

# Climate Water Loss Experiment - Plasma Osmolality Data Wrangling

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## Packages

```
`%nin%` = Negate(`%in%`)
if (!require("tidyverse")) install.packages("tidyverse")

## Loading required package: tidyverse

## -- Attaching packages ----- tidyverse 1.3.0 --

## v ggplot2 3.3.3    v purrr   0.3.4
## v tibble  3.0.6    v dplyr   1.0.2
## v tidyr   1.1.2    v stringr 1.4.0
## v readr   1.4.0    v forcats 0.5.0

## -- Conflicts ----- tidyverse_conflicts() --
## x dplyr::filter() masks stats::filter()
## x dplyr::lag()    masks stats::lag()
```

```
library("tidyverse") # workflow and plots
```

## Background and Goals

Blood was drawn from the postorbital sinus of adult male *Sceloporus occidentalis* between June - August 2021. After centrifuging and separating, plasma was run on a VAPRO vapor pressure osmometer in 1-4 replicates, when plasma volume allowed. In this R script, I check the distribution of replicates, omit outliers, and average remaining replicates. The final values will be more precise and accurate estimates of the true plasma osmolality for each lizard, and those values will be used in the capture\_analysis and experiment\_analysis R script files. Please refer to the published scientific journal article for full details.

## Load Data

```
osml_reps <- read.csv("./data/osmolality.csv",
  na.strings = c("", "NA"),
  header = TRUE
) %>%
  dplyr::mutate(date_blood_drawn = as.Date(date_blood_drawn,
    format = "%m/%d/%y"),
    date_osmom_run = as.Date(date_osmom_run,
    format = "%m/%d/%y"),
    time_osmom_run = as.POSIXct(time_osmom_run,
    format = "%H:%M"),
    individual_ID = as.factor(individual_ID),
    replicate_no = as.factor(replicate_no),
    osmolality_mmol_kg = as.numeric(osmolality_mmol_kg)
  )
summary(osml_reps)
```

```
##  date_blood_drawn    date_osmom_run    time_osmom_run
##  Min.      :2021-06-16  Min.      :2021-06-16  Min.      :2022-10-03 01:00:00
##  1st Qu.:2021-06-30  1st Qu.:2021-06-30  1st Qu.:2022-10-03 10:38:00
##  Median :2021-07-24  Median :2021-07-24  Median :2022-10-03 12:48:00
##  Mean   :2021-07-22  Mean   :2021-07-22  Mean   :2022-10-03 12:58:11
##  3rd Qu.:2021-08-16  3rd Qu.:2021-08-16  3rd Qu.:2022-10-03 15:14:00
##  Max.   :2021-09-01  Max.   :2021-09-02  Max.   :2022-10-03 21:22:00
##                                     NA's      :211
##  individual_ID  replicate_no  osmolality_mmol_kg  notes
##  339      :   13    1:544      Min.      :247.0    Mode:logical
##  206      :   12    2:531      1st Qu.:336.0    NA's:1484
##  207      :   12    3:406      Median   :351.0
##  208      :   12    4:  3      Mean     :357.3
##  211      :   12              3rd Qu.:370.0
##  214      :   12              Max.      :596.0
##  (Other):1411
```

## Check Data

### Dates

Blood was drawn on day 0, 4, 8, and 10 of the experiment. Create a list of the dates expected to have blood draw data, then determine whether I have dates outside those.

Trail 1: June 16-24 Trail 2: June 26 - July 4 Trial 3: July 20-28 Trial 4: August 8-16 Trial 5: August 22-30

```
      # trial 1
expected_dates <- as.Date(c("2021-06-16", "2021-06-20",
      "2021-06-24", "2021-06-26",
      # trial 2
      "2021-06-26", "2021-06-30",
      "2021-07-04", "2021-07-06",
      # trial 3
      "2021-07-20", "2021-07-24",
      "2021-07-28", "2021-07-30",
      # trial 4
      "2021-08-08", "2021-08-12",
      "2021-08-16", "2021-08-18",
      # trial 5
      "2021-08-22", "2021-08-26",
      "2021-08-30", "2021-09-01"))

# how many dates in our data do not match expected dates (should print zero)
length(osml_reps$date_blood_drawn[osml_reps$date_blood_drawn %nin% expected_dates]
)
```

```
## [1] 0
```

There are zero blood draw dates that are not in our expected list.

## Number of Blood Draws

Each lizard should have had their blood drawn on 4 different dates, unless they were taken out of the experiment early.

```
# get ID's of the individuals that completed treatment
individuals <- read.csv("./data/tmt_assignments.csv") %>%
  dplyr::select(conclusion, individual_ID) %>%
  dplyr::filter(conclusion == "complete") %>%
  mutate(individual_ID = as.factor(individual_ID),
         conclusion = as.factor(conclusion))
summary(individuals)
```

```
##      conclusion  individual_ID
## complete:134    201      : 1
##              202      : 1
##              203      : 1
##              204      : 1
##              205      : 1
##              206      : 1
##              (Other):128
```

```
# calculate the number of dates for each individual
osml_reps %>%
  dplyr::filter(individual_ID %in% individuals$individual_ID) %>%
  group_by(individual_ID, date_blood_drawn) %>%
  summarise(n()) %>%
  group_by(individual_ID) %>%
  summarise(n()) %>%
  arrange(n())
```

```
## `summarise()` regrouping output by 'individual_ID' (override with `.groups` argument)
## `summarise()` ungrouping output (override with `.groups` argument)
## # A tibble: 134 x 2
##   individual_ID `n()`
##   <fct>         <int>
## 1 201             4
## 2 202             4
## 3 203             4
## 4 204             4
## 5 205             4
## 6 206             4
## 7 207             4
## 8 208             4
## 9 209             4
## 10 210            4
## # ... with 124 more rows
```

Wahoo, every lizard has blood draws from 4 different dates.

## Replicates

Now, I will try to identify outliers within the replicates for a given individual on a given date. There must be at least 3 replicates to do this, so the first thing I need to do is figure out which individuals/dates have enough replicates, then subset my data to be only those individuals.

### Individuals w 3+ Replicates

```
# identify individuals with 3-4 reps
enuf_reps <- osml_reps %>%
  group_by(individual_ID, date_blood_drawn) %>%
  mutate(count = n()) %>%
  dplyr::filter(count > 2) %>%
  arrange(count)
enuf_reps

## # A tibble: 1,224 x 8
## # Groups:   individual_ID, date_blood_drawn [407]
##   date_blood_drawn date_osmom_run time_osmom_run individual_ID
##   <date>           <date>         <dtm>         <fct>
## 1 2021-06-16       2021-06-16       2022-10-03 11:00:00 201
## 2 2021-06-16       2021-06-16       2022-10-03 11:00:00 201
## 3 2021-06-16       2021-06-16       2022-10-03 11:00:00 201
## 4 2021-06-16       2021-06-16       2022-10-03 11:00:00 202
## 5 2021-06-16       2021-06-16       2022-10-03 11:00:00 202
## 6 2021-06-16       2021-06-16       2022-10-03 11:00:00 202
## 7 2021-06-16       2021-06-16       2022-10-03 11:10:00 203
## 8 2021-06-16       2021-06-16       2022-10-03 11:10:00 203
## 9 2021-06-16       2021-06-16       2022-10-03 11:10:00 203
## 10 2021-06-16      2021-06-16       2022-10-03 11:25:00 205
## # ... with 1,214 more rows, and 4 more variables: replicate_no <fct>,
## #   osmolality_mmol_kg <dbl>, notes <lgl>, count <int>
```

```

# identify individuals with 1-2 reps
not_reps <- osml_reps %>%
  group_by(individual_ID, date_blood_drawn) %>%
  mutate(count = n()) %>%
  dplyr::filter(count < 3) %>%
  arrange(count)
not_reps

## # A tibble: 260 x 8
## # Groups:   individual_ID, date_blood_drawn [136]
##   date_blood_drawn date_osmom_run time_osmom_run individual_ID
##   <date>           <date>         <dtm>         <fct>
## 1 2021-06-24       2021-06-24     2022-10-03 09:26:00 203
## 2 2021-06-24       2021-06-26       NA              204
## 3 2021-06-24       2021-06-26       NA              223
## 4 2021-06-26       2021-06-26     2022-10-03 09:47:00 201
## 5 2021-06-26       2021-06-26     2022-10-03 11:40:00 230
## 6 2021-07-04       2021-07-05     2022-10-03 10:03:00 227
## 7 2021-07-06       2021-07-06     2022-10-03 09:59:00 239
## 8 2021-07-28       2021-07-28     2022-10-03 10:32:00 259
## 9 2021-07-28       2021-07-28     2022-10-03 12:07:00 274
## 10 2021-07-30      2021-07-31     2022-10-03 13:55:00 291
## # ... with 250 more rows, and 4 more variables: replicate_no <fct>,
## #   osmolality_mmol_kg <dbl>, notes <lgl>, count <int>

# check total obs still add to original 1484
nrow(enuf_reps) + nrow(not_reps)

## [1] 1484

nrow(enuf_reps) + nrow(not_reps) == nrow(osml_reps)

## [1] TRUE

```

## Assess Variation

We want the Coefficient of Variation (CV) among our technical replicates to be small. We need to calculate it to identify whether there may be outliers.

```

CVs <- enuf_reps %>%
  group_by(individual_ID, date_blood_drawn) %>%
  summarise(mean = mean(osmolality_mmol_kg),
            SD = sd(osmolality_mmol_kg),
            CV = (SD/mean) *100,
            min = min(osmolality_mmol_kg),
            max = max(osmolality_mmol_kg),
            osml_range = max - min
            )

## `summarise()` regrouping output by 'individual_ID' (override with ``.groups` argument)
summary(CVs)

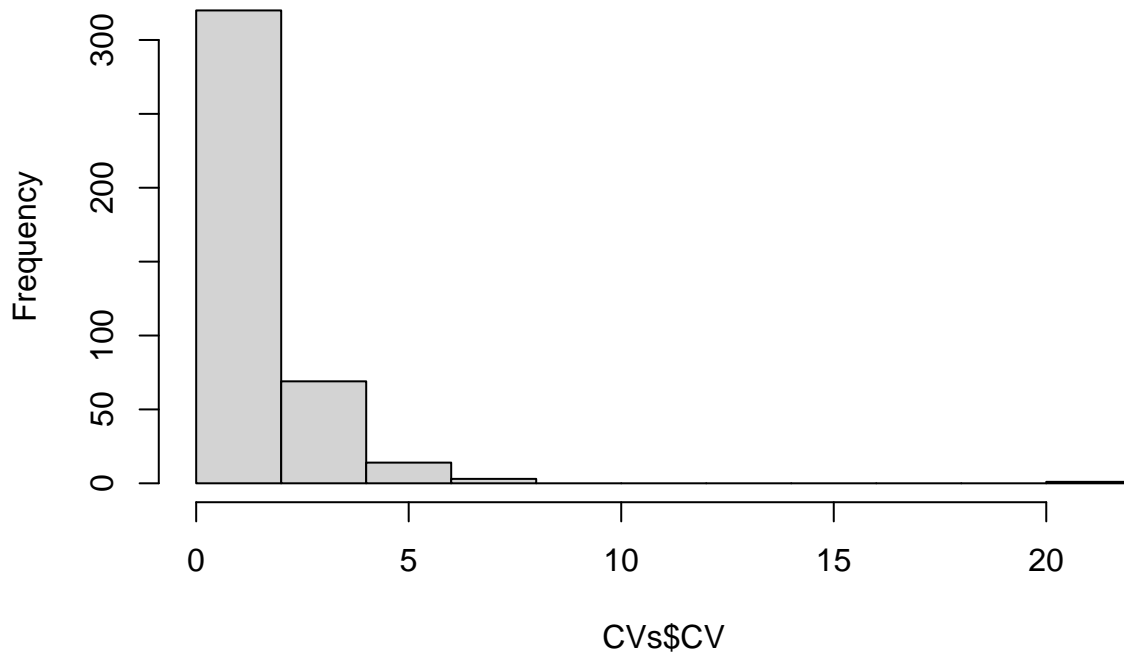
##   individual_ID date_blood_drawn      mean      SD
##   206      : 4   Min.   :2021-06-16   Min.   :295.3   Min.   : 0.000
##   207      : 4   1st Qu.:2021-06-30   1st Qu.:335.0   1st Qu.: 2.581
##   208      : 4   Median :2021-07-24   Median :350.0   Median : 4.359

```

```
## 211 : 4 Mean :2021-07-23 Mean :356.2 Mean : 5.596
## 214 : 4 3rd Qu.:2021-08-16 3rd Qu.:368.0 3rd Qu.: 6.658
## 220 : 4 Max. :2021-09-01 Max. :576.0 Max. :69.816
## (Other):383
## CV min max osml_range
## Min. : 0.0000 Min. :247.0 Min. :300.0 Min. : 0.00
## 1st Qu.: 0.7278 1st Qu.:330.5 1st Qu.:339.0 1st Qu.: 5.00
## Median : 1.2021 Median :345.0 Median :356.0 Median : 8.00
## Mean : 1.5627 Mean :351.1 Mean :361.7 Mean :10.66
## 3rd Qu.: 1.8746 3rd Qu.:364.0 3rd Qu.:374.0 3rd Qu.:12.50
## Max. :21.4600 Max. :566.0 Max. :596.0 Max. :134.00
##
```

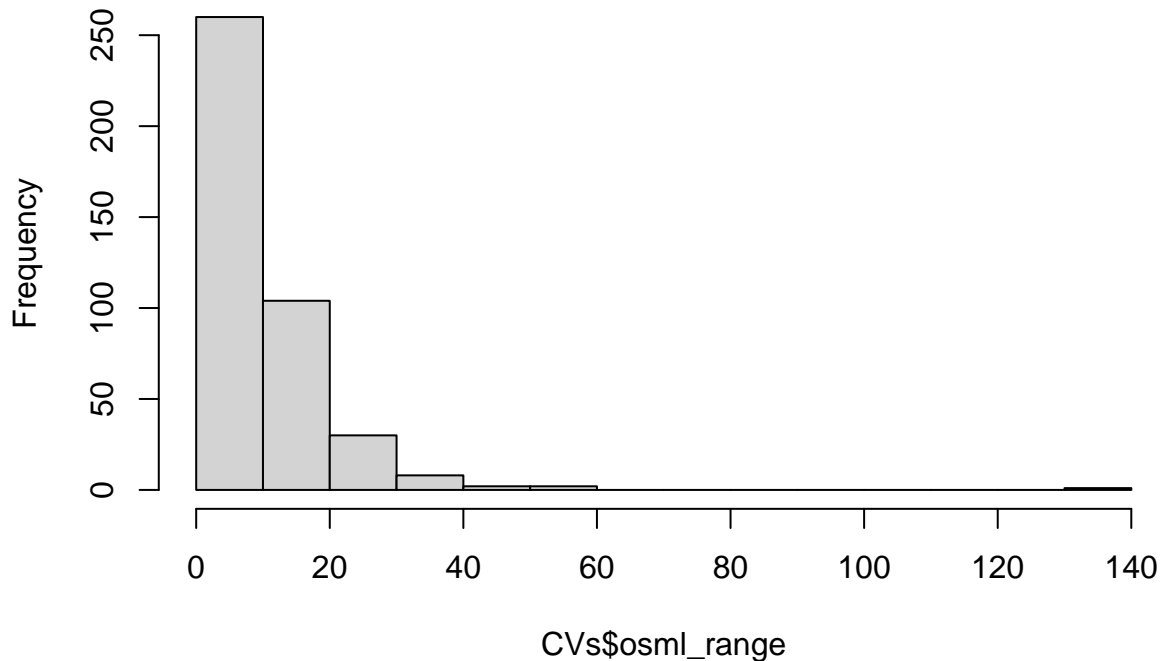
```
hist(CVs$CV)
```

**Histogram of CVs\$CV**



```
hist(CVs$osml_range)
```

## Histogram of CVs\$osml\_range



Ideally, CV would be <10-15%. If it's larger, and one of the replicates is very different than the others, we can assume that the replicates that are closer together are more reliable.

The CV is >10 for only one lizard on one date, so our replicates are already likely to accurately represent the true value. We don't want to lose accuracy by searching for precision, so we will only remove the one point driving the enormous CV value.

## Find Outliers Visually

```
# write function to find outliers for each individual on each date
find_outliers <- function(df) {

  # initiate a for loop to go through every who in df
  for(indiv_ch in unique(df$individual_ID)) {

    # select data for only the individual of interest
    df_sub <- df %>%
      dplyr::filter(individual_ID == as.numeric(indiv_ch))

    # make a boxplot
    ggplot(df_sub) +
      geom_boxplot(aes(x = as.factor(date_blood_drawn),
                       y = osmolality_mmol_kg,
                       fill = as.factor(date_blood_drawn))) +
      ggtitle(paste("Individual", indiv_ch)) +
      theme_classic() -> plot

    # print/save
    print(plot)
```

```

}

}

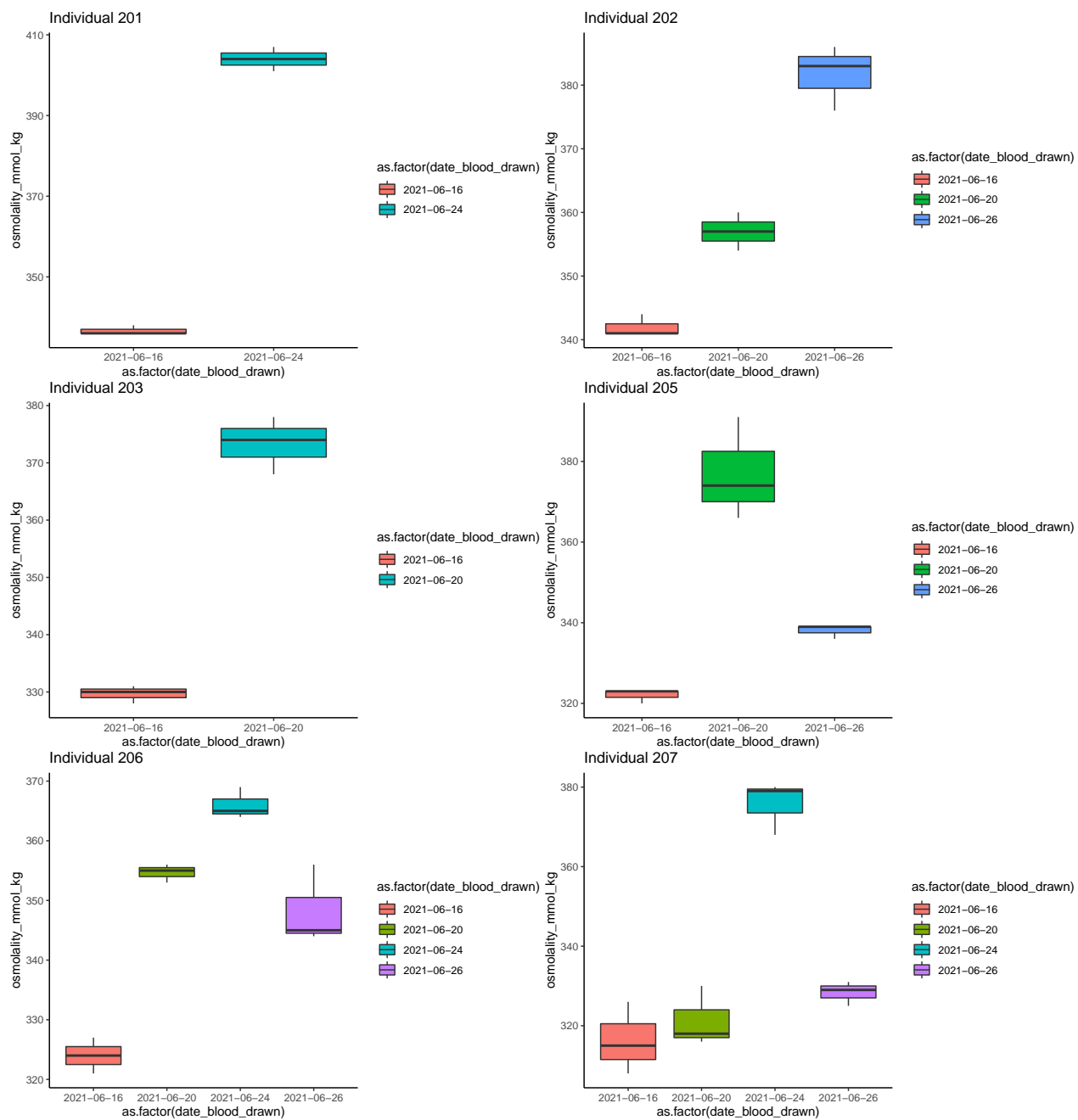
```

Now apply the function to the data:

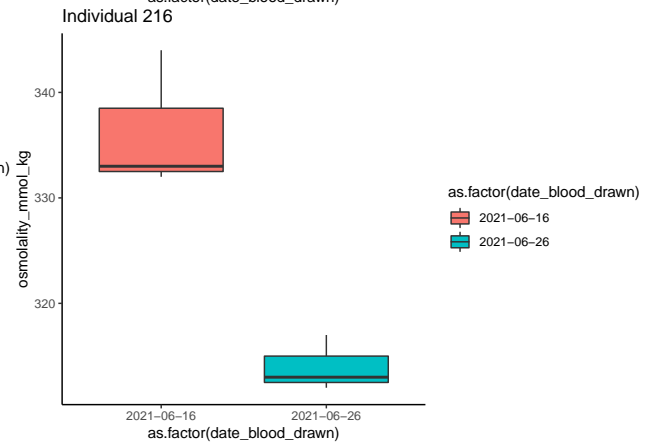
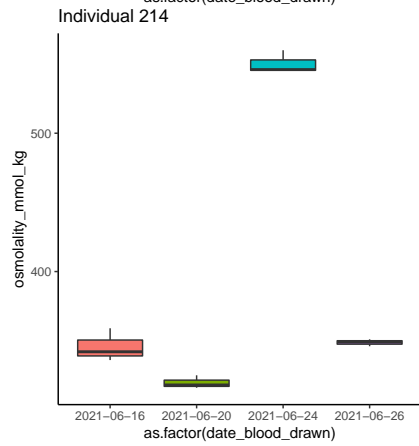
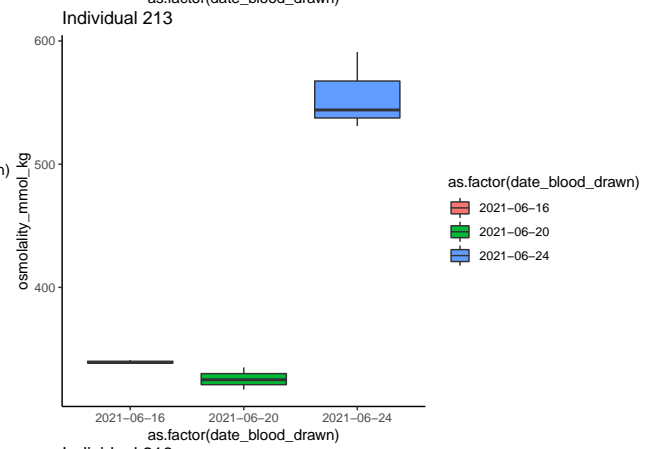
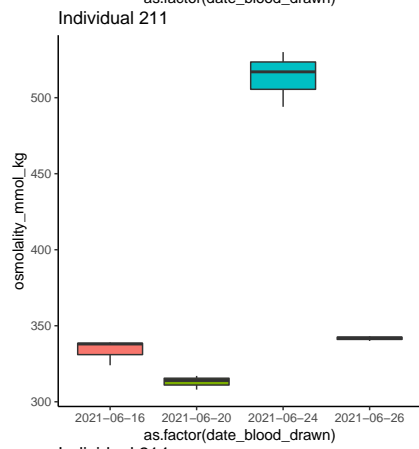
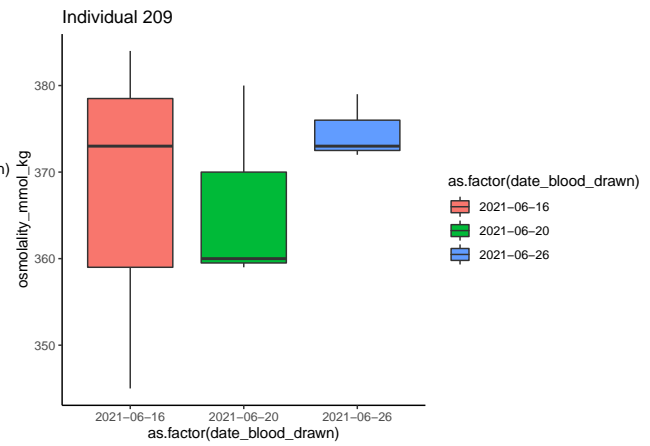
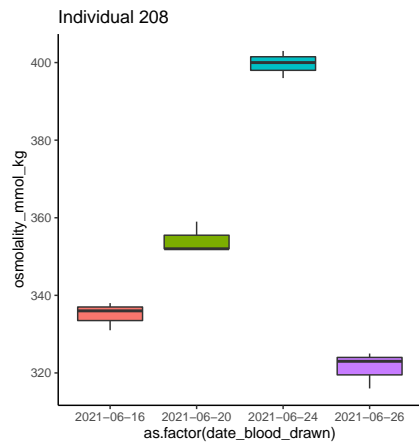
```

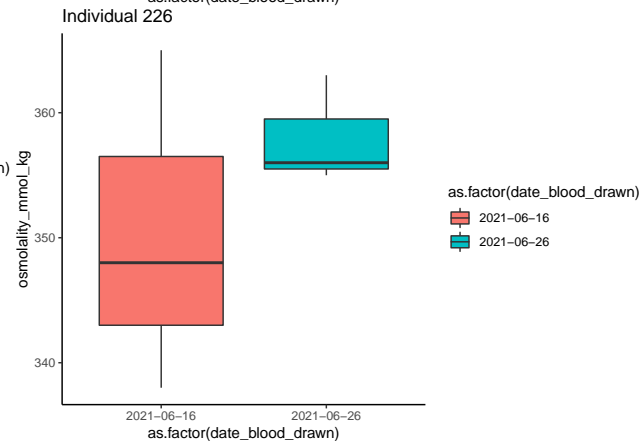
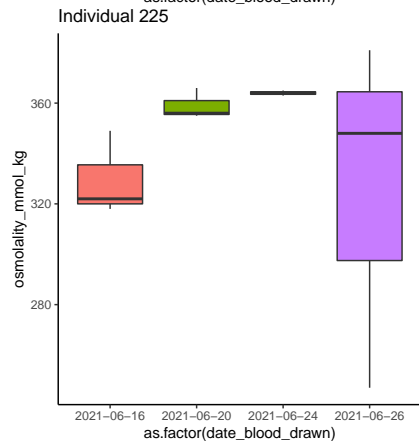
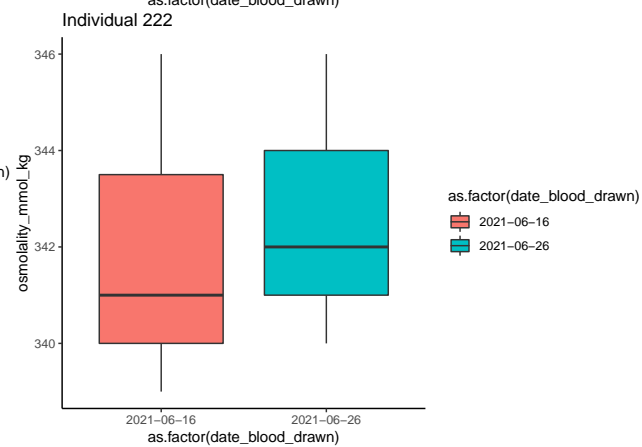
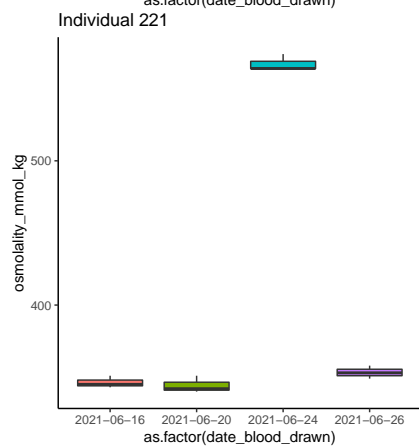
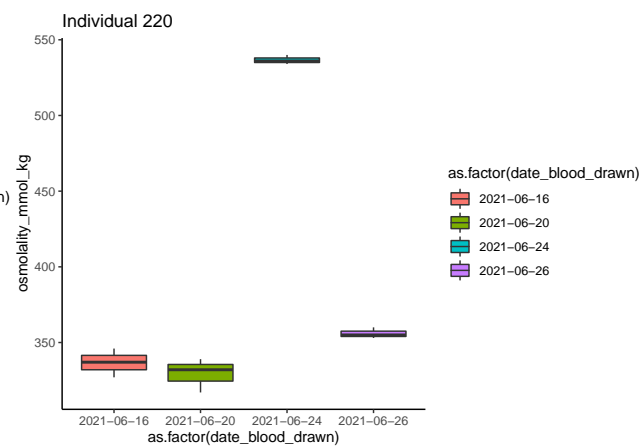
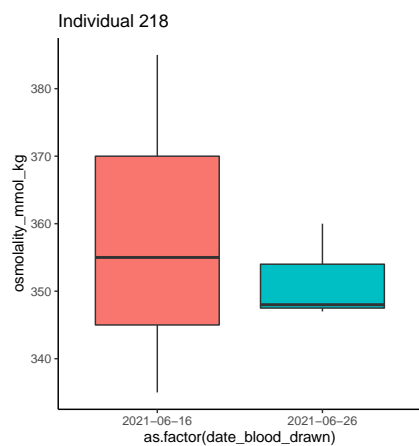
par(mfrow = c(71, 2))
find_outliers(enuf_reps)
par(mfrow = c(1, 1))

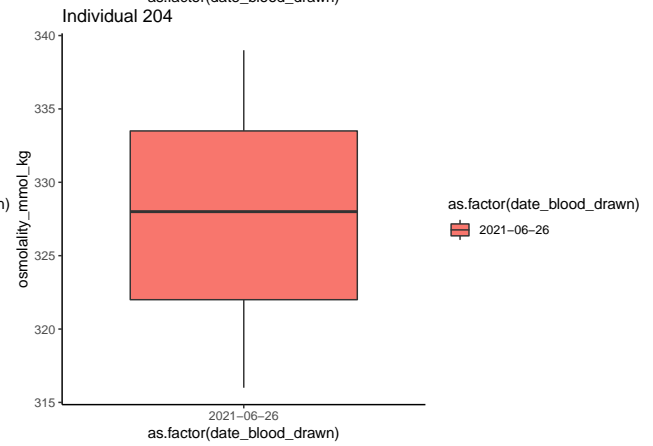
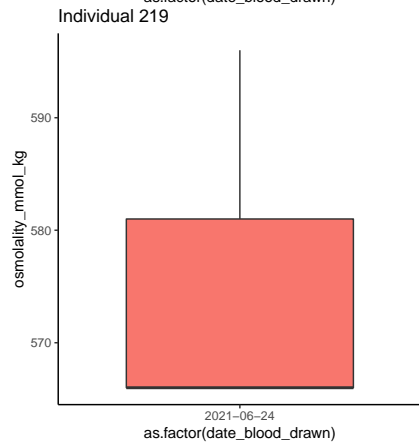
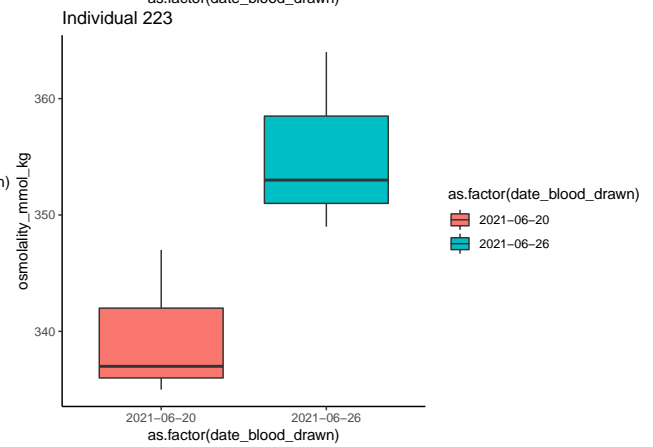
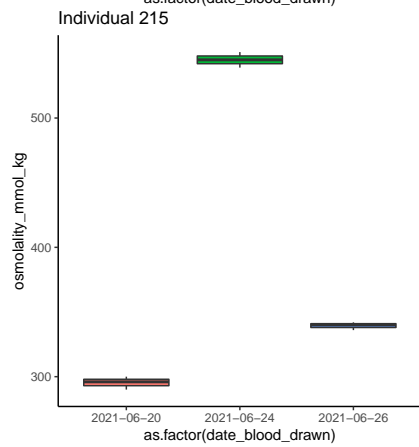
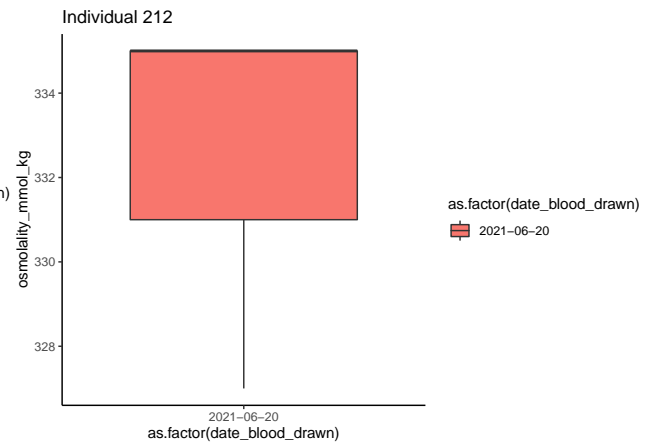
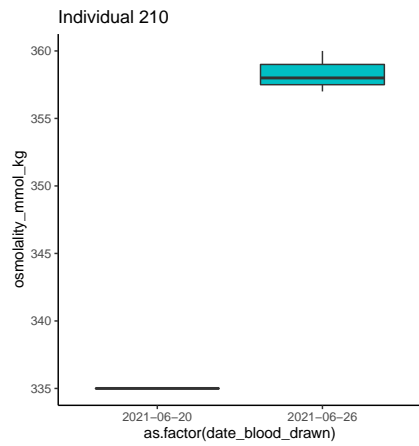
```

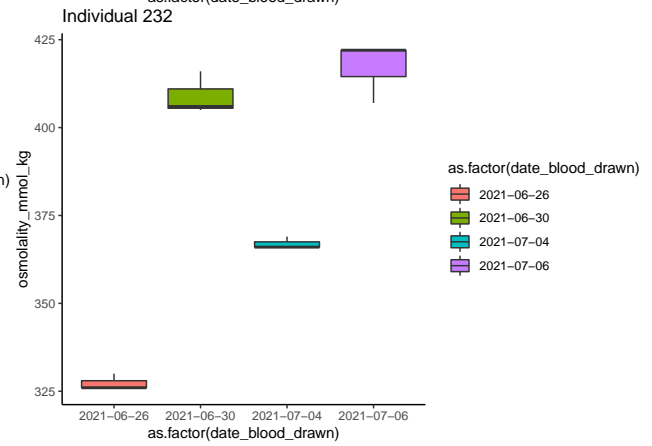
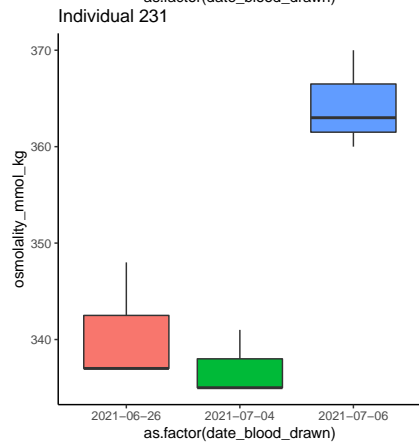
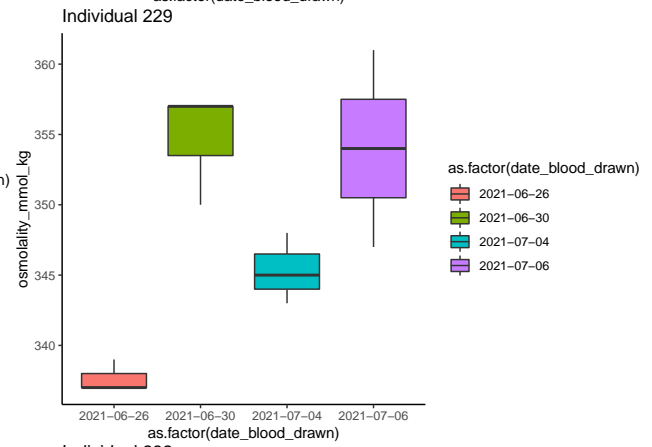
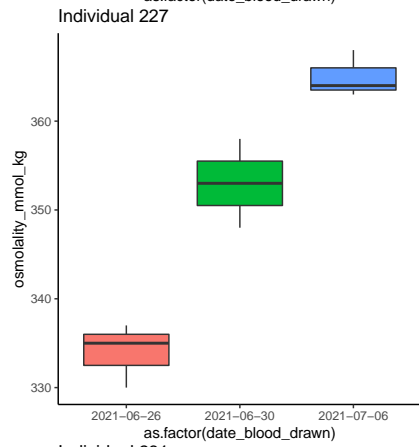
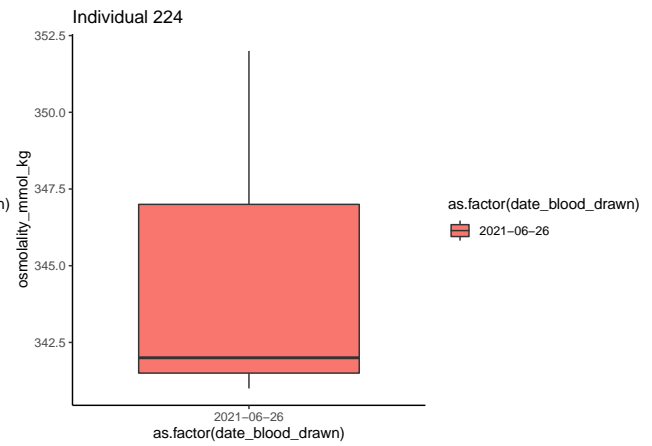
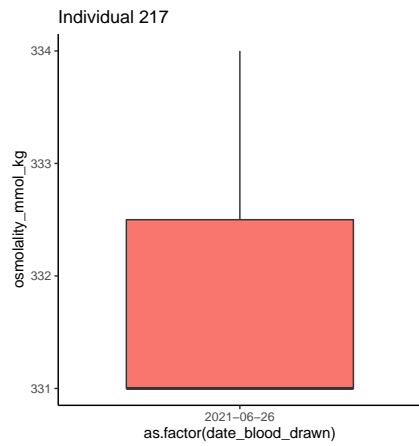


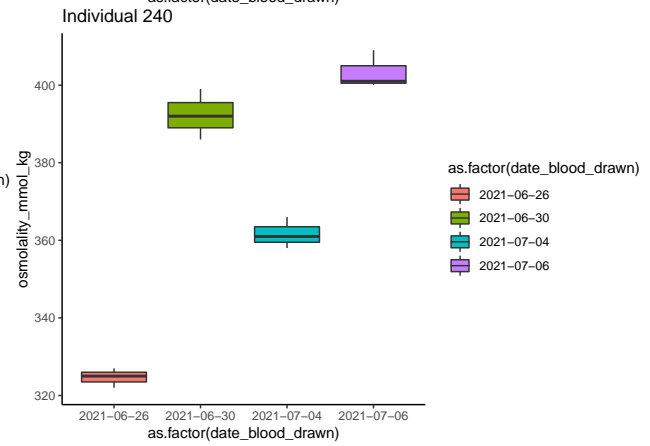
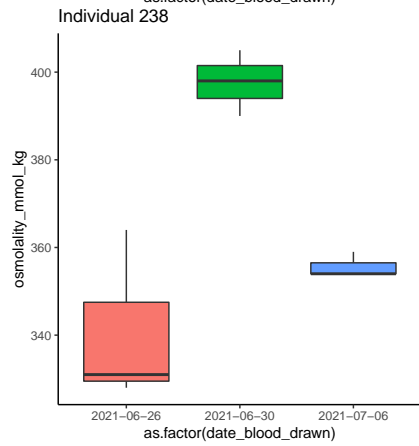
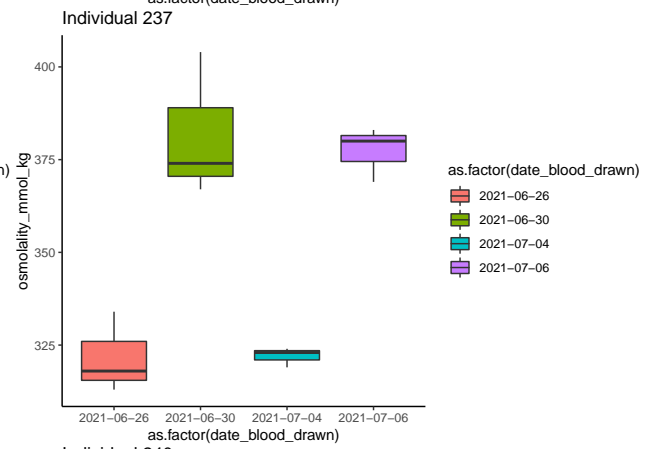
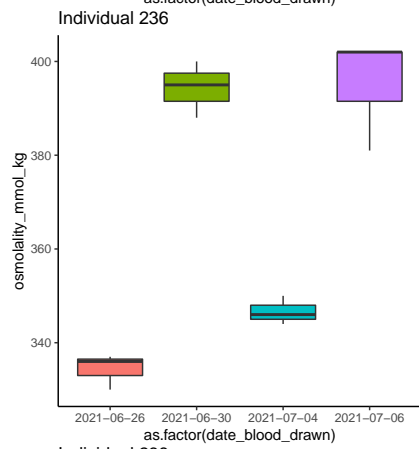
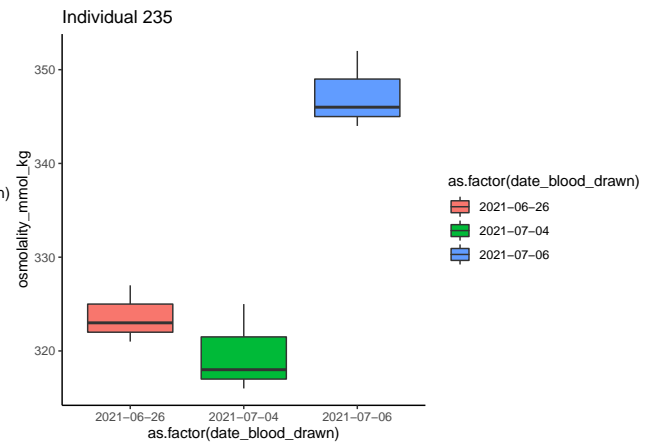
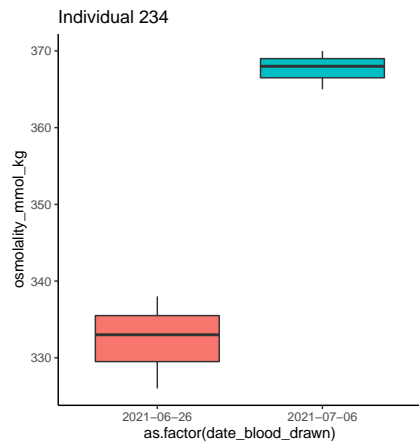


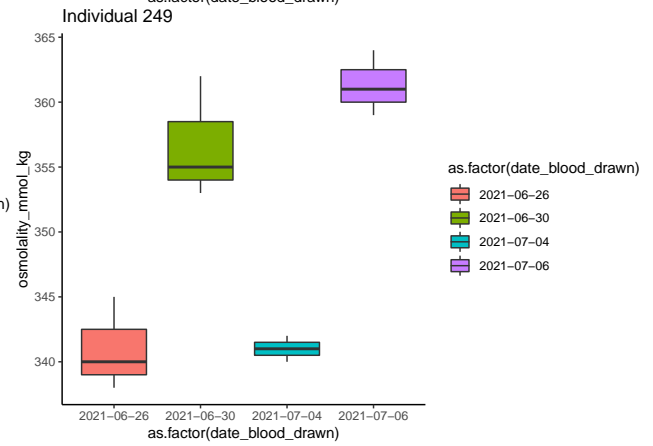
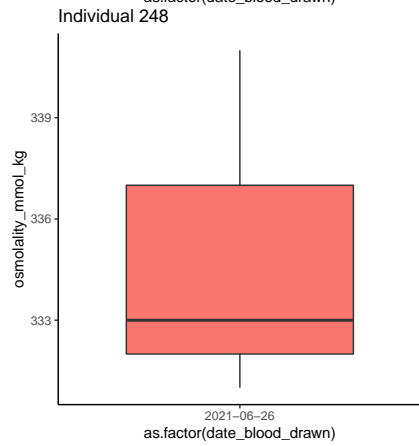
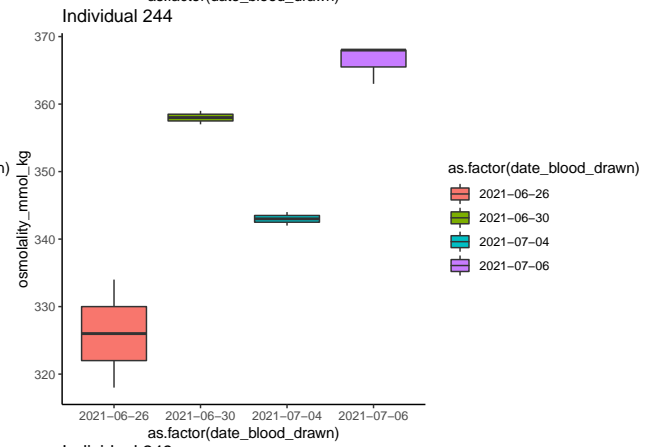
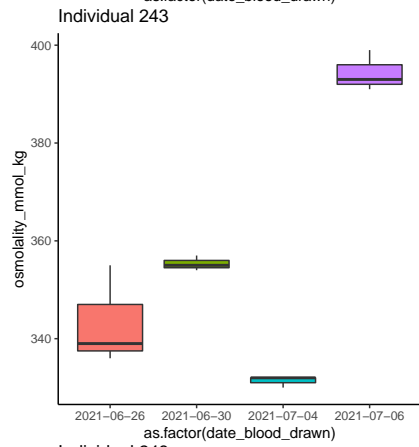
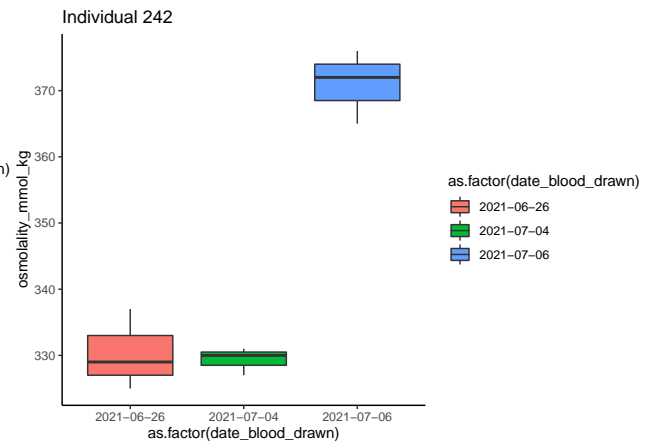
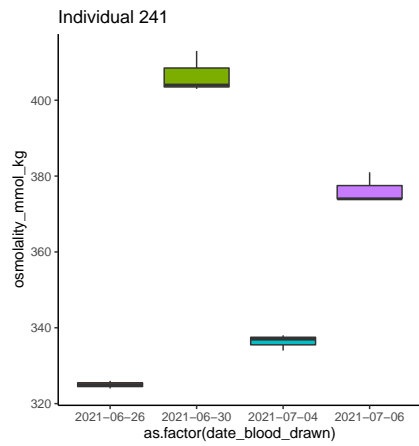


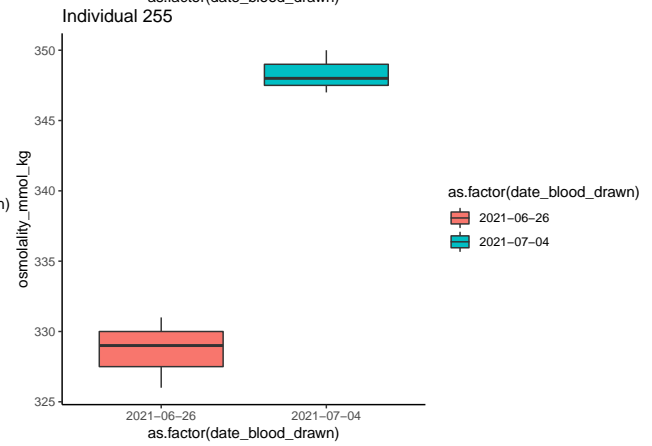
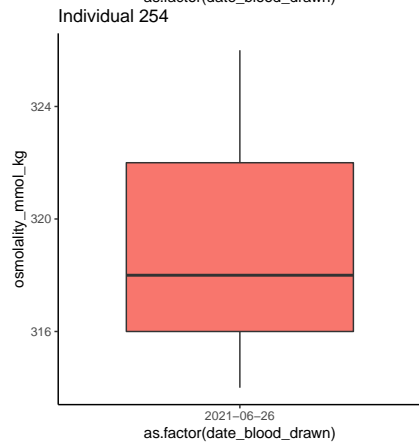
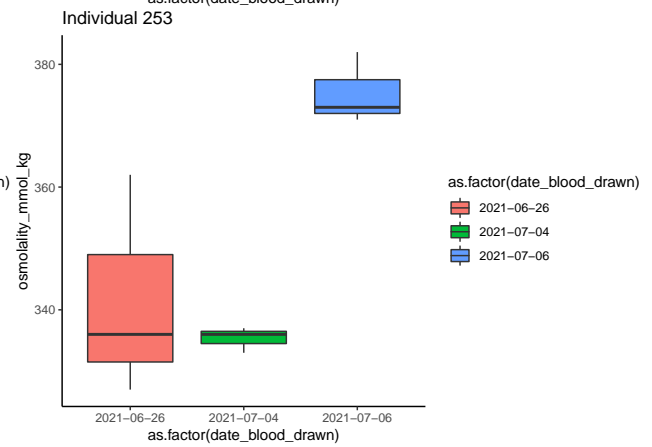
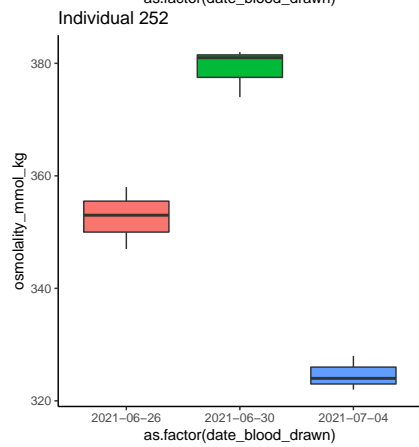
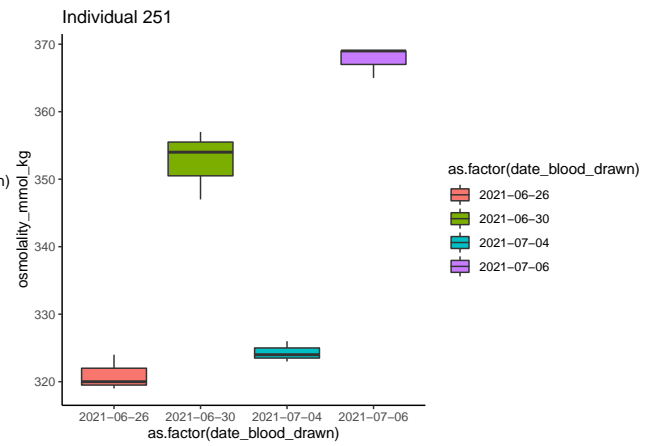
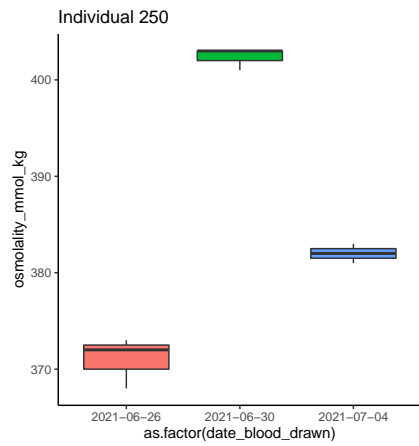


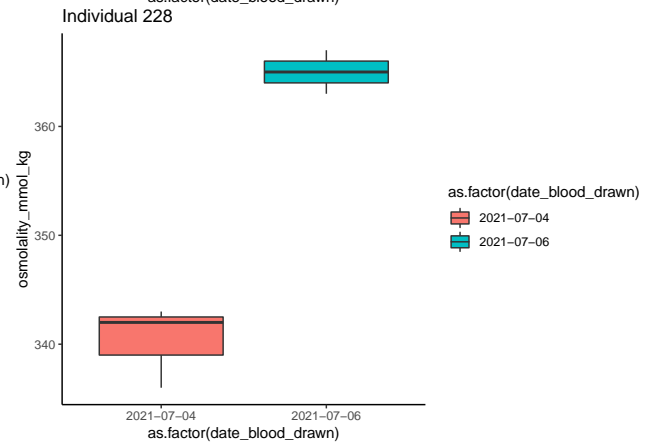
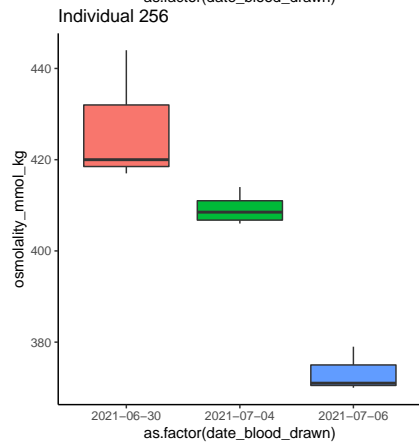
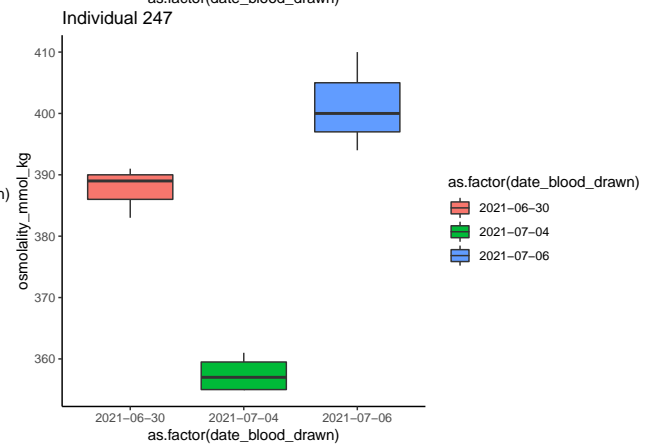
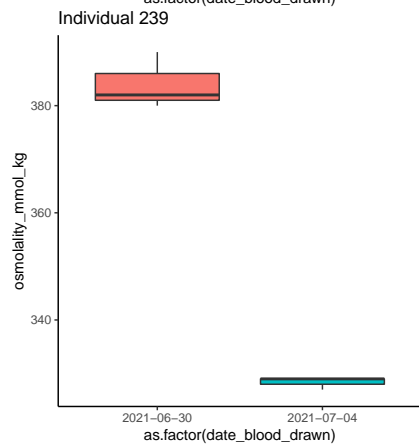
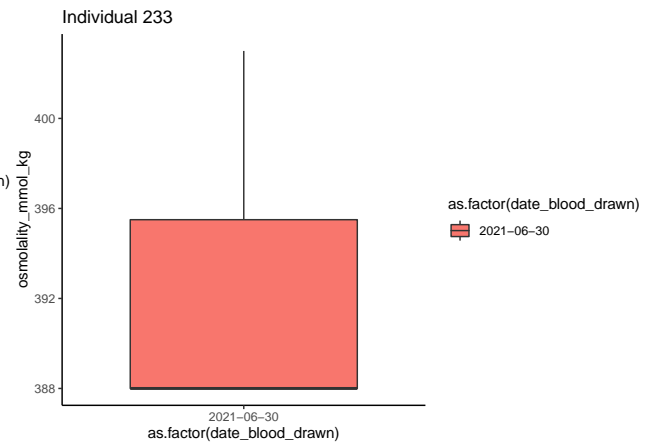
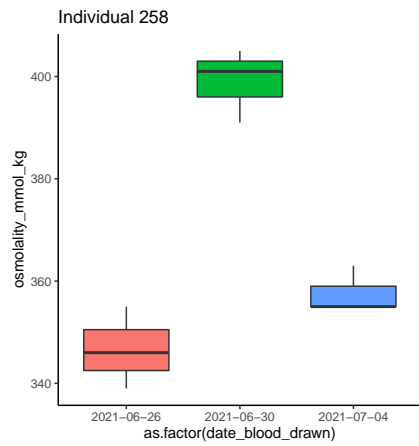




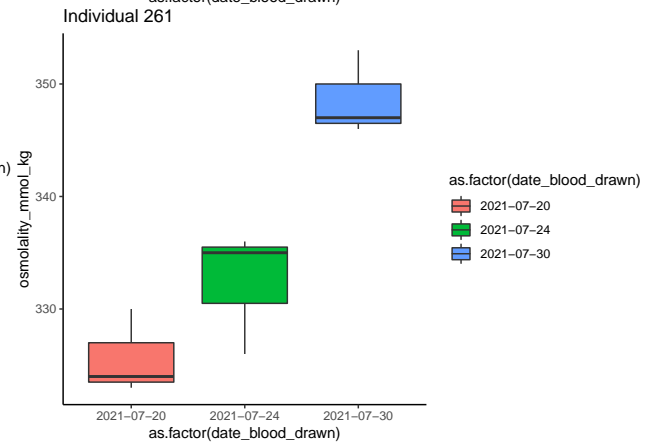
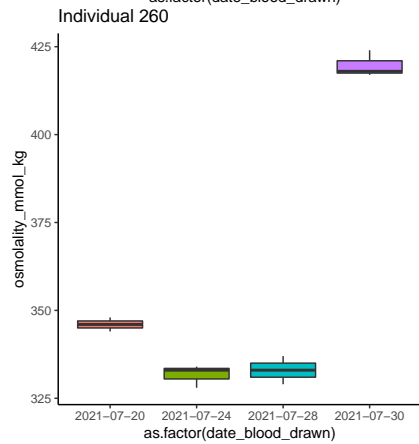
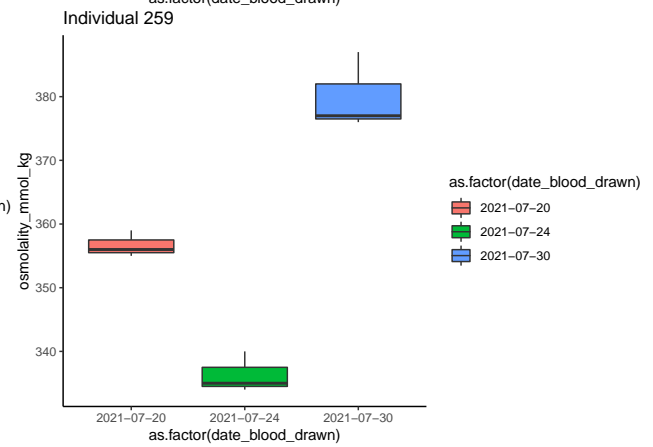
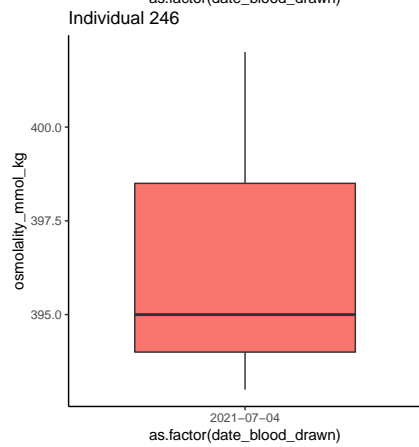
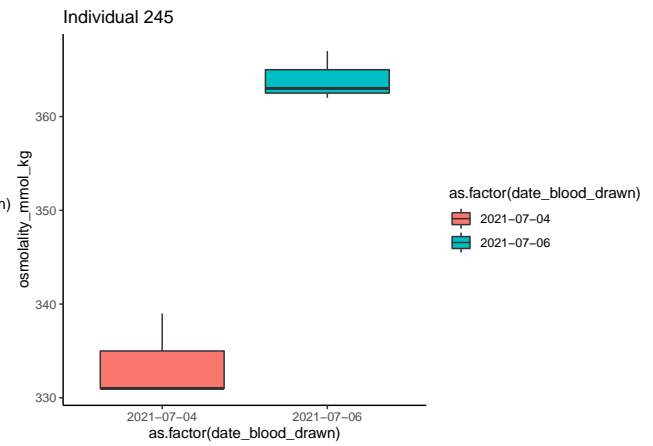
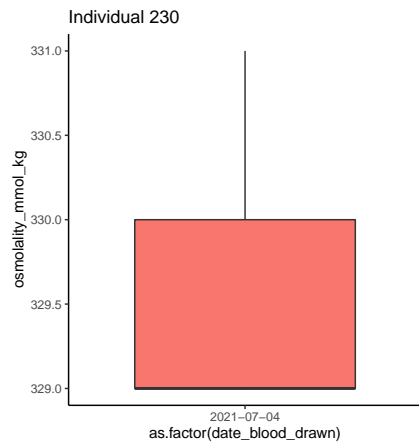


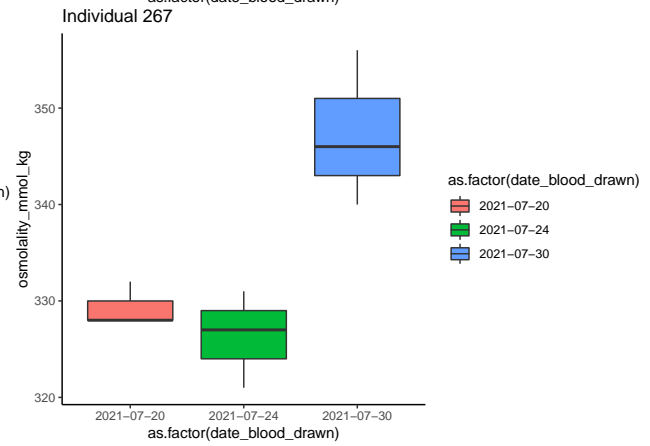
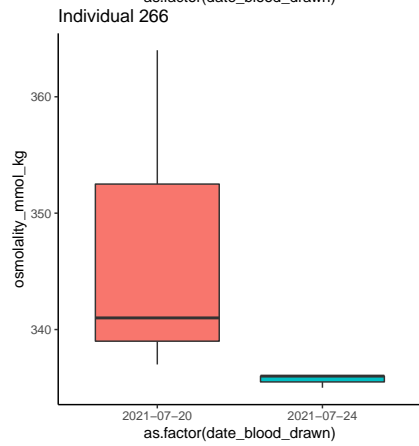
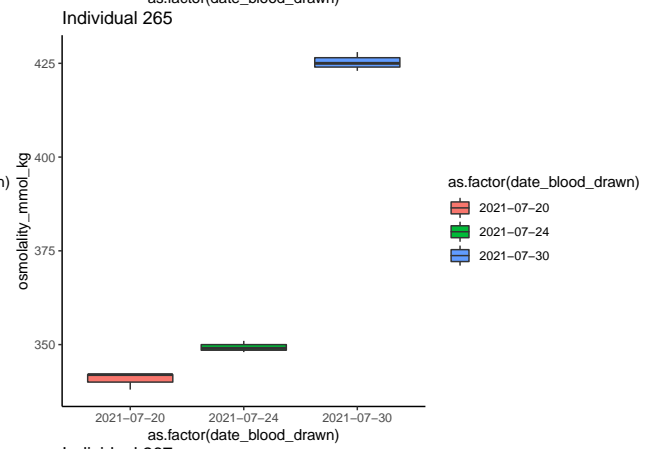
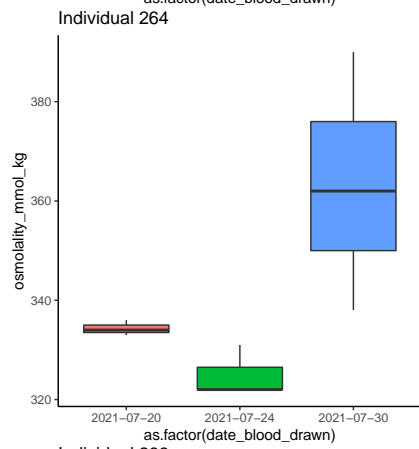
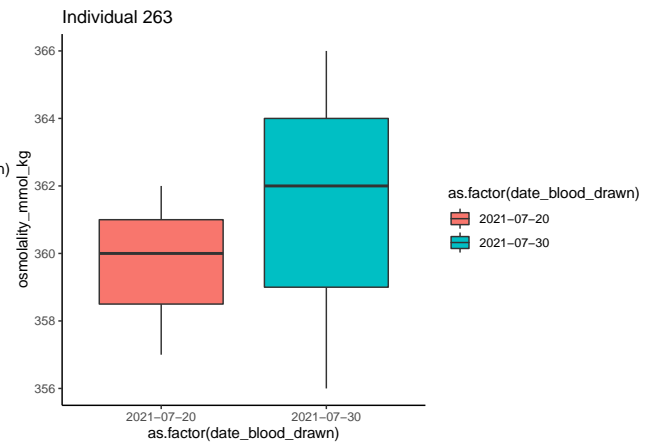
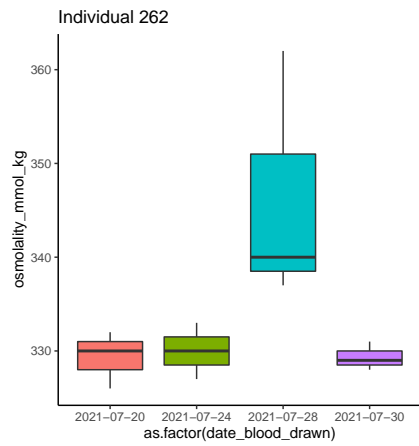


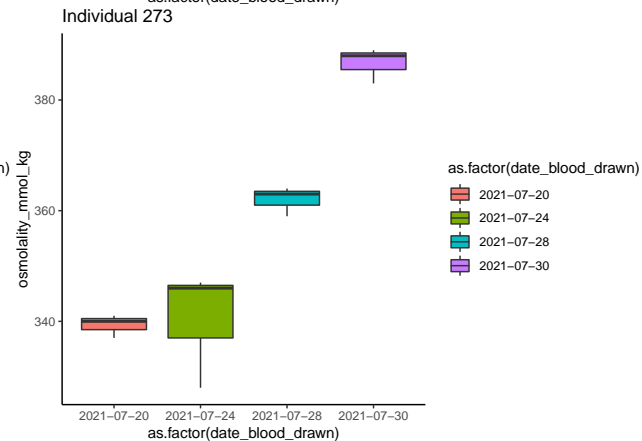
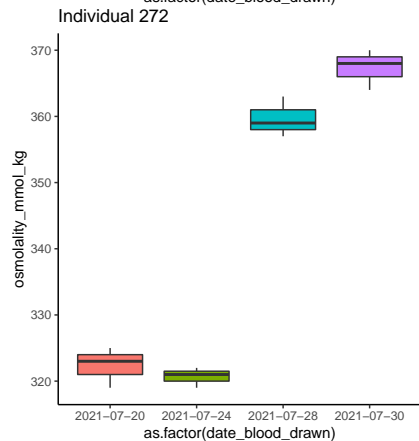
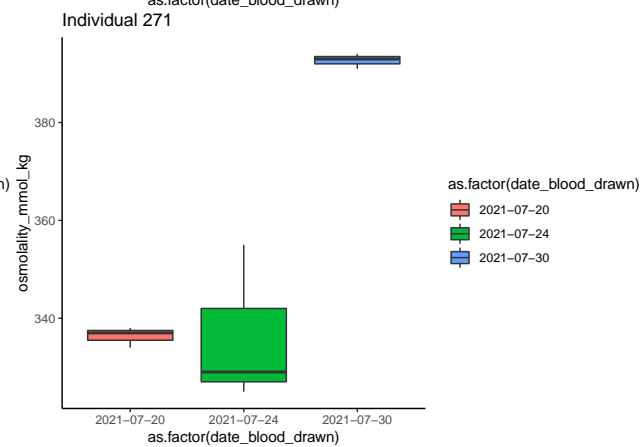
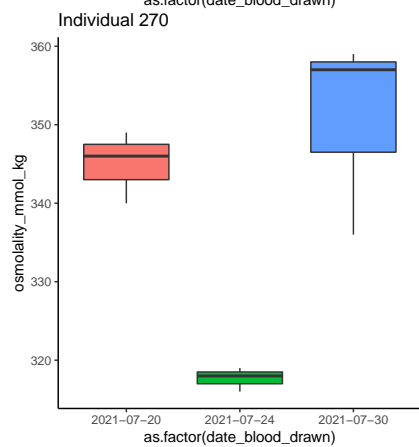
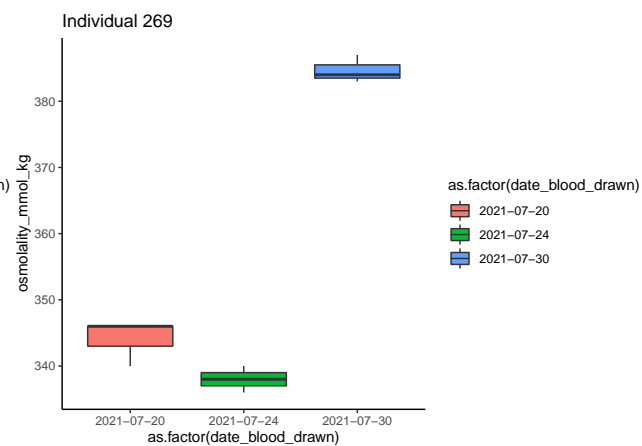
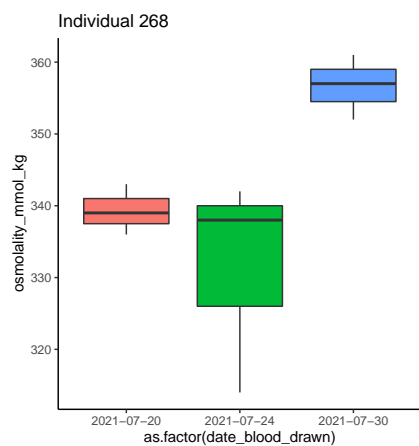


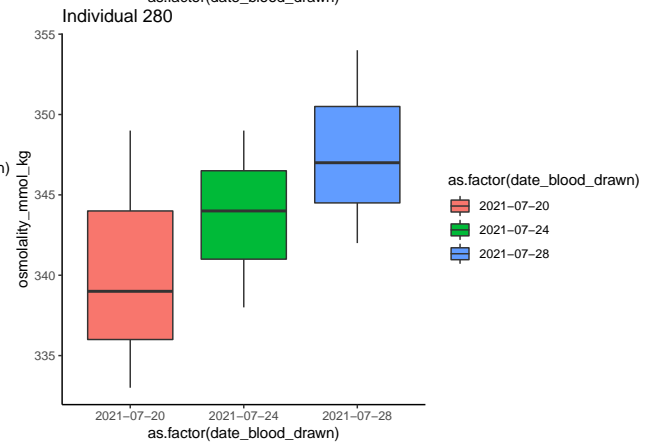
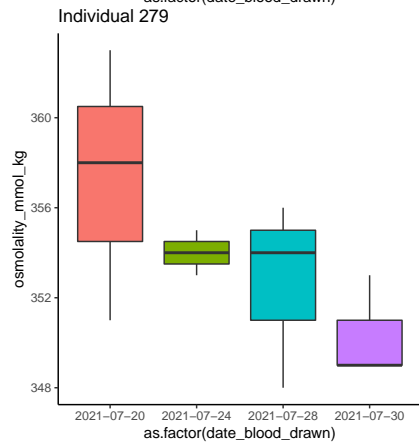
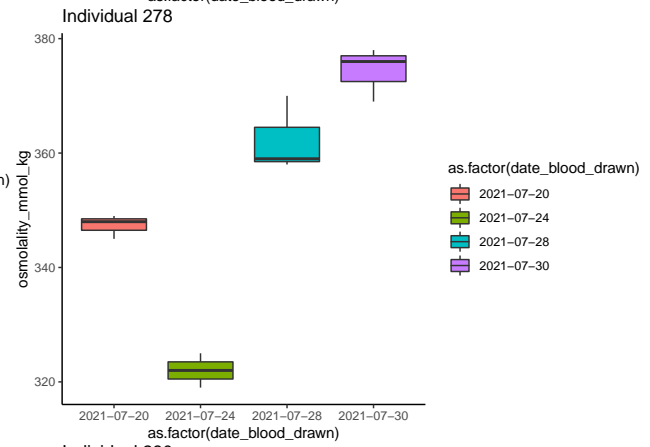
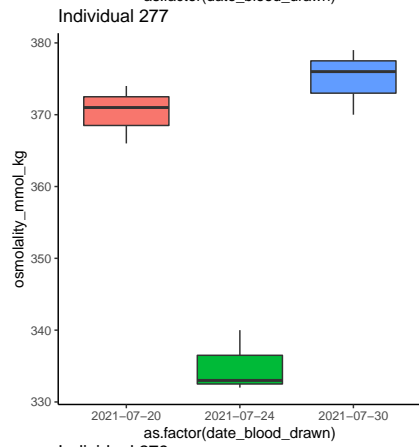
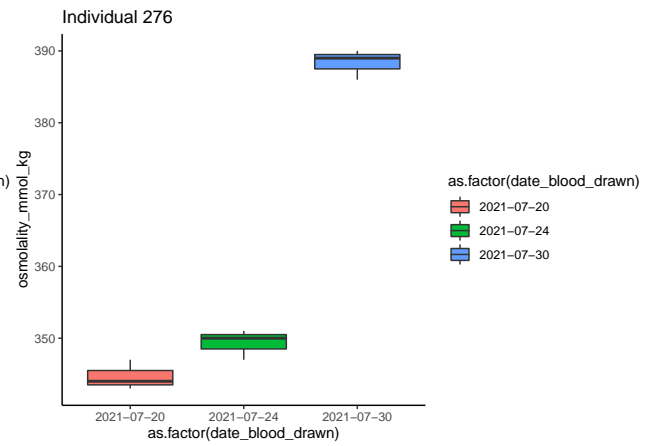
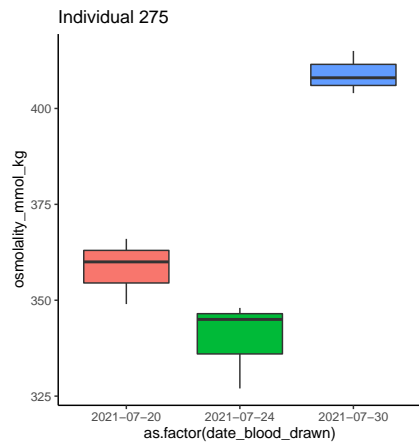


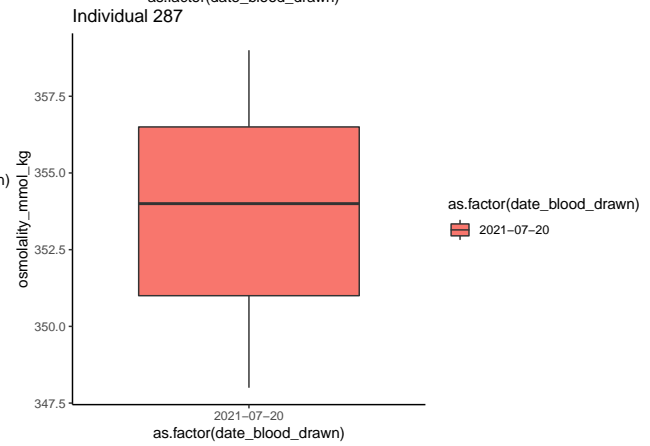
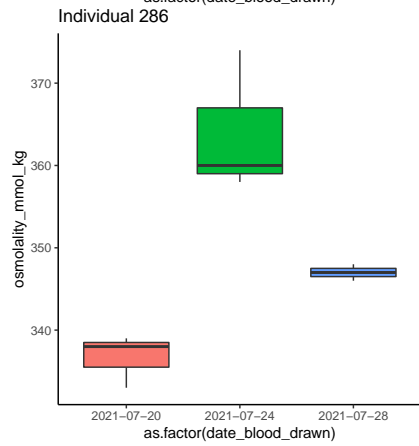
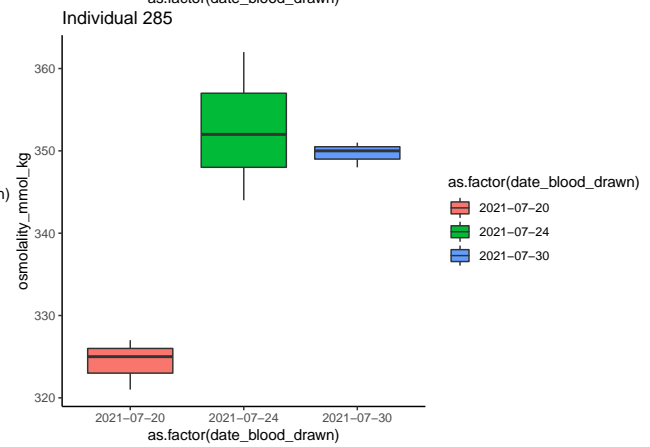
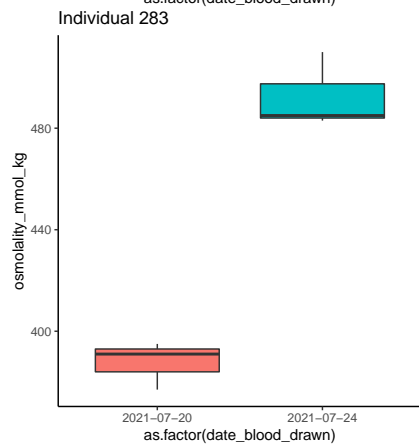
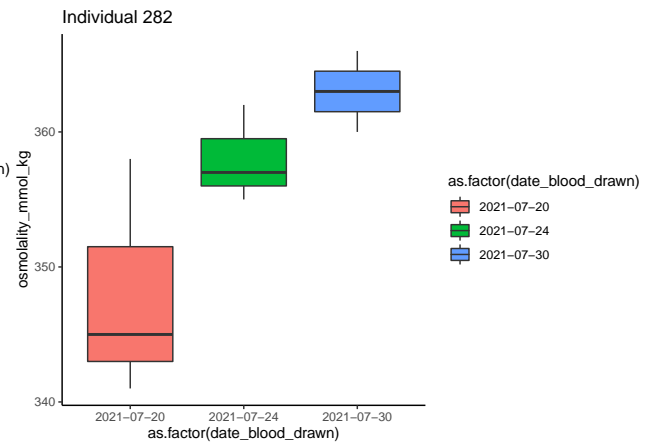
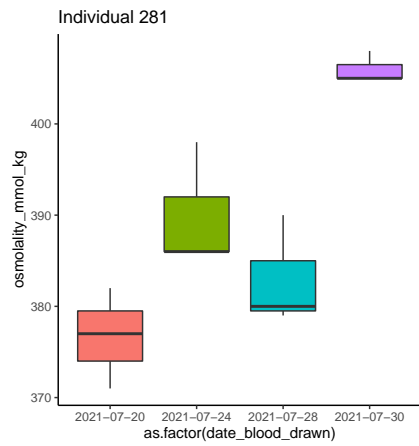


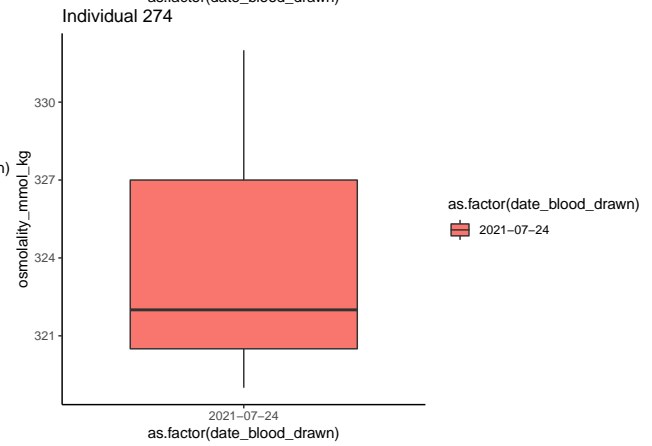
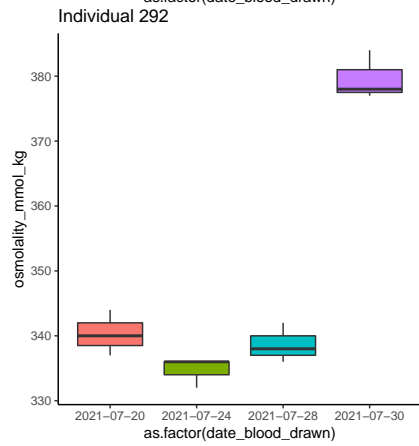
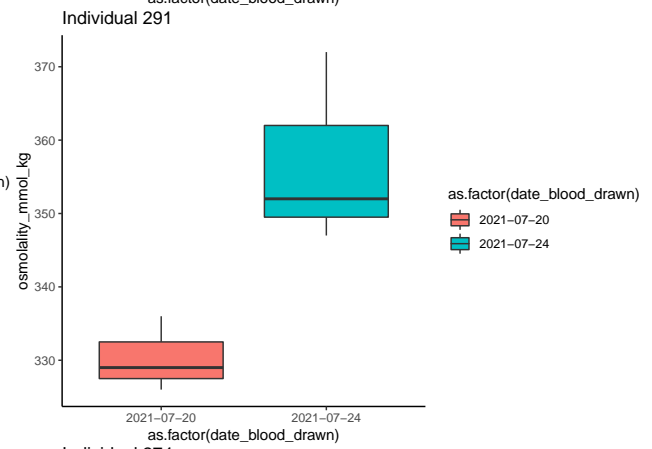
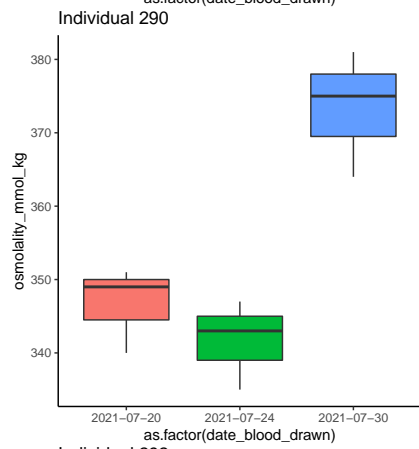
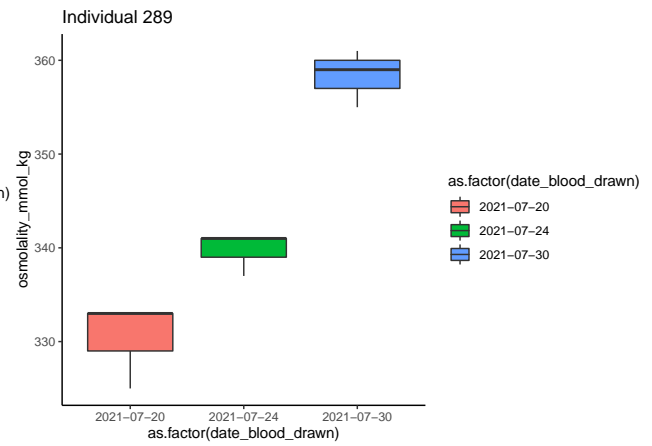
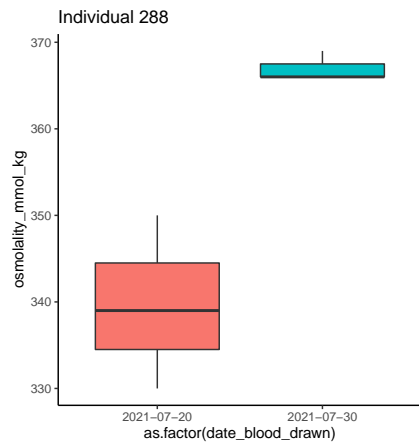


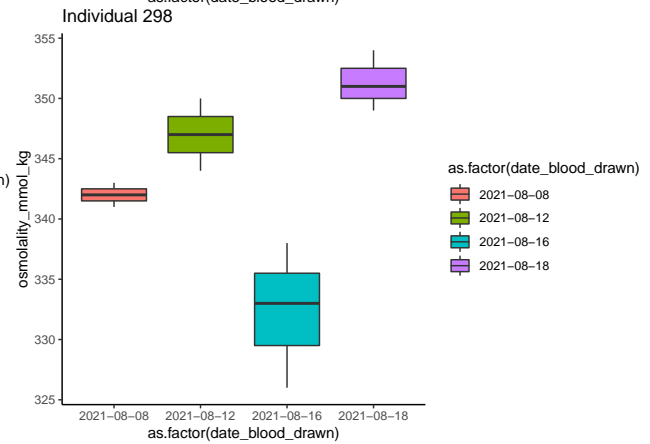
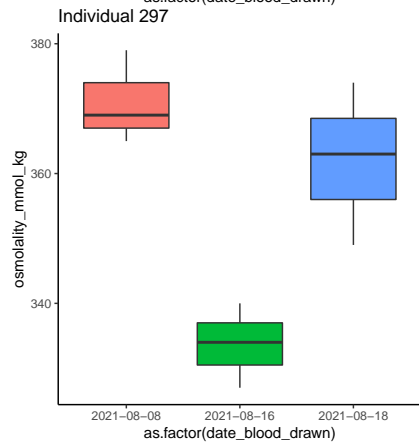
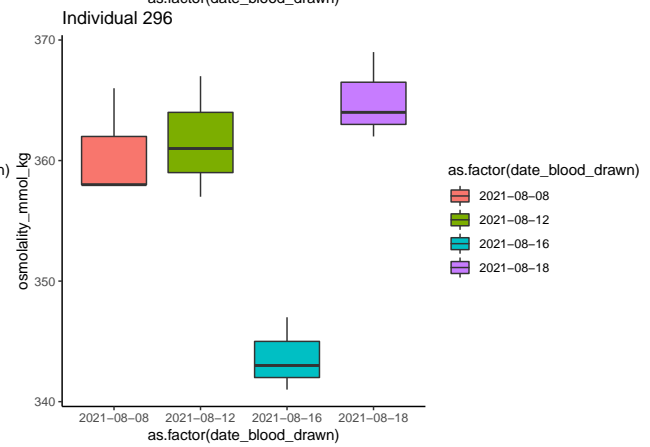
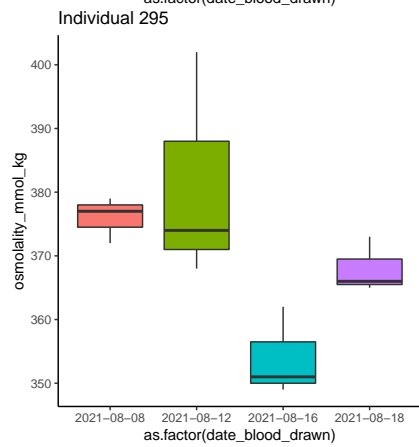
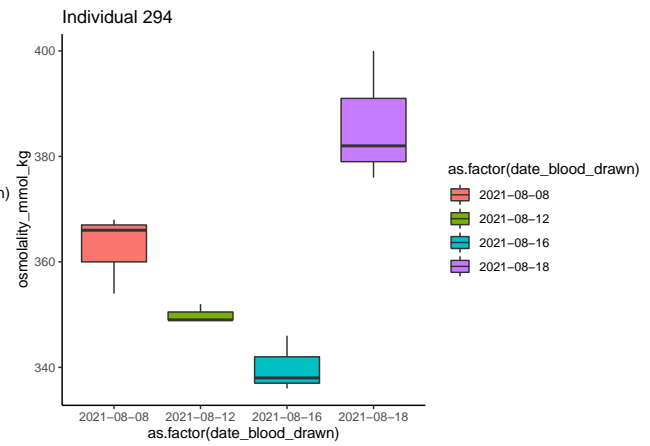
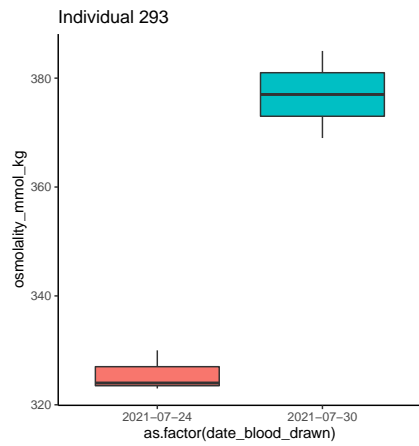


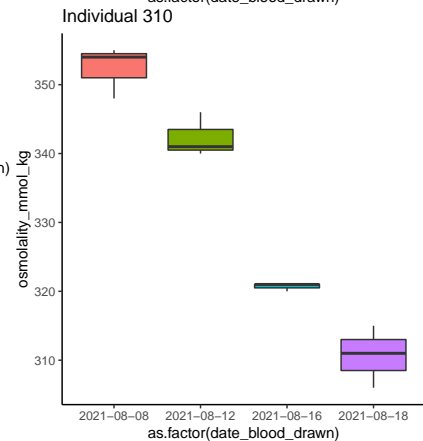
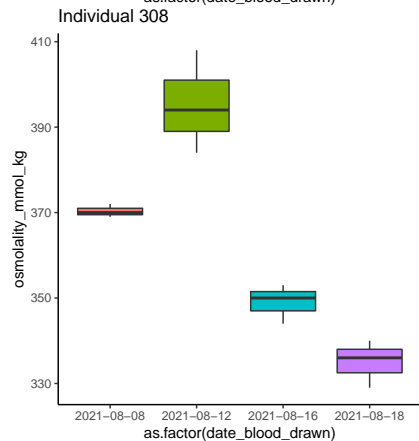
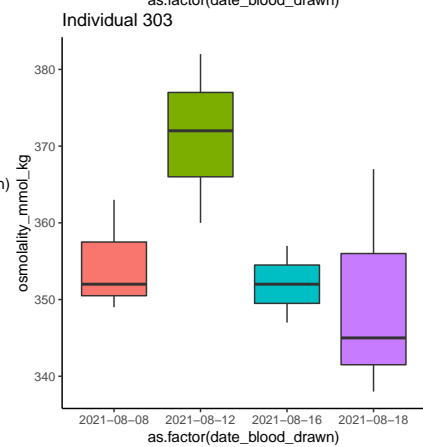
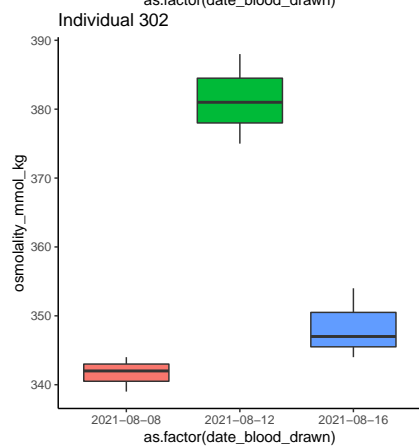
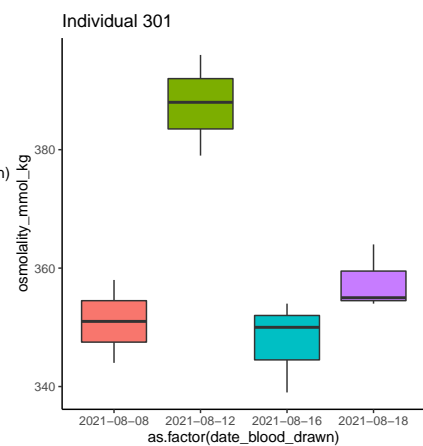
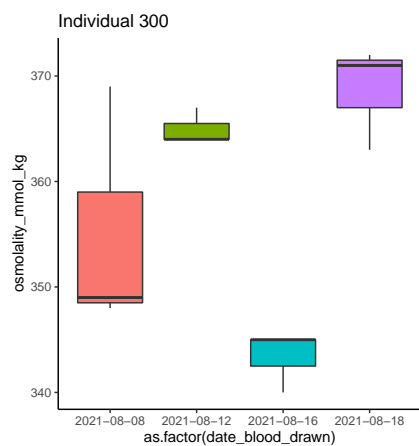




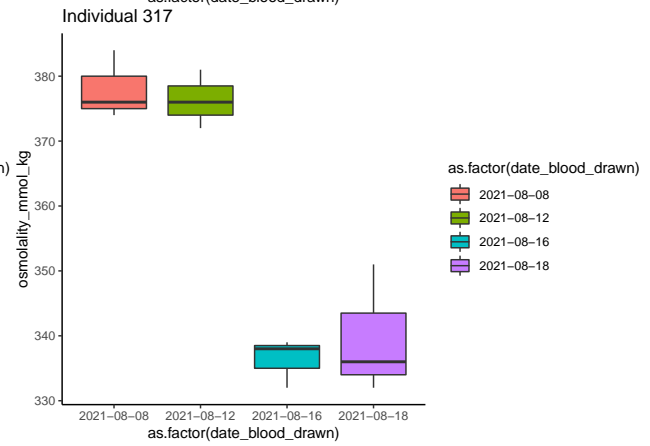
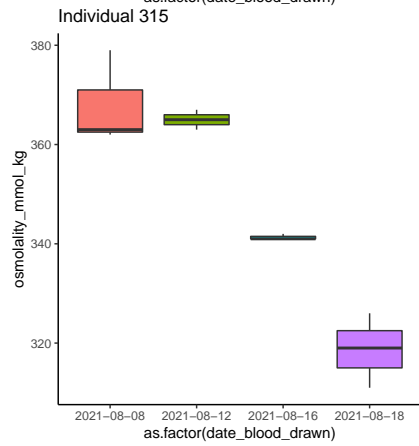
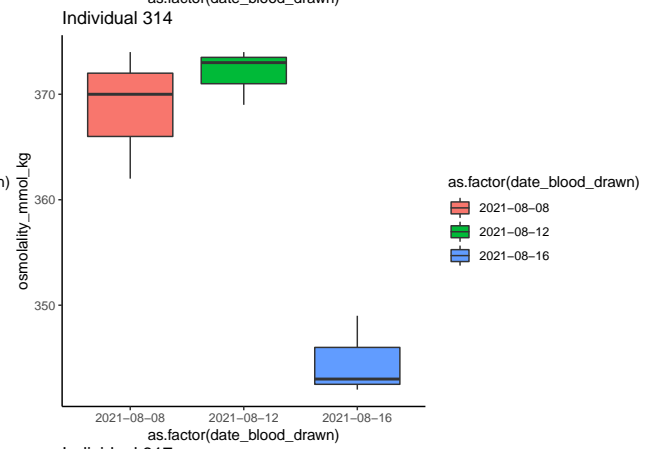
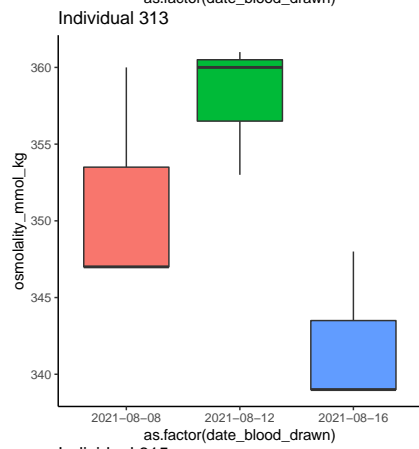
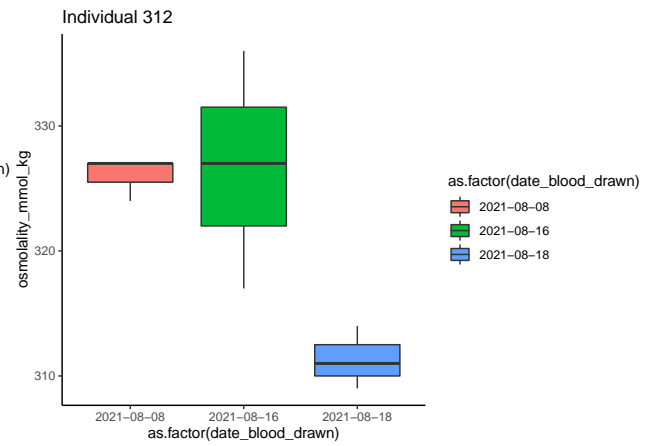
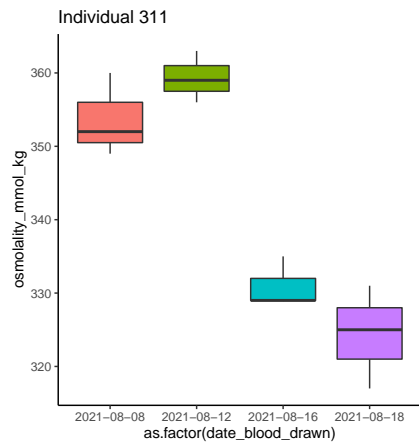


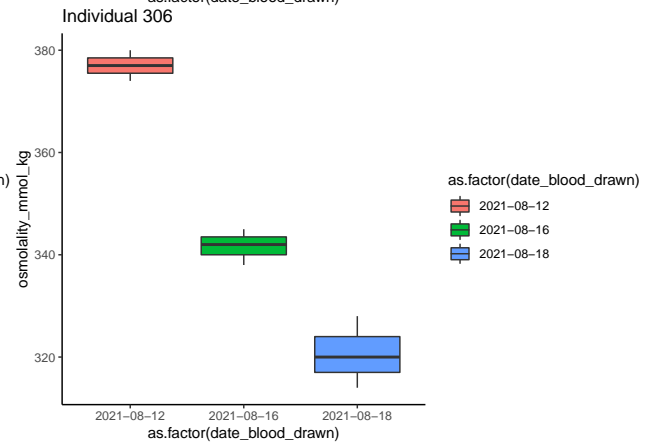
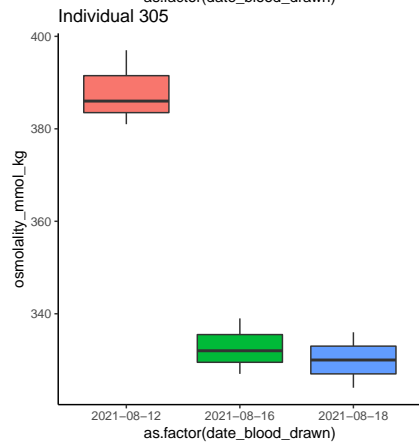
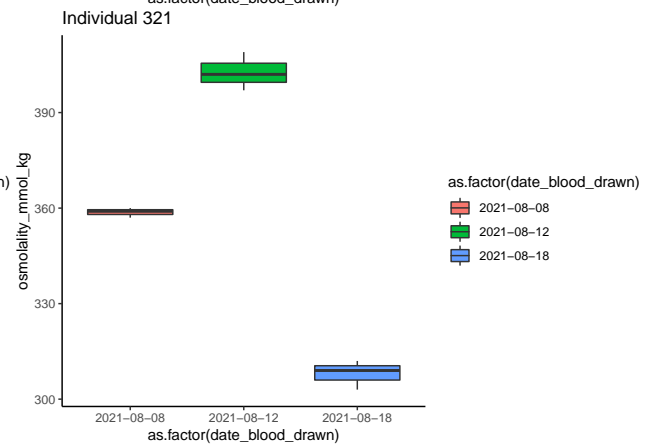
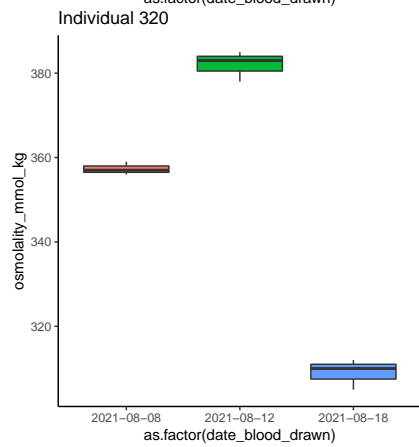
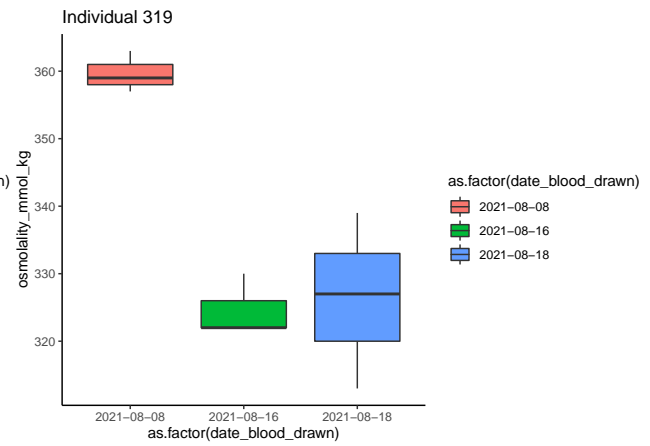
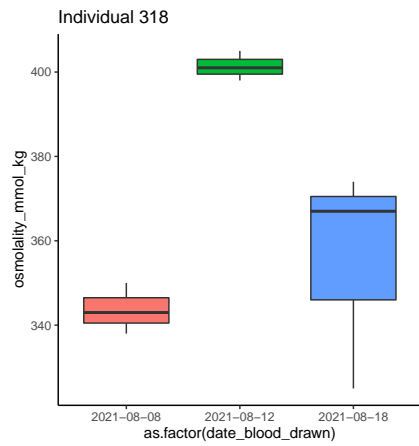


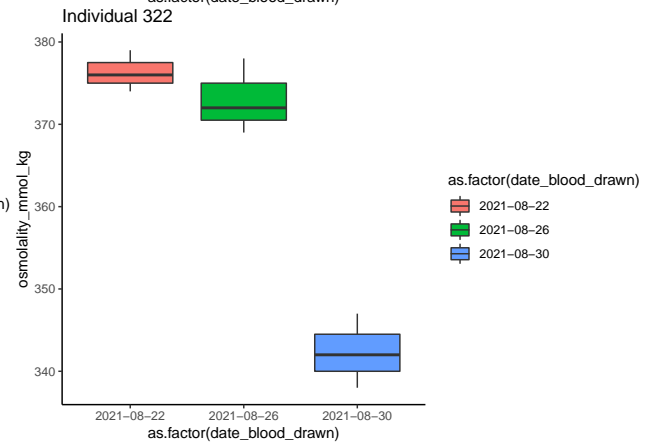
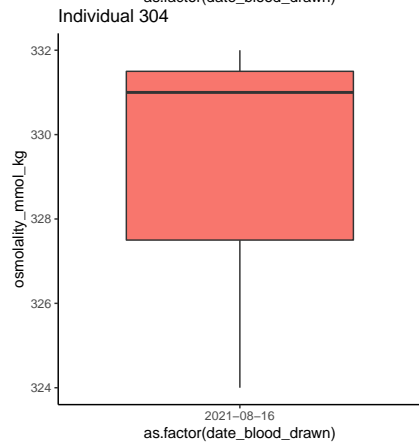
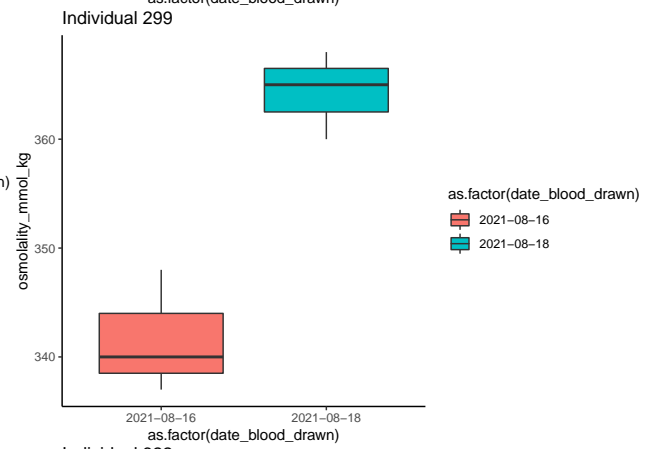
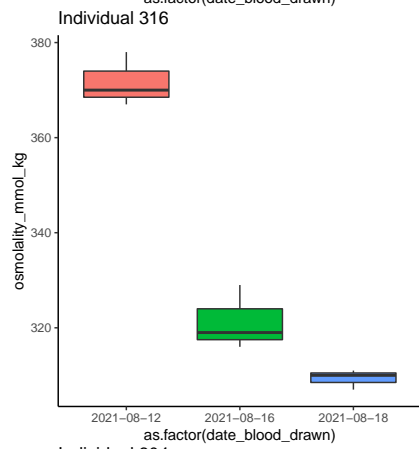
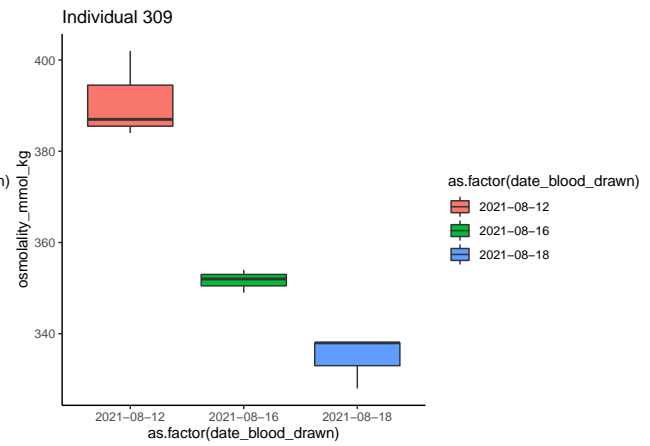
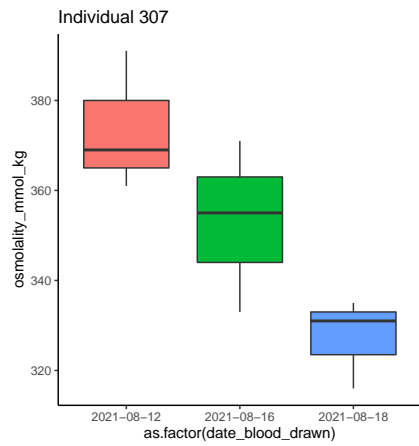


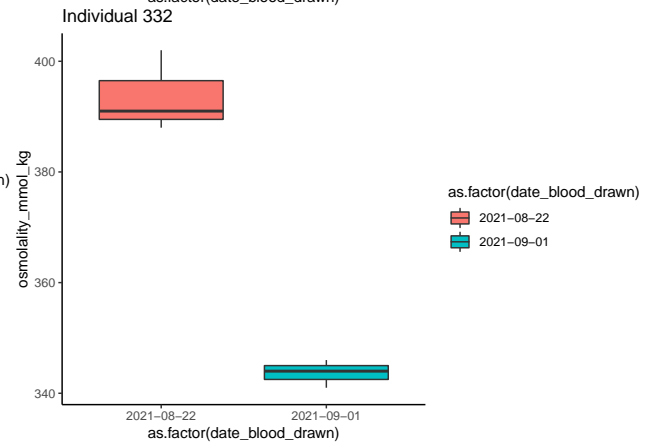
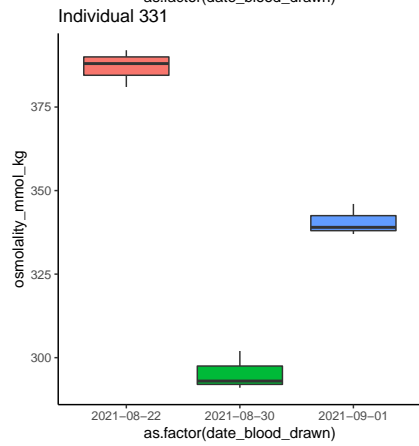
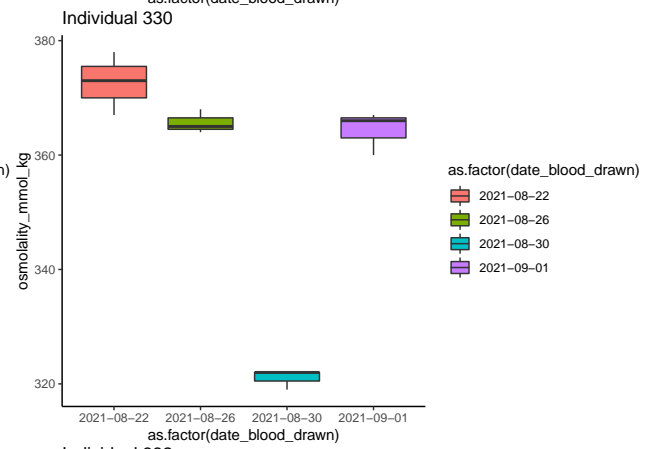
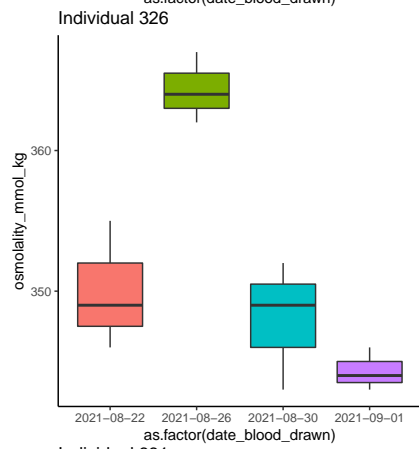
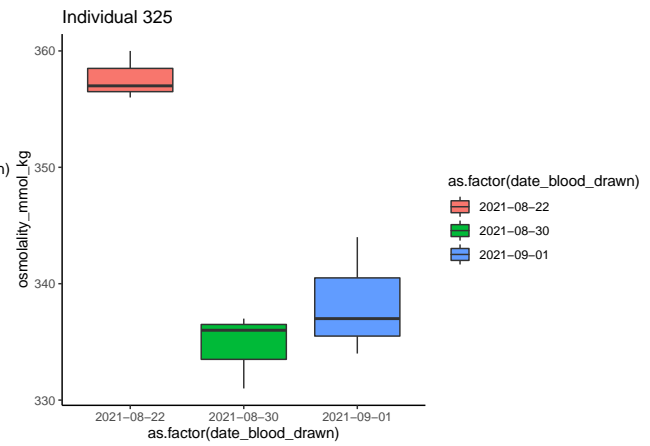
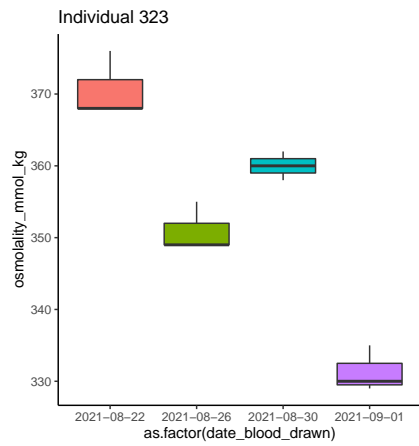


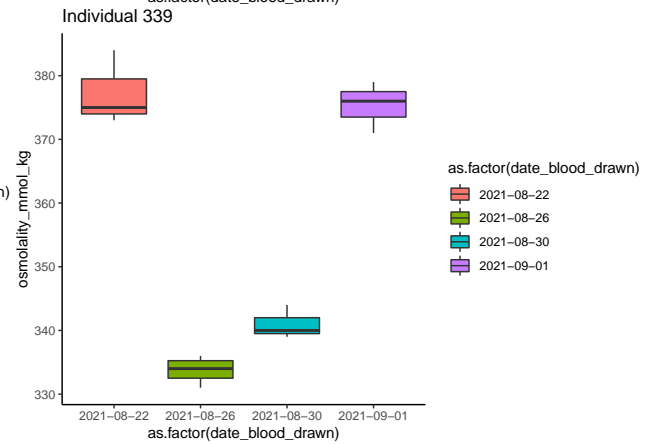
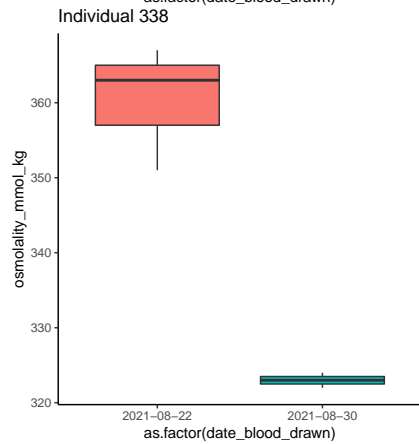
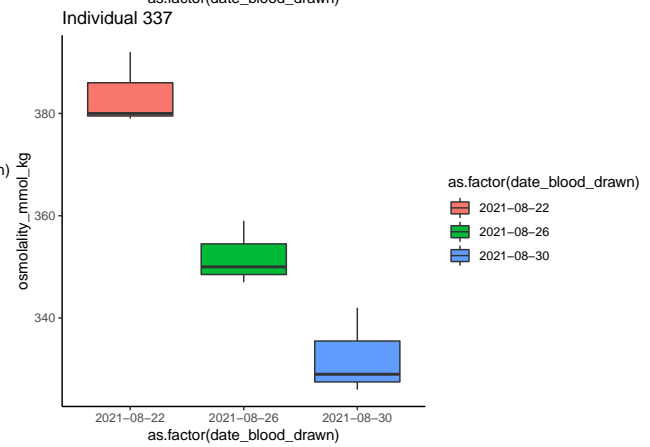
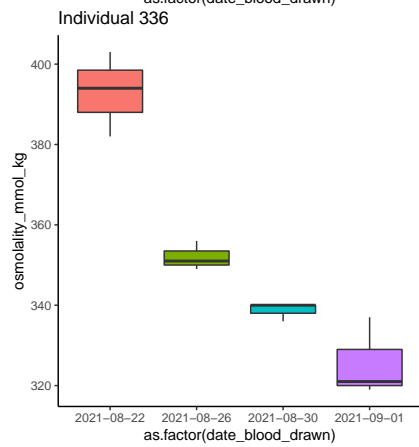
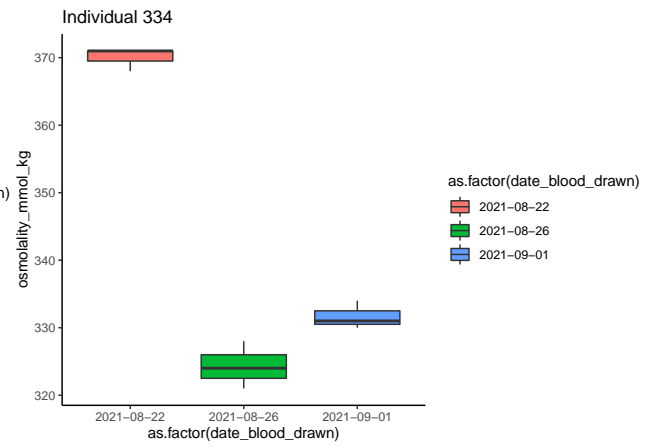
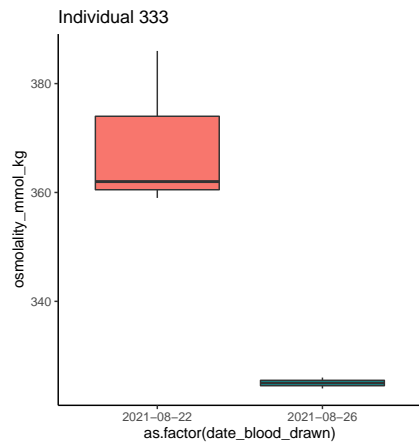


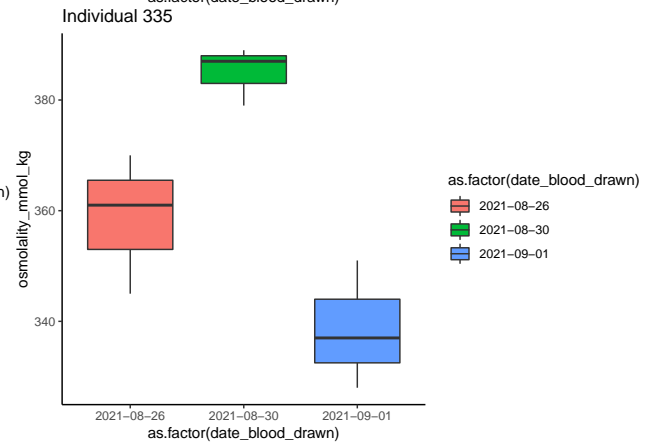
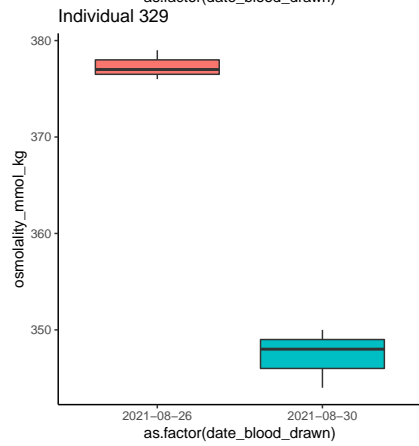
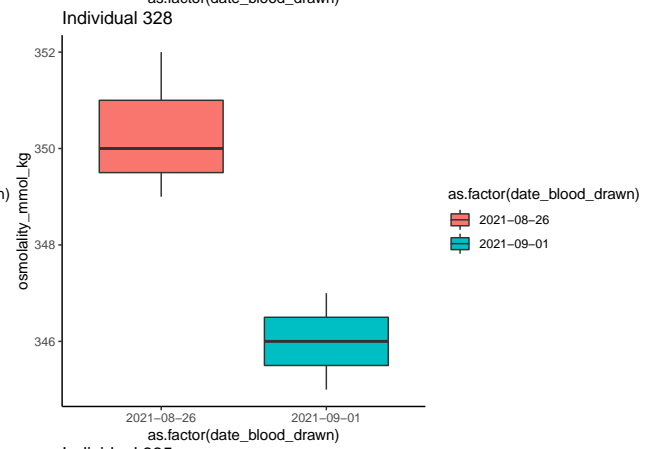
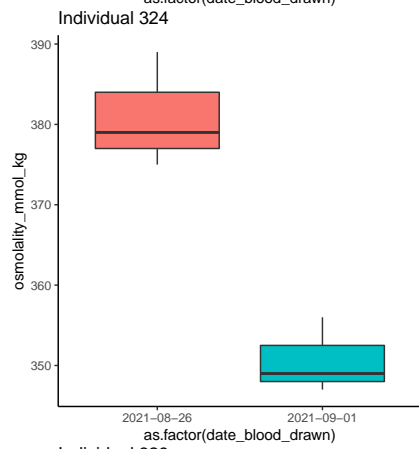
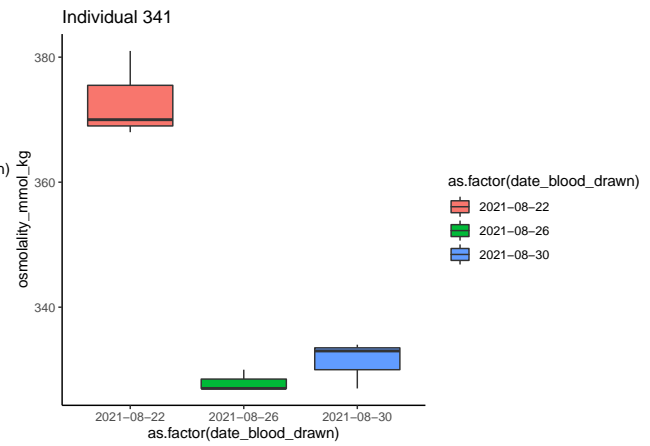
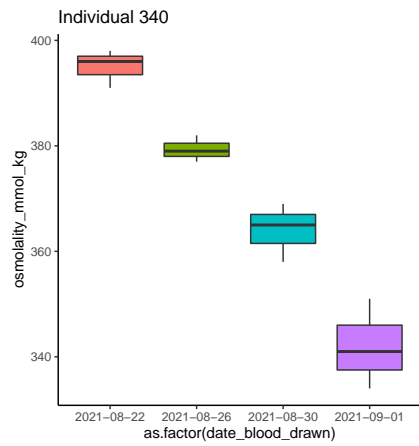


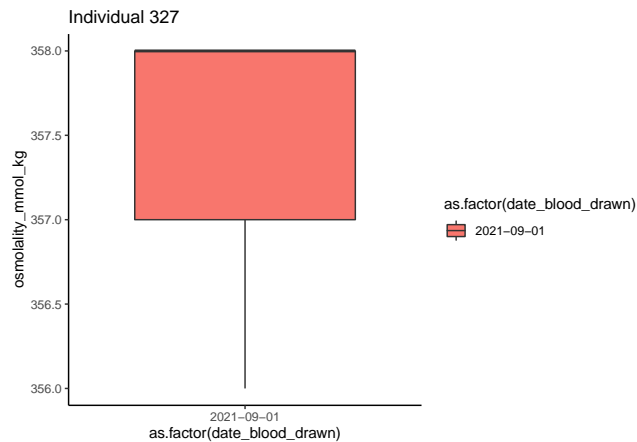












Based on boxplots, there are no outliers.

## Remove Outlier

Even though there are no outliers based on boxplot IQRs, we still want to find and omit the point severely increasing CV for one individual on one date, so find that one.

```
CVs %>% dplyr::filter(CV > 10)
```

```
## # A tibble: 1 x 8
## # Groups:   individual_ID [1]
##   individual_ID date_blood_drawn mean    SD    CV  min  max osml_range
##   <fct>         <date>         <dbl> <dbl> <dbl> <dbl> <dbl>      <dbl>
## 1 225          2021-06-26      325.  69.8  21.5  247  381        134
```

Determine which replicate of that group is an outlier and pulls the CV above our acceptable range (10-15%).

```
enuf_reps %>%
  dplyr::filter(individual_ID == 225 & date_blood_drawn == "2021-06-26")
```

```
## # A tibble: 3 x 8
## # Groups:   individual_ID, date_blood_drawn [1]
##   date_blood_drawn date_osmom_run time_osmom_run individual_ID replicate_no
##   <date>           <date>         <dtm>         <fct>         <fct>
## 1 2021-06-26      2021-06-26      2022-10-03 09:42:00 225          1
## 2 2021-06-26      2021-06-26      2022-10-03 09:42:00 225          2
## 3 2021-06-26      2021-06-26      2022-10-03 09:42:00 225          3
## # ... with 3 more variables: osmolality_mmol_kg <dbl>, notes <lgl>, count <int>
```

We should remove replicate 2 for individual 225 on June 26, which is >100 mmol/kg away from the closest values, and the other two values are within 35 mmol/kg.

```
enuf_reps_trimmed <- enuf_reps %>%
  # remove the one outlier
  dplyr::filter(!(individual_ID == 225 &
    date_blood_drawn == "2021-06-26" &
    replicate_no == 2))

# check
enuf_reps_trimmed %>%
  dplyr::filter(individual_ID == 225 &
    date_blood_drawn == "2021-06-26")
```

```
## # A tibble: 2 x 8
## # Groups:   individual_ID, date_blood_drawn [1]
##   date_blood_drawn date_osmom_run time_osmom_run   individual_ID replicate_no
##   <date>           <date>         <dtm>         <fct>         <fct>
## 1 2021-06-26       2021-06-26     2022-10-03 09:42:00 225           1
## 2 2021-06-26       2021-06-26     2022-10-03 09:42:00 225           3
## # ... with 3 more variables: osmolality_mmol_kg <dbl>, notes <lgl>, count <int>
```

## Average Remaining Replicates

Now that the single outlier has been removed from the technical replicates when there were enough replicates to identify them, I will rejoin the data for lizards with 1-2 (not\_reps) and 3-4 (enuf\_reps) replicates, then average the technical replicates for each lizard on each of their measurement dates.

```
osml_means <- not_reps %>%
  rbind(enuf_reps_trimmed) %>%
  group_by(date_blood_drawn, individual_ID) %>%
  summarise(osmolality_mmol_kg_mean = mean(osmolality_mmol_kg))
```

```
## `summarise()` regrouping output by 'date_blood_drawn' (override with `.`groups` argument)
```

## Other Cleaning

10 lizards in trial 1 have unreasonably high osmolality measurements on June 24, which we think are due to an osmometer technical error, as they were all from the same time period. The values are way too far outside the usual range to be trustworthy, and these measurements were taken just before the osmometer had to be recalibrated, so we will exclude them.

```
osml_means_clean <- osml_means %>%
  dplyr::filter(!(osmolality_mmol_kg_mean > 500))
```

## Export

```
write_rds(osml_means_clean, "./data/osml_means_clean.RDS")
```

## Reporting

We only removed one measurement that was an outlier within its technical replicate group.

We omitted 10 mean measurements (after taking tech rep mean) for lizards in trial 1 which had unreasonably high osmolality measurements on June 24 that can be confidently attributed to instrumental error.