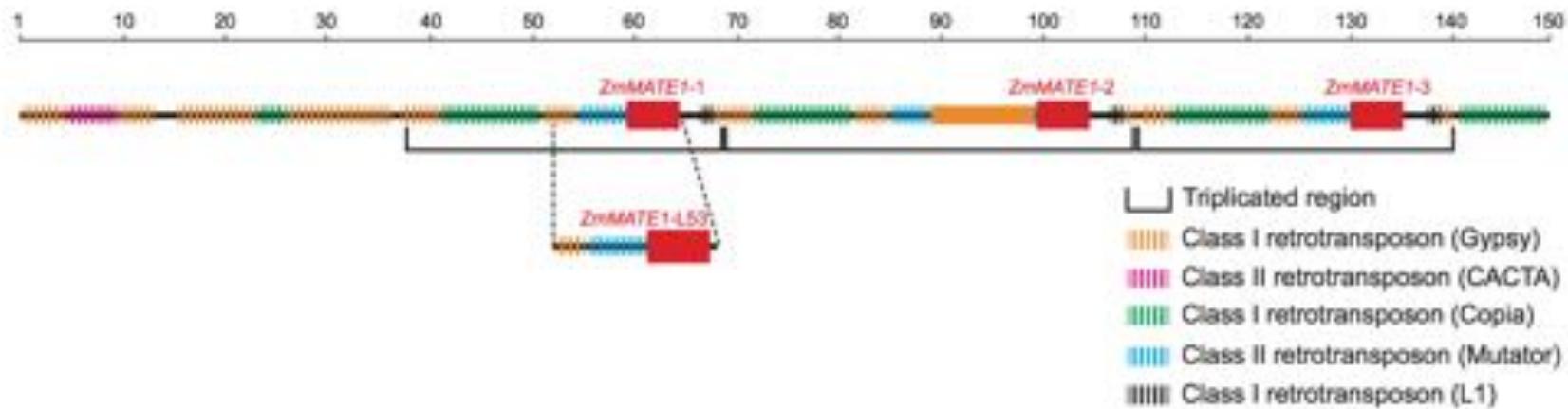


**Hybrid error correction and de novo assembly of single-molecule sequencing reads.**  
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# Long Read CNV Analysis

Aluminum tolerance in maize is important for drought resistance and protecting against nutrient deficiencies

- Segregating population localized a QTL on a BAC, but unable to genotype with Illumina sequencing because of high repeat content and GC skew
- Long read PacBio sequencing corrected by CCS reads revealed a triplication of the ZnMATE1 membrane transporter

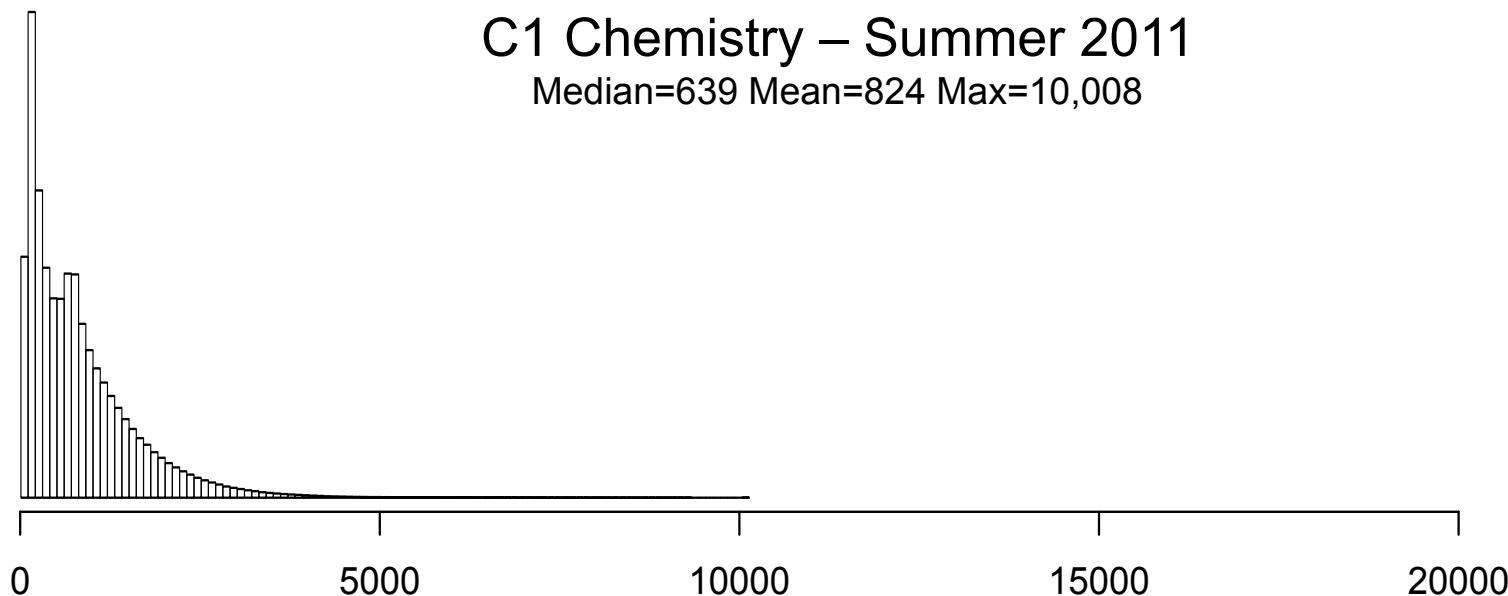


A rare gene copy-number variant that contributes to maize aluminum tolerance and adaptation to acid soils

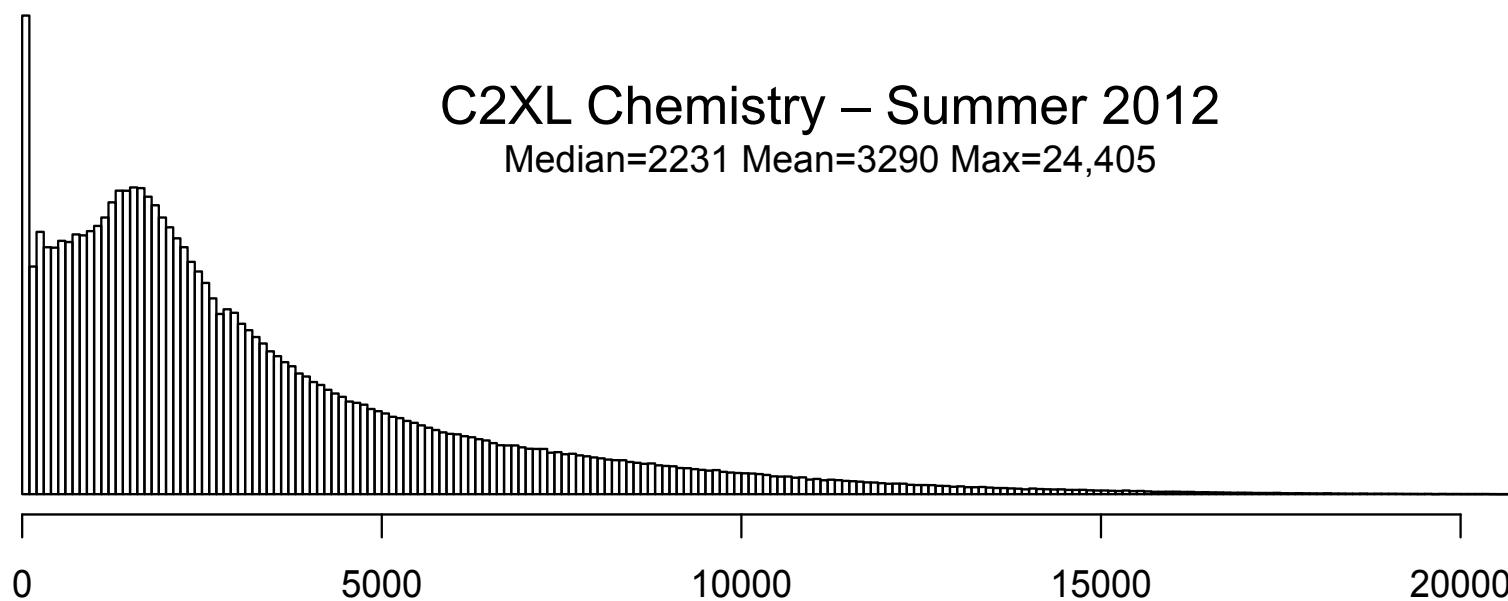
Maron, LG et al. (2013) PNAS doi: 10.1073/pnas.1220766110

# PacBio Long Read Rice Sequencing

C1 Chemistry – Summer 2011  
Median=639 Mean=824 Max=10,008

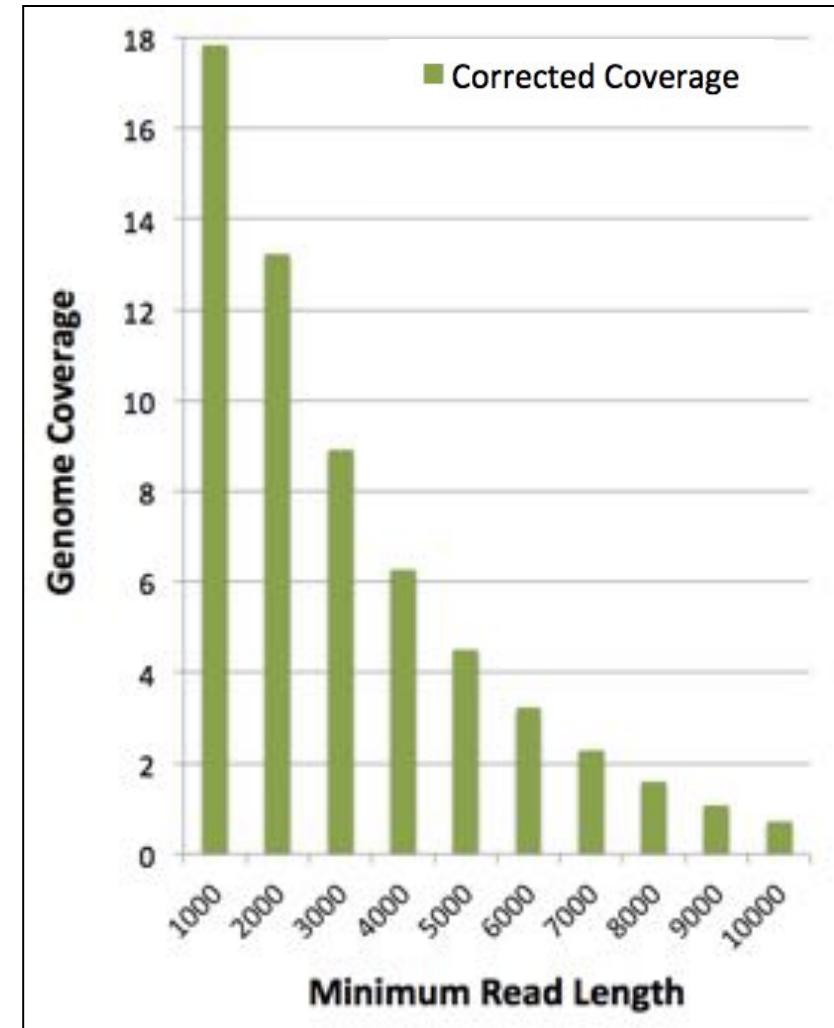


C2XL Chemistry – Summer 2012  
Median=2231 Mean=3290 Max=24,405



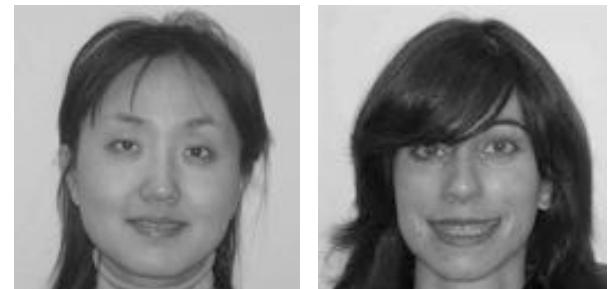
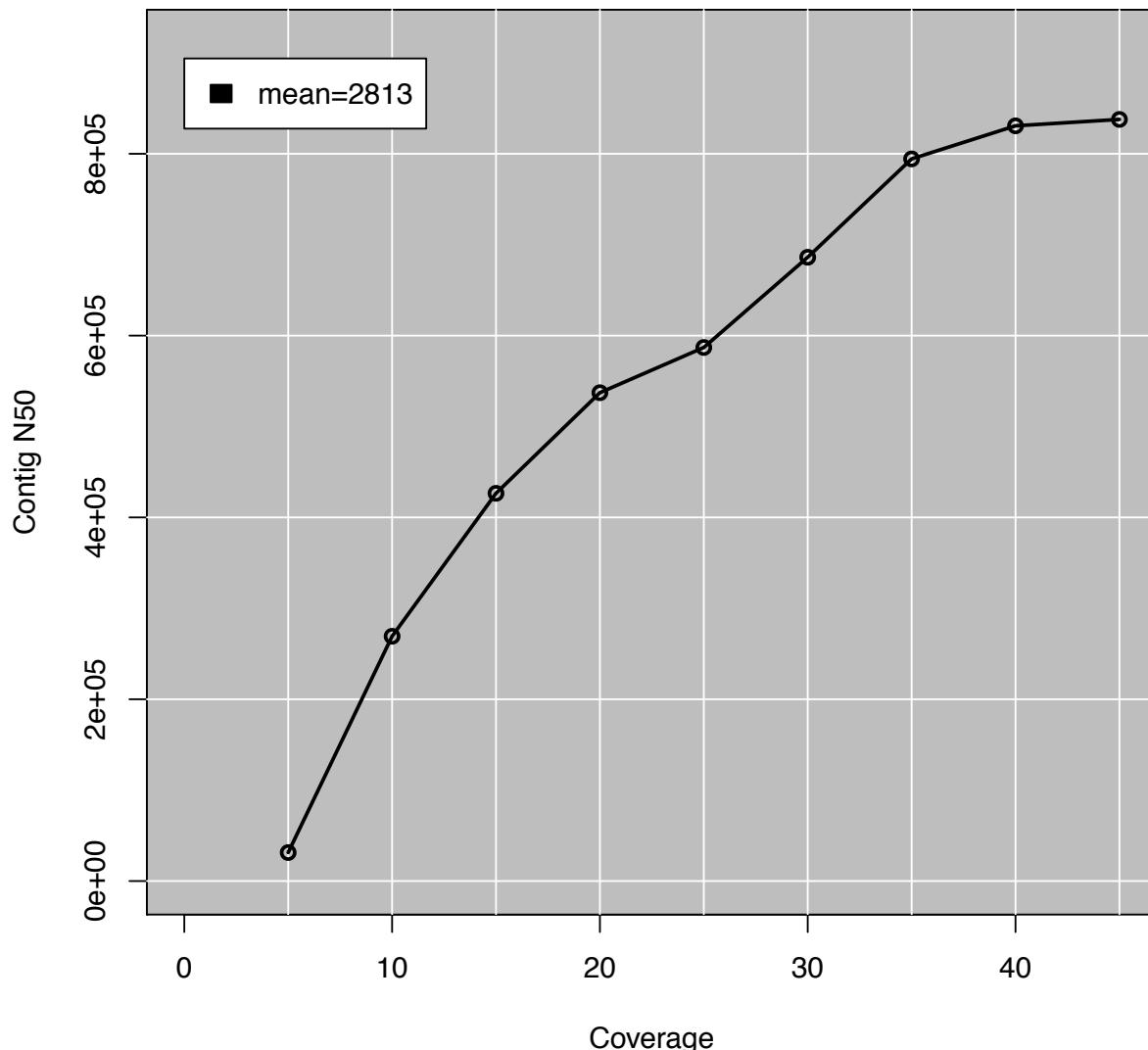
# Preliminary Rice Assemblies

Assembly	Contig NG50
HiSeq Fragments 50x 2x100bp @ 180	3,925
MiSeq Fragments 23x 459bp 8x 2x251bp @ 450	6,332
“ALLPATHS-recipe” 50x 2x100bp @ 180 36x 2x50bp @ 2100 51x 2x50bp @ 4800	18,248



In collaboration with McCombie & Ware labs @ CSHL

# Assembly Coverage Model



Simulate PacBio-like reads to predict how the assembly will improve as we add additional coverage

Only 8x coverage is needed to sequence every base in the genome, but 40x improves the chances repeats will be spanned by the longest reads

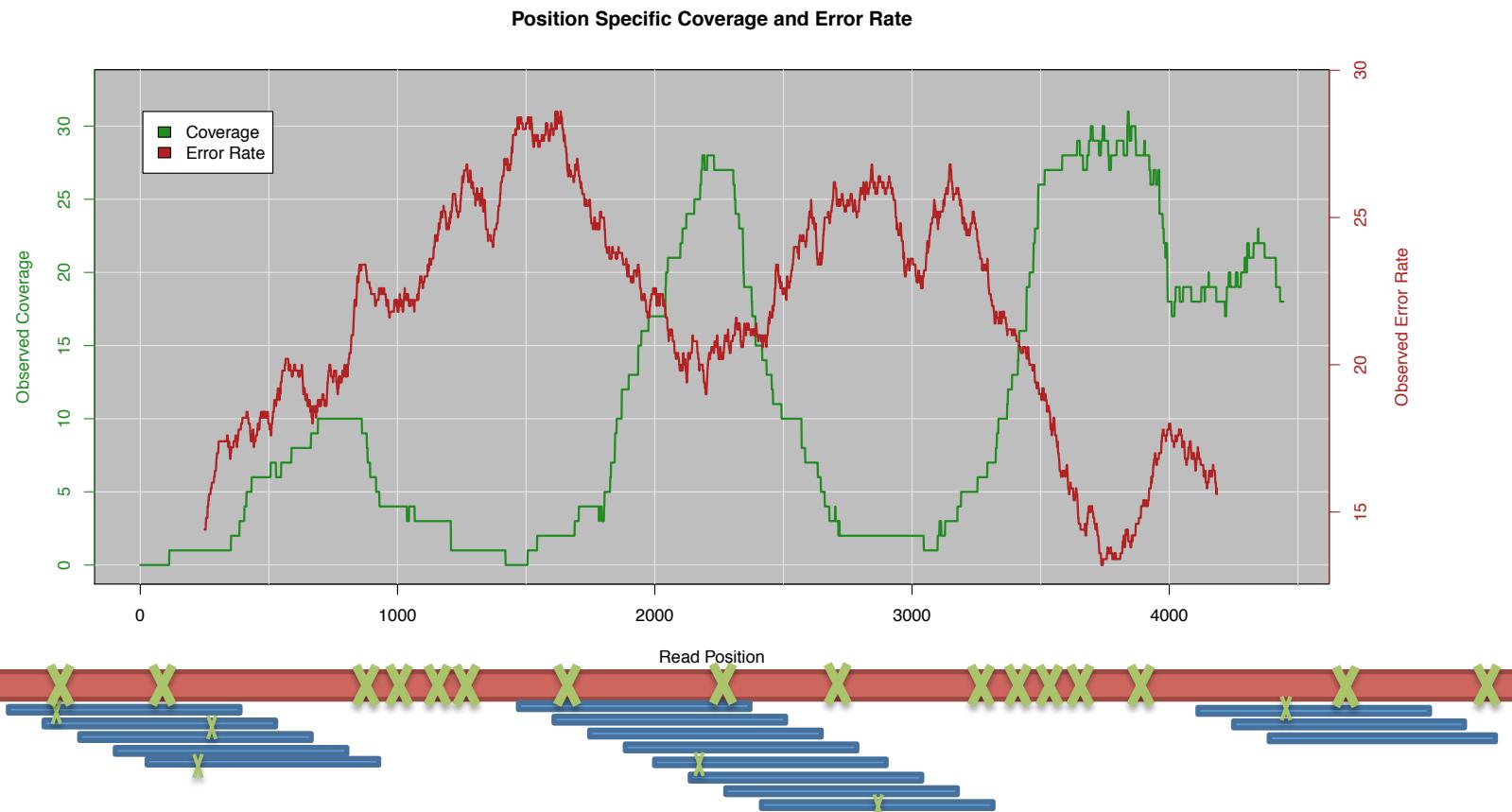
## Assembly complexity of long read sequencing

Lee, H\*, Gurtowski, J\*, Yoo, S, Marcus, S, McCombie, WR, Schatz MC et al. (2013) *In preparation*

# Enhanced PacBio Error Correction

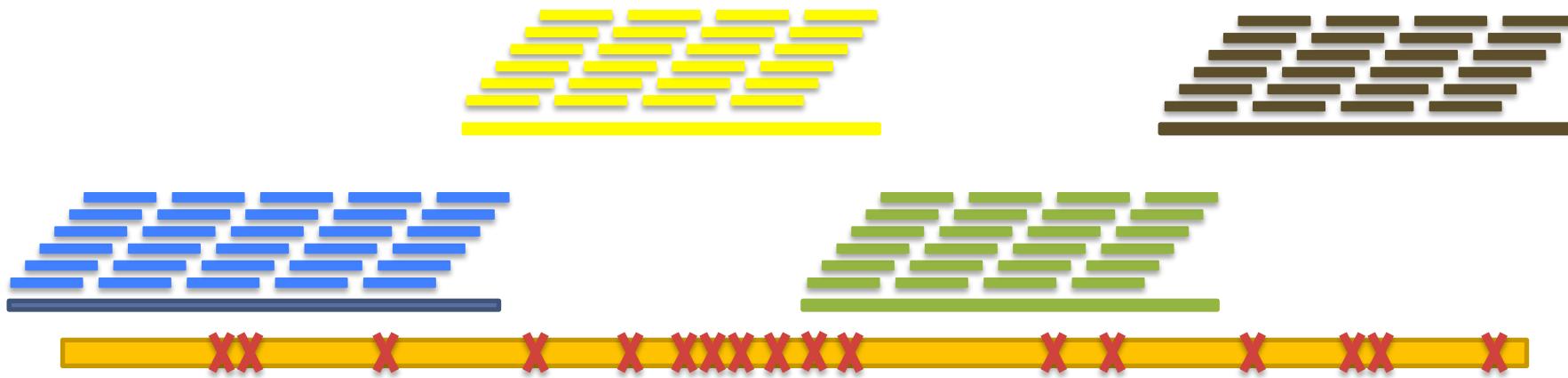
## PacBioToCA fails in complex regions

1. Simple Repeats – Kmer Frequency Too High to Seed Overlaps
2. Error Dense Regions – Difficult to compute overlaps with many errors
3. Extreme GC – Lacks Illumina Coverage



# Error Correction with pre-assembled Illumina reads

<https://github.com/jgurtowski/pbtools>



**Short Reads -> Assemble Unitigs -> Align & Select -> Error Correct**

**Unitigs:**

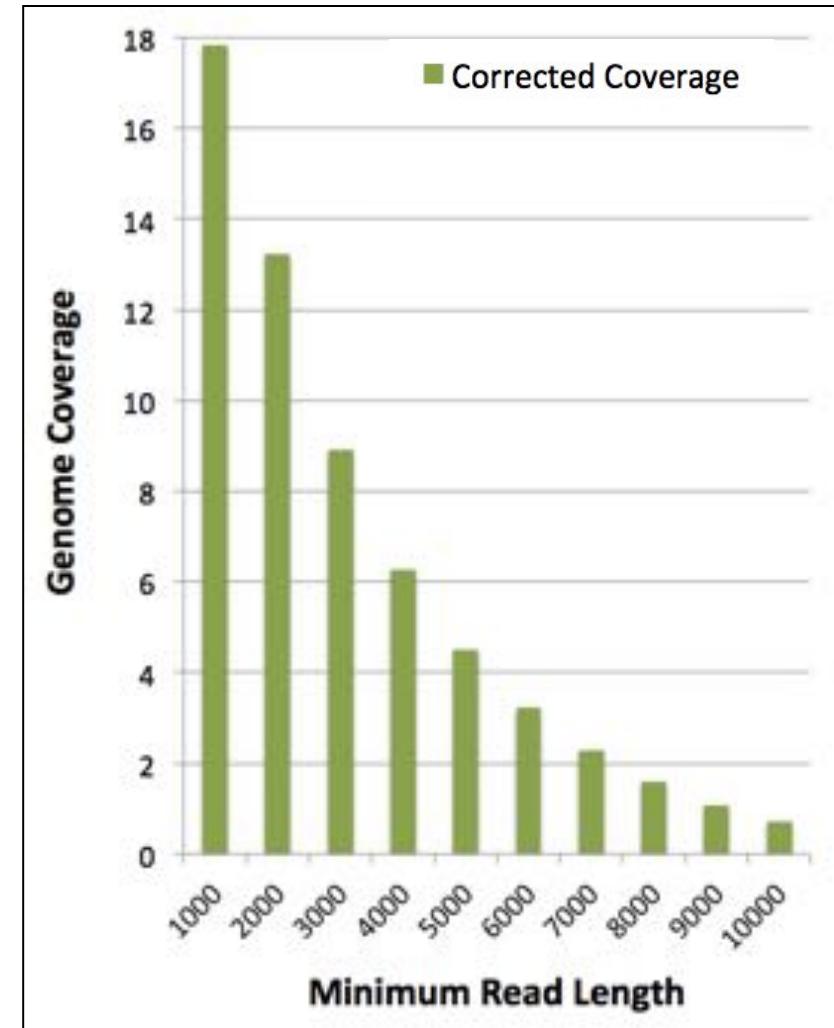
High quality contigs formed from unambiguous, unique overlaps of reads  
Each read is placed into a single unitig

Can Help us overcome:

- 1. Simple Repeats – Kmer Frequency Too High to Seed Overlaps**
- 2. Error Dense Regions – Difficult to compute overlaps with many errors**

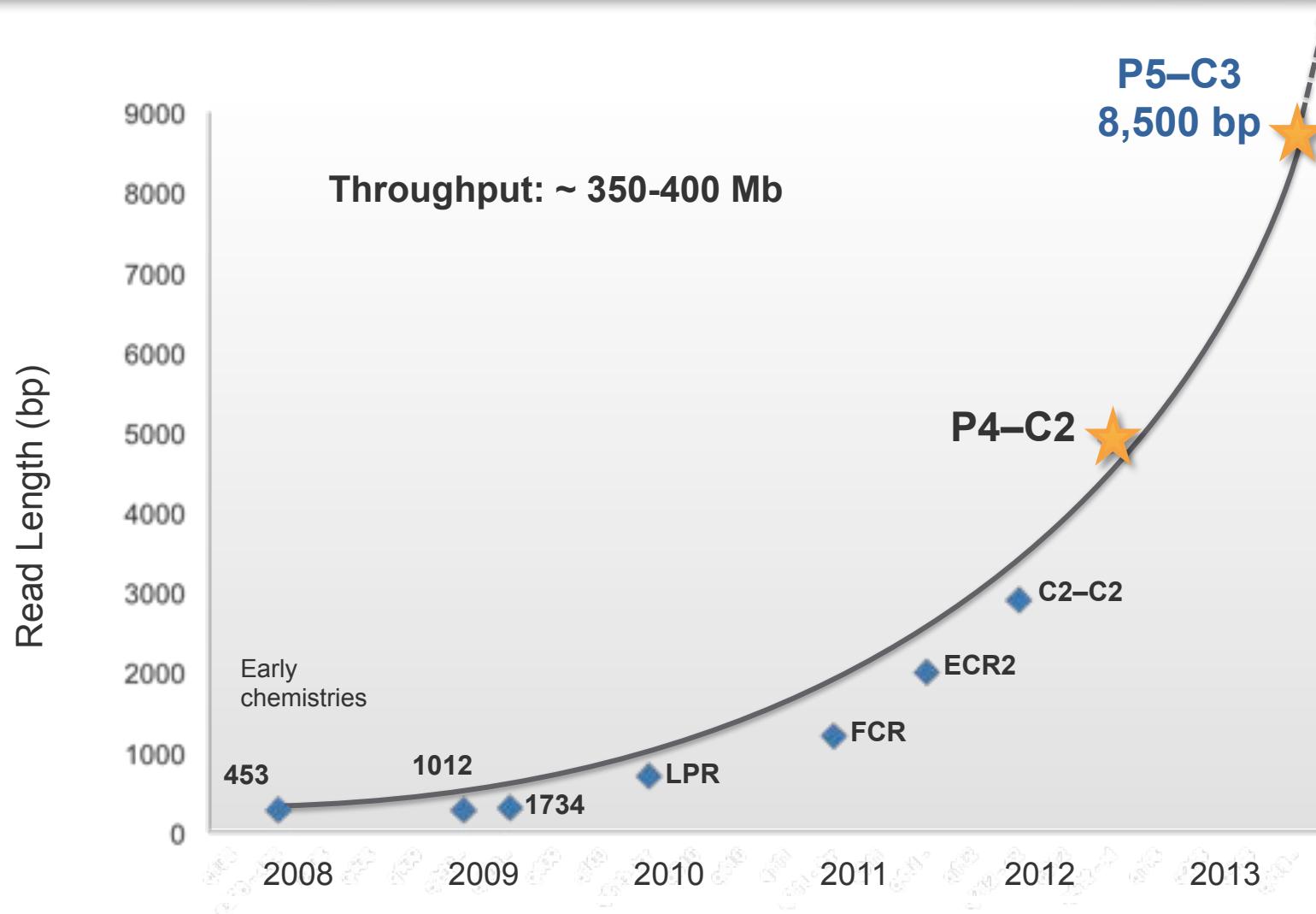
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“ALLPATHS-recipe” 50x 2x100bp @ 180 36x 2x50bp @ 2100 51x 2x50bp @ 4800	18,248
PBeCR Reads 19x @ 3500 ** MiSeq for correction	50,995



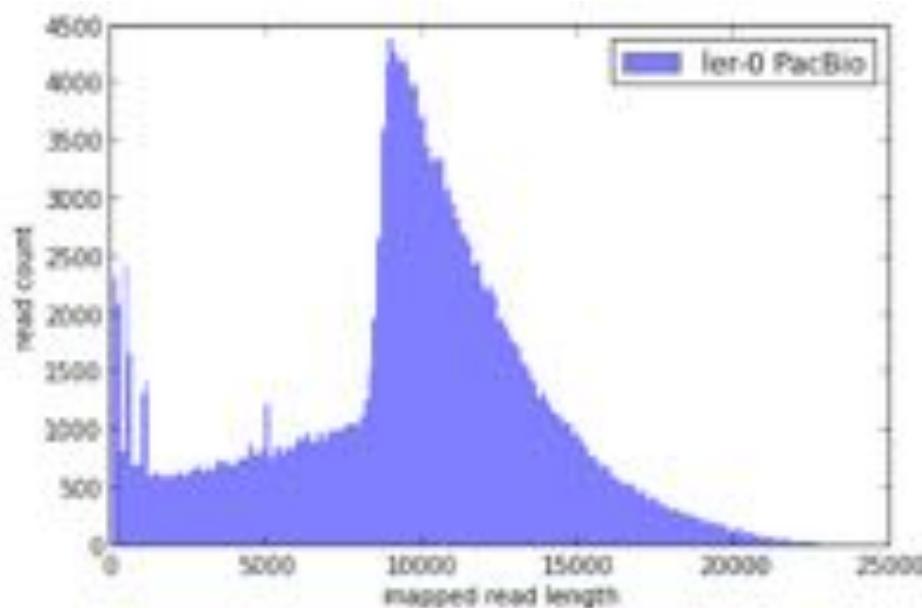
In collaboration with McCombie & Ware labs @ CSHL

# P5-C3 Chemistry Read Lengths



# De novo assembly of Arabidopsis

<http://blog.pacificbiosciences.com/2013/08/new-data-release-arabidopsis-assembly.html>



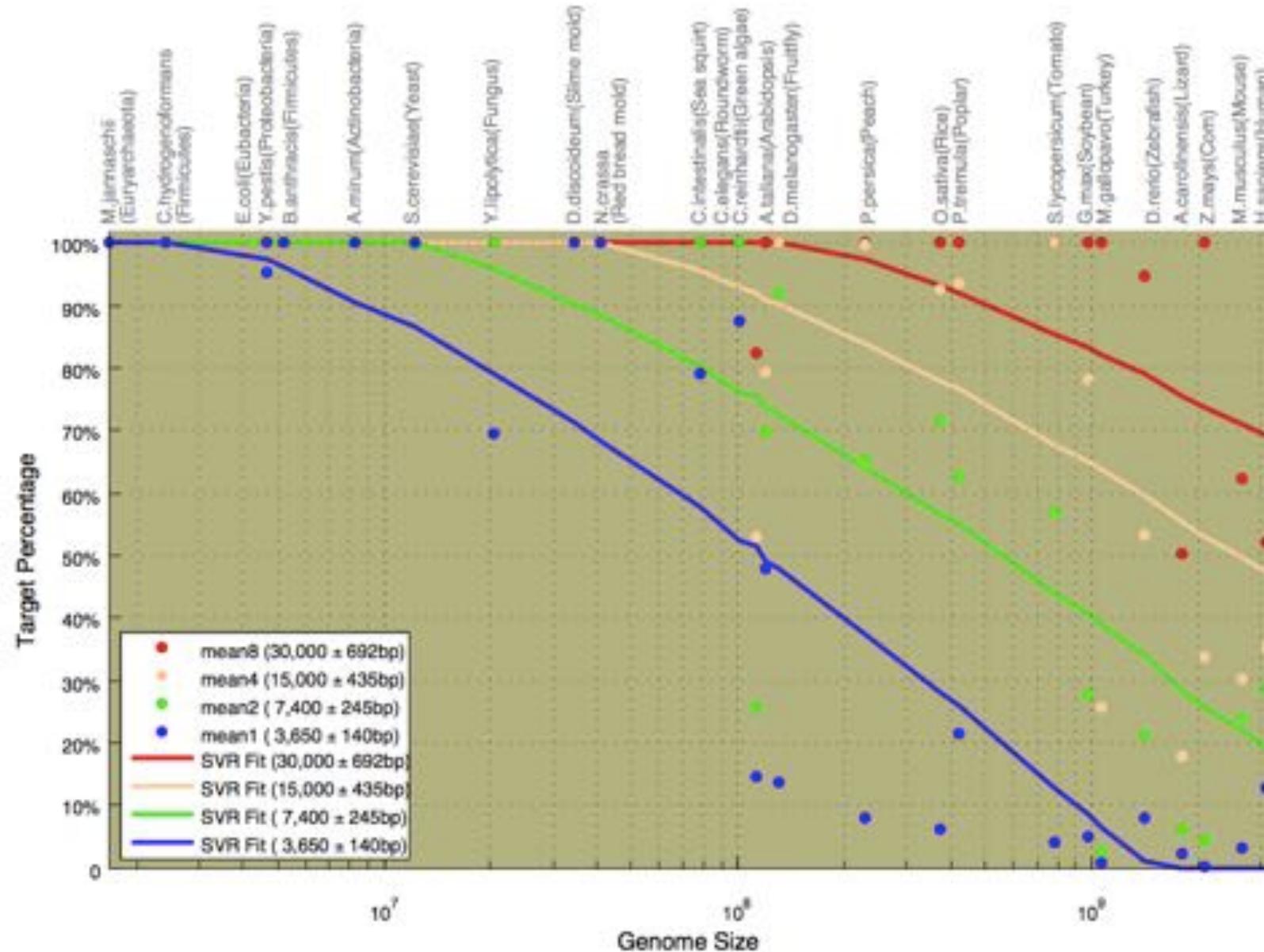
## *A. thaliana* Ler-0 sequenced at PacBio

- Sequenced using the latest P4 enzyme and C2 chemistry
- Size selection using an 8 Kb to 50 Kb elution window on a BluePippin™ device from Sage Science
- Total coverage >100x

Genome size: 124.6 Mb  
GC content: 33.92%  
Raw data: 11 Gb  
Assembly coverage: 15x over 9kbp

Sum of Contig Lengths: 149.5Mb  
Number of Contigs: 1788  
Max Contig Length: 12.4 Mb  
N50 Contig Length: 8.4 Mb

# Assembly Complexity of Long Reads





# Outline

- I. Read length & assembly complexity
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3. De novo indel mutations in autism

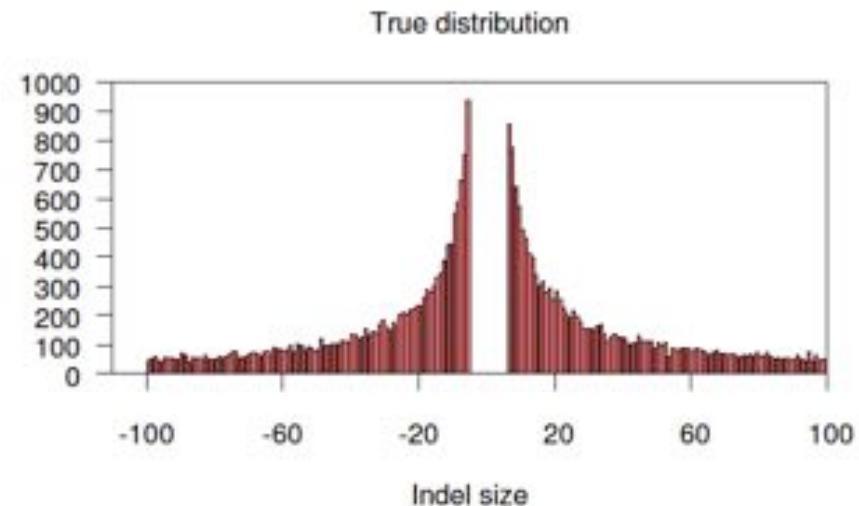
# Variation Detection Complexity

# SNPs + Short Indels

**High precision and sensitivity**

.. TTT**AGAATAG**-CGAGTGC .. .

||| ||| |||  
**AGAATAGGCGAG**

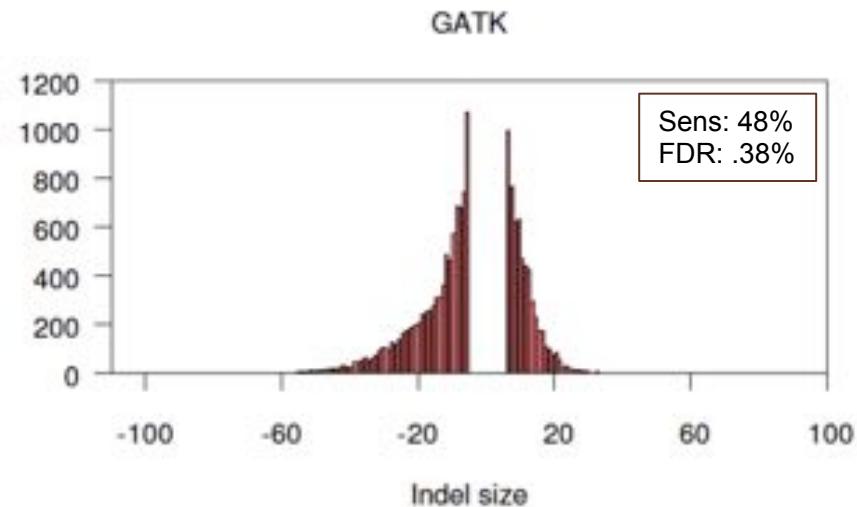


# “Long” Indels (>5bp)

## Reduced precision and sensitivity

..TTTAG-----AGTGC...

TTT **AATAGGC**  
**ATAGGCGAGTGC**



Analysis confounded by sequencing errors, localized repeats, allele biases, and mismapped reads

# Scalpel: Haplotype Microassembly

DNA sequence **micro-assembly** pipeline for accurate detection and validation of *de novo* mutations (SNPs, indels) within exome-capture data.



## Features

1. Combine **mapping** and **assembly**
2. Exhaustive search of **haplotypes**
3. **De novo** mutations



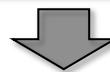
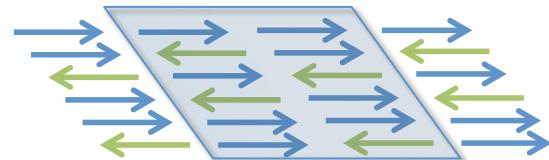
NRXN1 *de novo* SNP  
(auSSC12501 chr2:50724605)

**SCALPEL: Micro-assembly approach to accurately detect *de novo* and transmitted indel mutations within exome-Capture data**

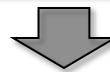
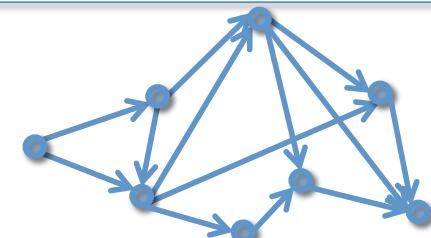
Narzisi, G, O'Rawe, J, Iossifov, I, Lee, Y, Wang, Z, Wu, Y, Lyon, G, Wigler, M, Schatz, MC (2013) *In preparation*

# Scalpel Pipeline

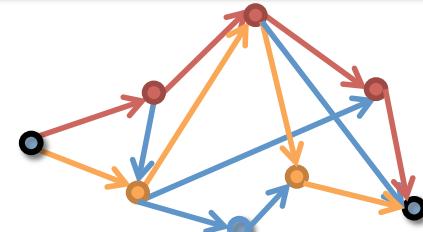
Extract reads mapping within the exon including (1) well-mapped reads, (2) soft-clipped reads, and (3) anchored pairs



Decompose reads into overlapping k-mers and construct de Bruijn graph from the reads



Find end-to-end haplotype paths spanning the region



Align assembled sequences to reference to detect mutations

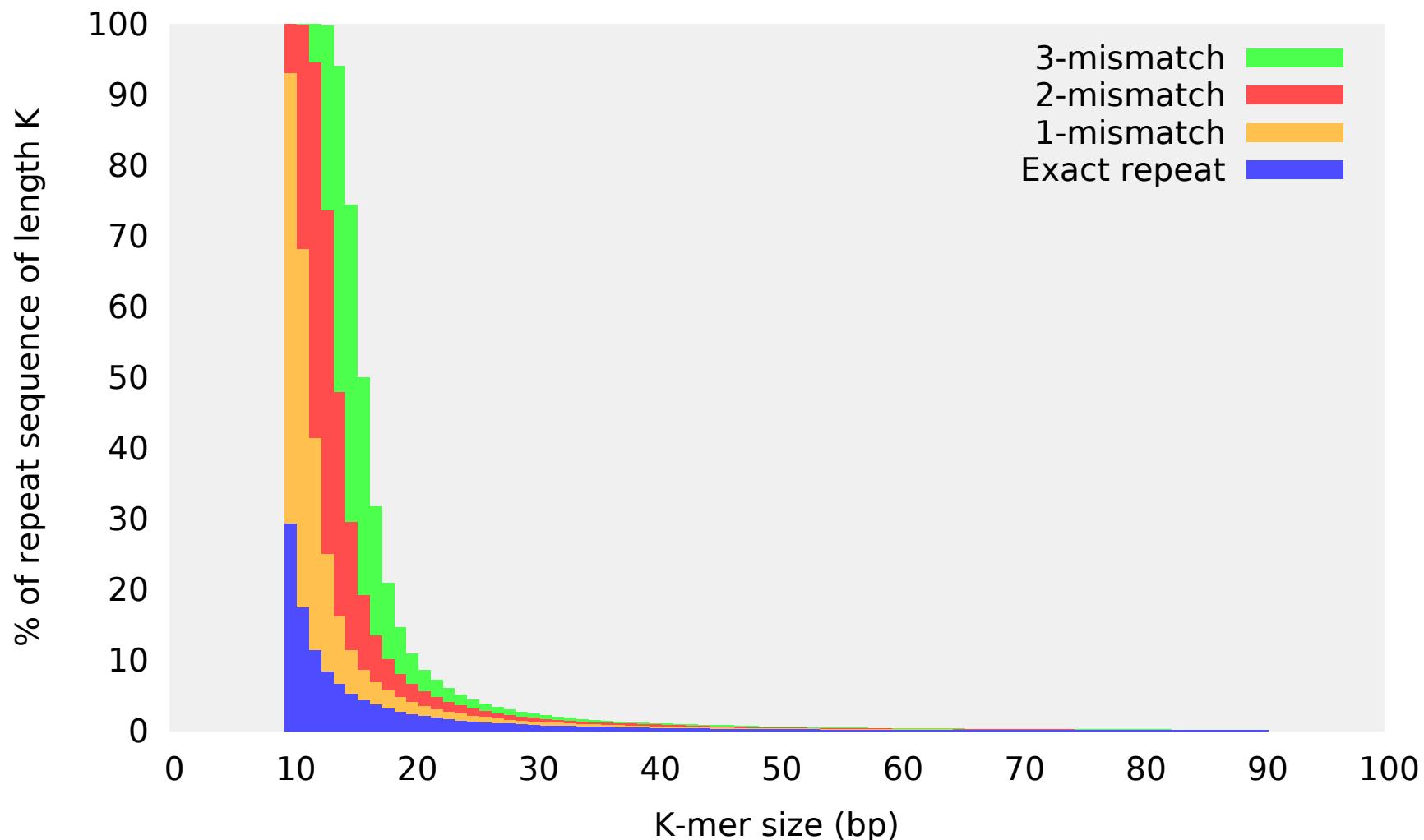


deletion

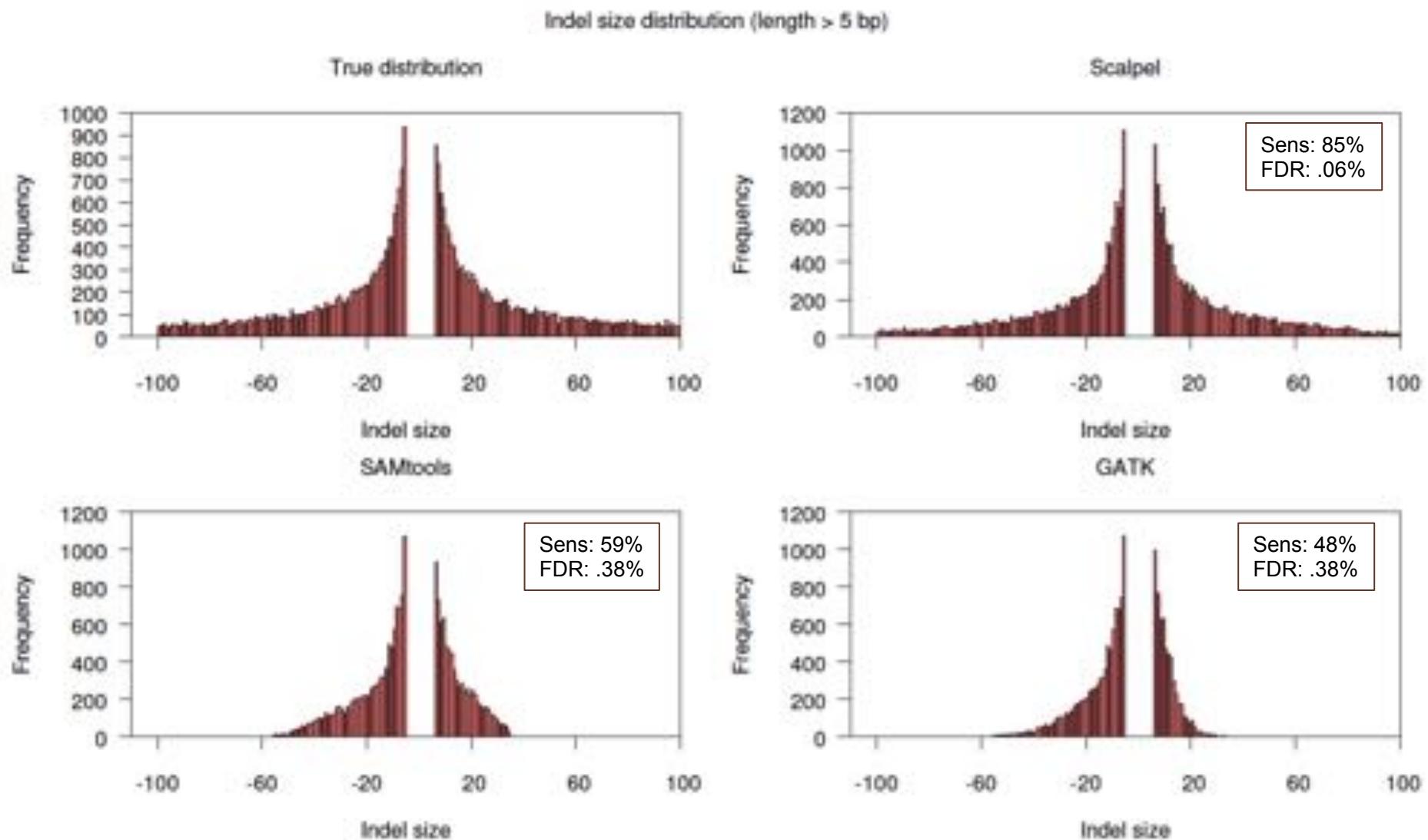
insertion

# Repeats in the Exome

Specificity Challenge: 30% of exons have a perfect 10bp or larger repeat  
Compute an on-the-fly analysis of repeat composition



# Simulation Analysis



Simulated 10,000 indels in a exome from a known log-normal distribution

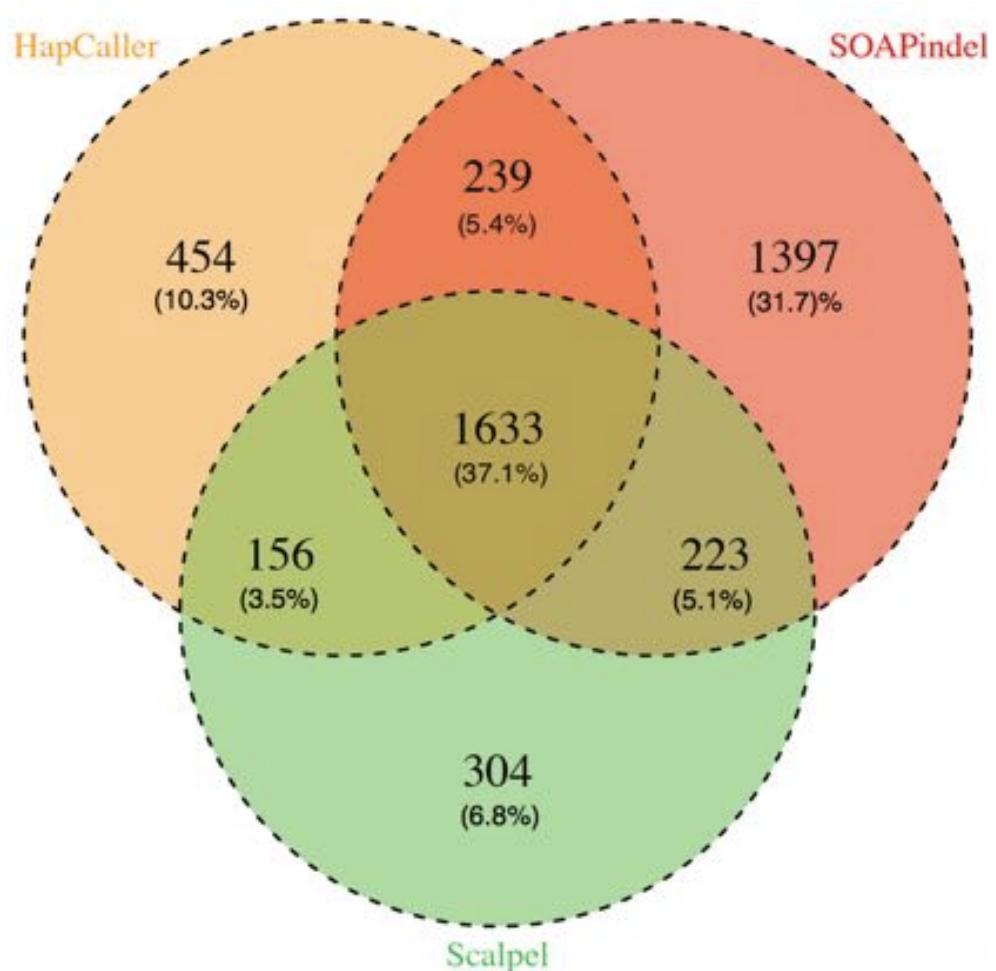
# Experimental Analysis & Validation

Selected one deep coverage exome for deep analysis

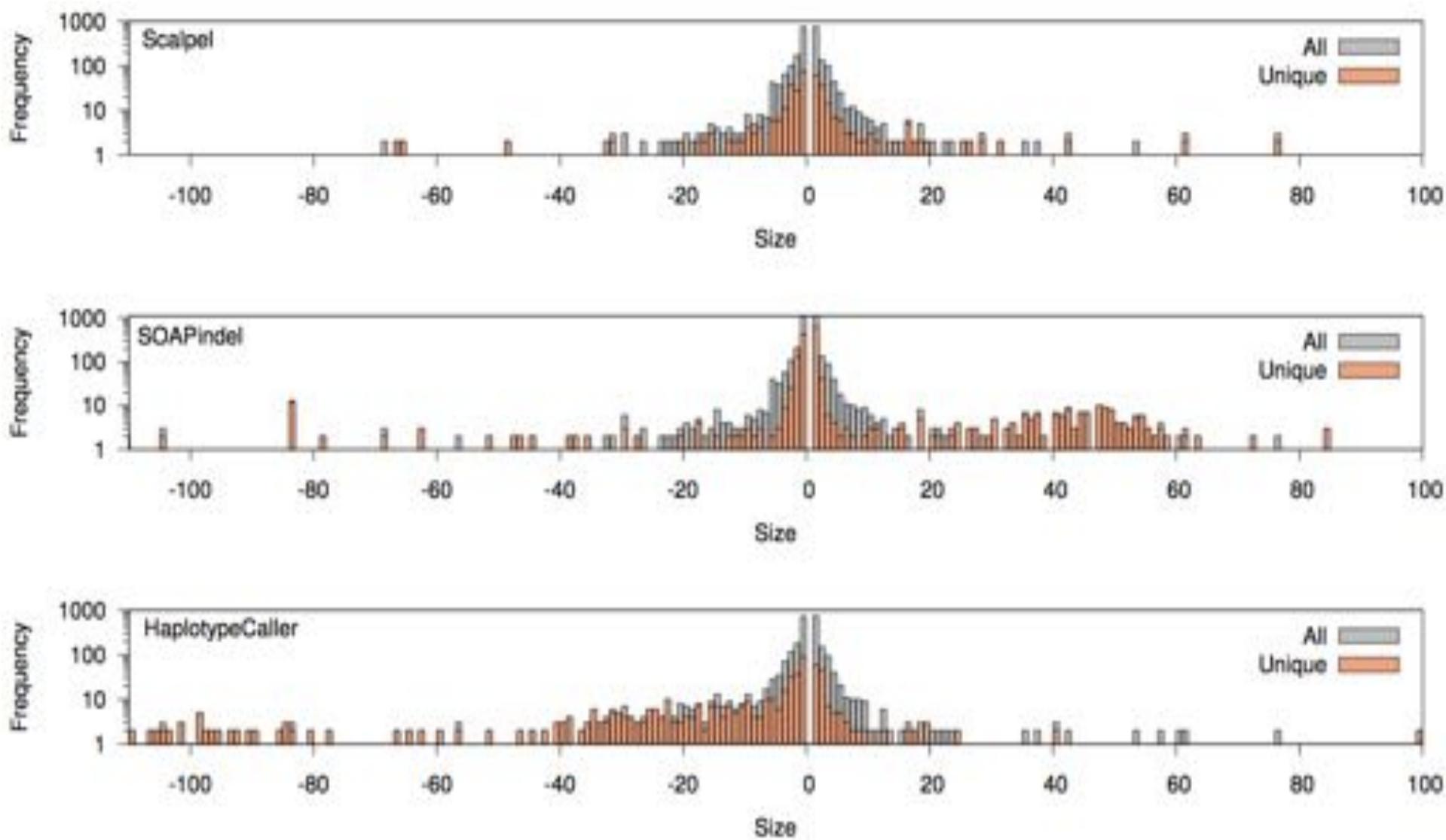
- Individual was diagnosed with ADHD (See Gholson for details)
- 80% of the target at >20x coverage
- Evaluated with Scalpel, SOAPindel, and GATK Haplotype Caller

1000 indels selected for validation

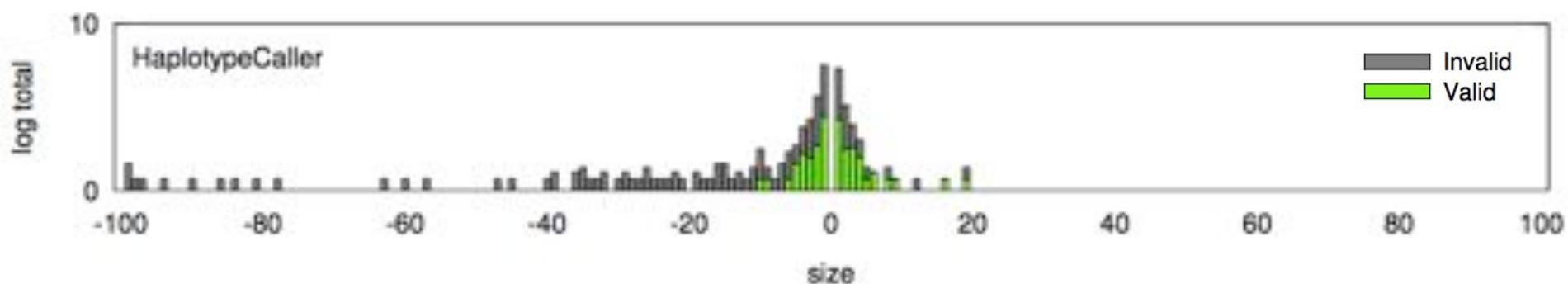
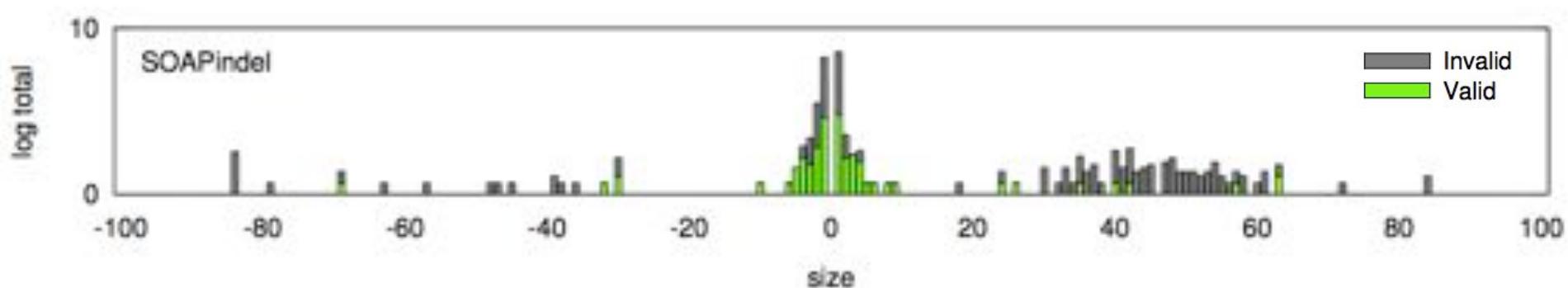
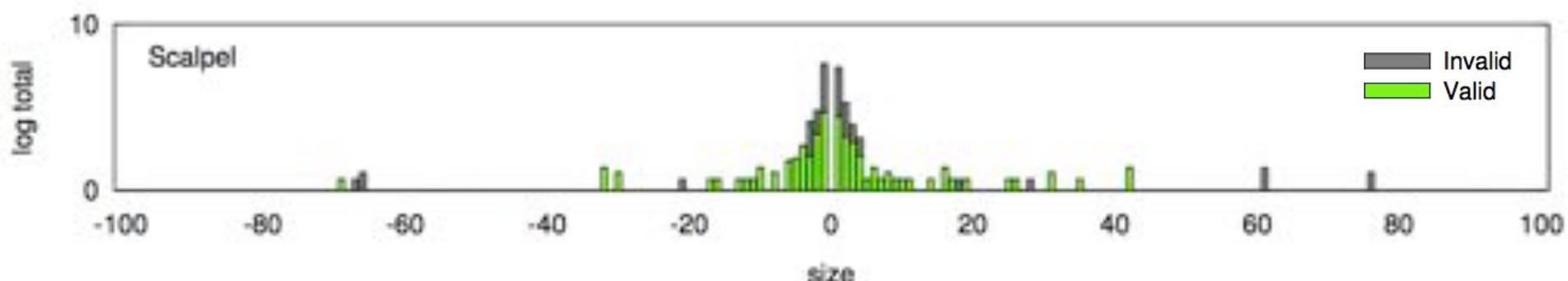
- 200 Scalpel
- 200 GATK Haplotype Caller
- 200 SOAPindel
- 200 within the intersection
- 200 long indels (>30bp)



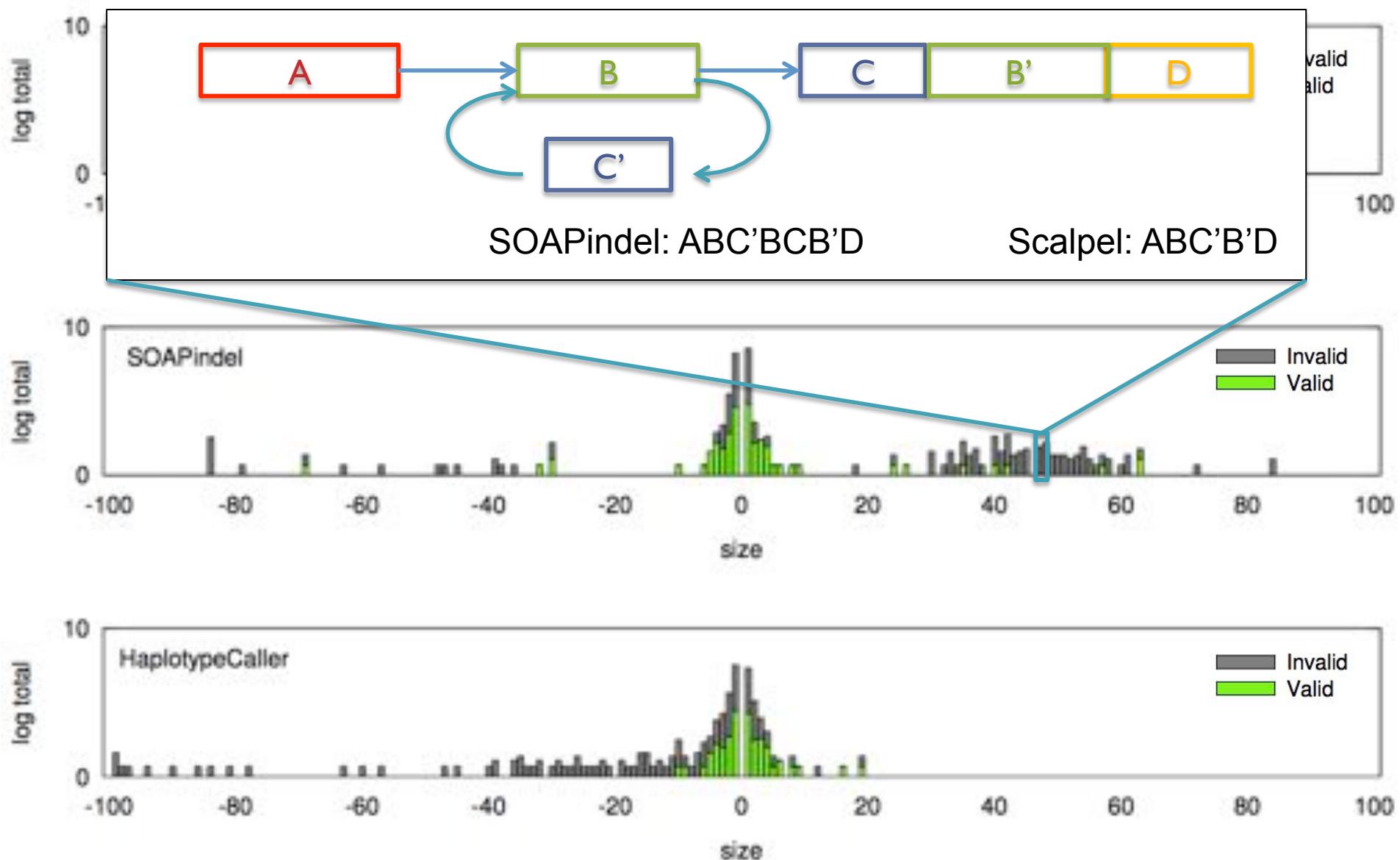
# Scalpel Indel Discovery



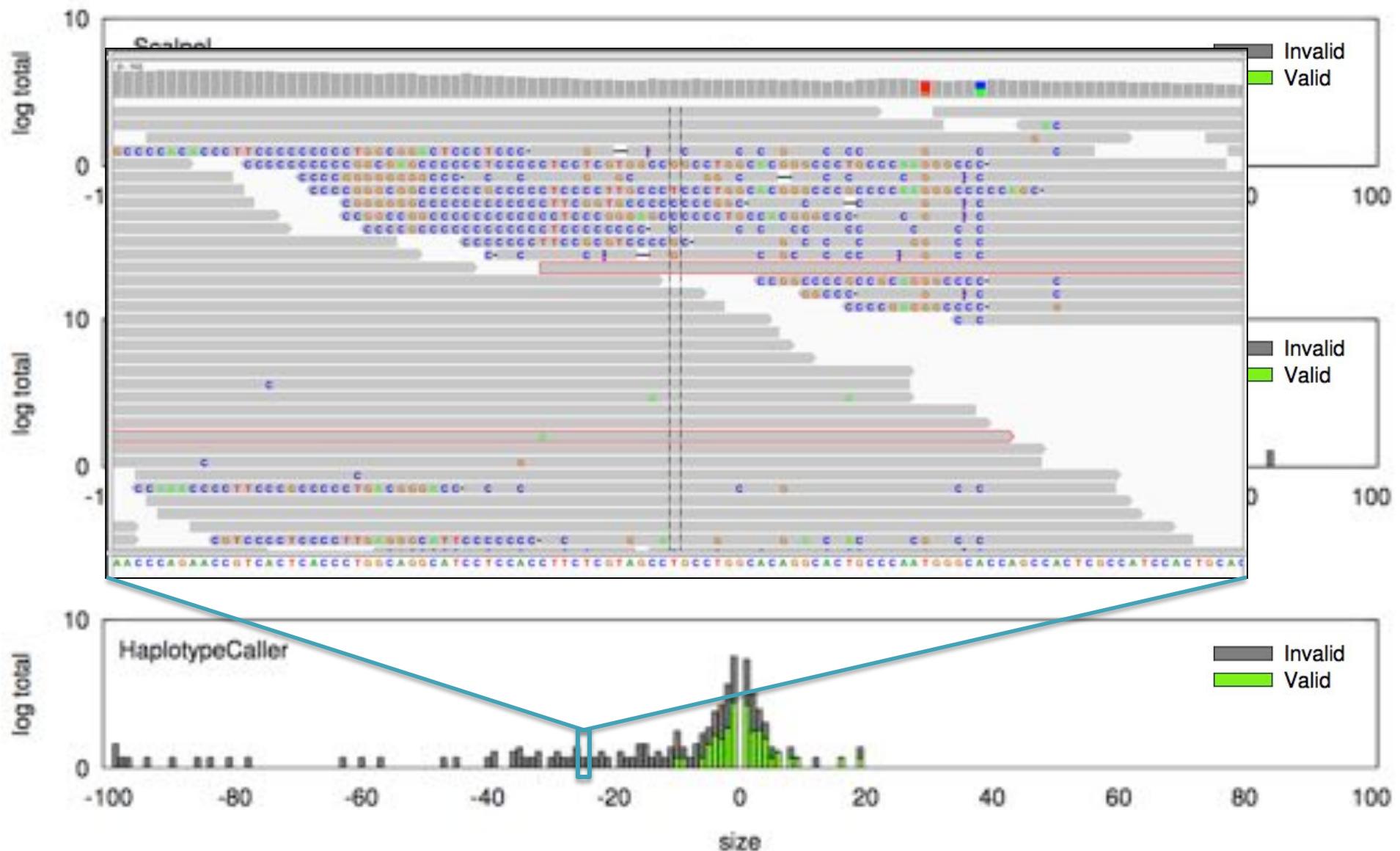
# Scalpel Indel Discovery



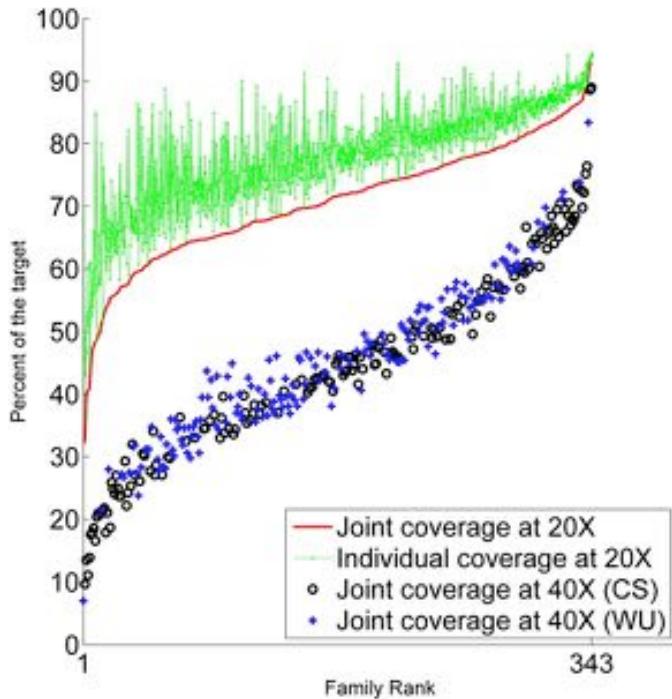
# Scalpel Indel Discovery



# Scalpel Indel Discovery



# Exome sequencing of the SSC



Last year saw 3 reports of >593 families from the Simons Simplex Collection

- Parents plus one child with autism and one non-autistic sibling
- All reported strong enrichment for de novo gene killing mutations (nonsense, frameshift, splice site mutations)
- Iossifov (343) and O’Roak (50) used GATK, Sanders (200) didn’t attempt to identify indels

## De novo gene disruptions in children on the autism spectrum

Iossifov et al. (2012) *Neuron*. 74:2 285-299

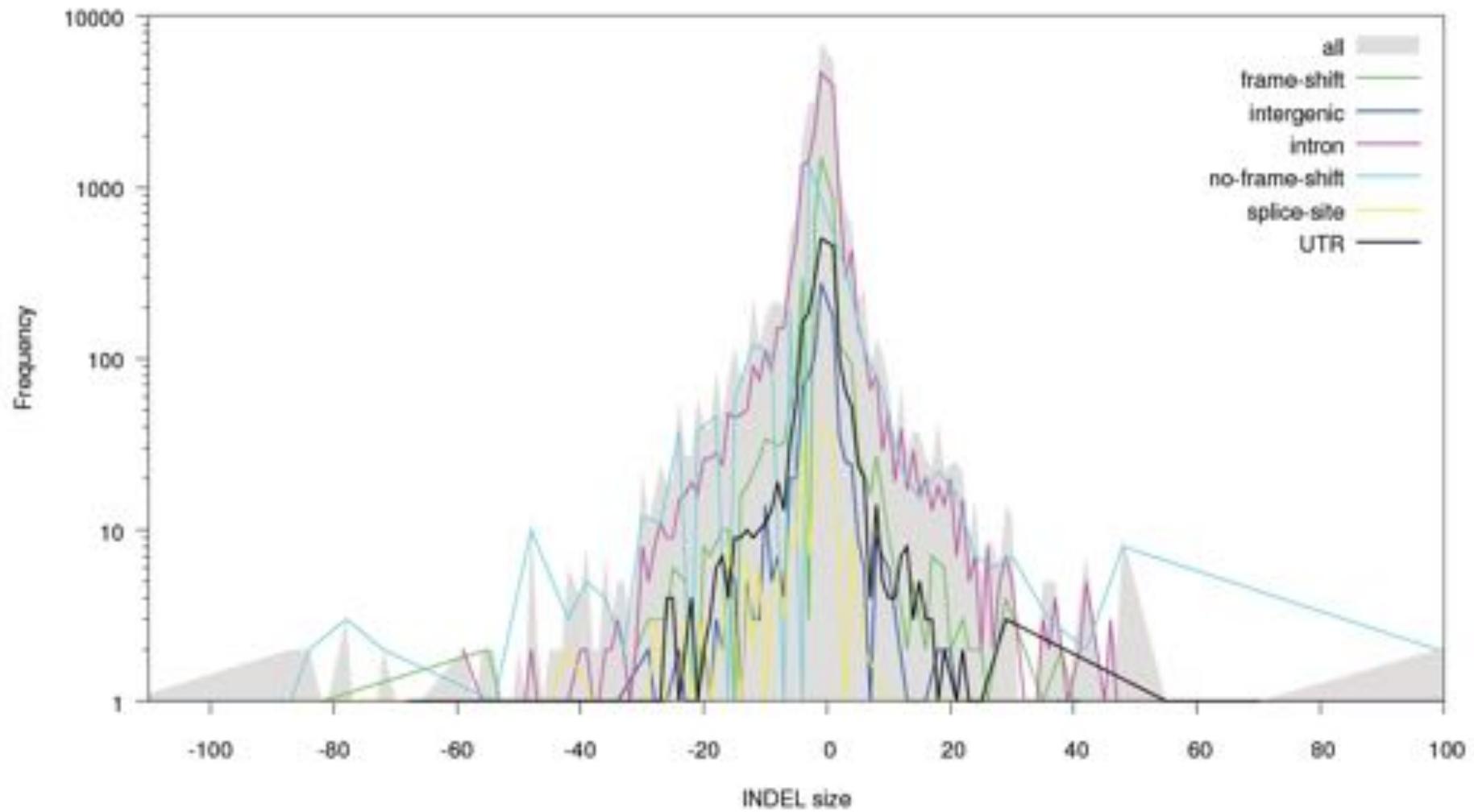
## De novo mutations revealed by whole-exome sequencing are strongly associated with autism

Sanders et al. (2012) *Nature*. 485, 237–241.

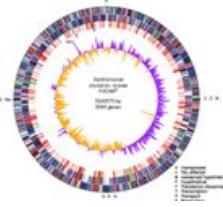
## Sporadic autism exomes reveal a highly interconnected protein network of de novo mutations

O’Roak et al. (2012) *Nature*. 485, 246–250.

# Revised Analysis of the SSC



Constructed database of >1M transmitted and de novo indels  
Strengthened enrichment for de novo frameshift mutations (35:16)  
Many new gene candidates identified, population analysis underway



# Summary



- Hybrid assembly let us combine the best characteristics of 2<sup>nd</sup> and 3<sup>rd</sup> gen sequencing
  - Long reads and good coverage are the keys to a good de novo assembly
  - Single contig de novo assemblies of entire microbial chromosomes are now routine; Single contig de novo assemblies of entire plant and animal chromosomes on the horizon
- Assembly is the missing link towards high accuracy indel mutation discovery
  - Allows the algorithm to break free from the expectations of the reference
  - Pinpointing de novo mutations require both high sensitivity and specificity
- We are starting to apply these technologies to discover significant biology that is otherwise impossible to measure

# Acknowledgements

## Schatz Lab

Giuseppe Narzisi  
Shoshana Marcus  
James Gurtowski  
Srividya  
Ramakrishnan  
Hayan Lee  
Rob Aboukhalil  
Mitch Bekritsky  
Charles Underwood  
Tyler Gavin  
Alejandro Wences  
Greg Vurture  
Eric Biggers  
Aspyn Palatnick

## CSHL

Hannon Lab  
Gingeras Lab  
Jackson Lab  
Iossifov Lab  
Levy Lab  
Lippman Lab  
Lyon Lab  
Martienssen Lab  
McCombie Lab  
Ware Lab  
Wigler Lab

IT Department





*See you at*

*Genome Informatics*

*Oct 30 – Nov 2*

<http://schatzlab.cshl.edu>  
@mike\_schatz